Effect of *Acacia nilotica* Fruit Extract on Serum Glucose and Lipid Concentrations in Alloxan-induced Diabetic Rats

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**Abstract:** The present investigation was performed to study the effects of *Acacia nilotica* Delile (Fabaceae) fruit extract on serum concentrations of total cholesterol, low-density lipoprotein cholesterol, high-density lipoprotein cholesterol, triglycerides and glucose in control and alloxan-induced diabetic rats. The rats were divided into four groups: Normal Control Rats (NC), normal control rats administered *A. nilotica* (NC+AN), Diabetic Control rats (DC) and diabetic rats administered *A. nilotica* (DC+AN). Each group comprised 10 to 14 rats. The methanolic extract *A. nilotica* fruit was orally administered at a concentration of 200 mg kg$^{-1}$ b.wt. for 5 consecutive weeks. The *A. nilotica* fruit extract significantly (p<0.005) decreased serum glucose levels in control rats after the third week. The serum glucose concentrations of diabetic rats following *A. nilotica* fruit extract administration did not change significantly. The *A. nilotica* extract showed a strong hypolipidemic effect on diabetic rats and significantly decreased serum levels of triglycerides and low-density lipoprotein cholesterol (p<0.02 and p<0.001, respectively). The present findings suggest that hypolipidemic effects following oral administration of *A. nilotica* fruit extract could be beneficial for treatment of diabetes-related complications and hyperlipidemia.

**Key words:** Antioxidants, diabetes complications, hyperlipidemia, low-density lipoprotein, triglycerides

**INTRODUCTION**

*Acacia nilotica* Delile (Fabaceae) is a spiny tree widely distributed in tropical and subtropical regions of Africa. In folk medicine, *A. nilotica* has been used for treating haemorrhages, tuberculosis, leprosy, colds and diarrhoea (New, 1984; Al-Mustafa and Dafallah, 2000). The efficacy of *A. nilotica* in the treatment of diabetes has been previously reported (Akhbar and Khan, 1985). The methanolic extract of *A. nilotica* was observed to cause a decrease in the concentration of plasma glucose, cholesterol, triglycerides and low density lipoprotein in alloxan-induced diabetic rabbits (Ahmad et al., 2008). Several reports have indicated a strong antioxidant potential of *A. nilotica* extracts in metal ion chelation and free radicals scavenging (Sultana et al., 2007; Singh et al., 2009).

To our knowledge, no report study the effect of *A. nilotica* fruit on diabetic animal models, however, the present study was conducted to investigate the effects of *A. nilotica* fruit extract on serum concentrations of Total Cholesterol (TC), Low-density Lipoprotein Cholesterol (LDL-C), High-density Lipoprotein Cholesterol (HDL-C), Triglycerides (TG) and glucose in alloxan-induced diabetic rats.

**MATERIALS AND METHODS**

**Chemicals and reagents:** Alloxan was obtained from CDH, Ltd. (New Delhi, India). Methanol and other chemicals were of analytical grade and were obtained from BDH (Poole, UK). Gallic acid, Folin-Ciocalteu reagent, quercetin, 2,2-azinobis (3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt (ABTS) and potassium persulfate were purchased from Sigma-Aldrich (St. Louis, MO, USA). Trolox (6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid) was obtained from Fluka Chemie AG (Buchs, Switzerland). Kits for the assays of (TC), (LDL-C), (HDL-C), (TG) and glucose were purchased from Human GmbH (Wiesbaden, Germany).

**Plant material and extraction:** *Acacia nilotica* fruit was collected from the western province of Sudan in April 2011 and identified by Dr. A. Almeligy, Department of Botany, King Saud University, Riyadh, Saudi Arabia. The *A. nilotica* fruit was immersed in liquid nitrogen prior to blending and 100 g of powder of the *A. nilotica* fruit was prepared with an electric blender. The methanol extract of the *A. nilotica* powder was obtained by dissolving the 100 g of *A. nilotica* powder in 900 mL methanol-water (4:1 v/v) at room temperature overnight using an orbital shaker. The filtrates were pooled and centrifuged at 8000 rpm for 10 min and the supernatant was concentrated under reduced pressure at 40°C using a rotary evaporator which yielded 22.3 g of *A. nilotica* methanol extract powder.

