Some Haematological Parameters of Inter-generic Hybrid of African catfish (Clarias anguillaris x Heterobranchus bidorsalis) Juveniles and Their Pure Lines in North Eastern Nigeria

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ABSTRACT
Haematology is important in order to know the state of health of fish species. Some haematological parameters of inter-generic hybrids African catfish (Clarias anguillaris x Heterobranchus bidorsalis) were investigated in North east Nigeria. The study aimed at investigating the haematological competence of the hybrids for survival in culture conditions in the region. Most fish farmers in the North east shade their ponds concrete of earthen due to stress as a result of harsh weather conditions in the region. Red Blood Cells (RBC) were significantly higher (2.46±0.03×10⁷ μL⁻¹) in hybrid Ca×Hb (Heteroclarias). However, MCV and MCH were significantly (p<0.05) higher (138.07±1.98 fl and 46.53±0.70 pg, respectively) in pure line of Heterobranchus bidorsalis. MCHC values of pure line C. anguillaris and their maternal hybrids #Ca×Hb♂ (30.03±1.96 and 35.23±0.62 g dL⁻¹, respectively) varied significantly (p<0.05) from those of Heteroclarias (34.57±0.33 g dL⁻¹). However, all the blood parameters of fish species recorded during this study were within the healthy ranges for fish. The results of this study show that Heteroclarias have more haematological competence for survival in culture conditions in the North east region.

Key words: Haematological parameters, inter-generic hybrids, Clarias anguillaris x Heterobranchus bidorsalis, weather conditions

INTRODUCTION
In aquaculture, production of hybrid fish plays important role in fast growth, survival and improvement of genetic traits. Inter-generic hybridization has been performed between African catfish viz: C. anguillaris×H. bidorsalis (Diyaware et al., 2010). Hybridizations between African catfish and Asian catfish; C. gariepinus×C. batrachus were also carried out (Sahoo et al., 2003) and C. gariepinus×Heterobranchus longifilis (Legendre et al., 1992; Nwadukwe, 1995), C. batrachus×C. gariepinus has been attempted by Rahman et al. (1995). Hybridization between Heteropneustes fossilis and C. gariepinus have been reported by Muthukumaran and Sukumaran (2005) while that of C. macrocephalus and C. gariepinus was documented by Na-Nakorn et al. (2004). Aluko and Ali (2001) successfully produced eight fast inter-generic hybrid fish from four African catfish species belonging to two genus Clarias and Heterobranchus.
Haematological indices in these hybrids critical are parameters for the evaluation physiological status and genetic resilience of the fish to resist bacterial infection, fungal and viral disease and withstand intensive culture conditions that compromises immunity. Response to these depends on fish species, age, sexual maturity of fish and diseases (Luskova, 1997; Golovina and Trombicky, 1989) as cited by Vosyliene (1999), Golovina (1996) and Zhiteneva et al. (1989). Haematological tests and analysis of serum constituents have yielded useful information for detection and diagnosis of metabolic disturbances and disease conditions in fishes (Jamalzadeh et al., 2009). Like in warm-blooded animals, changes in the blood parameters of fish, occurring from injuries or infections of tissues or organs, can be used to determine extend of the dysfunction or injuries organs or tissues. However, in fish, these parameters are more related to the response of the whole organism, i.e., to the effect on fish survival, reproduction and growth than discrete organ or tissue (Vosyliene, 1999).

In recent years, variation in haematological indices were used when clinical diagnosis of fish physiology was required to determine the effects of external stressors and toxic substances due to the close association between the circulatory system and the external environment (Cech et al., 1996; Wendelaar-Bonga, 1997). Weper (1991) also suggested that haematological and biochemical changes, growth rate and oxygen consumption of fish are used in determining the toxic effects of pollutants.

According to Fernandes and Mazon (2003), haematological parameters are closely related to the response of the animal to the environment, an indication that the environment where fish lives could exert some influence on the haematological characteristics (Kori-Siakpere, 1985). Sex of the fish may also influence the blood parameters. Smieszko (1960) showed that males consistently had higher packed cell volume values than the females and this has been proposed as means of sexing fish. Blood cell responses are important indicators of changes in the internal and/or external environment of animals (Adyemo, 2007). A number of haematological indices such as Haematocrit (Ht) Haemoglobin (Hb), Red Blood Cells (RBCs) are used to assess the functional status of the oxygen carrying capacity of bloodstream and have been used as indicators of the presence of metallic pollutants in aquatic environment (Shah and Altindag, 2004).

