# Specifying Human Platelet cAMP and cGMP Phosphodiesterase Inhibitory Activity of the Plants Used in Traditional Iranian Medicine for the Purpose of Erectile Dysfunction 

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#### Abstract

In the present study, cAMP and cGMP phosphodiesterase inhibitory (PDEI) activities of the ethanolic extracts of nineteen plants of Traditional Iranian Medicine (TIM) with aphrodisiac effects were investigated. The plants were extracted in a similar way and then three concentrations ( $0.1,1,5 \mathrm{mg} \mathrm{mL}^{-1}$ ) from each were tested for PDEI activity against control and sildenafil. Among plants tested, 8 including Allium cepa, Trigonella foenum-graecum, Brassica rapa, Alpinia officinarum, Tribulus terrestris, Urtica pilulifera, Linum usitatissimum and Zingiber zerumbet exhibited a significant dose-dependent cAMP-PDEI activity and 6 including A. cepa, A. officinarum, T. terrestris, L. usitatissimum, Withania somnifera and Z. zerumbet gave a remarkable dose-dependent cGMP-PDEI effects in comparison to control. Among tested herbs, Zingiber officinalis and Peganum harmala at dose of $5 \mathrm{mg} \mathrm{mL}^{-1}$ demonstrated better cGMP-PDEI in comparison to sildenafil. The results of this study give idea for discovery of safe and better drugs in management of erectile dysfunction.


Key words: Phosphodiesterase inhibitor, sildenafil, traditional Iranian medicine, cAMP, cGMP

## INTRODUCTION

Phosphodiesterases (PDEs) are a class of enzymes that exist in nearly all tissues regulating the second messengers cAMP and cGMP involving in many diverse physiological functions. PDE inhibitors (PDEIs) are used for management of clinical disorders such as dementia, depression, cardiovascular diseases, diabetes, chronic obstructive pulmonary diseases, colitis, toxicities, pulmonary hypertension and Erectile Dysfunction (ED) (Abdollahi et al., 2003a, b, c; Jeon et al., 2005; Khoshakhlagh et al., 2007; Milani et al., 2005). Inhibitors of PDE type 5 (PDE5I) i.e., sildenafil, vardenafil and tadalafil are now used in treatment of ED but have some common side effects like headache, flushing and blurred vision. As reviewed by Rahimi et al. (2010), there are many single and
combination herbal formulas in Traditional Iranian Medicine (TIM) for ED that needs experimental and clinical testing. The aim of the present study was to evaluate PDEI activity of the medicinal plants claimed to be beneficial for ED in TIM.

## MATERIALS AND METHODS

Materials: Different parts of nineteen plants that were selected on the basis of TIM data were prepared from the clinic of Traditional Medicine, Tehran University of Medical Science (TUMS), Tehran, Iran, in May 2010. Voucher specimens have been deposited in the Central Herbarium of Medicinal Plants (ACECR, Karaj, Iran). All chemicals or reagents needed in this study were purchased from Sigma-Aldrich Company (Germany) unless otherwise stated.

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Extraction method: Dried and semi powdered plants ( 20 g ) were extracted using ethanol $80 \%(3 \times 60 \mathrm{~mL})$ at room temperature for a week. After removal of the solvent in the vacuum at $30^{\circ} \mathrm{C}$, the residues were used for further analysis.