**Determination of antioxidants in *A. nilotica* fruit:** Using the aforementioned protocol, 5 g of powdered *A. nilotica* fruit was extracted with 50 mL methanol-water (4:1 v/v).
The final extract was dissolved in distilled water, the final volume was recorded and the suspended extract was divided into aliquots and kept in the dark at room temperature until determination of antioxidant properties.

**Total phenolics:** The concentration of total phenolics was determined using the method of Kim *et al.* (2003). The total phenolic content of *A. nilotica* fruit was expressed as grams of gallic acid per 100 g of *A. nilotica*.

**Total flavonoid:** The concentration of total flavonoid was measured with the aluminum chloride colorimetric method described by Chang *et al.* (2002). The total flavonoid content of *A. nilotica* fruit was expressed as milligrams of quercetin per 100 g of *A. nilotica*.

**Radical cation ABTS scavenging activity:** The ABTS free radical decolorization assay was performed as described by Re *et al.* (1999), using Trolox as a standard. The free radical scavenging activity of *A. nilotica* fruit extract was expressed as the Trolox equivalent antioxidant capacity mg Trolox equivalents per 100 g of *A. nilotica*.

**Animal groups and dosage:** Male Wistar rats (180-205 g) were obtained from the animal house at the College of Pharmacy, King Saud University. All animals were housed in cages with a 12/12 h light/dark cycle at 22°C. The animals were given rat chow and water ad libitum. Diabetes was induced via an intraperitoneal injection of alloxan (150 mg kg⁻¹ dissolved in 0.2 mL of physiological saline). The animals were kept under observation for 1 week prior to the start of treatment. All animal experiments were carried out in accordance with the King Saud University Ethical Committee Acts. Extract of *A. nilotica* was administered orally at a volume of 0.2 mL, equivalent to a concentration of 200 mg kg⁻¹ b.wt., using oral gavage. Supplementation with extract continued regularly on a daily basis over a period of 35 days.

The rats were divided into four groups: Normal Control rats (NC), normal control rats administered *A. nilotica* (NC+AN), Diabetic Control rats (DC) and diabetic rats administered *A. nilotica* (DC+AN). Each group comprised 10 to 14 rats. Body weights and food intake of all groups were recorded daily. Aliquots of blood samples from all groups were collected weekly in plain tubes. Serum samples were separated from cells by centrifugation at 5000×g in a refrigerated centrifuge.

**Biochemical assays:** Concentrations of the following biochemical parameters were determined in serum samples by standard enzymatic methods using Human kits on a Humalyzer 3000 (Human GmbH, Wiesbaden, Germany): TC (Richmond, 1973), LDL-C (Okada *et al.*, 1998), HDL-C (Issawa *et al.*, 1997), TG (Nagele *et al.*, 1984) and glucose (Barham and Trinder, 1972).

**Data analysis:** The data are presented as Means±SD. Comparisons between experimental and corresponding control rats were performed using Student’s *t*-test. The probability value *p*<0.05 was considered significant.

**RESULTS AND DISCUSSION**

**Body weights and food intake of control and alloxan diabetic rats administered *A. nilotica* fruit extract:**

Table 1 shows that gains in body weight and food intake in DC rats was significantly reduced (*p*<0.001) compared with those in NC rats. However, NC+AN and DC+AN did not show any significant changes with regard to these parameters.

