



Journal of Biological Sciences

ISSN 1727-3048

science
alert

ANSI*net*
an open access publisher
<http://ansinet.com>

Oxygen Transport in Amphibia: The Functional Properties of Hemoglobins from *Bufo bufo* and *Bufo viridis*

¹Maria Elisabetta Clementi, ²Francesco Misiti, ¹Federica Orsini, ¹Michela Pezzotti and ¹Bruno Giardina

¹CNR Institute Chimica del Riconoscimento Molecolare and Institute of Biochemistry and Clinical
Biochemistry, Faculty of Medicine, Catholic University Largo F. Vito 1, 00168 Rome, Italy

²Department of Health and Motor Sciences, University of Cassino, V.le Bonomi, 03043 Cassino (FR), Italy

Abstract: The oxygen binding properties of the hemoglobins from two toads, *Bufo bufo* and *Bufo viridis*, have been investigated as a function of protons, chloride ions, organic phosphates and temperature. Electrophoretic analysis of the hemolysates showed the presence of a main hemoglobin in each of the two species. We found that the hemoglobin from *Bufo bufo* shows at 20°C a slight Bohr effect which tends to increase in the presence of the different allosteric effectors (chloride ions and ATP). At 37°C, the effect of protons is completely abolished in all experimental conditions. The *Bufo viridis* Hb presents a Bohr effect slightly more pronounced (doubled with respect to *B. bufo*) which increases only with the simultaneous presence of modulators, both at 20 and at 37°C. Moreover the overall heats of oxygenation (expressed by ΔH values) result in the two amphibian hemoglobins much less exothermic than that of the human hemoglobin and in the case of *Bufo viridis* completely independent by organic phosphate (DPG). These particular features are very interesting because the two hemoglobins seem well adapted to the different habitats and physiological needs characterizing the two toads.

Key words: Amphibian hemoglobin, oxygen transport, overall oxygenation enthalpy (ΔH)

INTRODUCTION

Hemoglobins in their purified state exhibit a great deal of variation in terms of absolute affinities for oxygen and their susceptibility to control by effectors such as chloride, carbon dioxide, protons and organic phosphates. This is generally thought to be a reflection of both the variable oxygen tension in which organisms live and the variable oxygen demands of their respiring tissues.

The modulation of function induced by the effectors mentioned above has important physiological effects. As an example, at the tissue level, the decrease of oxygen affinity brought about by the increase in proton activity (alkaline Bohr effect) allows a more efficient unloading of oxygen and contributes to the neutralization of protons produced by CO₂.

Another important feature of the reaction of Hb with O₂ is its temperature dependence which is determined by the associated overall enthalpy change (ΔH).

Hence, a number of studies (di Prisco *et al.*, 1991; Giardina *et al.*, 1992; Clementi *et al.*, 1994) of Hb function at varying temperature revealed, in different species, a series of adaptive mechanisms which are based on the thermodynamic connection between the binding of

heterotropic effectors and the reaction with oxygen. From a physiological point of view, it may be important to outline that this thermodynamic approach is particularly interesting in the case of cold-blooded species such as amphibian.

Along these lines and in order to widen the scope of this emerging scheme, we have investigated the functional properties of the hemoglobin from two species of genus *Bufo*: *Bufo bufo* and *Bufo viridis*.

Bufo bufo is the so called common toad, is a widespread species and is present in moist meadows and woods, whereas *Bufo viridis* prefers xeric environments tolerating more dehydration, the salinity and higher temperatures.

The results obtained from the hemoglobins of *Bufo bufo* and *Bufo viridis*, put in evidence the different characteristics developed by the hemoproteins of these toads (concerning to the same genus) during evolution in relation to their different habitats in which they live.

MATERIALS AND METHODS

Adult toads of both sexes (ten individuals for each species) were captured during March to November in

Calabria. Blood samples were collected, in the presence of EDTA, from anaesthetized toads, through heart puncture. Values for chloride ions concentration was determined with a colorimetric method (Clorofix-Menarini); the assay for nucleotide derivatives (DPG and ATP) was performed with enzymatic kits (Sigma).

