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## Diversity, Pathogenicity and Toxicology of *A. niger*: An Important Spoilage Fungi

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### ABSTRACT

*Aspergillus niger*, a worldwide distributed member of ascomycotina, has been isolated from numerous habitats. *A. niger* is one of the fungi that has been labelled with the GRAS (generally recognized as safe) status from the US Food and Drug Administration. This dull or dark black looking fungus has several important products in fermentation industry. But due to cosmopolitan nature, human beings gets frequently exposed to spores and vegetative forms of *A. niger* present in air, on foodstuffs and others stored consumables products and suffers with allergic problems. *A. niger* may also produce certain mycotoxins which are hepatocarcinogenic, nephrogenic immunological in nature. In addition, this fungus is also causative agent for many rot diseases in plants. So, the present review article is an important step to understand the diversity, pathogenicity and toxicology of this important spoilage *A. niger*.

**Key words:** *Aspergillus niger*, distribution, industrial applications, pathogenicity and toxicology

### INTRODUCTION

*Aspergillus niger* (black mold), a filamentous ascomycete having ability of fast growth and pH tolerance is most important cosmopolitan fungi associated with postharvest decay of different substrates (Pitt and Hocking, 1997; Perfect *et al.*, 2009; Perrone *et al.*, 2007). This organism is a soil saprobe with a wide array of hydrolytic and oxidative enzymes involved in the breakdown of plant lignocelluloses. Because of their ability to produce extracellular organic acids some of them are commonly used in food industry. These features of *A. niger* enable them to cause decay of various organic substances including fruits, vegetables, nuts, beans, cereals, herbs, wood and herbal drugs. *A. niger* also plays a significant role in the global carbon cycle (Baker, 2006). Moreover, *A. niger* is one of the fungi that has been labelled with the GRAS (generally recognized as safe) status from the US Food and Drug Administration (Powell *et al.*, 1994). But instead of the safe categorization, *A. niger* has been found to be a opportunistic reason for infections of humans. If inhaled, in sufficient quantity it can cause severe lung problems i.e., aspergillosis in humans. It is also associated with various plant diseases resulting in huge economic loss. Beside animal and plant pathogen, *A. niger* is also reported to produce ochratoxin A and fumonisin B2 and aflatoxins (Abraca *et al.*, 1994; Schuster *et al.*, 2002; Noonimabc *et al.*, 2009; Al-Abdalall, 2009) in stored commodities, which seems to be very inevitable. Mycotoxins produced by *A. niger* are not only linked to discoloration, quality deterioration, reduction in commercial values but can also cause

several ailments of liver, kidney, nervous system, muscles, skin, respiratory organs, digestive tract, genital organs, etc. (Muntanola, 1987; Purchase, 1974; Durakovic *et al.*, 1989; Rai and Mehrotra, 2005; Truckesses and Scott, 2008). Therefore, the purpose of this review is to summarize the current knowledge like diversity, pathogenicity and toxicology about this important spoilage fungus.

**General characteristics:** *Aspergillus niger* is a versatile filamentous fungus found in soil, water, air, decaying plant material and large number of food and feeds all over the world (Pitt and Hocking, 1997). Raper and Fennell (1965) designated 15 species as comprising the *Aspergillus niger* group, which includes all of the aspergilli with black conidia, but now the concept of retention of the *A. niger* group based on black conidia seems dominant (Someren *et al.*, 1990).

*Aspergillus niger* is both a species and a group within the genus *Aspergillus*. The taxonomic description is as follows:

***A. niger* (Tiegh.) Speg.:**

Domain : Eukaryota  
Kingdom : Fungi  
Phylum : Ascomycota  
Subphylum : Pezizomycotina  
Class : Eurotiomycetes  
Order : Eurotiales  
Family : Trichocomaceae  
Genus : *Aspergillus*  
Species : *niger*

The major difference between *A. niger* and other species of *Aspergillus* is the production of carbon black or very dark brown spores from biseriate phialides (Raper and Fennell, 1965). Vegetative growth is very rapid on culture media with submerged mycelium. The hyphae are septate and hyaline more or less yellow in color. The colonies are black coloured and reverse usually colourless (Fig. 2a, b). Conidiophores mostly arise directly from substratum and are smooth, septate or nonseptate, varying greatly in length and diameter, i.e., 200-400×7-10 and 20 µ, respectively. Conidial heads are fuscous, blackish-brown to purple-brown or in every shade to carbonous black, varying from small, almost columnar masses of a few conidial chains to the common globes or radiate heads, up to 300, 500 µ, or 1000 µ long. Vesicle globose, commonly 20-50 µ up to 100 µ in diameter. Phialides typically in two series, (biseriate), thickly covering the vesicle, primary greatly varies in length, secondary 6-10×2-3 µ (Fig. 1). Conidia are globose, at first smooth, but later spinose with coloring substance, mostly 2.5-4 µ (Gilman, 2001).