Assay of platelet cAMP and cGMP PDEI activity: The human platelets as a rich source of PDE enzymes were purchased from Tehran blood transfusion organization and stored at $25^{\circ} \mathrm{C}$ in a shaker during the process of experiment. An aqueous stock solution of $40 \mathrm{mM} \mathrm{NaCl}, 0.1 \mathrm{mM} \mathrm{MnCl} 2,125 \mu \mathrm{M}$ phenol red were prepared and the pH of this solution adjusted at 8.0. The assay mixture $(200 \mu \mathrm{~L})$ for control contained $53 \mu \mathrm{~L}$ of stock solution and $49 \mu \mathrm{~L}$ of platelets. Then cAMP and cGMP ( 1.9 mM ) were added separately to the wells. The method is based on the absorbance level of the incorporating phenol red with titratable proton liberated from hydrolysis of cyclic nucleotide to corresponding 5'-phosphate ester by PDE (Frielle et al., 1979). The $49 \mu \mathrm{~L}$ of plant extract that was dissolved in dimethyl sulfoxide (DMSO) $10 \%$ was added to $53 \mu \mathrm{~L}$ of stock solution and $49 \mu \mathrm{~L}$ of platelets and incubated for 20 min at $25^{\circ} \mathrm{C} .49 \mu \mathrm{~L}$ of cAMP and cGMP ( 1.9 mM ) were separately added to the wells and then after 5 min the absorbance of the test samples were read at 560 nm using a fluorescence microplate ELISA reader. The extracts with various concentrations ranging from 0.1 to $5 \mathrm{mg} \mathrm{mL}^{-1}$ were tested. Each experiment was carried out in duplicate wells and the absorbance was read triplicate. Sildenafil was used as the standard solution and blank was $53 \mu \mathrm{~L}$ of stock solution and $49 \mu \mathrm{~L}$ of platelets with DMSO $10 \%$. Three concentrations $\left(0.1,1,5 \mathrm{mg} \mathrm{mL}^{-1}\right)$ of the extracts and sildenafil $(40,2,20 \mu \mathrm{M})$ were tested.

Statistical analysis: Mean and standard error values were determined for all the parameters and the results were expressed as Mean $\pm$ SEM. All data were analyzed using analysis of variance ANOVA followed by Newman Keuls. Differences between groups was considered significant when $\mathrm{p}=0.05$.

## RESULTS AND DISCUSSION

Nineteen plants of TIM with aphrodisiac effects were selected and extracted with ethanol. The extraction ratio for each plant was shown in Table 1. As shown in Fig. 1, significant dose-dependent cAMP-PDEI activity were observed from eight plants including Allium cepa, Trigonella foenum-graecum, Brassica rapa, Alpinia officinarum, Tribulus terrestris, Urtica pilulifera, Linum
usitatissimum and Zingiber zerumbet. They showed the most cAMP-PDEI activity at $0.1 \mathrm{mg} \mathrm{mL}^{-1}$, whereas the extracts of Zingiber officinalis and Peganum harmala demonstrated the most cAMP-PDEI effect at $5 \mathrm{mg} \mathrm{mL}^{-1}$. The rest of the extracts showed similar cAMP-PDEI effect in three tested concentrations in comparison to sildenafil. As shown in Fig. 2, six plants including Allium cepa, Alpinia officinarum, Tribulus terrestris, Linum usitatissimum, Withania somnifera and Zingiber zerumbet showed significant dose-dependent cGMP-PDEI activity. The most cGMP PDEI effect was seen with $0.1 \mathrm{mg} \mathrm{mL}^{-1}$ of these plants, while Zingiber officinalis and Peganum harmala had the most effect at $5 \mathrm{mg} \mathrm{mL}^{-1}$. The rest of the extracts showed the same cGMP-PDEI effect in the three concentrations used in comparison to sildenafil. In this study, a total of 19 plants were screened for assaying inhibitory potential on platelets cAMP and cGMP PDEs. In comparison to sildenafil, 10 plants showed better cAMP-PDEI effect and 8 plants exhibited better cGMP-PDEI effect. These extracts showed the most PDEI activity at $0.1 \mathrm{mg} \mathrm{mL}^{-1}$, whereas $Z$. officinalis and $P$. harmala showed a different pattern with the most PDEI effect at $5 \mathrm{mg} \mathrm{mL}^{-1}$ that might be related to antiplatelet activity of both plants (Guh et al., 1995; Im et al., 2009; Thomson et al., 2002). A. cepa and P. harmala have been previously shown for antiasthmatic effect (Duke, 2002; Wagner et al., 1990). A. cepa, B. rapa, L. usitatissimum, T. foenum-graecum, $U$. pilulifera, $W$. somnifera, Z. officinalis and $Z$. zerumbet have already shown antioxidant activity (Hu et al., 2007; Singh et al., 2009; Kim et al., 2006). A. cepa, A. officinarum, P. harmala, T. foenum-graecum, W. somnifera, Z. officinalis and Z. zerumbet have already shown anticancer or antitumor effects (Brown et al., 2009; Huang et al., 2005; Kaileh et al., 2007; Lamchouri et al., 1999; Singh et al., 2009; Sur et al., 2001; Yasukawa et al., 2008). A. cepa and Z. officinalis are currently used as cardiotonic and cardiovascular (Shoji et al., 1982; Park et al., 2009). A. officinarum and $P$. harmala are found as antispasm (Duke, 2002; Gruenwald et al., 2007). A. officinarum, T. foenum-graecum, $W$. somnifera, U. pilulifera, $Z$. officinalis and $Z$. zerumbet are used as antiinflammatory agents (Aimbire et al., 2007; Kavalali and Tuncel, 1997; Lee et al., 2009; Vyas et al., 2008; Zakaria et al., 2010a). P. harmala and T. terrestris have shown vasodilator property (Berrougui et al., 2006; Phillips et al., 2006). W. somnifera and Z. zerumbet exhibited immunomodulatory action (Rasool and Varalakshmi, 2006; Yob et al., 2011). P. harmala and Z. zerumbet were identified as analgesic (Farouk et al., 2008; Zakaria et al., 2010b). P. harmala, Z. officinalis and

