Hepatoprotective Activity of Coccinia grandis Leaves Against Carbon Tetrachloride Induced Hepatic Injury in Rats

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Abstract: Coccinia grandis Linn. (Cucurbitaceae) is a perennial branched handsome tendril climber, distributed throughout India. It has been used in folk medicine for the treatment of jaundice. The aim of this work was to study the hepatoprotective effect of crude ethanolic and aqueous extracts from the leaves of C. grandis against liver damage induced by CCl4 in rats. The ethanolic extract at an oral dose of 200 mg kg⁻¹ exhibited a significant (p<0.05) protective effect as shown by lowering serum levels of glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, alkaline phosphatase, total bilirubin and total cholesterol and increasing levels of total protein and albumin levels as compared to silymarin, the positive control. These biochemical observations were supported by histopathological examination of liver sections. The activity may be due to the presence of flavonoid compounds. The extracts showed no signs of acute toxicity up to a dose level of 2000 mg kg⁻¹. Thus it could be concluded that ethanolic extract of C. grandis leaves possesses significant hepatoprotective activity.

Keywords: Indian traditional medicine, extracts, protective, Cucurbitaceae, silymarin

INTRODUCTION

Liver plays a major role in detoxification and excretion of many endogenous and exogenous compounds. Any impairment to its functions may lead to many implications on one's health. Management of liver diseases is still a challenge to the modern scientific community (Reddy et al., 1992). Modern medicine has little to offer alleviation of hepatic ailments. Phytoconstituents remain to be a major contributor in the treatment of liver disorders. The growing concern for the identification of novel hepatoprotective agents from natural sources is evident from literature available on the same. The hepatoprotective activity of Amaranthus spinosus (Zeeshan et al., 2008), Hibiscus ecculentus Linn. (Sunilson et al., 2008) and Vitis negundo leaf extract (Tandon Vishal et al., 2008) are reported in the literature. The study of hepatoprotective role of pomegranate flowers (Celik et al., 2009), Baccaiside-A, a major constituent of Bacopa monniera (Sumathi and Nongbri, 2008) and aqueous extract of Solanum fastigiatum (Sabir and Rocha, 2008) bears further testimony to the quest of natural hepatoprotective agents.

Recent scientific studies revealed that the plants Trichosanthes cucumerina (Kumara et al., 2009) and Cucurbita pepo L. (Makri et al., 2008) both belonging to the Cucurbitaceae family possessed significant hepatoprotective activity. Coccinia grandis Linn, also, belonging to the family Cucurbitaceae and commonly known as kovai in tamil, is a perennial much branched handsome tendril climber, cultivated throughout India and also found wild. The medicinal use of the plant can be traced to ancient period where the juice of the roots and leaves were used in the treatment of diabetes mellitus, bronchitis, skin diseases, tongue-sores and ear ache (Geeta et al., 2007). The fruit is commonly eaten in Indian cuisine. The plant is ethnomedically used as anti inflammatory, wound healing, purgative, antiemic, antiulcer, antipyretic anticonvulsant and astringent (Warrier et al., 1995) and is traditionally used to treat jaundice by the people of Tirunelveli district, Tamil Nadu state. Literature survey revealed that few works have been carried out on this plant. The fruits have been reported to possess hepatoprotective activity (Vadivu et al., 2008). However, there has been no report on the hepatoprotective activity of the leaves of this plant. The
present study was thus undertaken to scientifically prove
the activity of the leaves against liver disorders.

MATERIALS AND METHODS

Plant material: Leaves of *Coccinia grandis* were
collected from Kadayanallur, Tirunelveli District of
Tamilnadu, India, in July 2005. The plant sample was
authenticated by Dr. P. Jayaraman, M.Sc., Ph.D., Director,
Plant Anatomy Research Center, Chennai, Tamil Nadu,
India. A voucher specimen of the collected plant sample
was also deposited in the Herbarium of Sankaralingam
Bhavanesswari College of Pharmacy, Sivakasi, Tamil Nadu
(SBCPHA/C12/M 0014).

