Effect of Dietary Fishmeal on Cell-Mediated Immune Response of Laying Hens

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Abstract: In this study, we examined the role of different amounts of dietary fishmeal, a component of chicken diet on cell-mediated immune response of chickens. Forty eight laying white Leghorn chickens were randomly divided into 4 groups and fed diets containing 0, 1.5, 3 or 6% fish-meal. Six chickens from each dietary group were euthanized 3 and 9 weeks later and the spleens were removed to isolate cells and assess immune response. Mitogenic response was determined by [3H]-thymidine incorporation and the cell subsets were established by flow cytometry. Three weeks of dietary treatment had no impact on Concanavalin A (Con A) or phytohemagglutinin (PHA) induced proliferation or cell subsets of splenic lymphocytes. However, 9 weeks post dietary treatment PHA-induced proliferation and CD4 cell population were elevated in 3% fishmeal fed hens compared to those fed the control (0% fishmeal) diet. Our results suggest that a moderate increase in dietary fish-meal may enhance the immune response in laying hens. This may prove useful in infections in which cell-mediated immunity plays a role.

Key words: Fishmeal, laying hens, cell-mediated immune response

Introduction

Fishmeal is a common reservoir of Salmonella and a leading vehicle for outbreaks. Consumption of raw or partially cooked Salmonella enteritidis (SE)-contaminated eggs has been recognized as a significant source of human salmonellosis (Hope et al., 2002). Salmonella is frequently found in the intestines of commercial broilers and laying hens. The organism colonizes primarily in the lower gastrointestinal tract of the chickens. Typically, these chickens appear healthy without any symptoms even though large numbers of Salmonella are excreted in their feces. One of the ways of eliminating or reducing the Salmonella contamination in the poultry products may be by improving the ability of the birds to avoid infection. Immunoenhancement could be one of the approaches since extensive mouse studies have shown the role for T cells, natural killer (NK) cells, macrophages, neutrophils and numerous cytokines in Salmonella resistance (Mastroeni et al., 2001; Mittrucker and Kaufmann, 2000). Our recent findings in which a live-attenuated vaccine improved cell-mediated immunity (CMI) and also reduced SE shedding suggested that the immune system plays a role in Salmonella clearance (Babu et al., 2004). Diet has been shown to play an important role in modulating the resistance to infection and the immune system (Fraser et al., 2000; Lesourd, 2004; Digby et al., 2003; Klasing, 1998). With regard to Salmonella infections, increased protein, lactose, zinc-methionine, lactulose and calcium phosphate, capsaicin, mannose or palm kernel meal have been shown to improve bacterial clearance (Otani et al., 2003; Corrier et al., 1990; Kidd et al., 1994; Bovee-Oudenhoven and Van Der Meer, 1997; McElroy et al., 1994; Allen et al., 1997). Similarly, there have been reports of negative effects of forced molting and high doses of n-3 and n-6 fatty acids on Salmonella clearance (Hoit, 2003; D’Ambola et al., 1991). Dietary fish oil was shown to decrease kuppfer cell phagocytosis and oxidative burst during the early part of Salmonella typhimurium infection as well as increase mortality rates of infected mice (Eicher and McVey, 1986; Anonymous, 1992; Chang et al., 1992). Fish oil has also been shown to be immunosuppressive at high concentrations and immunostimulatory at lower concentration in chickens (Wang et al., 2000; Korver and Klasing, 1997). Although fishmeal is a component of the commercial diet for laying hens (Gous and Swatson, 2000; Reddy et al., 1997; Ponce and Garnet, 2002; Aletor, 1990), its role on immune response has received little attention. Therefore, we investigated the impact of different levels of fishmeal on the functions of splenic lymphocytes from laying hens. Results from this study could help the poultry industry in optimizing the levels of fishmeal in the feed for improving the immune response.

Materials and Methods

Animals and diets: Forty eight specific pathogen-free White Leghorn laying hens were housed in individual laying wire cages and were provided feed and water ad libitum. The hens were randomly divided into 4 groups and were fed diets containing 0, 1.5%, 3% or 6% fishmeal. Composition of the diets is presented in
Table 1: Composition of Control and Test Diets

<table>
<thead>
<tr>
<th></th>
<th>Control</th>
<th>Test 1</th>
<th>Test 2</th>
<th>Test 3</th>
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</thead>
<tbody>
<tr>
<td>Ground corn</td>
<td>61.30%</td>
<td>61.30%</td>
<td>61.30%</td>
<td>61.30%</td>
</tr>
<tr>
<td>Soy Meal</td>
<td>21.5%</td>
<td>20.0%</td>
<td>18.50%</td>
<td>15.50%</td>
</tr>
<tr>
<td>Soy Oil</td>
<td>2%</td>
<td>2%</td>
<td>2%</td>
<td>2%</td>
</tr>
<tr>
<td>Protein Blend</td>
<td>5%</td>
<td>5%</td>
<td>5%</td>
<td>5%</td>
</tr>
<tr>
<td>LimeStone (Ca)</td>
<td>7%</td>
<td>7%</td>
<td>7%</td>
<td>7%</td>
</tr>
<tr>
<td>Alfaalfa Meal</td>
<td>2%</td>
<td>2%</td>
<td>2%</td>
<td>2%</td>
</tr>
<tr>
<td>Fish Meal</td>
<td>0.00%</td>
<td>1.50%</td>
<td>3%</td>
<td>6%</td>
</tr>
<tr>
<td>DL Methionine</td>
<td>0.50%</td>
<td>0.50%</td>
<td>0.50%</td>
<td>0.50%</td>
</tr>
<tr>
<td>Vitamin/Mineral Mix</td>
<td>0.50%</td>
<td>0.50%</td>
<td>0.50%</td>
<td>0.50%</td>
</tr>
<tr>
<td>NaCl</td>
<td>0.30%</td>
<td>0.30%</td>
<td>0.30%</td>
<td>0.30%</td>
</tr>
</tbody>
</table>

Table 1. The birds were euthanized by CO₂ asphyxiation after 3 and 9 weeks of dietary treatment. Spleens were removed immediately and used for isolating mononuclear cells (MNC).