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Table 1: The plants used in TIM for erection dysfunction

| Table 1: The plants used in |  | Voucher Family <br> number name | Part <br> used | Extraction <br> ratio (\%) | Persian <br> name | Chemical constituents |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |

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Table 1: Continued

| Scientific name | Voucher number | Family name | Part used | Extraction ratio (\%) | Persian name | Chemical constituents | Pharmacological effect |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Pegamum harmala L . | 1332 | Zygophylaceae | Seed | 6.63 | Esfand | Harmin, harmalin, harmane, harmalol Hemmateenejad et al. (2006), vasicine, vasicinon Pulpati et al. (2008) | Analgesic Farouk et al. (2008), vasorelaxant Berrougui et al. (2006) antispasmodic, aphrodiasiac, antiasmathic Duke (2002), anticancer Lamchouri et al. (1999), antiplatelet activity Im et al. (2009) |
| Pinusgeradiana Wall ex D. Don | 1333 | Pinaceae | Seed | 12.55 | Chalghouzeh |  |  |
| Tribulus terrestris L. | 1334 | Zygophyllaceae | Fruit | 13. $25 \%$ | Kharkhasak | Steroidal saponin Bedir et al. (2002), phenolic compound Ivanova et al. (2009), flavonoids Bhutani et al. (1969) | Diuretic DerMarderosian (2008), aphrodisiac Malviya et al. (2011), management of ED, Gauthaman and Ganesan (2008) vasodilator Phillips et al. (2006) |
| Trigonella foenum-graecum L . | 1335 | Papilionaceae | Seed | 13.8 | Shanbalile | Mucilages, protein, steroidal saponins, sterols, flavonoids, trigonellin, volatile oil Gruenwald et al. (2007) | Anti-inflammatory Vyas et al. (2008), antitumor Sur et al. (2001), antioxidant Genet et al. (2002) |
| Urica pilulifera L. | 1336 | Urticaceae | Seed | 7.80 | Anjoreh | - | Increasing sperm count and motility Irshaid and Mansi (2009) antioxidant Ozen etal. (2010), hypogly cemic activity Kavalali et al. (2003), anti-inflammatory Kavalali and Tuncel (1997) |
| Withania somnifera (L.) Dunal | 1337 | Solanaceae | Root | 8.85 | Bouzidan | Highly oxygenated steroid, alkaloids, flavonolgly cosides, phenolic acid DerMarderosian (2008) | Immunomudolatory Rasool and Varalakshm (2006), antioxidative Misra et al. (2009), aphrodisiac Malviya et al. (2011), anti-inflammatory, anticancer Kaileh et al. (2007) |
| Zingiber officinalis Roscoe | 1338 | Zingiberaceae | Rhizome | 16.1 | Zanjebil | Volatile oil, arylalkanes, gingerols, shogaols, gingerdiols, diarylheptanoids, starch Gruenwald et al. (2007), sesqiterpens DerMarderosian (2008), alkaloide Syamkumar et al. (2003) | enhancer of testosterone production DerMarderosian (2008), aphrodisiac Qureshi et al. (1989), antiinflammatory Aimbire et al. (2007), antioxidant Stoilova et al. (2007), antitumor Brown et al. (2009), cardiotonic Shoji et al. (1982), antithrombotic Thomson et al. (2002), antiplatelet activity Guh et al. (1995) |
| Zingiber zerumbet (L.) Sm. | 1339 | Zingiberaceae | Rhizome | 9.05 | Zorombad | Zerumbone Huang et al. (2005), essential oil Zakaria et al. (2010a), humulene, monoterpenes, sesquiterpenoids, flavonoids, aromatic compounds Yob et al. (2011) | Antitumor Huang et al. (2005), anti-inflammatory Zakaria et al. (2010a), antinociceptive Zakaria et al. (2010b),antipyretic, hepatoprotective,immunomodulatory, antiplatelet aggregation, antioxidant Yob et al. (2011) |