The genome size of *A. niger* is about 35.5 to 38.5 Mb composed of about 13,000 genes. Of these genes, about 8000 to 8500 genes have functional assignments. In addition, about 14,000 Open Reading Frames (ORF) were identified in the genome which could potentially encode a protein. The DNA sequence of *A. niger* consists of approximately 33.9 million base pairs. The possible function of 6500 genes could be established which is only about 45% of its total gene count. Electrophoretic karyotyping of *A. niger* allows the visualization of chromosomes separated into four separate bands. The chromosomal bands range from 3.5 to 6.6 Mb. The karyotype sequence that was obtained could be arranged into 19 separate supercontigs that correspond to eight linear chromosomes (Debets *et al.*, 1990).

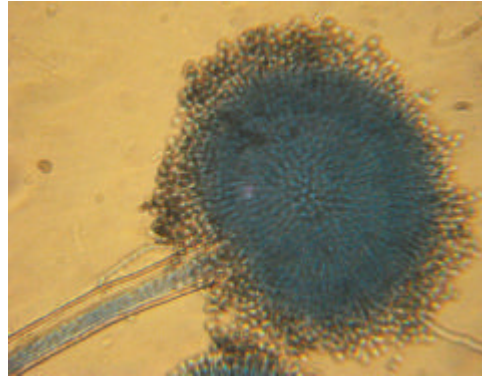


Fig. 1: Microscopic structure of *Aspergillus niger*

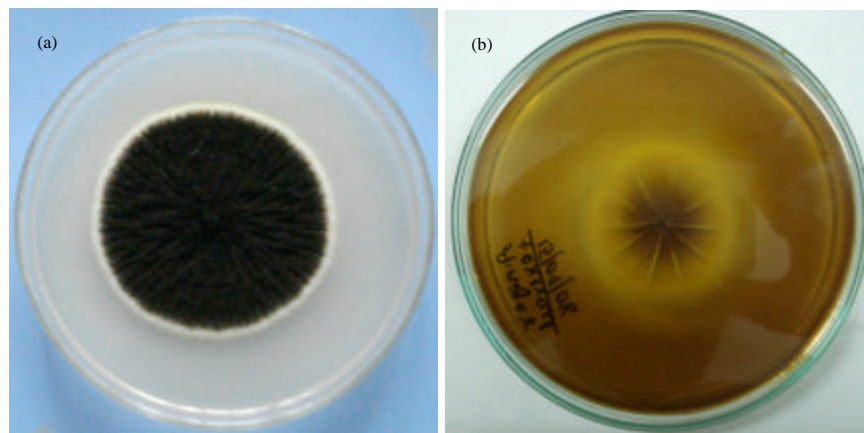


Fig. 2: Characteristics of *Aspergillus niger*: (a) Morphological and (b) Reverse side of colony

**Diversity and geographical distribution:** *Aspergillus niger* have the ability to grow in wide temperature (6-47°C) and pH range (1.4-9.8). The water activity limit for growth is 0.88, which is relatively high compared with other *Aspergillus* species. These abilities make ubiquitous occurrence of the species, with a higher frequency in warm and humid places (Palacios-Cabrera *et al.*, 2005). It can be found anywhere in and around of us (Kozakiewicz, 1989; Abarca *et al.*, 2004; Samson *et al.*, 2004). Although the main source of black aspergilli is soil (Khan *et al.*, 2007), it has also been isolated from various other sources like, air (Versar, 1991), food and food products (Agrawal *et al.*, 1980; Bennett and Klich, 1992; Mandeel, 2005; Essono *et al.*, 2007; Perrone *et al.*, 2007; Reddy *et al.*, 2009), herbs and herbal products (Gautam and Bhadauria, 2008; Gautam *et al.*, 2009; Sareen *et al.*, 2010; Gautam *et al.*, 2010, Avasthi *et al.*, 2010), fruits and fruits products (Magnoli *et al.*, 2003), etc. Not even as saprophytic fungi, *A. niger* is also isolated as parasitic/pathogenic fungi from onion (Narayana *et al.*, 2007), *Catharantus rosea* as endophytic fungi (Kharwar *et al.*, 2008) and from various other medicinally/commercially important plants (Table 1).