Preparation of extracts: The shade dried leaves of
*C. grandis* (500 g) were subjected to size reduction to
coarse powder. The powder was defatted with petroleum
ether (60-80°C) and then extracted with 95% ethyl alcohol
using soxhlet apparatus (Bose *et al.*, 2007) till exhaustion
for about 32 h. The total aqueous extract was also
prepared by the percolation method (Bugno *et al.*, 2007)
using water. Both the etherolic and aqueous extracts were
concentrated under vacuum to get the residues.
The percentage yields of etherolic extract and aqueous extracts were found to be 16.3% (w/w) and
22.13% (w/w), respectively. The etherolic extract was found to contain glycosides, triterpenoids, alkaloids and
flavonoids (Wagner and Bladt, 1996). Silymarin was used as
a positive control at an oral dose of 20 mg kg⁻¹
(Savita *et al.*, 1994). All the test suspensions are prepared
in vehicle, i.e., Tween-80.

Animals: Wistar albino rats of either sex, weighing
200-250 g, were obtained from the animal house of the
S.B. College of Pharmacy, Tamilnadu, India. The animals
were housed in groups of 6, in standard cages, at room
temperature (25±3°C), with 12h dark/12h light cycles,
food and water *ad libitum*. Twelve hours prior to the experiments they were transferred to the laboratory
and given only water *ad libitum*. The experiments were
approved by the Institutional Animal Ethical Committee
of the S.B. College of Pharmacy, Tamilnadu, India
(IAEC NO.SBCP/F.9 (f)/34b).

Toxicity studies: Acute toxicity study was performed for
etherolic and aqueous extracts in albino rats as per OECD
guidelines. The animals were fasted overnight and
provided only water, after which the extracts were
administered orally at the dose of 300 mg kg⁻¹ and
observed for 14 days. If mortality was observed in 2
out of 3 animals, the dose administered was assigned as
the toxic dose. If the mortality was observed in 1 animal,
the same dose was repeated again to confirm the toxic
dose. If mortality was not observed, the procedure was
repeated for a higher dose, i.e., 2000 mg kg⁻¹. One-
tenth of the maximum dose of the extract tested for acute
toxicity was selected for evaluation of hepatoprotective
activity, i.e., 200 mg kg⁻¹ (Handa and Sharma, 1990).

Carbon tetrachloride-induced hepatotoxicity in rats: Rats
were divided into 5 groups of 6 each, the control, CCl₄-
exposed, silymarin and two test groups. The control
group received oral vehicle treatment at 0, 24 and 48 h.
The animals in CCl₄-treated group received vehicle at 0 h
and at 24 h, followed by CCl₄ diluted with liquid paraffin
(1:1, i.p.) at a dose of 1.25 mL kg⁻¹, while at 48 h, these
animals received only vehicle. The test groups have
received the first dose of extracts at 0 h, the second dose
of extracts at 24 h, which was followed by a dose of CCl₄,
and the third dose of extracts at 48 h (Kumar and Mishra,
2005). The positive control group had received the first
dose of silymarin (20 mg kg⁻¹) at 0 h, at 24 h, the second
dose of silymarin followed by a dose of CCl₄, and at 48 h,
the third dose of silymarin. After 72 h, blood was collected
from all the groups sino-orbital puncture and allowed to
clot for the separation of serum. The serum was used for
estimation of biochemical parameters.

Glutamic oxaloacetic transaminase (SGOT) and
Glutamic pyruvic transaminase (SGPT) were estimated by
the method followed by Rajkapoor *et al.* (2006), alkaline
phosphatase (ALKP) by the method of McComb and
Bowers (1972), total bilirubin (TBL) by the method
followed by Rana *et al.* (2008), total cholesterol (CHL)
by the method of Richmond (1973), total protein (TPIN)
by the method followed by Peters (1968) and white albumin
(AlB) was estimated by Webster (1974) method. All the
determinations were carried out using standard kits by an
autoanlyser (300 TX, E. Merck-Micro Labs, Mumbai).