Fig. 1: Inhibitory effects of 10 extracts on cAMP-PDE activity in vitro. Values are expressed as Mean $\pm$ SE. ${ }^{* *}$ Significantly different from Sildenafil (Sild) groups at ( $\mathrm{p}<0.01$ )


Fig. 2: Inhibitory effects of 8 extracts on cGMP-PDE activity in vitro. Values are expressed as Mean $\pm$ SE. **Significantly different from Sildenafil (Sild) groups at ( $\mathrm{p}<0.01$ )
Z. zerumbet have shown antithrombotic and antiplatelet aggregation effects ( $\operatorname{Im}$ et al., 2009; Thomson et al., 2002; Guh et al., 1995; Yob et al., 2011). A. officinarum and $U$. pilulifera increase motility and count of sperm (DerMarderosian, 2008; Irshaid and Mansi, 2009). T. terrestris is currently used in management of ED (Gauthaman and Ganesan, 2008). All of these therapeutic effects could be relevant to PDEI properties (Rahimi et al., 2010). In our previous paper, it was indicated that main components of the plants with PDEI effects are flavonoids, alkaloids, saponins, lignans and coumarins (Rahimi et al., 2010) that are seen in all 19 plants tested here (Table 1). Among tested herbs, $Z$. officinalis and $P$. harmala at $5 \mathrm{mg} \mathrm{mL}^{-1}$ showed better cGMP-PDEI in comparison to sildenafil. Z. officinalis includes volatile oils, alkaloid and oleoresins (DerMarderosian, 2008; Gruenwald et al., 2007;

Syamkumar et al., 2003) while $P$. harmala contains alkaloid (Hemmateenejad et al., 2006; Pulpati et al., 2008).

## CONCLUSION

The present study for the first time reports PDEI activities of TIM plants that is notable in the field of drug discovery for ED or other relevant diseases. It is required to complete animal and then clinical profile of these extracts. Also, combinative use of these plants may reveal more efficiency in ED with respect to synergetic effect of constituents.

