Research Article

FICK-3 Score Combining Fibrosis-4, Insulin Resistance and Cytokeratin-18 in Predicting Non-alcoholic Steatohepatitis in NAFLD Egyptian Patients

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Abstract

Background and Objective: As liver biopsy had multiple procedure-related complications, the introduction of reliable noninvasive tests for accurate discrimination of NASH and liver fibrosis in patients with NAFLD are mandatory. The aim was to elucidate the diagnostic value of fibrosis-4 index (FIB-4), Homeostasis Model Assessment for Insulin Resistance (HOMA-IR) and Cytokeratin-18 fragments (CK18-Fs) in the prediction of NASH and liver fibrosis. Materials and Methods: One hundred Egyptian patients with NAFLD selected from outpatient’s clinics of Mansoura University Hospitals underwent histological examination through liver biopsy after approval and consent. The FIB-4, HOMA-IR, CK18-Fs (measured using a human ELISA Kit) and their combination in FICK-3 score were investigated for predicting NASH and liver fibrosis. Receiver operating characteristic (ROC) curves and multivariate logistic regression were analyzed. Results: In patients with NAFLD, the areas under the ROC were significantly high (AUC: 0.765, 0.700, 0.803, 0.835 for FIB-4, HOMA-IR, CK18-Fs, FICK-3 score, respectively, p = 0.05) displaying a highly statistically significant predictive ability for NASH. Significantly higher AUCs for these parameters were demonstrated predicting early-or advanced-stage liver fibrosis (AUC > 0.7, p ≤ 0.01). Also, the combined FICK-3 score (the sum of FIB-4 ≥ 1.46, HOMA-IR ≥ 2.11 and CK18-Fs ≥ 307U L⁻¹) had highly significant predictive values for NASH and liver fibrosis and had the best diagnostic accuracy at a cutoff value of 1 (AUC > 0.8, p ≤ 0.001). Contrasted with other diagnostic scores, FICK-3 had the best diagnostic accuracy for detection of fibrotic NASH (AUC = 0.954, p ≤ 0.001) and positively correlated with the histological features of NAFLD. Conclusion: The new combination FICK-3 score was a reliable and significant predictor for NASH and liver fibrosis in NAFLD Egyptian patients.

Key words: NASH, fibrosis-4, HOMA-IR, cytokeratin 18-Fs, FICK-3 score


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Data Availability: All relevant data are within the paper and its supporting information files.
INTRODUCTION

Non-alcoholic fatty liver disease (NAFLD) and non-alcoholic steatohepatitis (NASH) are now considered the most common chronic liver disease in western countries; its prevalence increased up to 30% in the United States with a trend towards further increase. Moreover, more recent data confirm that they play an equally important role in the middle east, far east, Africa, the Caribbean and Latin America. The NAFLD may have a benign course with just simple steatosis or progress to steatosis with inflammation, ballooning degeneration and pericellular fibrosis or Mallory’s hyaline (non-alcoholic steatohepatitis, NASH). The NASH is a progressive disease that requires medical intervention with a prevalence of 3-5% in the general adult populations. Up to 20% of patients with NASH will develop cirrhosis of the liver and 10% will suffer death related to their liver disease making NASH the second leading cause of liver transplantation.

The NAFLD is closely associated with metabolic syndrome (MetS) including diabetes mellitus, central obesity, dyslipidaemia and insulin resistance. The MetS is common in patients with NAFLD and is associated with an increased propensity to have NASH. Therefore, it has been suggested that the presence of MetS in patients with NAFLD may serve as a potential criterion for liver biopsy which is still the gold standard for diagnosis of simple hepatic steatosis and the only way to distinguish it from NASH.

Accurate differentiation of NASH from NAFLD and the evaluation of the presence of liver fibrosis determining prognosis and optimal therapy became an interesting area of research. Liver biopsy was postulated to be the most authoritative diagnostic approach identifying NASH and the consequent fibrosis in NAFLD patients. Nevertheless, it is local, invasive and limited by sampling error, misdiagnosis, inter-operational variability, cost and maneuver-related risk of mortality and morbidity. Serum aminotransferase and imaging techniques (ultrasound, CT, MRI) offered inadequate reliability in assessing NASH and liver fibrosis in NAFLD patients. Thus, clinical and biochemical parameters that would correlate with liver biopsy have to be searched for aiming at identifying non-invasive test(s) that may aid not only in the diagnosis of NAFLD but also for early prediction of NASH and monitoring disease progression and response to therapy.

