Chronic Administration of Tribulus terrestris Linn. Extract Improves Cardiac Function and Attenuates Myocardial Infarction in Rats

1Shreesh K. Ojha, 1Mukesh Nandave, 1Sachin Arora, 2Rajeev Narang, 3Amit K. Dinda and 1Dharamvir Singh Arya
1Department of Pharmacology, 2Department of Cardiology, 3Department of Pathology, All India Institute of Medical Sciences, New Delhi-110 029, India

Abstract: The present study was undertaken to evaluate the cardioprotective potential of hydro-alcoholic extract of Tribulus terrestris Linn. (Family: Zygophyllaceae), a traditional medicine used in Indian and Chinese systems of medicine. Wistar male albino rats weighing 150-200 g were randomly divided into three main experimental groups; sham (saline treated only), isoproterenol (ISP) control (saline and ISP) and Tribulus terrestris treatment groups (T. terrestris and ISP). Saline or T. terrestris extract 250 mg kg

INTRODUCTION

Among cardiovascular diseases (CVDs), myocardial infarction (MI) accounts for majority of death and disability and has high incidence rate in both developed and developing countries as seen over the past quarter century (Agrawal et al., 2006). Reduction of death incidences and prevention of myocardial infarction are of utmost importance. In spite of development in modern drugs, there is a growing interest in the use of herb based drugs from alternative medicine for long-term prevention of myocardial ischemia in high risk patients along with a realization that the use of herbal products can influence the course of heart diseases and may provides an
integrated approach of nutritional substances, which help in restoring and maintaining, the balanced body systems (Hertog et al., 1993; Dhar et al., 1968).

The plant *Tribulus terrestris* Linn, a deciduous tree of the Zygophyllaceae family, is an important herb from Indian and Chinese traditional medicine literature for the treatment of various diseases especially ischemic heart diseases (Warrier, 1994). Modern pharmacological studies have reported its hypoglycemic (Amin et al., 2006), nephroprotective (Kavitha and Jagesan, 2006), diuretic (Anand et al., 1994), aphrodisiac (Gauthaman et al., 2002) and immunomodulator (Bhattacharya et al., 2000) activities. *T. terrestris* extract standardized to contain saponins has been reported to dilate coronary arteries, increase coronary blood flow and scavenge free radicals in patients with ischemic heart diseases (Sharifi et al., 2003; Guo et al., 2005) and indicated for angina pectoris, myocardial infarction, congestive heart failure and stroke (Phillips et al., 2006; Guo et al., 2005). In addition to the antianginal activity, their antihypertensive effect has been demonstrated owing to its angiotensin converting enzyme (ACE) inhibitor property (Sharifi et al., 2003). Experimental studies also showed that ACE inhibitors administered chronically before acute MI might limit myocardial infarct size, improve cardiac function and prevent cardiac hypertrophy (Stauss et al., 1994). The extract of the plant contains a large amount of flavonoids, saponins, alkaloids and tannins, which possess significant antioxidant (Wang et al., 1997) and cardiac (Achenbach et al., 1996) activity. In addition to antioxidant constituents, it has been reported to consist of adaptogenic components which provide resistance against stress (Gauthaman et al., 2002). Several herbal antioxidants and adaptogens rich herbs have been shown to possess cardioprotective activity. However, so far very few scientific studies have been undertaken to evaluate the cardioprotective potential of *T. terrestris*. In addition, the exact mechanism of its cardioprotective effects with respect to the present knowledge of the pathophysiology of myocardial infarction has not been investigated. In view of the above, the present study was designed to evaluate the cardioprotective potential of lyophilized extract *T. terrestris* in isoproterenol (ISP)-induced myocardial infarction in rats.

We aimed to investigate the effects of chronic administration of the hydro-alcoholic extract of *T. terrestris* on: (i) hemodynamics e.g., heart rate (HR), systolic, diastolic and mean arterial blood pressure (SAP, DAP and MAP, respectively) (ii) left ventricular function e.g., left ventricular peak positive pressure change (rate of pressure development), left ventricular rate of peak negative pressure change (rate of pressure decline) [(+] and [–]) LV dp/dt] indicating the inotropic and lusitropic state of heart respectively and left ventricular end-diastolic pressure (LVEDP), a surrogate marker of preload (iii) endogenous antioxidant status e.g., reduced glutathione (GSH) and antioxidant enzymes, superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GSHP) (iv) lipid peroxidation marker e.g., malonaldehyde (MDA) (v) myocyte injury marker enzymes e.g., creatine phosphokinase-MB isoenzyme (CK-MB) and lactate dehydrogenase (LDH) (vi) histoarchitectural changes and (vii) ultramicroscopic changes in rats subjected to acute ischemic injury of heart following ISP-administration.