**Ecology:** *A. niger* is commonly isolated from soil, plant debris, air and indoor environments. In addition to producing extracellular enzymes and citric acid, *A. niger* is used for organic waste

Table 1: Diversity of *Aspergillus niger* on different substrates

Substrate	References
Soil	Gilman (2001), Thompson <i>et al.</i> (1994), Khan <i>et al.</i> (2007) and Morya and Yadav (2009)
Air	Versar (1991)
Water	Versar (1991)
Food	Agrawal <i>et al.</i> (1980), Bennett and Klich (1992), Mandeel (2005), Essono <i>et al.</i> (2007), Reddy <i>et al.</i> (2009) and Perrone <i>et al.</i> (2007)
Herbal drugs	Hitokoto <i>et al.</i> (1978), Roy and Chourasia (1990), Singh <i>et al.</i> (2008), Gautam and Bhadauria (2008, 2009), Gautam <i>et al.</i> (2009, 2010) and Sareen <i>et al.</i> (2010)
Fruits and juice	Bennett and Klich (1992), Battilani and Pietri (2002), Magnoli <i>et al.</i> (2003), Belli <i>et al.</i> (2004), Baiyewu <i>et al.</i> (2007)

management and biotransformation. The fungus is most commonly found in mesophilic environments such as decaying vegetation or soil and plants (Schuster *et al.*, 2002). *A. niger* is one of the fungi that have been labeled with the GRAS (generally recognized as safe) status according to the US Food and Drug Administration (Schuster *et al.*, 2002). The safe use of *A. niger* comes into existence from its use in the food industry for the production of many enzymes and acid proteases (Bennett, 1985; Ward, 1989). The annual production of citric acid by fermentation is now approximately 350,000 tons, using either *A. niger* or *Candida* yeast as the producing organisms. Citric acid fermentation using *A. niger* is carried out commercially in both surface culture and in submerged processes (Berry *et al.*, 1977; Ward, 1989).

By making use of industrial fermentation, *A. niger* produces many useful enzymes like amylase, amyloglucosidase, cellulases, glucoamylase, lactase, invertase, pectinases, etc. Glucoamylase is a useful enzyme used in the production of high fructose corn syrup and pectinases are used in cider and wine clarification.  $\alpha$ -galactosidase, an enzyme that breaks down certain complex sugars, is a component of Beano and other medications which the manufacturers claim can decrease flatulence. Another use for *A. niger* within the biotechnology industry is in the production of magnetic isotope-containing variants of biological macromolecules for NMR analysis (Staiano *et al.*, 2005).

Besides the production of useful enzymes, various strains of *A. niger* used in the industrial preparation of citric acid and gluconic acid, which have been assessed as acceptable for daily intake by the World Health Organisation (WHO) and Food and Drug Administration (FDA) (Schuster *et al.*, 2002). It is primarily used for the production of many enzymes such as amylase, amyloglucosidase, cellulases, lactase, invertase, pectinases and acid proteases (Bennett, 1985; Ward, 1989). In addition to production of enzymes and acids through fermentation, *A. niger* has some uses as the organism itself. Due to its ease of visualization and resistance to several antifungal agents, it is used to test the efficacy of preservative treatments (Jong and Gantt, 1987). Due to its exquisite sensitivity to micronutrient deficiencies, *A. niger* can be utilized for soil testing (Raper and Fennell, 1965). Besides, RNase produced by *A. niger* called *actibind* has antiangiogenic and anticarcinogenic characteristics (Schwartz *et al.*, 2007).

Other properties of this species include spoilage and production of secondary metabolites, such as aflatoxins, fumonisins and ochratoxins (Abraca *et al.*, 1994; Noonimabc *et al.*, 2009; Edwin *et al.*, 2010) that are toxic. The mycotoxin fumonisin B2 was recently found to be produced by *A. niger* (Noonimabc *et al.*, 2009). Metabolite production, involvement in spoilage of food and other commodities, simply being a pathogen makes the genome sequencing of this important fungus essential to biological applications (Takahashi *et al.*, 1991; May and Adams, 1997).

**Pathogenicity and toxicology:** *A. niger* is relatively harmless as compared to other filamentous fungi. Despite this fact, there have been some medical cases that have been accounted for, such as lung infections or ear infections in patients that have weakened immune system or an immune system that has been impaired by a disease or medical treatment (Schuster *et al.*, 2002; May and Adams, 1997). Besides human pathogenicity, *A. niger* can cause various plant diseases also.

***Aspergillus niger* as plant pathogenic fungi:** *A. niger* has been isolated from a variety of substrates but, these reports involve co-isolation with other perhaps more destructive microorganism or isolation from a stored product. The organism is considered as a strict saprophyte (Farr *et al.*, 1989). There are reports of *A. niger* being as plant pathogen (Fig. 3a, b, Table 2). This fungus can cause rotting of numerous fruits, vegetables and other food products, thus causing substantial economic loss. There are many examples of plant diseases caused by *A. niger*. Black rot of onions associated with *A. niger* is responsible for serious losses of onion bulbs in the field and storage (Narayana *et al.*, 2007). Other plant pathogenic reports of *A. niger* are, spoilage of mangos (Prakash and Raoof, 1989), grapes (Sharma and Vir, 1986), Tomatos (Sinha and Saxena, 1987), Shallot; stem rot of *Dracaena* (Abbasi and Aliabadi, 2008); root stalk rot of Sansevieria; and boll rot of Cotton; spoilage of cashew kernels, dates, figs, vanilla pods and dried prune (Bobbarala *et al.*, 2009). *A. niger* can induce a crown rot of peanuts due to *A. niger*-infected seed under specific hot, humid growth conditions (Anderegg *et al.*, 1976). Kharwar *et al.* (2008) isolated *A. niger* from *Catharanthes rosea* as an endophytic fungi which can alter its metabolite production.