Liver fibrosis is the only histological feature associated with long-term outcome of patients with NAFLD. So, diagnosis of liver fibrosis is the most important issue in clinical practice. Several noninvasive serologic markers have been developed to predict NASH and determine the degree of liver fibrosis. However, serum tests of aminotransferases, GGT, AST/ALT ratio, components of extracellular matrix (hyaluronic acid; HA) are not helpful or may have little or modest diagnostic/predictive ability in identifying the presence of liver fibrosis.

Multiple serum cytokines, TNF-α, IL-6, hsCRP, Adiponectin, resistin, Pentraxin-3 had been evaluated for predicting NASH yet with variable and inconsistent results. Several complex models and scores had been created with variable predictive ability in NASH, the majorities of such models and scores had not been externally validated and were obtained from certain populations, mostly morbidly obese patients.

On the other hand, several models and scores had been developed predicting NASH and liver fibrosis with reasonable diagnostic performance. Of the proposed models with consistent results; the BAAT score which incorporated BMI, Age, ALT, TG and yielded 100% sensitivity but not specific, NAFLD fibrosis score (NFS) which incorporated Age, DM, BMI, platelet count, albumin and AST/ALT ratio and has a great diagnostic accuracy for advanced fibrosis stages and BARD score incorporating BMI, AST/ALT Ratio and DM yet it had lower accuracy and worse performance than NFS.

Among the several investigated noninvasive biomarkers are the cytokeratin-18fragments (CK18-Fs) with consistent results in differentiating NASH from NAFLD. The CK18-Fs are marker of hepatocyte apoptosis and necrosis. The CK-18Fs are produced mainly by caspase3 and played crucial roles in the pathogenesis of chronic liver disease and NAFLD.

Fibrosis-4 index (FIB-4) has been developed mainly to predict liver fibrosis in viral hepatitis. Some studies investigated FIB-4 in patients with NAFLD. Some authors established new scoring systems as significant predictors of NASH in NAFLD patients such as FIC-22 score combining FIB-4 and CK18-Fs.

Despite the simplicity of different fibrosis score(s) based on clinical and biochemical laboratory results such as BAAT, NFS, BARD, FIB-4 and FIC-22, their diagnostic performance seems inadequate in accurate discrimination of disease stages. Moreover, the use of any of these tests or scores alone is likely not sufficient to rule in or rule out significant disease. Therefore, other non-invasive model(s) with robust analytical and clinical validation for detecting NASH and liver fibrosis are mandatory.

The aim of this study was to assess the reliability of a new scoring system (FICK-3) combining FIB-4 index, Insulin resistance (HOMA-IR) and Cytokeratin-18-fragments (CK-18Fs) as a predictor of NASH development and/or progression to liver fibrosis in Egyptian patients with NAFLD.
MATERIALS AND METHODS

Materials: A total of 150 adult consecutive outpatients older than 18 years with ultrasonographic evidence of fatty liver were conveniently selected from outpatient’s obesity clinics of Mansoura University Hospitals after a written informed conscious consent during the period from May, 2017 through 2019. The study was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice Guidelines and was approved by the Institutional Review Board of Mansoura Faculty of Medicine (MFM-IRB; R.19.06.536).

Inclusion criteria: Patients with ultrasonographic evidence of fatty liver and/or anthropometric and laboratory evidence of MetS according to the International Diabetes Federation (IDF) metabolic syndrome world wide definition: [at least three of the following: Waist circumference (≥94 cm in men and ≥80 cm in women), hypertension (systolic bp ≥135 mmHg and diastolic bp ≥ 85 mmHg), diabetes (FBS ≥100 mg dL⁻¹), hyperlipidemia (TG ≥150 mg dL⁻¹, LDL-C 40 mg dL⁻¹ in men and 50 mg dL⁻¹ in women)]¹⁸.