Red Blood Count (RBC), concentration of haemoglobin (MCH) and Haematocrit (Ht) have been reported to indicate secondary responses of an organism to irritants (Adyemo et al., 2008). Decrease in erythrocytes has been reported to be the major and reliable indicators of various sources of stress in fish (Rainza-Paiva et al., 2000; O’Neal and Weirich, 2001). White blood cells are the main defense cells of the animals. Decrease in these cells indicates vulnerability to stress and infection (Sunomono and Oyelola, 2008). Drop in red blood cells implies reduction in level of oxygen that is being carried to the tissues and the level of carbon dioxide returned to the lungs will also be reduced. Reduction in mean hemoglobin concentration and mean corpuscular hemoglobin concentration indicates anemia (Sunomono and Oyelola, 2008). According to MyDr (2006), packed cell volume or haematocrit, Red Blood Cell (RBC) and Mean Cell Haemoglobin Concentration (MCHC) are used for diagnosing anaemic condition while neutrophils and monocytes (differential counts) protects the body against bacterial invasion. Lymphocytes are involved in immune process, producing antibodies against foreign organism, protecting against microbes. Low RBC indicates malnutrition and platelets helps in diagnosing problems associated bleeding or bruising.

Hematological studies in fishes have assumed greater significance because these parameters could be used as an effective and sensitive index to monitor physiological and pathological changes induced by natural or anthropometric factors, such as bacteria and fungi infections or pollution of
water environment, pathogenicity of the organism. Osuigwe et al. (2005) documented some haematological changes in hybrid Heteroclarias fed raw and boiled jack beans seed meal. Effects of ascorbic acid on haematological parameters of C. gariepinus have been studies by Oluyemi et al. (2008). Rogers et al. (2003) concluded that, mechanism of lead toxicity occurs by ion regulatory disruption.

Few authors reported blood profile of Clariards not exposed to chemicals or feed or disease; Clarias burchiopogen (Kori-Siakpere and Egor, 1999), C. ishernensis (Kori-Siakpere, 1985), on hybrid Heteroclarias (H. bidorsalis and C. gariepinus) (Kori-Siakpere et al., 2005).

Changes in the blood characteristics of Clarias gariepinus caused by stress due to exposure to environmental pollutants, diseases or attack by pathogens have been studied extensively: (Onusiriuka and Ufodike, 2000; Ezeri, 2001; Gabriel et al., 2001; Rehulkka, 2002a, b). Maheswaran et al. (2008) studied effect of mercuric acid on blood indices of C. batrachus while Okechukwu et al. (2007) investigated the haematological indices of C. gariepinus to exposure to acute chlorpyrifos-ethyl. Ovie et al. (2007) reported effects of paraquat on blood indices. These indices have been employed in effective monitoring of the responses of the fish to stressors and thus its health status under adverse conditions.

Information on haematological profile of Clariards and their hybrids have not been fully documented. Haematological profiles of juvenile hybrids will reveal the possibility of the hybrids to withstand the harsh environmental condition that is likely to be faced by the species during rearing. There is also a need to establish baseline information on haematological profiles of our economically valued fish species for continual assessment of their health status and subsequent diagnosis of disease. The objective of this study is to compare the hematological parameters of hybrid African catfish (Clarias anguillaris × Heterobranchus bidorsalis) with view to assessing the ability of hybrids to thrive under culture conditions in north east Nigeria.

MATERIALS AND METHODS
Experimental fish: The study was conducted between March to August 2010 (6 month; dry and rainy season). Sixty each of juvenile hybrids of Clariabanchus (female C. anguillaris×male H. bidorsalis), Heteroclarias (female H. bidorsalis×male C. anguillaris) and their pure line C. anguillaris×C. anguillaris and H. bidorsalis×H. bidorsalis) were collected from Departmental polyethylene line fish pond and the blood samples collected immediately.