Table 2: Plant diseases caused by *Aspergillus niger*

Name of disease	Host	Reference
Black rot of onions	<i>Allium cepa</i> L. (Onion)	Narayana <i>et al.</i> (2007)
Crown rot of peanuts	<i>Pisum Sativum</i> L. (Peanut)	Anderegg <i>et al.</i> (1976)
Tuber rot of yam	<i>Dioscorea</i> sp. (Yam)	Awuah and Akraasi (2007)
Stem rot of <i>Dracaena</i>	<i>Dracaena sanderiana</i> Mast.	Abbasi and Aliabadi (2008)
Black mold rot of cherry	<i>Prunus avium</i> L.(Cherry)	Lewis <i>et al.</i> (1963)
Kernel rot of maize	<i>Zea mays</i> L. (Corn)	Palencia <i>et al.</i> (2010)
Fruit rot of grapes	<i>Vitis</i> sp. (Grape)	Sharma and Vir (1986)
Fruit rot of banana	<i>Musa</i> sp. (Banana)	Adebesin <i>et al.</i> (2009)
Rot of Tomatoes	<i>Solanum lycopersicum</i> L. (Tomato)	Sinha and Saxena (1987)
Mango rotting	<i>Mangifera indica</i> L. (Mango)	Prakash and Raoof (1989)

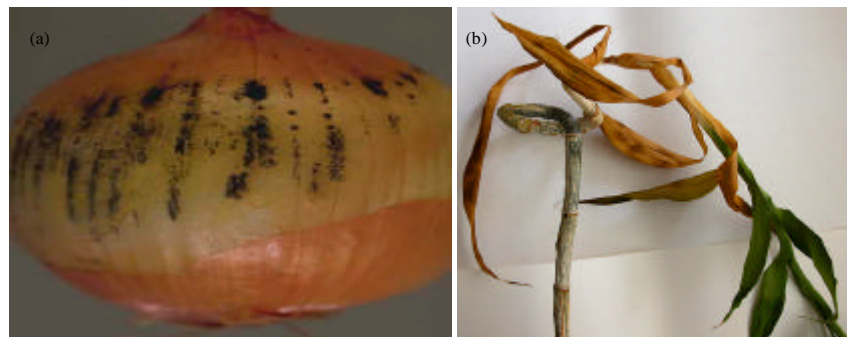


Fig. 3: Infection of *A. niger* in plants: (a) Black rot of onion and (b) Stem rot of *Dracaena sanderiana* (Abbasi and Aliabadi, 2008)

**Aspergillus niger as a human pathogen:** *Aspergillus niger* is believed to be most common storage fungi posing serious threat to contamination of stored commodities in tropical warm regions of the world. Food and herbal drug industries are very much suffering from *A. niger* and its mycotoxin contamination. It is studied that less than 10% of the *A. niger* strains were tested positive for ochratoxin A and fumonisins under conditions that were favorable (Schuster *et al.*, 2002). Living beings including humans, when contacted with *A. niger* and mycotoxins (ochratoxin A and fumonisins) usually through consumption may cause many negative effects, i.e., immunotoxicity, carcinogenicity and hepatotoxicity. The effects on animals include decrease in antibody responses, size reduction in immune organs and an alteration in the production of cytokine which are proteins and peptides specifically used in signaling. Poultry feed if contaminated by *A. niger* has major affect on the poultry industry. Different animals, such as chicken, turkey and ducks, are very prone to ochratoxin (Schuster *et al.*, 2002; May and Adams, 1997).

*Aspergillus niger* is commonly regarded as a pathogenic allergen generally associated with lung infections in individuals with weak immune system. Because the conidia and conidiophores are small, readily air borne, can easily be breathed in and cause deep or systemic mycosis (Kierownik, 1990) (Table 3). Ear is the location of *A. niger* infection (Fig. 4). Local lesions in both external and middle ear, as well as in post operative cavities, can create favourable conditions for fungal growth and subsequent otomycosis (Kaur *et al.*, 2000; Kurnatowski and Kilipiak, 2001). *A. niger* can produce a secondary metabolites include oxalic acids, kojic acids abundantly and cyclic pentapeptides having moderate to high acute toxicity (Ueno and Ueno, 1978). Oxalate crystals of oxalic acids produced by *A. niger* can cause pulmonary oxalosis (Nakagawa *et al.*, 1999) (Fig. 5).