**MATERIALS AND METHODS**

**Plant extract:** Standardized hydro-alcoholic lyophilized extract of whole plant of *Tribulus terrestris* was procured from Sanat Products Ltd., New Delhi, India. The identity of *Tribulus terrestris* was authenticated on the basis of routine pharmacognostical studies including organoleptic tests and macroscopic and microscopic observations. The voucher specimen of lyophilized extract of *T. terrestris* (No. TTS 104) has been deposited in Cardiovascular Laboratory, Department of Pharmacology, All India Institute of Medical Sciences, New Delhi, India for further reference. Phytochemical analysis of the extract was performed for saponin content determination. The total saponin content was 43.77% w/w and the extractive values of 1 g sample of *Tribulus terrestris* in water and 50% v/v methanol were 81.36 and 90.31%, respectively. The dose of *T. terrestris* (250 mg kg⁻¹) in the present study was selected on the basis of a pilot study assessing the antioxidant activity of *T. terrestris* in rat hearts (Ojha et al., 2006). The doses screened were 75, 150, 250 and 350 mg kg⁻¹ day⁻¹ and 250 mg kg⁻¹ of *T. terrestris* exhibited maximum antioxidant and anti-oxidative activity in hearts. Therefore, this dose was selected for further evaluation in the ISP-induced myocardial infarction incorporating biochemical, hemodynamic, histological and ultra-structural assessments.

**Chemicals:** All chemicals used in the study were of analytical grade and obtained from standard drug houses. Isoproterenol hydrochloride was obtained from Sigma Chemicals Co. (St. Louis, MO), USA.

**Animals:** Laboratory breed Wistar male albino rats weighing 150-200g were obtained from the Central Animal House Facility of All India Institute of Medical Sciences, New Delhi, India. The study protocol was reviewed and approved by the Institutional Animal Ethics Committee.
(227/IABC/03) and conducted in accordance with the Indian National Science Academy Guidelines for the use and care of experimental animals in research. The animals were acclimatized with the atmosphere of departmental animal house and housed under standard laboratory conditions of temperature at 25±2°C, relative humidity of 50±10% and light: dark cycle of 12 h photoperiod. All experiments were performed between 9.00 and 16.00 h. They were group housed in polypropylene cages (38×23×10 cm) with not more than four animals per cage and had free access to food pellets and tap water.

**Study design:** A total of thirty-nine rats were used in the study and equally divided into three main experimental groups as follows:

**Control group:** Rats of this group received oral saline once daily for 30 days and on days 29 and 30 administered 0.5 mL saline s.c. at an interval of 24 h.

**ISP group:** Rats of this group received oral saline once daily for 30 days and on days 29 and 30 administered ISP (85 mg kg⁻¹, s.c.) at an interval of 24 h.

**T. terrestris plus ISP group:** Rats of this group received *T. terrestris* extract 250 mg kg⁻¹ day⁻¹ for 30 days and on days 29 and 30 administered ISP (85 mg kg⁻¹, s.c.) at an interval of 24 h.

On day 31, 48 h after injection of ISP administration of saline, hemodynamic parameters were recorded.