Blood collection: Blood samples were collected from the fish through caudal pedunde puncture as described by (AQUALEX, 2004). Approximately 60 μL of heparin was drawn from EDTA bottle using 2 mL plastic syringe with 22 gauge hypodermic needle (Schmitt et al., 2007), 0.3-0.5 mL of blood was collected from each fish and deposited into bottle containing EDTA. The samples were transported in a cold pack to the haematology laboratory at Prof. Umaru Shehu specialist hospital Maiduguri for haematological analysis.

Haematological analysis: The haematological parameters were analyzed using automated haematological analyzer (Model: Sysmex KX-21N, Sysmex Cooperation, Kobe Japan). White Blood Cell (WBC) count was performed with WBC detector block using Direct Current (DC) detection method. Red Blood Cell (RBC) and Platelets (PL) were analyzed by Hydrodynamic focusing DC detection methods. Haemoglobin (Hb) levels were analyzed by Non-cyanide Sodium Lauryl Sulphate (SLS) method while Packed Cell Volume (PCV) was determined using Cumulative Pulse
Height Detection (CPHD) method. The blood sample with EDTA was mixed gently and probed with haematological analyzer and then start bottom was pressed. The haematological parameters were printed out immediately.

**Physicochemical parameters:** Water quality parameters such as dissolved oxygen, pH and Temperature during sample collection were collected were recorded using digital EC/TDS/pH kit (model: EC500 Meter SANXIN-China).

**Data analysis:** Haematological data obtained from all the treatments (cross combinations) were subjected to one way analysis of variance (ANOVA). Differences between the means were determined using Duncan’s multiple range tests, Duncan (1955) with SPSS.15 for windows at 95% confidence level (p = 0.05).

**RESULTS**

Table 1 shows mean blood profiles of juvenile inter-generic hybrid catfish *C. anguillaris* and *H. bidorsalis* and their pure line progenies in North east Nigeria. PCV, Hb, MCHC and platelet (PLT) were higher in pure line *C. anguillaris*. There were no significant differences between PCV, Hb and PLT values among the entire cross combination. MCHC values from *H. bidorsalis* (34.57±0.33 g dL⁻¹) and their maternal hybrids (33.67±0.44) were significantly (p<0.05) lower than those recorded in pure *C. anguillaris* (39.63±1.96 g dL⁻¹) and their maternal hybrids (35.23±0.61 g dL⁻¹). However, MCHC values of pure *H. bidorsalis* juveniles were not significantly (p>0.05) higher compared to the reciprocal hybrids. Similarly, MCHC values of pure *C. anguillaris* were significantly the same with to that of their maternal hybrids (*Clariabranchnus*).

**White blood cell (WBC):** The highest WBC was recorded in hybrid *Heteroclarias* followed by their maternal pure line *H. bidorsalis*, *C. anguillaris* and hybrid *Clariabranchnus*. There was no significant difference between the WBC values among the entire cross combination.

RBC counts were higher (2.46±0.03x10⁶ µL⁻¹) in *Heteroclarias* followed closely by *C. anguillaris* and the hybrid *Clariabranchnus* (2.42±0.12 and 2.18±0.10x10⁶ µL⁻¹) while *H. bidorsalis* (1.99±0.14x10⁶ µL⁻¹) was significantly (p<0.05) lower than the other cross combinations. The high RBC values recorded in *Heteroclarias* juveniles were not significantly different from that of *C. anguillaris* and *Clariabranchnus* (Table 1).

**Mean corpuscular volume (MCV):** The highest MCV were observed in *H. bidorsalis* (138.07±1.98 fl), followed by *C. anguillaris* (117.68±1.83 fl), *Clariabranchnus* (117.13±1.76 fl) and *Heteroclarias* (113.07±2.29 fl). There is no significant difference between MCV values obtained in *H. bidorsalis* compared to that of *C. anguillaris*. Similarly MCV values of *Clariabranchnus* and *Heteroclarias* were statistically (p>0.05) the same (Table 1).