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## REFERENCES

Abdollahi, M., A. Bahreini-Moghadam, B. Emami, F. Fooladian and K. Zafari, 2003a. Increasing intracellular cAMP and cGMP inhibits cadmiuminduced oxidative stress in rat submandibular saliva. Comp. Biochem. Physiol. C. Toxicol. Pharmacol., 135: 331-336.
Abdollahi, M., T.S. Chan, V. Subrahmanyam and P.J. O'Brien, 2003b. Effects of phosphodiesterase $3,4,5$ inhibitors on hepatocyte cAMP levels, glycogenolysis, gluconeogenesis and susceptibility to a mitochondrial toxin. Mol. Cell Biochem., 252: 205-211.
Abdollahi, M., F. Fooladian, B. Emami, K. Zafari and A. Bahreini-Moghadam, 2003c. Protection by sildenafil and theophylline of lead acetate-induced oxidative stress in rat submandibular gland and saliva. Hum. Exp. Toxicol., 22: 587-592.
Aimbire, F., S.C. Penna, M. Rodrigues, K.C. Rodrigues, R.A. Lopes-Martins and J.A.A. Sertie, 2007. Effect of hydroalcoholic extract of Zingiber officinalis rhizomes on LPS-induced rat airway hyperreactivity and lung inflammation. Prostag. Leukotr. Essent. Fatty Acids, 77: 129-138.
Al-Yahya, M.A., J.S. Mossa, A.M. Ageel and S. Rafatullah, 1994. Pharmacological and safety evaluation studies in Lepidium sativum L. seeds. Phytomedicine, 1: 155-159.
Bedir, E., I.A. Khan and L.A. Walker, 2002. Biologically active steroidal glycosides from Tribulus terrestris. Pharmazie, 57: 491-493.
Berrougui, H., C. Martin-Cordero, A. Khalil, M. Hmamouchia, A. Ettaib, E. Marhuenda and M.D. Herrera, 2006. Vasorelaxant effects of harmine and harmaline extracted from Peganum harmala L. seeds in isolated rat aorta. Pharmacol. Res., 54: 150-157.
Bhutani, S.P., S.S. Chibber and T.R. Seshadri, 1969. Flavonoids of the fruits and leaves of Tribulus terrestris: Constitution of tribuloside. Phytochemistry, 8: 299-303.
Blumenthal, M., A. Goldberg and J. Brinckmann, 2000. Herbal Medicine: Expanded Commission E Monographs. American Botanical Council, Austin, TX, pp: 519.
Brown, A.C., C. Shah, J. Liu, J.T.H. Pham, J.G. Zhang and M.R. Jadus, 2009. Ginger's (Zingiber officinale Roscoe) inhibition of rat colonic adenocarcinoma cells proliferation and angiogenesis in vitro. Phytother. Res., 23: 640-645.

Coruh, N., A.G.S. Celep, F. Ogokce and M. Iscan, 2007. Antioxidant capacities of Gundelia tournefortii L. extracts and inhibition of glutathione S-transferase activity. Food Chem., 100: 1249-1253.
DerMarderosian, A., 2008. The Review of Natural Products. 5th Edn., Williams and Wilkins, USA.
Duke, J.A., 2002. Handbook of Medicinal Herbs. 2nd Edn., CRCPress, BocaRaton, USA.,ISBN: 9780849312847, Pages: 870.
Erdogan-orhan, I and M. Kartal, 2011. Insights into research on phytochemistry and biological activities of Prunus armeniaca L. (apricot). Food Res. Int., 44: 1238-1243.
Farouk, L., A. Laroubi, R. Aboufatima, A. Benharref and A. Chait, 2008. Evaluation of the analgesic effect of alkaloid extract of Peganum harmala L: possible mechanisms involved. J. Ethnopharmacol., 115: 449-454.
Frielle, T., A.A. Crimaldi and C.J. Coffee, 1979. A continuous spectrophotometric assay for cyclic $3^{\prime}, 5$ '-nucleotide phosphodiesterase. Anal. Biochem., 97: 239-247.
Fu-hua, L., 1987. Cardiac glycosides in traditional Chinese medicine. J. Huazhong Univ. Sci. Technol., 7: 195-201.
Gambhir, S.S., S.P. Sen, A.K. Sanyal and P.K. Das, 1979. Antispasmodic activity of the tertiary base of Daucus carota, Linn. seeds. Indian J. Physiol. Pharmacol., 23: 225-228.
Gauthaman, K and A.P. Ganesan, 2008. The hormonal effects of Tribulus terrestris and its role in the management of male erectile dysfunction an evaluation using primates, rabbit and rat. Phytomedicine, 15: 44-54.
Genet, S., R.K. Kale and N.Z. Baquer, 2002. Alterations in antioxidant enzymes and oxidative damage in experimental diabetic rat tissues: effect of vanadate and fenugreek (Trigonella foenum graecum). Mol. Cell. Biochem., 236: 7-12.
Gilani, A.H., F. Shaheen, S.A. Saeed, S. Bibi, Irfanullah, M. Sadiq and S. Faizi, 2000. Hypotensive action of coumarin glycosides from Daucus carota. Phytomedicine, 7: 423-426.
Gruenwald, J., T. Brendle and C. Jaenicke, 2007. PDR for Herbal Medicine. 4th Edn., Thomson Healthcare, USA., ISBN: 9781563636783 , Pages: 1026.
Guh, J.H., F.N. Ko, T.T. Jong and C.M. Teng, 1995. Antiplatelet effect of gingerol isolated from Zingiber officinale. J. Pharm. Pharmacol., 47: 329-332.