Exclusion criteria: Presence of possible secondary causes of fatty liver disease; alcohol abuse, intestinal malabsorption, drug-or toxin-induced liver steatosis (e.g., amiodarone, tamoxifen, glucocorticoids, synthetic estrogens, diltiazem, aspirin, methotrexate, highly active antiretroviral therapy), viral hepatitis and autoimmune liver diseases. Presence of contraindication to do liver biopsy; coagulopathy, decompensated chronic liver disease, patients refused to sign consent, low platelet count <50000 mm⁻³, INR >1.5. Others; advanced chronic or psychiatric illness, extremes of age (Age <18 year and age >70 year), renal impairment, pregnancy, endocardial and cardiopulmonary diseases, cardiac failure (ejection fraction <50%).

Methods: Initially, all participants were submitted to thorough history taking with special emphasis on age, lifestyle and symptoms suggestive of liver disease (fatigue, malaise, jaundice, abdominal discomfort, hepatosplenomegaly).

Anthropometric measurements: Body weight, standing height and waist circumference (WC) were measured to the nearest 0.1 kg, 0.1 cm, 0.1 cm, respectively. The BMI (kg m⁻²) calculated as body weight in kilograms divided by the square of their height in meters. The patient was considered non-obese when the BMI <25 kg m⁻², Overweight when BMI 25-29.9, class I mild obese when BMI 30-34.9 class II moderate obese when BMI 35-39.9 and class III morbid obese when BMI >40 kg m⁻².

Laboratory biochemical evaluations: Venous blood samples were obtained from each subject after a minimum of 10 h of fasting. Serum tubes were centrifuged at 3000 for 15 min at 4 °C, aliquoted and stored at -70 °C until assayed. White blood cell (WBC) count, hemoglobin, hematocrit, platelet count, serum albumin, bilirubin, serum transaminases, gamma glutamyltransferase (GGT), alkaline phosphatase (ALP), erythrocyte sedimentation rate (ESR) and high sensitive C-reactive protein (hs-CRP) were tested. Prothrombin time, INR, serum creatinine, ANA, serum iron, viral serological markers, fasting and 2 h postprandial blood glucose, serum total cholesterol, triglycerides and high-density lipoprotein-cholesterol (HDL-c) were determined by enzymatic methods with a chemistry analyzer (Hitachi747, Japan, Tokyo).

Cytokinin 18-fragments (CK18-Fs) levels were measured using a human CK-18-Fs/KRT18 (Cytokerin 18) ELISA Kit (catalog number: E-EL-H2072) following the manufacturer’s guidelines (Elabscience, Houston, Texas, USA). Detection range: 6.25-400 mIU mL⁻¹). The intra-assay and inter-assay coefficient variation were 6.05 and 5.75%, respectively, sensitivity 3.75 mIU mL⁻¹. The FIB-4 index was calculated to all patients using the following equation¹⁹:

\[
FIB-4 = \frac{\text{Age (year)} \times \text{AST (U L}^{-1})}{\text{Platelets (10}^9 \text{ L}^{-1}) \times \sqrt{\text{ALT (U L}^{-1})}}
\]

The aspartate aminotransferase (AST)-to-platelet ratio index (APRI) was calculated using this equation²⁰:

\[
\text{APRI} = \left[ \frac{\text{AST (U L}^{-1})}{\text{AST (upper limit of normal (U L}^{-1})}} \right] \times \frac{\text{Platelets (10}^9 \text{ L}^{-1})}{100}
\]

The plasma insulin was measured by immunoradiometric assay (INS-IRMA; Catalog# KIP1251, DiAsource ImmunoAssays S.A. - Rue du Bosquet, 2-B-1348 Louvain-la-Neuve - Belgium). The minimum detectable concentration was 1 mU mL⁻¹ and the inter-assay and intra-assay coefficients of variation (CVs) were 6.3 and 2.4%, respectively. The insulin sensitivity and/or resistance were calculated with the homeostasis model assessment (HOMA-IR) for insulin resistance²¹:

\[
\text{HOMA-IR} = \frac{\text{Fasting glucose (mg dL}^{-1}) \times \text{Fasting insulin (µU mL}^{-1})}{405}
\]
The FICK-3 score was calculated as the unweighted sum of the points obtained from the following three variables; 
FIB-4 >1.45, HOMA-IR≥2.11 and Cytokeratin 18-fragment 
CK18-Fs≥307U mL⁻¹; one point for each variable.