**Other risks:** Apart from the human and plant pathogenic effects, there are so many risks/problems being associated with *A. niger*. One of the most important one is its ability to grow on a

Table 3: Human diseases caused by *Aspergillus niger*

Name of disease	Target organ	Reference
Weak immune system	Any organ of the body	Louthrenoo <i>et al.</i> (1990)
Systemic mycosis	External body part	Louthrenoo <i>et al.</i> (1990)
Ear infection	External auditory system	Padhye (1982), Walsh and Pizzo (1988)
Aspergillosis	Lungs	Ueno and Ueno (1978), Bennett (1979), Richard <i>et al.</i> (2008)
Asthma and allergic alveolitis	Respiratory tract	Edwards and AlZubaidy (1977)

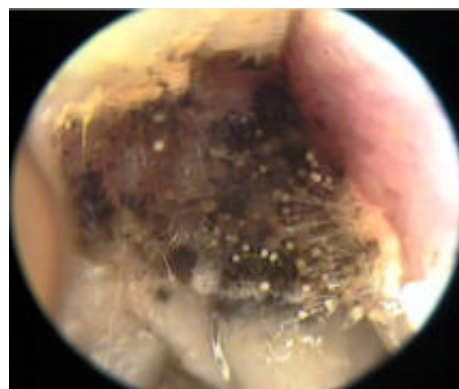


Fig. 4: Ear infection by *A. niger* in external auditory

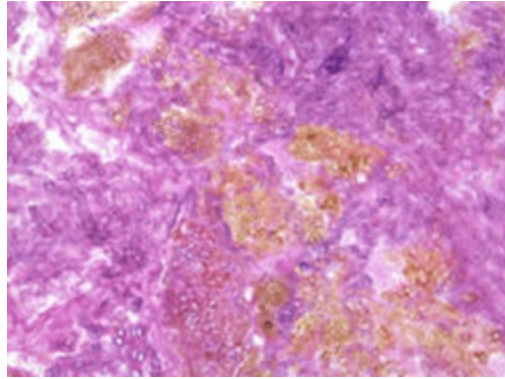


Fig. 5: Pulmonary oxalosis due to fungus ball of *Aspergillus niger*. The band of necrosis is due to the diffusion of oxalate crystals (Richard *et al.*, 2008)

variety of substrates, causing deterioration of materials. This spoilage or deterioration not only reduce the quality of the substrate but also alter its active components and commercial value. For example, *A. niger* causes economic losses due to spoilage of bakery products, fruit, herbal drugs and vegetables. *A. niger* also damages surface layers of wood, raw cotton fibers and many other materials. However, because *A. niger* is already ubiquitous, the increased environmental burden of *A. niger* due to release from commercial facilities is probably negligible. Thus, the baseline risk of materials damage by *A. niger* will not be affected by the use of *A. niger* in commercial facilities.

## CONCLUSION

*Aspergillus niger* is found anywhere in and around of us. In industrial fermentation, *A. niger* produces many useful enzymes like amylase, amyloglucosidase, cellulases, glucoamylase, lactase, invertase, pectinases. This enhances the importance of fungi in food and drug industries. In addition to beneficial aspects, human beings get frequently exposed to *A. niger* spores and vegetative forms present in air and on foodstuffs and suffers with allergic problems whereas, specific strains may produce mycotoxins, elicit allergy and carcinogenic responses to lungs, kidney and liver. Although, limited instances of adverse effects seems to be associated with a limited number of strains of *A. niger*. With proper characterization of different strains, industrial exploration of this important fungus can be increased and potential adverse effects can be avoided.

## REFERENCES

- Abarca, M.L., F. Accensi, J. Cano and F.J. Cabanes, 2004. Taxonomy and significance of black aspergilli. *Antonie Van Leeuwenhoek*, 86: 33-49.
- Abbasi, M. and F. Aliabadi, 2008. First report of stem rot of *Dracaena* caused by *Aspergillus niger* in Iran. *Plant Health Progress*, 10.1094/PHP-2008-0212-01-BR
- Abraca, M.L., G. Bragulat and F.J. Cabanes, 1994. Ochratoxin A production by strains of *Aspergillus flavus* var. *niger*. *Applied Environ. Microbiol.*, 60: 2650-2652.
- Adebesin, A.A., C.A. Odebode and A.M. Ayodele, 2009. Control of postharvest rots of banana fruits by conidia and culture filtrates of *Trichoderma asperellum*. *J. Plant Prot. Res.*, 49: 303-308.
- Agrawal, G.P., M.K. Thakur and S. Awasthi, 1980. Studies on the wheat grain storage on Madhya Pradesh fungi associated with different varieties of freshly harvested wheat grains. *Nat. Acad. Sci. Lett.*, 3: 195-197.