Histopathological studies: One animal from each of the
treated groups showing maximum activity as indicated by
improved biochemical parameters was used for this
purpose. The animals were sacrificed and the abdomen
was cut open to remove the liver. The liver was fixed in
Bouin's solution (mixture of 75 mL of saturated picric acid,
25 mL of 40% formaldehyde and 5mL of glacial acetic acid)
for 12 h and then embedded in paraffin using
conventional methods (Galigher and Kozloff, 1971). They
were then cut into 5 μm thick sections and stained using
haematxylin-cosin dye and finally mounted in
diphenylxylene. The sections were observed under a
microscope for histopathological changes in liver
architecture and their photomicrographs were taken.
Statistical analysis: The Mean±SEM were calculated for each parameter. The statistical analysis of the results was carried out with a SPSS 9.0 program and based on an Analysis of Variance (ANOVA) followed by the Dunne’s test.

RESULTS AND DISCUSSION

The ethanolic and aqueous extracts did not produce any mortality up to 2000 mg kg⁻¹ and were considered as safe (OECD, 1998). Carbon tetrachloride (CCL₄) intoxication in normal rats elevated the levels of SGOT, SGPT, ALKF, TBL, and CHL, whereas the levels of TPTN and ALB were observed significantly decreased indicating acute hepatic cellular damage and biliary obstruction (Table 1, Fig. 2).

The rats treated with ethanolic extract of C. grandis and silymarin, experienced a significant decrease in the levels of SGOT, SGPT, ALKF, TBL, and CHL which were elevated after administration of CCL₄, while a significant increase in TPTN and ALB levels were observed in these animals (Table 1). The rats treated with the aqueous extract showed significant decrease in the levels of SGOT and CHL and an increase in the levels of ALB. Histopathological examination of liver sections of the control group showed normal cellular architecture with distinct hepatic cells, sinusoidal spaces and a central vein (Fig. 1). Disarrangement of normal hepatic cells with intense central fibrosis and vacuolization of periportal vein was observed in liver of CCL₄-intoxicated rats (Fig. 2). The liver sections of the rat treated with ethanolic extract and intoxicated with CCL₄ (Fig. 3), showed less vacuole formation and absence of necrosis and no visible changes were observed as compared to silymarin (Fig. 5), which further corroborated the protective effect of the extract. Although, liver sections of rats treated with aqueous extract and intoxicated with CCL₄ (Fig. 4) showed less visible changes, it was not as prominent as that of ethanolic extract-treated rat sections.

In Indian system of medicine certain herbs are claimed to provide relief against liver disorders. The claimed therapeutic reputation has to be verified in a scientific manner. In the present study, the leaves of one such herb Coccinia grandis was taken for the evaluation of hepatoprotective activity. The ethanolic extract of Coccinia grandis leaves possessed significant (p<0.05) hepatoprotective effect in the CCL₄ model of intoxication in rats. Preliminary phytochemical investigation on the extracts showed the presence of glycosides, terpenoids, alkaloids and flavonoids in the ethanolic extract. According to these results, it may be hypothesized that flavonoids, which are present in the ethanolic extract, could be responsible for the hepatoprotective activity. The hepatotoxicity of CCL₄ has been attributed to the formation of the highly reactive trichloromethyl free radicals, which attack polyunsaturated fatty acids. CCL₄ produces hepatotoxicity by altering the liver microsomal membranes in experimental animals (Aschock et al., 2001). The effect of CCL₄ is generally observed after 24 h of its administration. Hence, the withdrawal of blood for biochemical parameters should be carried out only after 24 h of CCL₄ intoxication.

![Fig 1: Normal rat liver section](image1.jpg)

![Fig 2: Liver section of rat intoxicated with CCL₄](image2.jpg)

Table 1: Effect of oral administration of Coccinia grandis on CCL₄-induced liver damage indices