**Histopathological examination of liver biopsy specimens:**
Ultrasound-guided percutaneous tru-cut needle liver biopsy 
was performed in selected patients after consent at 
the seventh or eighth intercostal space in the midaxillary line. 
A core of liver tissue was obtained while the patient holds his 
breath in expiration. A sample about 1.5 cm long and 1-2 mm wide 
is considered adequate for histopathological assessment. 
The sample was preserved in 10% formalin. The biopsy 
specimens taken were embedded in Paraffin blocks, sectioned 
and stained with hematoxylin and eosin for conventional 
histopathological examination and Masson’s trichrome for 
assessment of liver fibrosis. The specimens were analyzed by 
two independent well-trained histopathologists, who were 
blinded to the diagnosis, clinical data, the treatment and 
subject group identity. Diagnosis and staging of NASH were 
based upon the previously described histological criteria⁶. 
Types of NAFLD may be NAFL (Type1: Fat accumulation only, 
Type 2: Fat accumulation and lobular inflammation) or NASH 
(Type 3: Fat accumulation and ballooning degeneration, 
Type 4: Fat accumulation, ballooning degeneration and either 
Mallory-Denk bodies or fibrosis). Stages of fibrosis may be 
(F0: normal connective tissue or no fibrosis, F1: zone3 
pericellular/perivenular fibrosis (a,b,c: mild, moderate, severe), 
F2: Zone 3 perisinusoidal, pericellular or periportal fibrosis, 
F3: Bridging or septal fibrosis, F4: Cirrhosis).

The NAFLD activity score (NAS) was developed to 
provide a numerical score for patients who most likely have 
NASH. Accordingly, NAS is calculated as the unweighted 
sum of the separate scores for steatosis (0-3 points), 
lobular inflammation (0-3 points) and hepatocellular 
ballooning (0-2 points). Definitive-NASH is described when 
NAS ≥5, while the Non-NASH score is NAS <3. The NAS of (3-4) 
points is defined as borderline-NASH. Fibrotic NASH is 
defined as NAS ≥4 and fibrosis stage ≥F2. This system was 
developed mainly to evaluate the disease activity and 
monitor the histological response with treatment, rather than 
NASH diagnosis².

**Statistical analysis:** Statistical analysis was carried out 
using IBM-SPSS software version 21.0 (SPSS Inc, Chicago, IL). 
The normality of quantitative data was tested using 
Kolmogorov-Smirnov and Shapiro-Wilk’s test with data 
being normally distributed if p>0.05. Qualitative data were 
expressed as number and percentage. Quantitative data were 
expressed as mean±standard deviation (SD) if normally 
distributed or median and interquartile range (IQR) if not. 
Chi-Square test, Fisher’s exact test and Chi-Square test with 
Bonferroni adjustment were used for comparison when 
indicated. Independent-Samples t-test was used if quantitative 
data were normally distributed and the non-parametric alternative 
Mann-Whitney U test was used if not. Pearson’s or Spearman’s correlations 
were used for correlations between two normally or two abnormally 
distributed scales, respectively. Multivariate logistic regression 
analysis was used to detect the significant “independent” 
predictors of NASH with their OR (95% CI). Receiver operating 
characteristic (ROC) curve analysis was used to detect the 
diagnostic accuracy, predictive performance and a diagnostic 
cutoff value.

**RESULTS**

In this study, 150 patients with NAFLD (diagnosed by 
the presence of fatty liver on US) with a mean age of 
47.12±3.8 years were recruited and evaluated by laboratory, 
biochemical and histopathological methods for the presence or 
absence of NASH. Only 100 patients agreed, signed a 
written consent and fit the criteria to do the diagnostic liver 
biopsy. Histologically, 66 out of 100 patients (66%) were 
diagnosed as NASH and the remainders were diagnosed as 
simple steatosis or NAFL.