- Al-Abdalall, A.H.A., 2009. Production of aflatoxins by *Aspergillus flavus* and *Aspergillus niger* strains isolated from seeds of pulses. J. Food Agric. Environ., 7: 33-39.
- Anderegg, R.J., K. Biemann, G. Buechi and M. Cushman, 1976. Malformin C, a new metabolite of *Aspergillus niger*. J. Amer. Chem. Soc., 98: 3365-3370.
- Avasthi, S., A.K. Gautam and R. Bhadauria, 2010. Antifungal activity of some commonly used spices against *A. niger*: A potential application in the control of a spoilage fungus. Biol. Forum Int. J., 2: 53-55.
- Awuah, R.T. and K.O. Akraasi, 2007. Suppression of rot of yam caused by *Aspergillus niger* with a yam *Rhizobacterium*. Afr. Crop Sci. Conf. Proc., 8: 875-879.
- Baiyewu, R.A., N. A. Amusa, O.A. Ayoola and O.O. Babalola, 2007. Survey of the post harvest diseases and aflatoxin contamination of marketed pawpaw fruit (*Carica papaya* L.) in South Western Nigeria. Afr. J. Agric. Res., 2: 178-181.
- Baker, S., 2006. *Aspergillus niger* genomics: Past, present and into the future. Med. Mycol., 44: 17-21.
- Battilani, P. and A. Pietri, 2002. Ochratoxin A in grapes and wine. Eur. J. Plant Patho., 108: 639-643.
- Belli, N., E. Pardo, S. Marín, G. Farre, A.J. Ramos and V. Sanchis, 2004. Occurrence of ochratoxin A and toxigenic potential of fungal isolates from Spanish grapes. J. Sci. Food Agric., 84: 541-546.
- Bennett, J.E., 1979. Aspergillosis. In: Cecil Textbook of Medicine, Beeson, P., W. McDermott and J. Wyngaarden (Eds.). W.B. Saunders, Philadelphia, pp: 546-547.
- Bennett, J.W., 1985. Molds, Manufacturing and Molecular Genetics. In: Molecular Genetics of Filamentous Fungi, Timberlake, W.E. (Ed.). Alan R. Liss Inc., New York.
- Bennett, J.W. and M.A. Klich, 1992. *Aspergillus*: Biology and Industrial Applications. Butterworth Heinemann Publication, USA., ISBN: 0-7506-9124-7.
- Berry, D., A. Chmiel and Z. Al Obaidi, 1977. Citric acid Production by *Aspergillus niger*. In: Genetics and Physiology of *Aspergillus*, Smith, J.E. and J.A. Pateman (Eds.). Academic Press, New York.
- Bobbarala, V., P.K. Katikala, K.C. Naidu and S. Penumajji, 2009. Antifungal activity of selected plants extracts against phytopathogenic fungi *Aspergillus niger*. Ind. J. Sc. Tech., 20: 87-90.
- Debets, A., E. Holub, K. Swart, H. van den Broek and C. Bos, 1990. An electrophoretic karyotype of *Aspergillus niger*. Mol. Gen. Genet., 224: 264-268.
- Durakovic, S., J. Galic and P. Pajnovic, 1989. Toxic and cancer metabolites of moulds in food and fodder. Hrana I. Ishrana, 30: 71-100.
- Edwards, J.H. and T.S. AlZubaidy, 1977. Medical Aspects. In: Genetics and Physiology of *Aspergillus*, Smith, J.E. and J.A. Pateman (Eds.). Academic Press, New York.
- Edwin, R., I. Palencia, D.M. Hinton and C.W. Bacon, 2010. The black *Aspergillus* species of maize and peanuts and their potential for mycotoxin production. Toxins, 2: 399-416.
- Essono, G., M. Ayodele, A. Akoa, J. Foko, S. Olemb and J. Gock, 2007. *Aspergillus* species on cassava chips in storage in rural areas of southern Cameroon: Their relationship with storage duration, moisture content and processing methods. Afr. J. Microbiol., 1: 001-008.
- Farr, D.F., G.F. Bills, G.P. Chamuris and A.Y. Rossman, 1989. Fungi on Plants and Plant Products in the United States. APS Press, St. Paul, MN..
- Gautam, A.K. and R. Bhadauria, 2008. Occurrence of toxigenic moulds and mycotoxins in ayurvedic medicine *Trifla Churn*. J. Myco. Plant Path., 38: 664-666.