**Induction of myocardial infarction and hemodynamic assessment:** Animals of all the experimental groups were anesthetized intraperitoneally with pentobarbital sodium (60 mg kg⁻¹). Atropine (4 mg kg⁻¹) was administered along with the anesthetic agent to maintain the heart rate especially during the surgery and to reduce tracheobronchial secretions. The body temperature was monitored and maintained at 37°C during the surgical period. The neck was opened with a ventral midline incision to perform tracheostomy and rats were ventilated with room air from a positive pressure ventilator (Inco, India) using compressed air at a rate of 90 strokes/min and a tidal volume of 10 mL kg⁻¹. Ventilator setting and PO₂ were adjusted as needed to maintain the arterial blood gas parameters within the physiological range. The left jugular vein was cannulated with polyethylene tube for continuous infusion of 0.9% saline solution. The right carotid artery was cannulated and the cannula was filled with heparinized saline. The cannula was connected with Cardiosys CO-101 (Experimentia, Hungary) using a pressure transducer and the signal was amplified by means of an amplifier for the measurement of SAP, DAP, MAP and HR. The left thoracotomy was performed at the fifth intercostal space on left side and the heart was exposed. A sterile metal cannula (1.5 mm bore) was introduced into the cavity of the left ventricle from the posterior apical region of the heart for measuring left ventricular dynamics such as (+) LV dP/dt, (-) LV dP/dt and LVEDP. The cannula was connected to a pressure transducer (Gould Statham P23ID, USA) through a pressure-recording catheter on the Polygraph (Grass 7D, USA). After the stabilization time of 10 min the tracings has been recorded polygraph paper following baseline measurements at different sensitivity and speed.

**Biochemical studies:** Hearts stored in liquid nitrogen were brought to room temperature and weighed. A 10% homogenate of whole heart was prepared in 50 mM phosphate buffer (pH 7.4) and an aliquot of 0.5 mL was used for the assay of MDA (Ohkawa et al., 1979) and reduced GSH (Moron et al., 1979). Remaining the homogenate was centrifuged at 7000 rpm for 15 min and the supernatant was used for estimation of the following biochemical parameters. SOD (Misra and Fridovich, 1976), CAT (Aebi, 1974), GSHPx (Paglia and Valentine, 1967) and protein (Lowry et al., 1951). Myocardial injury markers, CK-MB isoenzyme (Lamprecht et al., 1974) and LDH (Cabaud and Wroblewski, 1958) were estimated spectrophotometrically.

**Histopathological studies (Light microscopy):** For histopathological studies, the hearts fixed in 10% buffered neutral formalin solution were cut in to four segments from apex to bottom to visualize myocardial lesions at different levels. The segments of the tissue fixed tissues were embedded in paraffin wax and serial semi thin sections of 4 μm thickness were cut. After hematoxylin and eosin staining, for histoarchitectural evaluation, these sections were examined under light microscope (Nikon, Tokyo, Japan) and photomicrographs were taken. Representative area images were captured in an image analysis system. The Image Analyzer consisted of BX-50 Research Microscope (Olympus, Japan), Coollight 10 bit Digital Camera (Media Cybernetic, USA) with an image analysis software Image Plus Pro (Media Cybernetic, USA). The slides were evaluated for myonecrosis, inflammatory cell infiltration and edema.
Ultrastructural studies (Transmission electron microscopy): For ultrastructural study, at the end of experiment small pieces of myocardial tissue (approximately 1-2 mm in thickness) were immediately fixed in ice-cold Karnovsky's fixative. The tissues were then washed in phosphate buffer (0.1 M, pH 7.4) and post fixed for 2 h in 1% osmium tetroxide in the same buffer at 4°C. The specimens were then washed in phosphate buffer, dehydrated with graded acetone and then embedded in araldite CY 212 to make tissue blocks. The semi thin as well as ultra thin sections (80-100 nm) were cut by an ultramicrotome (Ultracut E, Retschert, Austria). The sections were stained with uranyl acetate and lead acetate and examined under transmission electron microscope (Morgagni 268D, Fei Co., The Netherlands) operated at 60 KV.

Statistical analysis: The statistical analysis was carried out by SPPS statistical package version 10. In tables, figures and text the data are expressed as mean±SD. Differences between groups were examined for statistical significance using one-way analysis of variance (ANOVA) followed by Bonferroni Multiple Range post-hoc analysis. p<0.05 was considered as statistical significance level.

RESULTS

There was no significant difference in body weight of the treated rats, when compared with sham control, either at the beginning or at end of the study period.

ISP induced myocardial necrosis produced a significant reduction in activities of SOD, CAT and GSHPx (Table 1) as compared to sham control. Treatment with T. terrestris significantly prevented fall in enzyme activities and increased the activities of SOD, CAT and GSHPx as compared to ISP control.

A significant decrease in myocardial GSH level and increase in lipid peroxidation product MDA, was observed in the ISP control group in comparison to sham control (Table 2). However, T. terrestris extract significantly prevented lipid peroxidation and decline of GSH level and as compared to ISP control.