**Mean cell haemoglobin (MCH):** MCH were observed to be higher in *H. bidorsalis* (46.53±0.70 pg), followed by pure *C. anguillaris* with (42.65±1.96) and *Heteroclarias*. MCH value of *H. bidorsalis* is significantly (p<0.05) different from the rest of the cross combinations while MCH values of *C. anguillaris* and that of their maternal hybrids (*Heteroclarias*) are statistically (p>0.05) the same (Table 1).
Table 1: Blood profiles of juvenile inter-generic hybrid catfish (C. anguillaris and H. bidorsalis) and their pure lines progenies in North east Nigeria

<table>
<thead>
<tr>
<th>Parameters</th>
<th>C. anguillaris</th>
<th>Clariasbranuchs</th>
<th>H. bidorsalis</th>
<th>Heteroclarias</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCV (%)</td>
<td>28.56±2.71a</td>
<td>25.57±1.12a</td>
<td>27.50±1.47a</td>
<td>27.87±1.02a</td>
</tr>
<tr>
<td>Hb (g dL⁻¹)</td>
<td>10.33±0.62b</td>
<td>9.09±0.26b</td>
<td>9.27±0.58b</td>
<td>9.63±0.28b</td>
</tr>
<tr>
<td>WBC (&gt;10⁶ µL⁻¹)</td>
<td>188.98±8.88a</td>
<td>181.53±6.57a</td>
<td>192.87±8.29a</td>
<td>193.70±3.26a</td>
</tr>
<tr>
<td>RBC (&gt;10⁸ µL⁻¹)</td>
<td>2.42±0.12a</td>
<td>2.18±0.10a</td>
<td>1.99±0.14a</td>
<td>2.46±0.08a</td>
</tr>
<tr>
<td>MCV (fl)</td>
<td>117.68±4.83a</td>
<td>117.13±1.76a</td>
<td>138.07±1.98a</td>
<td>113.07±2.59a</td>
</tr>
<tr>
<td>MCH (pg)</td>
<td>42.65±1.99a</td>
<td>41.30±1.15a</td>
<td>46.53±0.79a</td>
<td>39.10±0.61a</td>
</tr>
<tr>
<td>MCHC (g dL⁻¹)</td>
<td>39.03±1.96a</td>
<td>37.23±0.61a</td>
<td>33.67±0.44a</td>
<td>34.57±0.33a</td>
</tr>
<tr>
<td>PLT (&gt;10⁸ µL⁻¹)</td>
<td>19.26±4.78a</td>
<td>17.33±4.81a</td>
<td>10.67±8.17a</td>
<td>16.67±4.63a</td>
</tr>
<tr>
<td>LYM (&gt;10⁶ µL⁻¹)</td>
<td>185.65±5.13a</td>
<td>177.97±13.61a</td>
<td>187.00±13.86a</td>
<td>190.40±3.43a</td>
</tr>
<tr>
<td>LYM (%)</td>
<td>98.23±0.22a</td>
<td>98.03±0.29a</td>
<td>97.37±0.80a</td>
<td>98.30±0.12a</td>
</tr>
</tbody>
</table>

Means±SEM in the same row having different superscript are significantly different at p<0.05. PCV: Packed cell volume, Hb: Haemoglobin, WBC: White blood cells, RBC: Red blood cells, MCV: Mean corpuscular volume, MCH: Mean cell haemoglobin, MCHC: Mean cell haemoglobin concentration, PLT: Platelet count, LYM: Lymphocytes

Table 2: Water quality parameters of the pond during experimental fish collection

<table>
<thead>
<tr>
<th>Fish species</th>
<th>Temperature (°C)</th>
<th>pH</th>
<th>DO (mg L⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>C. anguillaris</td>
<td>30.40±0.12</td>
<td>7.74±0.11</td>
<td>4.50±0.38</td>
</tr>
<tr>
<td>Clariasbranuchs</td>
<td>30.10±0.00</td>
<td>7.85±0.30</td>
<td>5.04±0.64</td>
</tr>
<tr>
<td>H. bidorsalis</td>
<td>29.63±0.90</td>
<td>7.49±0.35</td>
<td>4.36±0.42</td>
</tr>
<tr>
<td>Heteroclarias</td>
<td>30.13±0.29</td>
<td>7.67±0.41</td>
<td>5.14±0.82</td>
</tr>
</tbody>
</table>

Lymphocytes: Lymphocytes values were higher (98.30±0.12±10⁶ µL⁻¹) in Heteroclarias, followed closely by 98.23±0.22±10⁶ µL⁻¹ and 98.03±0.29±10⁶ µL⁻¹ observed in that of pure H. bidorsalis and Clariasbranuchs juveniles, respectively. There were no significant difference (p>0.05) among the LYm values of the entire cross combination. Percentage LYM were also high in hybrid Heteroclarias. Significant difference did not exist between all the percentages LYM for all the treatments (Table 1).