Haghi, G., A. Hatami and R. Arshi, 2011. Distribution of caffeic acid derivatives in Gundelia tournefortii L . food chem., 124: 1029-1035.
Hemmateenejad, B., A. Abbaspour, H. Maghami, R. Miri and M.R. Panjehshahin, 2006. Partial least squaresbased multivariate spectral calibration method for simultaneous determination of $\beta$-carboline derivatives in Peganum harmala seed extracts. Anal. Chim. Acta, 575: 290-299.
Hu, C., Y.V. Yuan and D.D. Kitts, 2007. Antioxidant activities of the flaxseed lignin secoisolariciresinol diglucoside, its aglycone secoisolariciresinol and the mammalian lignansenterodiol and enterolactone in vitro. Food Chem. Toxicol., 45: 2219-2227.
Huang, G.C., T.Y. Chien, L.G. Chen and C.C. Wang, 2005. Antitumor effects of zerumbone from Zingiber zerumbet in p-388dl cells in vitro and in vivo. Planta Med., 71: 219-224.
Im, J.H., Y.R. Jin, J.J. Lee, J.Y. Yu and X.H. Han et al., 2009. Antiplatelet activity of $\beta$-carboline alkaloids from Perganum harmala: A possible mechanism through inhibiting PLC $\beta 2$ phosphorylation. Vascul. Pharmacol., 50: 147-152.
Irshaid, F. and K. Mansi, 2009. Effects of leaf extract of Urtica pilulifera L. on male reproductive system of streptozotocin-diabetic rats. Am. J. Pharmacol. Toxicol., 4: 22-28.
Ivanova, A., J. Serly, D. Dinchev, I. Ocsovszki, I. Kostova and J. Molnar, 2009. Screening of some saponins and phenolic components of Tribulus terrestris and Smilax excelsa as MDR modulators. In Vivo, 23: 545-550.
Jeon, Y.H., Y.S. Heo, C.M. Kim, Y.L. Hyun, T.G. Lee, S. Ro and J.M. Cho, 2005. Phosphodiesterase: Overview of protein structures, potential therapeutic applications and recent progress in drug development. Cell Mol. Life Sci., 62: 1198-1220.
Kaileh, M., W.V. Berghe, T. Boone, E. Essawi, T. and G. Haegeman, 2007. Screening of indigenous Palestinian medicinal plants for potential anti-inflammatory and cytotoxic activity. J. Ethnopharmacol., 113: 510-516.

Kavalali, G. and H. Tuncel, 1997. Anti-inflammatory activities of Urtica pilulifera. Int. J. Pharmacol., 35: 138-140.
Kavalali, G., H. Tuncel, S. Goksel and M.H. Hatemi, 2003. Hypoglycemic activity of Urtica pilulifera in streptozotocin-diabetic rats. J. Ethnopharmacol., 84: 241-245.

Khoshakhlagh, P., M. Bahrololoumi-Shapourabadi, A. Mohammadirad, L. Ashtaral-Nakhai, B. Minaie and M. Abdollahi, 2007. Beneficial effect of phosphodiesterase-5 inhibitor in experimental inflammatory bowel disease: Molecular evidence for involvement of oxidative stress. Toxicol. Mech. Methods, 17: 281-288.
Kim, Y.H., Y.W. Kim, Y.J. Oh, N.I. Back and S.A. Chung et al., 2006. Protective effect of the ethanol extract of the roots of Brassica rapa on cisplatin-induced nephrotoxicity in LLC-PK 1 cells and rats. Biol. Pharm. Bull., 29: 2436-2441.
Koocheki, A., S.A. Mortazavi, F. Shahidi, S.M.A. Razavi, R. Kadkhodaee and J.M. Milani, 2010. Optimization of mucilage extraction from Qodume shirazi seed (Alyssum homolocarpum) using response surface methodology. J. Food Process. Eng., 33: 861-882.
Kuhn, M.A. and D. Winston, 2000. Herbal Therapy and Supplements a Scientific and Traditional Approach. Lippincott, New York, pp: 2-15.
Lamchouri, F., A. Settaf, Y. Cherrah, M. Zemzami and B. Lyoussi et al., 1999. Antitumour principles from Peganum harmala seeds. Therapie, 54: 753-758.
Lee, J.S., K.A. Kim, S.H. Jeong, S.G. Lee, H.J. Park, N.J. Kim and S. Lim, 2009. Anti-inflammatory, anti-nociceptive, and anti-psychiatric effects by the rhizomes of Alpinia officinarum on complete Freund's adjuvant-induced arthritis in rats. J. Ethnopharmacol., 126: 258-264.