We observed a significant decrease in the myocardial enzyme activities, CK-MB isoenzyme and LDH (Table 3) in the rat myocardium of ISP control group as compared to sham control. Treatment with T. terrestris extract significantly prevented loss of CK-MB isoenzyme and LDH enzymes and restored both in the myocardium.

A significant fall in SAP, DAP and MAP was observed in the ISP control group as compared to sham control (Fig. 1). T. terrestris treatment significantly increased MAP with a slight but insignificant improvement in SAP and DAP in comparison to sham control. Subsequent to ISP administration, a significant fall in HR was observed as compared to sham control (Fig. 2). T. terrestris treatment significantly improved HR in comparison with ISP control.

ISP administration resulted in left ventricular dysfunction, as indicated by a significant fall in (+) and (-) LVDp/dt (Fig. 3) and increase in LVEDP as compared to sham control (Fig. 4). Myocardial relaxation (lusitropic state) denoted by (-) LVDp/dt was significantly restored by T. terrestris treatment as compared with ISP control (Fig. 3). Additionally, T. terrestris treatment markedly reduced LVEDP as compared to ISP control (Fig. 4).

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>SOD (U/mg protein)</th>
<th>CAT (U/mg protein)</th>
<th>GSHPx (μmol mg^-1 protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham</td>
<td>7.94±2.90</td>
<td>21.90±4.10</td>
<td>0.53±0.12</td>
</tr>
<tr>
<td>ISP control</td>
<td>3.40±1.40*</td>
<td>11.90±2.00*</td>
<td>0.19±0.08*</td>
</tr>
<tr>
<td>T. terrestris 250 mg kg^-1</td>
<td>6.84±1.95*</td>
<td>24.87±7.11*</td>
<td>0.38±0.09*</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Experimental group</th>
<th>MDA (μmol mg^-1 protein)</th>
<th>GSH (μmol mg^-1 protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sham</td>
<td>66.50±14.30</td>
<td>1.86±0.69</td>
</tr>
<tr>
<td>ISP control</td>
<td>99.00±9.50*</td>
<td>1.22±0.50*</td>
</tr>
<tr>
<td>T. terrestris 250 mg kg^-1</td>
<td>12.48±10.70*</td>
<td>2.97±0.61*</td>
</tr>
</tbody>
</table>

* p<0.05 vs sham control; * p<0.05 vs ISP control. Each value represents mean±SD of six readings (n = 6)
Fig. 2: Changes in heart rate in different experimental groups, *: p<0.05 vs sham control; #: p<0.05 vs ISP control. Each value represents mean±SD of six readings (n = 6)

Fig. 3: Changes in (+) and (-) LV dP/dt in different experimental groups, *: p<0.05 vs sham control; #: p<0.05 vs ISP control. Each value represents mean±SD of six readings (n = 6)

Fig. 4: Changes in left ventricular end-diastolic pressure in experimental groups, *: p<0.05 vs sham control; #: p<0.05 vs ISP control. Each value represents mean±SD of six readings (n = 6)

On histopathological examination depicted in Fig. 5A-C, we observed normal architecture of the heart in sham control (Fig. 5A). However, significant myocyte membrane damage with extensive myonecrosis, fibroblastic proliferation and infiltration of inflammatory cells was observed in ISP control hearts (Fig. 5B) in comparison

Fig. 5: Photomicrograph of heart tissue (H and E X 100) (A) Sham group showing normal architecture. Endocardium and pericardium are seen within normal limits without infiltration of inflammatory cells (B) ISP group showing focal myonecrosis with myophagocytosis and lymphocytic infiltration. Vascular changes and prominent edema along with chronic inflammatory cells were seen in subendocardium (C) Tribulus terrestris treated group administered showing lesser infiltration of inflammatory cells and myonecrosis
with sham control. *T. terrestris* treatment extract prevented myonecrosis (Fig. 5C) as demonstrated by significant reduction in the infiltration of inflammatory cells, vacuolar changes as well as edema as compared to the ISP control group.

Electron microscopic examination of the myocardium revealed normal appearance of cell organelles in sham control rats showing sarcomere and abundant mitochondria (Fig. 6A). In ISP control group, the myocardial damage was marked by significant disruption of myofibrils (Fig. 6B). In addition, the myocardial tissue of ISP control group revealed several intracytoplasmic vacuoles and lipid droplets. Other ultrastructural changes include the appearance of small and irregular mitochondria with loss of cristae (Fig. 6B). However, treatment of animals with *T. terrestris* showed structural protection of the myocardium with respect to ISP control group (Fig. 6C).