The demographic data, clinical and biochemical scores, 
the histopathological features in all studied patients were 
demonstrated in Table 1. The frequency of NASH among 
patients with NAFLD was significantly higher than the 
frequency of simple steatosis (NAFL) (p = 0.041). Also, female 
patients had statistical significant higher frequency of NASH 
than male patients. Patients with NASH had statistical 
significant higher values than NAFL patients as regards BMI, 
GGT, transaminases, hsCRP, MetS, glycated hemoglobin, 
platelet count, fasting insulin, HOMA-IR and 
Cytokeratin 18-fragments (p≤0.05 in all). Moreover, NASH 
patients had statistical significant higher values of FIB-4 index 
and APRI score (p≤0.01). However, No significant difference 
was noticed between the two groups as regards hemoglobin, 
s. bilirubin, fasting glucose (p = 0.05). Indeed, NASH patients 
had significantly higher frequency of liver fibrosis and 
significantly higher values of NAS and biochemical scores 
(FICK-3, FIC-22 and MACK-3) than NAFL patients (p = 0.001).
Table 1: Demographic data, clinical, biochemical scores and histopathological features in all studied groups

<table>
<thead>
<tr>
<th>Parameters</th>
<th>All biopsied patients (100)</th>
<th>NASH (66)</th>
<th>NAFL (34)</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Frequency</td>
<td>100</td>
<td>66 (66%)</td>
<td>34.0 (34%)</td>
<td>0.041</td>
</tr>
<tr>
<td>Age (years): Mean±SD</td>
<td>47.12±3.8</td>
<td>46.31±4.9</td>
<td>39.89±3.1</td>
<td>0.047</td>
</tr>
<tr>
<td>BMI (kg m⁻²)</td>
<td>30.8±4.02</td>
<td>32.33±4.22</td>
<td>28.±3.14</td>
<td>0.015</td>
</tr>
<tr>
<td>Gender (female/male)</td>
<td>62/38</td>
<td>43/23</td>
<td>19/15</td>
<td>0.027</td>
</tr>
<tr>
<td>Hemoglobin (g dl⁻¹)</td>
<td>13.12±2.66</td>
<td>12.41±2.34</td>
<td>13.21±1.65</td>
<td>0.289</td>
</tr>
<tr>
<td>Platelets (10³/mm³)</td>
<td>230.3±49.4</td>
<td>220±46.6</td>
<td>254±51.3</td>
<td>0.041</td>
</tr>
<tr>
<td>AST (IU L⁻¹)</td>
<td>69±29.5</td>
<td>78.1±25.4</td>
<td>61±33.2</td>
<td>0.017</td>
</tr>
<tr>
<td>Albumin (g dl⁻¹)</td>
<td>4.01±0.4</td>
<td>3.23±.03</td>
<td>3.6±0.2</td>
<td>0.041</td>
</tr>
<tr>