Isolation of mononuclear cells from spleen: Spleens were washed and minced in HBSS and the residual tissue was removed using a cell strainer. Single cell suspension from the spleens was overlayed onto Histopaque (density 1.077 g/ml) and centrifuged at about 400xg for 30 min at room temperature. Mononuclear cells obtained from the interface were washed twice with HBSS and suspended in RPMI 1640 medium containing 5% heat inactivated fetal calf serum (Life Technologies, Rockville, MD). Cell concentrations were adjusted to 8x10⁶ cells/ml.

Mitogenic reactivity in vitro: Aliquots (50 μl) of splenic MNC suspended in RPMI 1640 complete medium with 5% fetal calf serum were placed in 96 well plates. Another aliquot (50 μl) of either RPMI 1640 complete medium or medium with 3 different concentrations of Con A (0.4, 0.8 or 1.6 μg/ml) or PHA (0.8, 1.6 or 3.2 μg/ml). After 48 hours of incubation in air-5% CO₂ at 41°C, 1μCi ³H-thymidine was added to each well. Cultures were incubated for an additional 24 hours. Cells were harvested using a 96 well automatic harvester and counted in a microbeta counter. Data are presented as cpm from stimulated cultures after subtracting the cpm from non-stimulated cultures.

Antibodies and flow cytometric analysis: Analysis of splenic lymphocyte populations was done by FACS analysis using an EPICS Elite flow cytometer (Beckman/Coulter, Miami FL). Splenic lymphocytes were suspended in phosphate buffered saline (PBS) containing 2% heat inactivated fetal bovine serum and 0.05% sodium azide (FACS diluent) at a concentration of 10⁷ cells per ml. Monoclonal antibodies specific for chicken cell surface antigens were added to 50μl of FACS diluent in wells of a 96 well microtiter plate to achieve a predetermined optimal final concentration (0.1-1.0 μg/50μl). Ten microliters of spleen cell suspension were added to each well. Antibodies (Southern Biotechnology Associates, Inc, Birmingham, AL) used for immunofluorescent staining were directed against chicken T lymphocytes (CD3, CD4, CD8). Direct FITC or R-phycocerythrin conjugated antibodies were used for staining. Matched isotype control antibody conjugates were also used to determine background staining. Immunofluorescent staining took place for 30 minutes at 4°C. Samples were then washed twice and re-suspended in 100μl FACS diluent. FACS analysis was conducted on the viable lymphocyte population as determined by forward light scatter versus 90 degree light scatter gating. Five thousand cells were analyzed for each antibody combination.

Statistical analysis: Data pertaining to proliferation and cell populations were compared between the control and the experimental groups by conducting one-way analysis of variance (ANOVA) followed by pair-wise multiple comparison procedures (Tukey test). These statistical analyses were done using Sigma Stat v2.03. Differences between the treatment groups were considered statistically significant at P < 0.05.

Results and Discussion
In this study, we tested the impact of dietary fishmeal ranging from 0 to 6% (by weight) on cell-mediated immune parameters, such as splenic lymphocyte proliferation and cell subsets. This range of fishmeal included the amount present in commercial chicken feeds (1.5%) and at the highest level (6%), it did not cause any off flavor or taste as evidenced by no negative impact on food consumption or health of the hens (data not shown). Fishmeal has been used as a protein source for growing chicks for over three decades (Schumaier and McGinnis, 1969; Reddy et al., 1997; Ponce and Gernat, 2002). Although dietary fishmeal has been shown to improve growth, performance and resistance to parasitic infections in chickens and young sheep, (Reddy et al., 1997; Aleto, 1990; van Houtert et al., 1995), its impact on the immune response has received little attention. Therefore, we assessed the role of moderate increase in dietary fishmeal on the splenic
In addition to increased proliferation of splenic lymphocytes, we observed a significant increase in the percentage of splenic CD4 cells from hens that were fed 3% fishmeal for 9 weeks (Fig. 2). Enhanced splenic lymphocyte proliferation to PHA in 3% fishmeal fed hens (and the lack there of in the 6% fishmeal fed hens) could be partially attributed to the increase in CD4 cells (Kasirnker and Tokrasiwin, 1999), which are known to support lymphocyte activation, possibly by increased IL-2 production (Quade and Roth, 1999; Abdalla et al., 2003). In addition to their role in lymphocyte activation, the CD4 cells are known to play a role in protection against intracellular pathogens by producing macrophage activating factors and by generating a response to Salmonella flagellar protein (Mastroeni et al., 1992; Kaufmann 1993; McSorley et al., 2000). Involvement of CD4 cells in protection against Salmonella infection was also established by abrogation of Salmonella typhimurium clearance from the spleens by in vivo treatment of mice with anti-CD4 monoclonal antibody (Nauci, 1990). In general, role of cell-mediated immunity (CMI) in Salmonella elimination has been established in rodents (Ugrinovic et al., 2003; Bumann, 2003), and one of our recent studies showed an association between improved CMI and reduced Salmonella enteritidis (SE) shedding in young vaccine
treated chickens (Babu et al., 2004).
Overall, our results showed that moderate levels of
dietary fishmeal had the potential to improve the
immune response of laying hens. Upon further
exploring, if there is a positive correlation between
dietary fishmeal and SE clearance, it may be used in
addition to other dietary regimens to reduce the
incidence of Salmonella infection in chickens.

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