**DISCUSSION**

*Trihulus terrestris* has significant cardioprotective activity as shown by its mitigating effects on ISP-induced hemodynamic, biochemical, histopathological and ultrastructural perturbations. In the present study, ISP, a β-adrenergic agonist was used to produce myocardial necrosis and interstitial fibrosis at its supra-maximal dosages in a multi-step manner. In addition, isoproterenol causes substrate loss, depletion of energy reserves, hypoxia due to myocardial hyperactivity, increased inotropic activity, coronary hypotension, a high-oxygen demand and production of free radicals resulting from oxidative metabolism of catecholamine (Tappia *et al.*, 2001). Free radical mediated peroxidation of membrane phospholipids and consequent changes in membrane permeability are the primary reasons for cardiotoxicity induced by ISP (Rathore *et al.*, 1998; Blasig *et al.*, 1984). ISP-induced oxidative stress also depresses the sarcoplasmic Ca²⁺ transport and results in the development of intracellular Ca²⁺ overload and ventricular dysfunction (Tappia *et al.*, 2001). The oxidative stress may be exerted through quinine metabolites of ISP which react with oxygen to produce superoxide anions and others reactive oxygen species and interfere with SOD, glutathione reductase and ATP pumps (Rathore *et al.*, 1998; Remiao *et al.*, 2000). Experimental and clinical studies have demonstrated that therapeutic interventions having antioxidant or free radical scavenging activity may exert cardioprotective effects against oxidative stress associated with various cardiovascular diseases.
including ischemic heart disease (Bandopadhyay et al., 2004). Hence, medicinal herbs or natural products possessing antioxidant constituents has recently gained a great deal of scientific interest (Gupta et al., 2004; Mohanty et al., 2004; Karthikeyan et al., 2003) as it is expected to offer better protection against oxidative stress associated diseases through cellular adaptive mechanism and may provide a lead for promotion of therapeutic agent especially of natural origin.

In the present study, chronic administration of *T. terrestris* extract maintained the antioxidant activity of the endogenous antioxidants as evidenced by increase in the levels of reduced GSH and activities of endogenous antioxidant enzymes such as SOD, CAT and GSH-Px along with decrease in the level of lipid peroxidation product, MDA. The depletion in GSH levels along with increased plasma lipid peroxides (measured as TBARS or thiobarbituric acid-reactive substances) has been observed during myocardial ischemia and associated with the consumption of endogenous antioxidant and enzymes such as SOD, CAT and GSH-Px. These antioxidant enzymes are known to remove the ISP-induced generation of toxic oxygen species by their peroxidative and free radical scavenging activity. In addition, an increase in MDA levels was observed in the heart tissue following ISP administration. The myocardial necrosis observed in the animals receiving ISP can also be attributed to peroxidative damage as it has been previously reported that ISP generates lipid peroxide (Blasig et al., 1984).

The fall in activity of GSH-Px in the ISP group might be co-related with decreased availability of its substrate, GSH. Moreover, due to impairment in both enzymatic and non-enzymatic antioxidant defense mechanisms, it is possible that the free radicals may not be effectively neutralized, thereby rendering the myocardium susceptible to lipid peroxidation. The increased GSH levels observed in the *T. terrestris* groups in the present study may be due to its enhanced synthesis, as antioxidant compounds have been shown to increase glutathione reductase activity, which maintains GSH in its reduced state (Mohanty et al., 2004). The observations of our study are in conformity with earlier reports that have demonstrated the modulation of synthesis of cellular antioxidants by treatment with natural products (Gupta et al., 2004; Mohanty et al., 2004; Karthikeyan et al., 2003).

In addition to the antioxidant parameters, the estimation of CK-MB isoenzyme and LDH levels, both of which are useful parameters for assessing myocardial injury due to disintegration of contractile elements and sarcoplasmic reticulum. ISP challenge in the present study produced significant depletion of myocardial CK-MB isoenzyme and LDH. *T. terrestris* treatment prevented the leakage of these enzymes and restored their activities as compared to the ISP control. The observation that *T. terrestris* treatment significantly restored CK-MB isoenzyme and LDH levels is suggestive of its cardioprotective effect.