<td>Bilirubin (mg dl⁻¹)</td>
<td>0.7±0.25</td>
<td>1.3±.08</td>
<td>0.99±0.4</td>
<td>0.098</td>
</tr>
<tr>
<td>ALT (IU L⁻¹)</td>
<td>75.53±22.3</td>
<td>85.98±15.2</td>
<td>67.53±31.5</td>
<td>0.014</td>
</tr>
<tr>
<td>GGT (U L⁻¹)</td>
<td>68.8±7.4</td>
<td>72.7±8.3</td>
<td>46.8±2.6</td>
<td>0.015</td>
</tr>
<tr>
<td>hsCRP</td>
<td>61.8±7.4</td>
<td>64.7±8.3</td>
<td>56.8±2.6</td>
<td>0.013</td>
</tr>
<tr>
<td>MetS (yes/no)</td>
<td>59/41</td>
<td>39/27</td>
<td>20/14</td>
<td>0.036</td>
</tr>
<tr>
<td>Triglycerides (mg dl⁻¹)</td>
<td>125±26.4</td>
<td>142±34.2</td>
<td>111±24.1</td>
<td>0.039</td>
</tr>
<tr>
<td>Glycated hemoglobin (%)</td>
<td>6.71±1.4</td>
<td>7.51±1.6</td>
<td>6.1±1.5</td>
<td>0.035</td>
</tr>
<tr>
<td>Fasting blood sugar (mg dl⁻¹)</td>
<td>131±26.4</td>
<td>142±28.3</td>
<td>109±24.2</td>
<td>0.090</td>
</tr>
<tr>
<td>Fasting insulin (mU ml⁻¹)</td>
<td>13.57±23</td>
<td>17.3±3.1</td>
<td>11.2±2.4</td>
<td>0.017</td>
</tr>
<tr>
<td>NAFLD activity score (NAS)</td>
<td>3.71±1.3</td>
<td>4.51±1.5</td>
<td>2.8±1.6</td>
<td>0.028</td>
</tr>
<tr>
<td>Type of NAFLD (type1/2/3/4)</td>
<td>21/40/30/9</td>
<td>9/18/30/9</td>
<td>12/22/0</td>
<td>0.013</td>
</tr>
<tr>
<td>Fibrosis stage (F0/1/2/3/4)</td>
<td>28/26/29/10/7</td>
<td>14/19/16/10/7</td>
<td>14/7/3/0/0</td>
<td>0.001</td>
</tr>
<tr>
<td>APRI score</td>
<td>0.74±0.17</td>
<td>0.98±0.15</td>
<td>0.64±0.04</td>
<td>0.009</td>
</tr>
<tr>
<td>FIB-4 index</td>
<td>1.63±0.36</td>
<td>1.86±0.26</td>
<td>1.27±0.12</td>
<td>0.002</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>4.1±1.2</td>
<td>4.8±1.3</td>
<td>2.6±1.03</td>
<td>0.014</td>
</tr>
<tr>
<td>Cytokeratin 18-Fs (U L⁻¹)</td>
<td>596±205.5</td>
<td>831.1±123.4</td>
<td>445.5±150.2</td>
<td>0.001</td>
</tr>
<tr>
<td>FICK-3 score (0/1/2/3)</td>
<td>15/33/40/12</td>
<td>6/8/40/12</td>
<td>9/25/0/0</td>
<td>-</td>
</tr>
<tr>
<td>MACK-3 score</td>
<td>1.61±1.1</td>
<td>1.97±1.16</td>
<td>1.47±0.92</td>
<td>0.0001</td>
</tr>
<tr>
<td>FIC-22 score</td>
<td>0.54±0.41</td>
<td>0.71±0.26</td>
<td>0.29±0.14</td>
<td>0.000</td>
</tr>
<tr>
<td></td>
<td>0.94±0.46</td>
<td>1.16±0.69</td>
<td>0.86±0.35</td>
<td>0.000</td>
</tr>
</tbody>
</table>