Li, J., X. Liu, F. Dong, J. Xu, Y. Zheng and W. Shan, 2010. Determination of the volatile composition in essential oil of Descurainia sophia (L.) webb ex prantl (Flixweed) by gas chromatography/mass spectrometry (GC/MS). Molecules., 15: 233-240.
Malviya, N., S. Jain, V.B. Gupta and S. Vyas, 2011. Recent studies on aphrodisiac herbs for the management of male sexual dysfunction-a review. Acta Pol. Pharm., 68: 3-8.
Mei-fang, M., L. Wen-hai and L. Jie, 2008. Experimental Research on Pharmacology and Acute Toxicology of Descurainia Sophia (L.) Webb ex Prantl Processed. Chin. Arch. Traditional Chin. Med.
Milani, E., S. Nikfar, R. Khorasani, M.J. Zamani and M. Abdollahi, 2005. Reduction of diabetes-induced oxidative stress by phosphodiestrase inhibitors in rats. Comp. Biochem. Physiol. Part C, 140: 251-255.
Misra, D.S., R. Maiti and D. Ghosh, 2009. Protection of swimming-induced oxidative stress in some vital organs by the treatment of composite extract of Withania somnifera, Ocimum sanctum and Zingiber officinalis in male rat. Afr. J. Tradit. Complement. Altern. Med., 6: 534-543.

Ozen, T., Z. Collu and H. Korkmaz, 2010. Antioxidant properties of Urtica pilulifera root, seed, flower and leaf extract. J. Med. Food., 13: 1224-1231.
Park, S., M.Y. Kim, D.H. Lee, S.H. Lee and E.J. Baik et al., 2009. Methanolic extract of onion (Allium cepa) attenuates ischemia/hypoxia-induced apoptosis in cardiomyocytes via antioxidant effect. Eur. J. Nutr., 48: 235-242.
Phillips, O.A., K.T. Mathew and M.A. Oriowo, 2006. Antihypertensive and vasodilator effects of methanolic and aqueous extracts of Tribulus terrestris in rats. J. Ethnopharmacol., 104: 351-355.
Pulpati, H., Y.S. Biradar and M. Rajani, 2008. High-performance thin-layer chromatography densitometric method for the quantification of harmine, harmaline, vasicine and vasicinone in Peganum harmala. J. AOAC Int., 91: 1179-1185.
Qureshi, S., A.H. Shah, M. Tariq and A.M. Ageel, 1989. Studies on herbal aphrodisiacs used in Arab system of medicine. Am. J. Chin. Med., 17: 57-63.
Rahimi, R., S. Ghiasi, H. Azimi, S. Fakhari and M. Abdollahi, 2010. A review of the herbal phosphodiesterase inhibitors: Future perspective of new drugs. Cytokine, 49: 123-129.
Rasool, M. and P. Varalakshmi, 2006. Immunomodulatory role of Withania somnifera root powder on experimental induced Inflammation: An in vivo and in vitro study. Vascular Pharmacol., 44: 406-410.
Shoji, N., A. Iwasa, T. Takemoto, Y. Ishida and Y. Ohizumi, 1982. Cardiotonic principles of ginger (Zingiber officinale Roscoe). J. Pharm. Sci., 71: 1174-1175.
Singh, B.N., B.R. Singh, R.L. Singh, D. Prakash and D.P. Sing et al., 2009. Polyphenolics from various extracts/fractions of red onion (Allium cepa) peel with potent antioxidant and antimutagenic activities. Food Chem. Toxicol., 47: 1161-1167.
Stoilova, I., A. Krastanov, A. Stoyanova, P. Denev and S. Gargova, 2007. Antioxidant activity of a ginger extract (Zingiber officinale). Food Chem., 102: 764-770.
Sun, K., X. Li, J.M. Liu, J.H. Wang, W. Li and Y. Sha, 2005. A novel sulphur glycoside from the seeds of Descurainia sophia (L.). J. Asian Nat. Prod. Rese., 7: 853-856.
Sur, P., M. Das, A. Gomes, J.R. Vedasiromoni, N.P. Sahu, S. Banerjee, R.M. Sharma and D.K. Ganguly, 2001. Trigonella foenum graecum (Fenugreek) seed extract as an antineoplastic agent. Phytother. Res., 15: 257-259.
Syamkumar, S., B. Lowarence and B. Sasikumar, 2003. Isolation and amplification of DNA from rhizomes of turmeric and ginger. Plant Mol. Biol. Rep., 21: 171-171.