**Antioxidant activity of *A. nilotica* fruit extract:** The percentage concentration of total phenolics, total flavonoid and Trolox equivalent of *A. nilotica* fruit extracts are shown in Table 2. A high concentration of total phenolics was observed (30.65±2.2 g per 100 g) in the *A. nilotica* fruit extract. Present results are in agreement with previous reports in which considerable amounts of total phenolics were reported in bark extracts of *A. nilotica* (Sultana *et al.*, 2007; Singh *et al.*, 2010). The total flavonoid (as quercetin equivalent) percentage concentration in *A. nilotica* fruit extract was

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>NC</th>
<th>NC+AN</th>
<th>DC</th>
<th>D+AN</th>
</tr>
</thead>
<tbody>
<tr>
<td>Initial b.wt. (g)</td>
<td>181.0±7.96 (12)</td>
<td>188.0±9.35 (14)</td>
<td>194.0±7.86 (14)</td>
<td>202.0±9.00 (12)</td>
</tr>
<tr>
<td>Final b.wt. (g)</td>
<td>324.0±10.74 (12)</td>
<td>335.0±10.10 (14)</td>
<td>367.0±8.54 (10)*</td>
<td>327.0±11.28 (10)</td>
</tr>
<tr>
<td>Food intake (g day⁻¹)</td>
<td>22.7±2.11 (10)</td>
<td>22.2±3.61 (12)</td>
<td>18.2±2.15 (12)*</td>
<td>20.8±1.97 (12)</td>
</tr>
</tbody>
</table>

Data are expressed as Means±SD, with number of rats given in parentheses. As compared with matched groups, *p*<0.001 (student *t*-test)

<table>
<thead>
<tr>
<th>Analyte</th>
<th>Total phenolics gallic acid (g/100 g)</th>
<th>Total flavonoid quercetin (mg/100g)</th>
<th>Trolox equivalent (mg/100g)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>A. nilotica</em> fruit extract</td>
<td>30.65±2.2</td>
<td>227.21±5.7</td>
<td>84.13±3.1</td>
</tr>
</tbody>
</table>

Data are Mean±SD (n = 3). All calculations made on dry mass basis.
Table 3: Effect of the *A. nilotica* fruit extract on serum levels of glucose, total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL-C) and high-density lipoprotein cholesterol (HDL-C) in normal control rats for 5 weeks

<table>
<thead>
<tr>
<th>Groups</th>
<th>Weeks</th>
<th>Glucose (mmol L⁻¹)</th>
<th>TC (mmol L⁻¹)</th>
<th>TG (mmol L⁻¹)</th>
<th>LDL-C (mmol L⁻¹)</th>
<th>HDL-C (mmol L⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>NC</td>
<td>0</td>
<td>6.44±1.19</td>
<td>1.18±0.23</td>
<td>0.83±0.08</td>
<td>0.47±0.16</td>
<td>0.43±0.12</td>
</tr>
<tr>
<td>NC+AN</td>
<td>1</td>
<td>5.11±0.42**</td>
<td>1.39±0.16</td>
<td>0.66±0.01***</td>
<td>0.44±0.05</td>
<td>0.51±0.12</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>4.37±0.53**</td>
<td>1.47±0.09**</td>
<td>0.66±0.10*</td>
<td>0.42±0.03</td>
<td>0.46±0.14</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>5.36±0.11**</td>
<td>1.22±0.05</td>
<td>0.69±0.05*</td>
<td>0.40±0.05</td>
<td>0.58±0.13***</td>
</tr>
</tbody>
</table>

Data are expressed as Mean±SD, with number of rats given in parentheses. As compared with matched groups, *p<0.05, **p<0.02, ***p<0.005. (student t test)

Table 4: Effect of the *A. nilotica* fruit extract on serum levels of glucose, total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL-C) and high-density lipoprotein cholesterol (HDL-C) in alloxan-induced diabetic rats for 5 weeks