The red cells were washed three times with iso-osmotic NaCl solution by centrifugation at 1000 g and the packed cells lysed by adding 2 volumes of cold hypotonic buffer in the presence of 10 μ M Triton X-100. The stroma and the nuclei were removed by centrifugation at 12000 g for 30 min. Electrophoretic analysis of hemolysates was performed both with cellulose acetate alkaline electrophoresis and with IEF (isoelectrofocusing), a 2% ampholine in the pH range 5.5-8.5 after treatment of the samples with KCN. Purification of principal components was performed using ion-exchange chromatography on a CM-column equilibrated with 10 mM PO_4 buffer. The homogeneity of the isolated fractions was checked by cellulose acetate electrophoresis.

Stripped hemoglobins were obtained by passing first through a Sephadex G-25 column equilibrated with 0.01 M Tris buffer, pH = 8, containing 0.1 M NaCl and then through a column of mixed-bed ion-exchange resin (Bio-Rad AG 501 \times 8).

Concentrated stock solutions of ATP and DPG were prepared by dissolving powder (Sigma) in Hepes buffer.

Oxygen binding isotherms have been determined by the tonometric method (Giardina and Amiconi, 1981), in the absence and presence of allosteric effectors, between 15 and 37°C. The overall oxygenation enthalpy corrected for the solubilization heat of O₂ ($\Delta H = -3$ kcal. mol⁻¹), has been calculated from the integrated Van't Hoff equation:

$$\Delta H = - 4.574 [(T_1 T_2) / (T_1 - T_2)] \Delta \log P_{50} / 1000, \text{ kcal. mol}^{-1},$$

where P₅₀ is the partial pressure of the ligand at which 50% of hemes is oxygenated. Over the temperature range explored (15 to 37°C) Van't Hoff plots were linear within the experimental error. An average standard deviation of $\pm 8\%$ for values of P₅₀ and of $\pm 15\%$ for ΔH values was calculated.

Under all the experimental conditions, the ferric derivative of Hb from two toads (met-Hbs) were always lower than 3%.

RESULTS

Table 1 shows the hematocrit values, plasmatic concentration of chloride ions and the intra-erythrocytic concentration of ATP (Adenosine-5'-Triphosphate) and DPG (2,3-diphosphoglyceric acid) calculated for *Bufo bufo* and *Bufo viridis* blood.

Table 1: Means and standard deviations (obtained on 10 observations for each species) for hematocrit, plasmatic concentration of chloride ions and intra-erythrocytic concentration of DPG and ATP in *Bufo bufo* and *Bufo viridis* blood

Parameters	<i>Bufo bufo</i>	<i>Bufo viridis</i>
Hematocrit (%)	30 \pm 8	28 \pm 8
Cl ⁻ (mEq L ⁻¹)	125 \pm 10	129 \pm 25
ATP (μ mol L ⁻¹ erythrocytes)	0.350 \pm 0.050	0.329 \pm 0.050
DPG (μ mol L ⁻¹ erythrocytes)	0.150 \pm 0.030	0.125 \pm 0.030

There are no statistical differences between the values of two toads; however it is evident that the organic phosphate present in great amount in red cells is Adenosine-5'-Triphosphate. Our functional data have been performed in the presence of both ATP alone and of ATP plus DPG and the obtained values of oxygen affinity were identical. Thus, the reported results are those obtained in the presence of Adenosine-5'-Triphosphate alone.

The electrophoretic analysis of the hemolysate from *Bufo bufo* and *Bufo viridis* has pointed out the presence of a principal hemoglobin component, characterized by mobility very similar to that of human HbA (data not shown) and purified as reported in the methods section.

Figure 1 shows the Bohr effect of *Bufo bufo* hemoglobin investigated within the pH range of 6.3-8.2 both in the absence and presence of chloride ions and ATP, at 20°C (panel A) and 37°C (panel B). At 20°C, both in the presence and in the absence of chloride ions, the Bohr effect is slightly hinted; instead, the presence of physiological organic phosphate, amplifies strongly the effect of the protons.

The situation is different at 37°C: In fact, in all examined conditions, the effect of protons is completely abolished and the values of P₅₀ are identical over the entire pH range.