As described earlier, hemodynamic parameters were also incorporated into the experimental design for a better understanding of the co-relation between biochemical and functional changes in the myocardium subjected to ISP-induced damage. Previous studies have shown that ISP mediated oxidative stress could progresses to myocardial necrosis which leads to cardiac dysfunction characterized by increased end-diastolic volume and pressure and left ventricular wall thickness (Grimm et al., 1998; Teerlink et al., 1994) and these changes are significantly prevented by antioxidants. In the present study, the favorable modulation of hemodynamic and left ventricular function by *T. terrestris* treatment also supports the previous observations it may modify the course of action and help in retaining the function of heart to normal level.

In the ISP control group, myocardial dysfunction was clearly evident by a significant fall in SAP, DAP and MAP, HR, LV (±) and (−) dP/dt along with a rise in LVEDP, which might be ascribed to its ISP-induced myocardial infarction. The left ventricular rate of negative pressure change was more markedly depressed indicating a more diastolic dysfunction per se which may result in the persistence of elevated LVEDP. It is speculated that deteriorating myocardial contractile status following ISP-induced necrosis might be responsible for the significant fall in MAP. Normally a fall in MAP reflexly increases HR and myocardial contractility. However, none of these effects were observed in the present study, suggesting impairment in the conducting system of the heart following ISP-induced myocardial necrosis. *T. terrestris* treatment in our study significantly increases MAP and HR as compared to the isoproterenol control group. Moreover, *T. terrestris* appeared to improve left ventricular function as evidenced by significant preservation of both inotropic and lusitropic functions of the heart along with correction of elevated LVEDP. Preservation of cardiac reflexes resulting in increased heart rate and ventricular function to maintain cardiac output may be on account of myocardial salvage, produced by *T. terrestris*.

The histopathological examination of myocardium in ISP control animals showed presence of focal
myonecrosis with myophagocytosis and lymphocytic infiltration in sub-endocardial region indicative of infarct like lesions similar to previous studies (Grimm et al., 1998; Teerlink et al., 1994). Treatment with T. terrestris extract preserved the normal histoarchitecture and ultrastructure of the myocardium as evidenced by reduced myonecrosis and lesser infiltration of inflammatory cells. Taken together, the biochemical and histopathological results of the present study demonstrate the cardioprotective effects of T. terrestris. Additionally, ultrastructural perturbations in ISP administered rat hearts further confirmed the injured state of myocardium. Restoration of myocardial CK-MB activity, along with preservation of myofibrils and mitochondrial morphology is indicative of cytoprotective activity of T. terrestris.

Recently, adaptogens are demonstrated to confer resistance against a variety of stresses and aid in myocardial adaptation. Various experimental studies have demonstrated the adaptogenecity i.e., increased nonspecific resistance to stress of medicinal herbs is considered as a function of antioxidant components which may produce myocardial adaptation (Bhattacharya and Muruganandam, 2003; Lei and Chiu, 1986). The major active constituents of T. terrestris are flavonoids and steroidal saponins, lignanamides, protodioscin and tribulosins (Wang et al., 1997; Achenbach et al., 1996). It has been reported that protodioscin and tribulosins are the adaptogenic components present in T. terrestris (Gauthaman et al., 2002). The myocardial adaptation against ischemic stress may be mediated through augmentation of cellular tissue antioxidant enzymes such as SOD, CAT and GSH-Px and stress proteins. Although, the exact mechanism of such myocardial adaptation is not known. Further, flavonoids have been shown to exhibit wide spectrum of biological activities including free radical scavenging, antioxidant and anti-ischemic activities (Hertog et al., 1993). Therefore, it could be possible that T. terrestris may have exerted its cardioprotective effects partly due to its antioxidant as well as adaptogenic constituents.

In summary, the present study demonstrated that multiple mechanisms may be responsible for the cardioprotective effect of Tribulus terrestris. Improved hemodynamics and left ventricular function along with improved endogenous myocardial antioxidant status and myocardial adaptability assures its cardioprotective potential in ISP-induced myocardial necrosis. The histopathological and ultrastructural examination, further confirmed its cardioprotective effects through preservation of cell organelle and myocyte membrane integrity. In conclusions, the observations of present study provide a scientific basis for cardioprotective effect of Tribulus terrestris during myocardial ischemia and demonstrate its therapeutic potential in the treatment of ischemic heart disease.

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