Water quality parameters: Temperature in the ponds where the fish sampled collected were between 29.43±0.90-30.40±0.12°C, pH 7.49±0.35-7.85±0.30 while dissolved oxygen was between 4.36±0.42-5.14±0.82 mg L⁻¹ (Table 2).

DISCUSSION

Water quality parameters recorded in the ponds where the experimental fish were collected were within the range for fish culture recommended by Boyd (1981).

Packed cell volume: PCV ranges of juvenile hybrids and pure line African catfish (C. anguillaris×H. bidorsalis) recorded in this study fall within normal values of 20-35% (Erondu et al., 1993) and 22-40% (Bhaskar and Rao, 1989). PCV values of hybrids recorded in this study were lower than that of Heteroclarias juvenile (38.40%) (Kori-Siakpere and Ubogu, 2003) and that of control juvenile C. gariepinus (39.00%) (Ogunji et al., 2005). The difference in the blood profiles could be due to variation in environmental factors (Fernandes and Mazon, 2003) that
haematological characteristics are closely related to the response of the animal to its environment. Accordingly, the environment where the fish lives could exert some influence on the haematological characteristics of that species (Kori-Siakpere, 1985). However, the PCV values recorded in this study are higher than those recorded by Ochang et al. (2007) for control juveniles of *C. gariepinus* (22.00%) during trials on growth performance, body composition, haematology and product quality of the African catfish (*C. gariepinus*) fed diets with palm oil.

**Haemoglobin concentration:** Hb reported in this study for the entire cross combinations (9.00-10.33 g dL$^{-1}$) is approximate to 10.63 g dL$^{-1}$ reported by Osuigwe et al. (2005) for controlled juvenile hybrid between *H. longifilis*×*C. gariepinus* but higher than (15.31 g dL$^{-1}$) documented by Kori-Siakpere and Ubogu (2008) for juvenile hybrid as well as 13.00 g dL$^{-1}$ recorded from *C. gariepinus* juvenile by Ogunji et al. (2005). Similarly, 27.00 g dL$^{-1}$ reported by Sunomonu and Oyelola (2008) varied with the result of this study. This variation may be due to difference in the species and environment where the fish lived. The high level of Hb recorded in pure line juveniles *C. anguillaris* indicates that this trait of high Hb might have been inherited from the paternal parents by the hybrids *Heteroclarias*.

**White blood cell:** The high (181.53±3.57-193.70±3.26×10$^3$ µL$^{-1}$) WBC recorded during this study are higher than those reported by most authors viz., 18.8×10$^6$ µL$^{-1}$ (Ogunji et al., 2005), 37.78×10$^6$ µL$^{-1}$ for wild adult *C. gariepinus* Gabriel et al. (2001) while 8.42×10$^6$ µL$^{-1}$ for juvenile *Heteroclarias* was documented by Osuigwe et al. (2005). Bunni (2010) observed 49.73×10$^6$ µL$^{-1}$ wild Clariabranth (C. gariepinus×H. bidorsalis), 16.51×10$^6$ µL$^{-1}$ for adult *C. anguillaris* and 9.04×10$^6$ µL$^{-1}$ for *C. macromystax* in North east Nigeria. However, the high WBC (183.70×10$^6$ µL$^{-1}$) observed in ♀Hb×♂Ca indicates stronger immune system toward invasion by foreign organism, prevents infection and at least transport and distributes antibodies in immune response as suggested by Sunomonu and Oyelola (2008). The increase in the WBC may be due to environmental stresses as a result of drastic fluctuations in the weather conditions of the North east Nigeria (semi arid zone). Stress-mediated condition may trigger the release of more white blood cells into the blood stream. MyDr (2006), reported that an abnormal WBC counts can indicates many possible medical conditions. This may suggest that the *H. bidorsalis* and their maternal hybrids may be affected by the harsh weather conditions of North east region during intensive aquaculture.