Figure 2 shows the Bohr effect of *Bufo viridis* Hb in the presence and in the absence both of chloride ions and ATP at 20°C (panel A) and at 37°C (panel B). As evident, *Bufo viridis* hemoglobin, reveals a major sensitivity to protons, showing at 20°C, a Bohr effect, in the presence and in the absence of Cl⁻, higher than that showed by *Bufo bufo* Hb. The effect of protons, in the presence both of chloride ions and ATP, is similar in amplitude to that displayed by the other toad Hb in the same conditions. At 37°C the situation is completely different respect to *Bufo bufo* Hb: in fact *Bufo viridis* hemoglobin shows a strong pH effect that reaches the maximum values when the experiments are performed in the presence both of chloride ions and organic phosphates.

These results are better evidenced by values (reported in Table 2) of Bohr coefficients ($\Delta \log P_{50} / \Delta \text{pH}$), calculated for all experimental condition in a pH range between 6.6 and 8.0. In the same table the corresponding

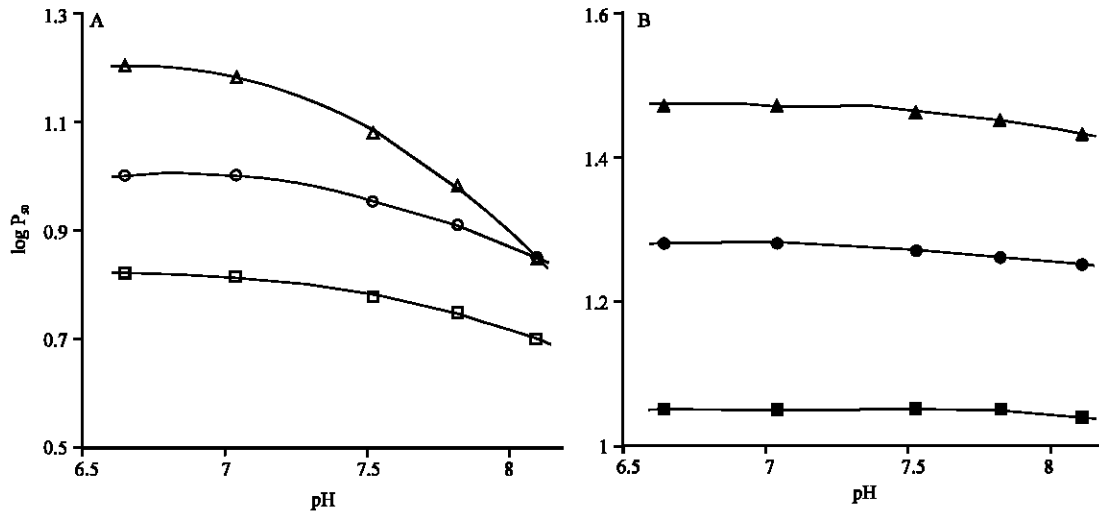


Fig. 1: Effect of pH on the oxygen affinity (in terms of $\log P_{50}$) of *Bufo bufo* hemoglobin at 20°C (panel A) and 37°C (panel B). The experiments were performed in the absence (squares) and presence (circles) of 0.1 M chloride ions and in the presence (triangles) of 10 mM ATP. Conditions: 0.1 M Mes or HEPES or TAPS buffer