Thomson, M., K.K. Al-Qattan, S.M. Al-Sawan, M.A. Alnaqeeb, I. Khan and M. Ali, 2002. The use of ginger (Zingiber officinale Rosc.) as a potential anti-inflammatory and antithrombotic agent. Prostaglandins Leukot Essent Fatty Acids, 67: 475-478.
Vasudevan, M., K.K. Gunnam and M. Parle, 2006. Antinociceptive and anti-inflammatory properties of daucus carota seeds extract. J. Health Sci., 52: 598-606.
Vyas, S., R.P. Agrawal, P. Solanki and P. Trivedi, 2008. Analgesic and anti-inflammatory activities of Trigonella foenum-graecum (seed) extract. Acta Pol. Pharm., 65: 473-476.
Wagner, H., W. Dorsch, T. Bayer, W. Breu and F. Willer, 1990. Antiasthmatic effects of onions: Inhibition of 5-lipoxygenase and cyclooxygenase in vitro by thiosulfinates and cepaenes. Prostag. Leukotr. Essent. Fatty Acids, 39: 59-62.
Waldschlager, J., C. Bergemann, W. Ruth, U. Effmert and U. Jeschke et al., 2005. Flax-seed extracts with phytoestrogenic effects on a hormone receptorpositive tumour cell line. Anticancer Res., 25: 1817-1822.
Yasukawa, K., Y. Sun, S. Kitanaka, N. Tomizawa, M. Miura and S. Motohashi, 2008. Inhibitory effect of the rhizomes of Alpinia officinarum on TPA-induced inflammation and tumor promotion in two-stage carcinogenesis in mouse skin. J. Nat. Med., 62: 374-378.
Yob, N.J., S.M. Jofrry, M.M. Affandi, L.K. Teh, M.Z. Salleh and Z.A. Zakaria, 2011. Zingiber zerumbet (L.) smith: A review of its ethnomedicinal, chemical and pharmacological uses. Evid. Based. Compl. Alt. Med., 10.1155/2011/543216.
Yurt, B. and I. Celik, 2011. Hepatoprotective effect and antioxidant role of sun, sulphited-dried apricot (Prunus armeniaca L.) and its kernel against ethanolinduced oxidative stress in rats. Food Chem. Toxicol., 49: 508-513.
Zakaria, Z.A., A.S. Mohamad, M.S. Ahmad, A.F. Mokhtar and D.A. Israf, N.H. Lajis and M.R. Sulaiman, 2010a. Preliminary analysis of the anti-inflammatory activity of essential oils of Zingiber zerumbet. Biol. Res. Nurs., 13: 425-432.
Zakaria, Z.A., S. Mohamad, C.T. Chear, Y.Y. Wong, D.A. Israf and M.R. Sulaiman, 2010b. Antiinflammatory and antinociceptive activities of Zingiber zerumbet methanol extract in experimental model systems. Med. Princ. Pract., 19: 287-294.