- Gautam, A.K. and R. Bhadauria, 2009. Fungal contamination of few important herbal fruit samples. Internet J. Nut. Wellness, USA. Vol. 8.
- Gautam, A.K., S. Sharma and R. Bhadauria, 2009. Detection of toxigenic fungi and mycotoxins in medicinally important powdered herbal drugs. Internet J. Microbiol., Vol. 7.
- Gautam, A.K., S. Avasthi, A. Sharma and R. Bhadauria, 2010. Efficacy of raw materials of triphala churn against *A. niger* and potential of clove extract as herbal fungitoxicant. Biol. Med., 2: 1-9.
- Gilman, J.C., 2001. A Manual of Soil Fungi. 2nd Edn., Biotech Books, New Delhi.
- Hitokoto, H., S. Morozumi, T. Wauke, S. Saka and H. Kurata, 1978. Fungal contamination and mycotoxin detection of powdered herbal drugs. Applied Environ. Microbiol., 36: 252-256.
- Jong, S.C. and M.J. Gantt, 1987. Catalogue of Fungi and Yeasts. 17th Edn., American Type Culture Collection, Rockville, MD.
- Kaur, R., N. Mittal, M. Kakkar, A.K. Aggarwal and M.D. Mathur, 2000. Otomycosis: A clinicomycologic study. Ear Nose Throat J., 79: 606-609.
- Khan, M.R., M.A. Anwer and F.A. Mohiddin, 2007. Molecular diversity in *Aspergillus* isolates collected from pigeon pea field in Aligarh region. Environ. Bio. Conser., 12: 59-64.
- Kharwar, R.N., V.C. Verma, G. Strobel and D. Ezra, 2008. The endophytic fungal complex of *Catharanthus roseus* (L.) G. Don. Curr. Sci., 95: 228-233.
- Kierownik, 1990. Pulmonary aspergillosis caused by *Aspergillus niger*. Pneumonol. Pol., 58: 328-333.
- Kozakiewicz, Z., 1989. *Aspergillus* species on stored products. Mycol. Pap., 161: 1-188.
- Kurnatowski, P. and A. Kilipiak, 2001. Otomycosis: Prevalence, clinical symptoms, therapeutic procedure. Mycoses, 45: 472-479.
- Lewis, J.C., C.F. Pierson and M.J. Powers, 1963. Fungi Associated with softening of bisulfite-brined Cherries. Applied Microbiol., 11: 93-99.
- Louthrenoo, W., Y.S. Park, L. Philippe and H.R. Schumacher, 1990. Localized peripheral calcium oxalate crystal deposition caused by *Aspergillus niger* infection. J. Rheumato., 17: 407-412.
- Magnoli, C., M. Violante, M. Combina, G. Palacio and A. Dalcerro, 2003. Mycoflora and ochratoxin: A producing strains of *Aspergillus* section *Nigri* in wine grapes in Argentina. Lett. Applied Microbiol., 37: 179-184.
- Mandeel, Q.A., 2005. Fungal contamination of some imported species. Mycopath., 159: 291-298.
- May, G. and T. Adams, 1997. The Importance of fungi to Man. Genome. Res., 7: 1041-1044.
- Morya, V. and D. Yadav, 2009. Isolation and screening of different isolates of *Aspergillus* for amylases production. Internet J. Microbiol., Vol. 7.
- Muntanola, M., 1987. General Mycology. NIRO. Knjez Evne Novine., Beograd, pp: 257-269.
- Nakagawa, Y., K. Shimazu, M. Ebihara and K. Nakagawa, 1999. *Aspergillus niger* pneumonia with fatal pulmonary oxalosis. J. Infect. Chemother., 5: 97-100.
- Narayana, K.J.P., M. Srikanth, M. Vijayalakshmi and N. Lakshmi, 2007. Toxic spectrum of *Aspergillus niger* causing black mold rot of onions. Res. J. Microbiol., 2: 881-884.
- Noonimabc, P., W. Mahakarnchanakulb, K.F. Nielsend, J.C. Frisvadd and R.A. Samsona, 2009. Fumonisin B2 production by *Aspergillus niger* in Thai coffee beans. Food Addit. Contam., 26: 94-100.
- Padhye, A.A., 1982. Fungi Pathogenic to Man and Animals. In: CRC Handbook of Microbiology, Laskin, A. and H.A. Lechevalier (Eds.). CRC Press, West Palm Beach, FL.