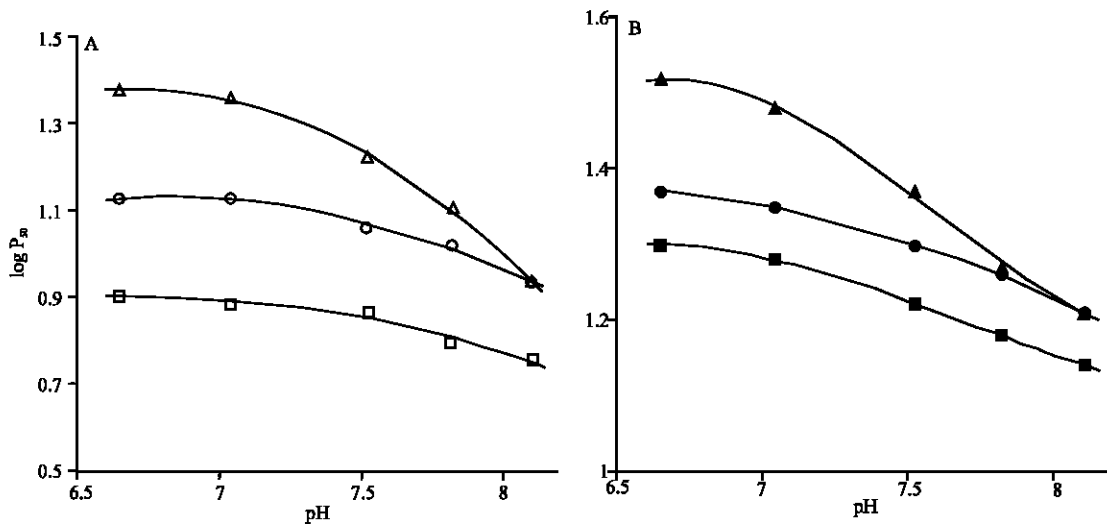


Fig. 2: Effect of pH on the oxygen affinity (in terms of $\log P_{50}$) of *Bufo viridis* hemoglobin at 20°C (panel A) and 37°C (panel B). The experiments were performed in the absence (squares) and presence (circles) of 0.1 M chloride ions and in the presence (triangles) of 10 mM ATP. Conditions: 0.1 M Mes or HEPES or TAPS buffer

Table 2: Bohr coefficients (expressed as $\Delta \log P_{50} / \Delta \text{pH}(6.6-8)$) for *Bufo bufo* and *Bufo viridis* Hbs and human hemoglobins (HbA), in different experimental conditions. The utilized organic phosphate was ATP for toad Hbs and DPG for human Hb. The data are means (\pm SD) obtained on 10 observations, for each species. The experiments were performed in HEPES buffer 0.1

Conditions	<i>Bufo bufo</i> Hb	<i>Bufo viridis</i> Hb	HbA
Stripped			
20°C	-0.082 \pm 0.010	-0.172 \pm 0.060	-0.45 \pm 0.040
37°C	-0.010 \pm 0.002	-0.110 \pm 0.040	-0.28 \pm 0.030
100 mM NaCl			
20°C	-0.103 \pm 0.030	-0.172 \pm 0.050	-0.58 \pm 0.050
37°C	-0.020 \pm 0.020	-0.110 \pm 0.040	-0.31 \pm 0.030
100 mM NaCl + 10 mM Organic phosphate			
20°C	-0.240 \pm 0.070	-0.275 \pm 0.050	-0.70 \pm 0.050
37°C	-0.024 \pm 0.030	-0.213 \pm 0.055	-0.40 \pm 0.020

values obtained for human hemoglobin, representing an homeothermic animal. As evidenced, the low temperatures and the presence of organic phosphates increase the values of Bohr coefficient also if in a different mode in the three Hbs, showing a common regulatory mechanism both in ecto- than homeothermic animals.

Another characteristic of the hemoglobins from the two toads concerns their response to changes in temperature. Figure 3 shows the effect of temperature and the influence of pH on the overall oxygenation enthalpy (ΔH expressed as Kcal/mole of Oxygen) for *Bufo bufo* (panel A) and *Bufo viridis* (panel B) both in the presence and absence of chloride ions and organic phosphates. As