<table>
<thead>
<tr>
<th>Groups</th>
<th>SGOT (U/L)</th>
<th>SGPT (U/L)</th>
<th>ALKF (mg/dL)</th>
<th>TBL (mg/dL)</th>
<th>CHL (mg/dL)</th>
<th>TPTN (mg/dL)</th>
<th>ALB (mg/dL)</th>
</tr>
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<tbody>
<tr>
<td>Central</td>
<td>50.24±3.24</td>
<td>49.20±3.35</td>
<td>342.10±1.24</td>
<td>0.23±0.13</td>
<td>7.08±0.12</td>
<td>1.02±0.12</td>
<td></td>
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<tr>
<td>CCL₄</td>
<td>487.78±5.72</td>
<td>332.01±2.32</td>
<td>471.02±7.26</td>
<td>2.64±0.21</td>
<td>478.20±13</td>
<td>1.26±0.78</td>
<td></td>
</tr>
<tr>
<td>Silymarin</td>
<td>102.30±1.23</td>
<td>62.32±0.12</td>
<td>351.28±4.27</td>
<td>0.24±0.02</td>
<td>69.42±3.59</td>
<td>2.7±0.32</td>
<td>2.76±0.32</td>
</tr>
<tr>
<td>Aqueous extract</td>
<td>158.20±3.2</td>
<td>223.18±4.2</td>
<td>598.24±9.12</td>
<td>1.88±0.04</td>
<td>60.91±2.22</td>
<td>2.32±0.24</td>
<td>2.52±0.12</td>
</tr>
<tr>
<td>Ethanol extract</td>
<td>114.81±2.72</td>
<td>85.61±1.32</td>
<td>362.22±4.32</td>
<td>0.56±0.21</td>
<td>45.81±4.55</td>
<td>6.12±0.46</td>
<td>2.52±0.12</td>
</tr>
</tbody>
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Values are Mean±SEM. *Significant reduction compared to CCL₄ (p<0.05). **Significant increase compared to CCL₄ (p<0.05).
failure and a decrease in protein synthesis and the accumulation of triglycerides. Intoxication with CCl₄ also resulted in inhibition of bile acids synthesis from cholesterol which is synthesized in liver or derived from plasma lipids, leading to an increase in cholesterol levels. Suppression of cholesterol levels by CCl₄ suggests the inhibition of the synthesis of bile acids from cholesterol which was reversed by the extract. A reduction in the levels of SGOT and SGPT towards the normal value is an indication of revival of plasma membrane as well as repair of hepatic tissue damage caused by CCl₄. A reduction of ALKP levels with concurrent decrease in the raised bilirubin level suggests the stabilization of biliary function which had been adversely affected by injury due to CCl₄. Protein and albumin levels were raised by the extract suggesting the stabilization of endoplasmic reticulum leading to protein synthesis.

The protective effect exhibited by the ethanolic extract was similar to that due to silymarin treatment. Histological examination of the liver sections revealed that the normal liver architecture was disturbed by hepatotoxin intoxication. In the sections obtained from the rats treated with ethanolic extract and intoxicated with CCl₄, the normal cellular architecture was retained as compared to silymarin, thereby confirming the protective effect of the extract. Although, the less visible changes were observed in the liver sections of the rats intoxicated with CCl₄ and subsequently treated with aqueous extract (200 mg kg⁻¹) and intoxicated with CCl₄, the intensity was less compared to ethanolic extract-treated rat sections. Most of the previous studies are reported on the hepatoprotective activity of the fruits of C. grandis (Vadivelu et al., 2008; Swamy et al., 2007). The only literature available for the same activity using the leaves extract is reported by Unnamaneswari and Tapas Kumar (2008), in which the hepatic damage is induced in mice using Paracetamol (2 g kg⁻¹). However, in the present study the hepatotoxicity was induced by CCl₄, in which the changes associated with liver damage are similar to that of acute viral hepatitis (Sreelatha et al., 2009). The ethanolic extract of C. grandis leaves at a dose of 200 mg kg⁻¹ showed significant hepatoprotective activity in the latter model. Preceding studies revealed the presence of Cucurbitacin glucosides in members of the Cucurbitaceae family and are well known for its hepatoprotective activity (Tehiah et al., 2007) which may support the findings of our study. The results obtained validate the folklore claim of the plant being used for the treatment of jaundice.

**CONCLUSION**

From this investigation it is evident that, the ethanolic extract of the leaves of Cocculus grandis
possessed significant hepatoprotective activity against CCl4 intoxication in rats. Further detailed studies may confirm the utility profile of this drug.

ACKNOWLEDGMENTS

Authors are grateful to Dato Prof. Dr. Ishak Bin Tambi Kechik, Vice Chancellor and Dato. Edmund Santhara, CEO, Masterskill University College of Health Sciences, Malaysia, for their encouragement and support.

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