**Red blood cell:** Mean RBC values obtained *Heteroclarias, C. anguillaris* and Clariabranth (2.46, 242 and 2.18×10$^6$ µL$^{-1}$, respectively) in this study are higher (1.63×10$^{12}$) than reported by Kori-Siakpere and Ubogu (2008), 1.43×10$^{12}$; Osuigwe et al. (2005) and 1.77×10$^6$ mm$^3$ (Maheswaran et al., 2008). Higher RBC indicated that the level of oxygen that will be transported to the tissue and the level of carbon dioxide returned to the lungs will also increase thus efficient oxygen supply, survival and resistance to environmental conditions. However, Ogunji et al. (2005) observed as high as 1.80×10$^6$ mm$^3$ RBC from juvenile *C. gariepinus* in Zaria, Nigeria while Sunomonu and Oyelola (2008), reported 298.50×10$^{12}$ µL$^{-1}$ from juvenile *C. gariepinus* also from in Ilorin Nigeria. These values disagree with the results obtained in this study. The variation in the RBC could be due variation in the ecological conditions as suggested by Orun et al. (2003) that blood parameters are influenced by water temperature and oxygen concentration.

**Mean corpuscular volume:** MCV is an estimate of the volume of red blood cells. The mean MCV values observed in this study are lower than 240.18 fl recorded for juvenile hybrid African catfish
reported by Kori-Siakpere and Ubogu (2008) for juveniles *Heteroclarias* and 200.93 fL for *C. gariepinus* fingerlings (Gbore et al., 2006). It is higher than 96.62 fL for *C. gariepinus* fingerlings (Ochang et al., 2007). However, the high MCV may be due the high concentration of haemoglobin in the red blood cell.

**Mean cell haemoglobin:** The MCH values observed in this study are higher than earlier reports 24.24 pg *C. gariepinus* juveniles (Omitoyin, 2007) and 33.10 pg (Ochang et al., 2007), respectively. Kori-Siakpere and Ubogu (2008) reported higher MCH for juvenile *Heteroclarias* while, Gbore et al. (2006) reported 51.50 pg which contrary to what was obtained this study. This explains that the red cell enlargement due probably to nutritional deficiency of folic acid or Vitamin B12. In another word higher MCH indicates good volume of haemoglobin which indicates good oxygen transportation in the blood stream for healthy well being of the fish.

**Mean cell haemoglobin concentration:** MCHC values recorded among the entire cross combination are within the range recommended by Bhaskar and Rao (1989) for healthy fish, except for σCa, ηCa which is slightly higher than the rest of the fish species. MCHC for *Clariabranchus* (35.27 g dL⁻¹) very close to (35.47 g dL⁻¹) for *Heteroclarias* juvenile reported by Kori-Siakpere and Ubogu (2008).

**Platelet count:** Platelet values recorded in this study are lower than 132×10⁶ µL⁻¹ for juvenile *C. gariepinus* reported by Sunomonu and Oyelola (2008) and 175.92×10⁶ µL⁻¹ for *Salotherodon melanotheron* adult (Akinrotimi et al., 2007). The high platelet values observed in *C. anguillaris* and their maternal hybrid indicates that the fish species are likely to withstand and heal from bruises that might have been acquired during fighting and prevention of excessive bleeding via enactment of rapid clotting at injury site.

**Lymphocytes (LYM):** The higher lymphocytes observed in hybrid *Heteroclarias* and pure *C. anguillaris* suggested immunity for these two compared to the other cross combinations though there was no significant difference between the lymphocytes values among the entire cross combinations. Hence have the potentials of thriving well in the harsh conditions of North east Nigeria. The lymphocytes in this study are higher than 33.00% for juvenile *C. gariepinus* (Adeyemo, 2007) and 82.8% for juvenile *C. gariepinus* reported by Yaji and Auta (2007).

**CONCLUSION**

Based on this study, *Heteroclarias* may have more haematological competence for survival in culture conditions in the North east region since RBC was significantly higher in the *Heteroclarias* compared to the entire cross combination. The high PCV, Hb, MCHC and PLT observed in pure line *Clarias anguillaris* concluded that, they are harder than both their maternal and paternal hybrids. However, since most of the blood profiles of the entire cross combinations fall with the desired blood profiles of healthy fish this indicates that fish hybrids can withstand the culture conditions in region compared to their pure parental progenies.

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