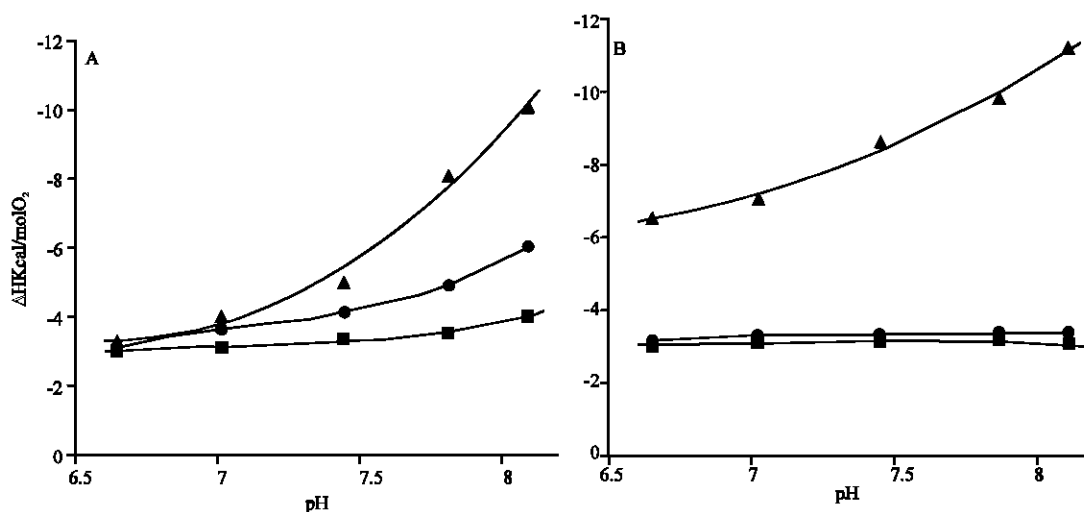


Fig. 3: Overall ΔH values (expressed as Kcal/mol of oxygen) as a function of pH, for *Bufo bufo* (panel A) and *Bufo viridis* (panel B) hemoglobins. The experiments were performed in the absence (squares) and presence (circles) of 0.1 M chloride ions and in the presence (triangles) both of 0.1 M chloride ions and 10 mM ATP. The values are calculated from van't Hoff equation, by using the data obtained from oxygen equilibria experiments and are corrected for the heat contribution of oxygen in solution (-3 Kcal/mol). Conditions: 0.1 M Mes or Hepes or Taps buffer

evident both the hemoglobins, present ΔH values obtained without and plus chloride strongly exothermic.

When the experiments were carried out in the presence of ATP, the temperature effect is markedly increased in particular for *Bufo viridis* Hb. This effect is the contrary of that observed for human Hb where the presence of organic phosphates decreases the effect of temperature, reducing the ΔH value from -11 (in stripped condition) to -9 Kcal/mol of oxygen (data not shown).

DISCUSSION

Oxygen carriers are one of the most interesting systems for studying the interrelationships between environmental conditions and molecular evolution. Hemoglobin (Hb), being a direct link between the exterior and body requirements, has experienced a major evolutionary pressure to adapt and modify its functional features. In order to ensure an adequate supply of oxygen to the entire organism, hemoglobins have developed a number of complex regulatory mechanisms that involve binding of allosteric effectors (i.e., protons, chloride ions, organic phosphates and CO_2) and the effect of temperature. Based on these considerations, the study of the functional properties of hemoglobin from two toads *Bufo bufo* and *Bufo viridis*, adapted to different environments, may be considered an interesting model to study the molecular strategies adopted by two ectothermic animals in relation to different habitats. In this

regard, the hemoglobins from the two toads showed a slight Bohr effect indicating a slight sensitivity of oxygen affinity to changes in blood pH; this was particularly true for *Bufo bufo* Hb at 37°C. Moreover, the two Hbs are very sensitive to the effect of both chloride ions and organic phosphates. These phenomena can not be considered unique to these species of amphibians. For example, the Bohr effect of *Rana temporaria* is strongly pronounced (between 0.36 and 0.51) and chloride ions and ATP have no significant effect on oxygen affinity (Bardgard *et al.*, 1997). The same characteristics are present also in other anuran amphibians Hbs such as *Rana esculenta* (Sinha, 1983) and *Xenopus laevis* (Brunori *et al.*, 1985): all these hemoglobins, in N-terminus of beta chains, possess a glycine (Kleinschmidt and Sgouros, 1987) which is considered partially responsible for the Bohr effect (Perutz *et al.*, 1969): in the case of *Bufo bufo* instead, this position is occupied by a residue of Valine (Caffin *et al.*, 1969). As the ATP effect is concerned, the beta chains of frog hemoglobin lack of the first six amino acid residues, thus there is no His β 2 lining the organic phosphate binding pocket: these residues instead are present in toad hemoglobins (Brittain, 1991).

However the maximum sensitivity to pH changes of *Bufo bufo* and *Bufo viridis* Hbs, is present (in the presence of chloride ions and ATP) at 20°C and at 37°C respectively: it is important to note that *Bufo bufo* prefers environments around at 20°C while *Bufo viridis* is adapted better at high temperatures. In any case, a low Bohr effect,

may be considered as a protective mechanism to modulate the altered blood parameters, such as O₂ and CO₂ and urea concentration that the toads manifest during aestivation periods in which metabolic pathways are very depressed (Jokumsen and Weber, 1980).