- Palacios-Cabrera, H., MH, Taniwaki, J.M. Hashimoto and H.C. De Menezes, 2005. Growth of *Aspergillus ochraceus*, *A. carbonarius* and *A. niger* on culture media at different water activities and temperature. Braz. J. Microbiol., 36: 24-28.
- Palencia, E.R., D.M. Hinton and C.W. Bacon, 2010. The black *Aspergillus* species of maize and peanuts and their potential for mycotoxin production. Toxins, 2: 399-416.
- Perfect, J.R., G.M. Cox, J.Y. Lee, S.W. Chapman and C.A. Kauffman *et al.*, 2009. The impact of culture isolation of *Aspergillus* species: A hospital-based survey of aspergillosis. Clin. Infect. Dis., 33: 1824-1833.
- Perrone, G., A. Susca, G. Cozzi, K. Ehrlich and J. Vargas *et al.*, 2007. Biodiversity of *Aspergillus* species in some important agricultural products. Stud. Mycol., 59: 53-66.
- Pitt, J.I. and A.D. Hocking, 1997. Fungi and Food Spoilage. 2nd Edn., Springer, New York, ISBN-10: 0834213060, pp: 593.
- Powell, K.A., A. Renwick and J.F. Peberdy, 1994. The genus *Aspergillus*, from taxonomy and genetics to industrial application. Plenum Press, New York.
- Prakash, O. and M.A. Raoof, 1989. Control of mango fruit decay with post harvest application of various chemicals against black rot, stem end rot and anthracnose disease. Int. J. Trop. Plant Dis., 6: 99-106.
- Purchase, I.F.H., 1974. Mycotoxins. Elsevier, Amsterdam, pp: 1-28.
- Rai, V. and S. Mehrotra, 2005. Toxic contaminants in herbal drugs. Environ. News Arch., 11: 1-3.
- Raper, K.B. and D.I. Fennell, 1965. The genus *Aspergillus*. Williams and Wilkins Company, Baltimore, MD.
- Reddy, K.R.N., C.S. Reddy and K. Muralidharan, 2009. Detection of *Aspergillus* spp. and aflatoxin B1 in rice in India. Food Microbiol., 26: 27-31.
- Richard, L., M.D. Kradin, J. Eugene and M.D. Mark, 2008. The pathology of pulmonary disorders due to *Aspergillus* sp. Arch. Path. Lab. Med., 132: 606-614.
- Roy, A.K. and H.K. Chourasia, 1990. Mycoflora, mycotoxin producibility and mycotoxins in traditional herbal drugs from India. J. Genet. Applied Microbiol., 36: 295-302.
- Samson, R.A., J.A.M.P. Houbraken, A.F.A. Kuijpers, J.M. Frank and J.C. Frisvad, 2004. New ochratoxin or sclerotium producing species in *Aspergillus* section Nigri. Stud. Mycol., 50: 45-61.
- Sareen, A., R. Ahirwar, A. Gautam and R. Bhadauria, 2010. Fungal contamination of some common medicinal plant samples of Himachal Pradesh. Sci. Cult., 76: 118-120.
- Schuster, E., N. Dunn-Coleman, J. Frisvad and P. van Dijck, 2002. On the safety of *Aspergillus niger*: A review. Applied Microbiol. Biotech., 59: 426-435.
- Schwartz, B., O. Shoseyov, V.O. Melnikova, M. McCarty and M. Leslie *et al.*, 2007. ACTIBIND, a T2 RNase, competes with angiogenin and inhibits human melanoma growth, angiogenesis and metastasis. Cancer Res., 67: 5258-5266.
- Sharma, R.C. and D. Vir, 1986. Post harvest diseases of grapes and studies on their control with benzimidazole derivatives and other fungicides. Pesticides, 20: 1415-1415.
- Singh, P., B. Srivastava, A. Kumar and N.K. Dubey, 2008. Fungal contamination of raw materials of some herbal drugs and recommendation of *Cinnamomum camphora* oil as herbal fungitoxicant. Micro. Eco., 56: 555-560.
- Sinha, P. and S.K. Saxena, 1987. Effect of treating tomatoes with leaf extract of *Lantana camara* on development of fruit rot caused by *A. niger* in presence of *Drosophila busckii*. Indian J. Exp. Biol., 25: 143-144.

- Somerén, K., H.C.M. Kester, R.A. Samson and J. Visser, 1990. Variations in Pectolytic Enzymes of the Black *Aspergilli*: A Biochemical and Genetic Approach. In: Modern Concepts in *Penicillium* and *Aspergillus* Classification, Samson, R.A. and J.I. Pitt (Eds.). Plenum Press, New York.
- Staiano, M., P. Bazzicalupo, M. Rossi and S. DAuria, 2005. Glucose biosensors as models for the development of advanced protein-based biosensors. *Mol. BioSyst.*, 1: 354-362.
- Takahashi, K., H. Inoue, K. Sakai, T. Kohama and S. Kitahara *et al.*, 1991. The primary structure of *Aspergillus niger* acid proteinase A. *J. Biol. Chem.*, 266: 19480-19483.
- Thompson, L., M.A. Castrillon, M. Delgado and M. Garcia, 1994. Isolation of several species of the genus *Aspergillus* from soil of intrahospital ornamental plants. *Revista Medica de Chile*, 122: 1367-1371.
- Truckesses, M.W. and P.M. Scott, 2008. Mycotoxins in botanicals and dried fruits: A review. *Addit. Cont.*, 25: 181-192.
- Ueno, Y. and I. Ueno, 1978. Toxicology and Biochemistry of Mycotoxins. In: Toxicology, Biochemistry and Pathology of Mycotoxins, Uruguchi, K. and M. Yamazaki (Eds.). John Wiley and Sons, Halstead Press, New York.
- Versar, 1991. Screening Level Exposure Assessment of *Aspergillus* Species for 5(h)(4) Exemption Under the Proposed Biotech Rule. U.S. Environmental Protection Agency, Washington, D.C.
- Walsh, T.J. and P.A. Pizzo, 1988. Nosocomial fungal infections: A classification for hospital-acquired fungal infections and mycoses arising from endogenous flora or reactivation. *Ann. Rev. Microbiol.*, 42: 517-545.
- Ward, O.P., 1989. Fermentation Biotechnology. Prentice Hall, Englewood Cliffs, New Jersey.