Data were presented as Mean±Standard deviation (M±SD), NASH: Nonalcoholic steatohepatitis, NAFL: Nonalcoholic fatty liver, P: Probability value comparing NASH and NAFL, BMI: Body mass index, hsCRP: High sensitive C-reactive protein, MetS: Metabolic syndrome, ALT: Alanine transaminase, AST: Aspartate transaminase, GGT: Gamma glutamyl transferase, HOMA-IR: Homeostasis model assessment for insulin resistance, APRI: Aspartate aminotransferase-to-platelet ratio, CK18-Fs: Cytokeratin 18-fragments, FIB-4: Fibrosis-4, FICK-3 score: Sum of FIB-4, HOMA-IR and Cytokeratin 18-Fs, MACK-3: homa, Ast and Cytokeratin-18, FIC-22: FIB-4 and Cytokeratin-18, P: Probability value, Significance at p<0.05

Table 2: Diagnostic accuracies of different parameters predicting nonalcoholic steatohepatitis (NASH) in patients with nonalcoholic fatty liver disease (NAFLD)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>AUC</th>
<th>95% CI</th>
<th>p-value</th>
<th>SE</th>
<th>Cutoff values</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Accuracy (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>FIB-4 index</td>
<td>0.739</td>
<td>0.596-0.882</td>
<td>0.005</td>
<td>0.072</td>
<td>1.45</td>
<td>90.9</td>
<td>89.5</td>
<td>92.2</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>0.700</td>
<td>0.533-0.838</td>
<td>0.039</td>
<td>0.077</td>
<td>2.11</td>
<td>85.1</td>
<td>76.9</td>
<td>79.2</td>
</tr>
<tr>
<td>CK18-Fs (U L⁻¹)</td>
<td>0.768</td>
<td>0.613-0.924</td>
<td>0.001</td>
<td>0.079</td>
<td>307</td>
<td>89.7</td>
<td>87.4</td>
<td>90.7</td>
</tr>
<tr>
<td>FICK-3</td>
<td>0.817</td>
<td>0.696-0.939</td>
<td>0.000</td>
<td>0.062</td>
<td>1.00</td>
<td>95.6</td>
<td>91.1</td>
<td>94.6</td>
</tr>
</tbody>
</table>

95%CI: 95% confidence interval, P: Probability value and significance at p<0.05, SE: Standard error, AUC: Area under the curve, HOMA-IR: Homeostasis model assessment for insulin resistance, CK18-Fs: Cytokeratin 18-fragments, FIB-4: Fibrosis-4, FICK-3 score: Sum of FIB-4, HOMA-IR and Cytokeratin 18-Fs

The diagnostic performances and accuracies of the different parameters predicting the presence or absence of NASH in patients with NAFLD were demonstrated in Table 2. FIB-4 index, HOMA-IR, CK18-Fs and the new combination score FICK-3 were significant predictors of NASH in patients with NAFLD (AUC 95%CI: 0.739 (0.596-0.882), 0.700 (0.533-0.838), 0.768 (0.613-0.924), 0.817 (0.696-0.939), respectively; p=0.05). In the combined FICK-3 score (at a cutoff value of 1 point) the AUC 0.8 denoted great significant predictive value of NASH and had the best diagnostic accuracy (94.6%). Also, the sensitivity, specificity and diagnostic accuracy were highly significant in FIB-4 (90.9, 89.5 and 92.2%, respectively), HOMA-IR (85.1, 76.9 and 79.2%), CK18-Fs (89.7, 87.4 and 90.7%, respectively) and FICK-3 score (95.6, 91.1 and 94.6%, respectively), p<0.05 for all. Areas under the curve (AUC) values for these risk factors were ≥0.7, displaying statistically significant predictive values (p<0.05).