With respect to the overall heats of oxygenation (ΔH) of *Bufo bufo* and *Bufo viridis* Hbs, these are very little exothermic in particular in the absence of organic phosphates evidencing a low sensitivity to temperature. The more exothermic values of ΔH are observed in the presence of ATP especially at alkaline pH values. This result is completely opposite to that found for human Hb and for other mammalian Hbs where the liberation of organic phosphates is an endothermic reaction whereby the apparent overall heat of oxygenation, in the presence of DPG is less and less exothermic with respect to stripped conditions. Moreover it is possible to note that enthalpy values in the presence of ATP are pH dependent: this variation of ΔH is probably due to the increasing endothermic contribution of the Bohr protons, which cancels some of the heat released upon O₂ binding. In any case, in para-physiological conditions (presence of chloride ions and ATP at pH between 6.8 and 7.6) the ΔH values of both the toads are sufficiently exothermic as expected for etherothermic animals. This property is interpretable as a mechanism of adaptation in relation to the sudden variations of body temperature that these cold blood animals present seasonally, representing an example of molecular adaptation to specific respiratory requirements. Moreover the smaller variation on ΔH from pH observed in the two toads Hbs might be indicative on an energy saving mechanism of loading and unloading oxygen with respect to human Hb, but the differences observed between the *Bufo bufo* and *Bufo viridis* Hbs are not so much significant.

REFERENCES

- Bardgard, A., A. Fago, H. Malte and R.E. Weber, 1997. Oxygen binding and aggregation of hemoglobin from the common European frog, *Rana temporaria*. Comp. Biochem. Physiol. B Biochem. Mol. Biol., 117: 225-231.
- Brittain, T., 1991. Cooperativity and allosteric regulation in non-mammalian vertebrate hemoglobins. Comp. Biochem. Physiol. B., 99: 731-740.
- Brunori, M., S.G. Condo, A. Bellelli B. Giardin and G. Micheli, 1985. Tadpole *Xenopus laevis* hemoglobin. Correlation between structure and functional. J. Mol. Biol., 181: 327-329.
- Caffin, J.P., J.P. Chauvet and R. Acher, 1969. Les hemoglobines des amphibiens: Separation et caracterisation preliminaire des chaines d'une hemoglobine du crapaud *Bufo bufo*. FEBS Lett., 5: 196-198.
- Clementi, M.E., S.G. Condo, M. Castagnola and B. Giardin, 1994. Hemoglobin function under extreme life conditions. Eur. J. Biochem., 223: 309-317.
- di Prisco, G., S.G. Condò, M. Taniburrini and B. Giardina, 1991. Oxygen transport in extreme environment. Trends Biochem. Sci., 16: 471-474.
- Giardina, B. and G. Amiconi, 1981. Measurement of binding of gaseous and nongaseous ligands to hemoglobin by conventional spectrophotometric procedures. Meth. Enzymol., 76: 417-427.
- Giardina, B., A. Galtieri, A. Lania, P. Ascenz and A. Desideri *et al.*, 1992. Reduced sensitivity of O₂ transport to allosteric effectors and temperature in loggerhead sea turtle hemoglobin: Functional and spectroscopic study. Biochim. Biophys. Acta, 1159: 129-133.
- Jokumsen, A. and R.E. Weber, 1980. Hemoglobin-oxygen binding properties in the blood of *Xenopus laevis*, with special reference to the influences of aestivation and of temperature and salinity acclimatation. J. Exp. Biol., 86: 19-37.
- Kleinschmidt, T. and J.G. Sgouros, 1987. Hemoglobin sequences. Biol. Chem. Hoppe-Seyler, 368: 579-615.
- Perutz, M.F., H. Muirhead, L. Mazzarella, R.A. Crowther, J. Greer and J.V. Kilmartin, 1969. Identification of residues responsible for the alkaline Bohr effect in hemoglobin. Nature, 222: 1240-1243.
- Sinha, R.C., 1983. Haematological studies on the prewintering and wintering frog, *Rana esculenta*. Comp. Biochem. Physiol. A., 74: 311-314.