<table>
<thead>
<tr>
<th>Groups</th>
<th>Weeks</th>
<th>Glucose (mmol L⁻¹)</th>
<th>TC (mmol L⁻¹)</th>
<th>TG (mmol L⁻¹)</th>
<th>LDL-C (mmol L⁻¹)</th>
<th>HDL-C (mmol L⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>DC</td>
<td>0</td>
<td>25.88±3.71</td>
<td>1.67±0.19</td>
<td>1.10±0.17</td>
<td>0.70±0.14</td>
<td>0.59±0.15</td>
</tr>
<tr>
<td>D+AN</td>
<td>1</td>
<td>24.53±3.42</td>
<td>1.49±0.25*</td>
<td>0.88±0.09*</td>
<td>0.46±0.02***</td>
<td>0.59±0.13</td>
</tr>
<tr>
<td></td>
<td>2</td>
<td>23.72±5.05</td>
<td>1.55±0.29</td>
<td>0.82±0.10*</td>
<td>0.36±0.05***</td>
<td>0.46±0.15</td>
</tr>
<tr>
<td></td>
<td>3</td>
<td>21.58±3.12</td>
<td>1.47±0.12</td>
<td>0.78±0.16**</td>
<td>0.37±0.07***</td>
<td>0.54±0.14</td>
</tr>
</tbody>
</table>

Data are expressed as Mean±SD. As compared with matched groups, *p<0.05, **p<0.02, ***p<0.001. (student t-test)

227.21±5.7 mg per 100 g in the present study; however, Sultana *et al.* (2007) previously reported that the percentage concentration of total flavonoid (as catechin equivalent) in bark extract of *A. nilotica* was 3.2±0.12 g per 100 g. The difference in the total flavonoid concentration was because Sultana *et al.* (2007) used the bark of the plant, whereas the present study used the fruit of the plant. Furthermore, Sultana *et al.* (2007) expressed total flavonoid as catechin equivalent; however, in the present study, total flavonoid were expressed as quercetin equivalent.

**Effect of A. nilotica fruit extract on serum concentration of glucose and lipids of control and alloxan diabetic rats:**

Table 3 shows that the serum glucose concentration of NC+AN rats was significantly decreased compared with that of the NC rats. The decrease in serum glucose level was observed starting from the second week of supplementation (p<0.02) and continued until the third and fifth weeks (p<0.005 and p<0.02, respectively). On the other hand, no significant effect was noticed in the DC+AN rats throughout the study (Table 4), although the serum glucose concentration was decreased from 25.88 to 21.5 mmol L⁻¹ during the fifth week of *A. nilotica* administration. However, this decrease was not significant. Asad *et al.* (2011) reported that the *A. nilotica* leaf extract at a dose of 300 mg kg⁻¹ b.wt. produced hypoglycemic and antiplatelet aggregation activity in streptozotocin-induced diabetic rats and that the effects were comparable to those of gliburide. In another report by Ahmadd *et al.* (2008) aqueous methanolic extract of *A. nilotica* pods at a dose of 400 mg kg⁻¹ b.wt. caused a decrease in blood glucose levels in alloxan-induced diabetic rabbits.

![Control](image1.png) ![Diabetic](image2.png)

**Fig. 1:** Effect of *A. nilotica* fruit extract on serum LDL-C levels of control and alloxan-induced diabetic rats

Although no clear effect was noticed in serum TC concentrations of NC and DC rats, significant (p<0.001) decreases in the second (37%) and fourth (47%) weeks were observed in the serum LDL-C concentration of DC+AN (Fig. 1, Table 4). This beneficial effect of *A. nilotica* extract on the LDL-C could be due to the presence of a high concentration of polyphenol compounds, which are known to have antioxidant properties. This finding on the effect of *A. nilotica* extract on LDL-C is comparable to that in the report of Lerman *et al.* (2010) which suggested that individuals with a high risk of developing cardiovascular diseases may benefit from a medical food containing procyanidins from *A. nilotica* because of the observed decreases in their LDL-C levels. A significant decrease in the serum TG level was also observed in NC+AN and DC+AN rats (Table 3 and 4). In NC+AN rats, the serum levels of TG decreased by 28% during the second week of
CONCLUSION

The findings of the present investigation suggest the possibility of utilizing *A. nilotica* fruit extract in a pharmaceutical preparation that could help in the treatment of diabetes related-complications and hyperlipidemia.

ACKNOWLEDGMENT

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REFERENCES