Table 3 demonstrated the AUROC values of different parameters that predicted the presence/absence of liver fibrosis and its grade in patients with NASH. The FIB-4 index, HOMA-IR, CK18-Fs and FICK-3 score had statistically significant predictive values for the presence of early-stage liver fibrosis (F3-2) in NASH patients with NAFLD (AUC 95% CI: 0.765 (0.621-0.968), 0.700 (0.532-0.893), 0.768 (0.613-0.924), 0.817 (0.696-0.939), respectively; p<0.05).
Table 3: Diagnostic accuracies of different parameters in predicting liver fibrosis (LF) in patients with nonalcoholic steatohepatitis (NASH)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>AUC</th>
<th>95% CI</th>
<th>p-value</th>
<th>SE</th>
<th>Cutoff values</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Accuracy (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>F1-2</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FIB-4 index</td>
<td>0.765</td>
<td>0.621-0.968</td>
<td>0.006</td>
<td>0.073</td>
<td>≥1.45</td>
<td>92.90</td>
<td>91.10</td>
<td>92.00</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>0.700</td>
<td>0.532-0.893</td>
<td>0.033</td>
<td>0.078</td>
<td>≥2.11</td>
<td>63.10</td>
<td>71.40</td>
<td>72.20</td>
</tr>
<tr>
<td>CK18-Fs (U L⁻¹)</td>
<td>0.803</td>
<td>0.650-0.956</td>
<td>0.002</td>
<td>0.079</td>
<td>≥307</td>
<td>90.60</td>
<td>82.30</td>
<td>87.60</td>
</tr>
<tr>
<td>FICT-3</td>
<td>0.835</td>
<td>0.561-0.903</td>
<td>0.000</td>
<td>0.062</td>
<td>≥1.00</td>
<td>92.60</td>
<td>88.40</td>
<td>95.410</td>
</tr>
<tr>
<td><strong>F3-4</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FIB-4 index</td>
<td>0.782</td>
<td>0.645-0.920</td>
<td>0.003</td>
<td>0.070</td>
<td>≥1.45</td>
<td>93.40</td>
<td>87.20</td>
<td>89.10</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>0.706</td>
<td>0.543-0.869</td>
<td>0.028</td>
<td>0.083</td>
<td>≥2.11</td>
<td>83.10</td>
<td>77.80</td>
<td>81.40</td>
</tr>
<tr>
<td>CK18-Fs (U L⁻¹)</td>
<td>0.825</td>
<td>0.677-0.972</td>
<td>0.001</td>
<td>0.075</td>
<td>≥307</td>
<td>91.20</td>
<td>84.70</td>
<td>87.30</td>
</tr>
<tr>
<td>FICT-3</td>
<td>0.907</td>
<td>0.822-0.991</td>
<td>0.000</td>
<td>0.043</td>
<td>≥1.00</td>
<td>94.10</td>
<td>91.10</td>
<td>96.77</td>
</tr>
<tr>
<td><strong>F2-4</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>FICT-3</td>
<td>0.954</td>
<td>0.917-0.992</td>
<td>0.000</td>
<td>0.019</td>
<td>≥1.00</td>
<td>94.74</td>
<td>96.77</td>
<td>95.65</td>
</tr>
<tr>
<td>MACK-3</td>
<td>0.837</td>
<td>0.758-0.915</td>
<td>0.000</td>
<td>0.040</td>
<td>≥0.134</td>
<td>94.32</td>
<td>92.06</td>
<td>93.38</td>
</tr>
<tr>
<td>FICT-22</td>
<td>0.803</td>
<td>0.719-0.887</td>
<td>0.000</td>
<td>0.043</td>
<td>≥1.00</td>
<td>91.40</td>
<td>89.81</td>
<td>94.70</td>
</tr>
</tbody>
</table>

Table 4: Univariate and multivariate analysis for the independent predicting factors of nonalcoholic steatohepatitis in patients with nonalcoholic fatty liver disease (NAFLD)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Cutoff values</th>
<th>p-value</th>
<th>OR (95% CI)</th>
<th>p-value</th>
<th>OR (95% CI)</th>
</tr>
</thead>
<tbody>
<tr>
<td>BMI (kg m⁻²)</td>
<td>≥27.4</td>
<td>0.016</td>
<td>1.56 (1.05-2.91)</td>
<td>0.161</td>
<td>2.39 (1.92-3.78)</td>
</tr>
<tr>
<td>Albumin (g dl⁻¹)</td>
<td>≥4.5</td>
<td>0.040</td>
<td>1.03 (1.01-1.55)</td>
<td>0.294</td>
<td>1.47 (0.981-2.82)</td>
</tr>
<tr>
<td>hsCRP</td>
<td>≥54.0</td>
<td>0.033</td>
<td>1.20 (0.87-1.86)</td>
<td>0.325</td>
<td>1.19 (0.79-2.89)</td>
</tr>
<tr>
<td>GGT (U L⁻¹)</td>
<td>≥57.0</td>
<td>0.025</td>
<td>0.89 (0.45-1.83)</td>
<td>0.512</td>
<td>1.84 (0.950-2.66)</td>
</tr>
<tr>
<td>FIB-4 index</td>
<td>T.45</td>
<td>0.001</td>
<td>0.68 (1.10-1.88)</td>
<td>&lt;0.001</td>
<td>2.26 (1.19-4.63)</td>
</tr>
<tr>
<td>HOMA-IR</td>
<td>≥2.11</td>
<td>0.01</td>
<td>2.38 (1.94-4.12)</td>
<td>&lt;0.001</td>
<td>1.47 (1.01-4.50)</td>
</tr>
<tr>
<td>CK18-Fs (U L⁻¹)</td>
<td>≥307</td>
<td>0.001</td>
<td>1.46 (0.76-2.55)</td>
<td>&lt;0.001</td>
<td>1.98 (0.97-3.33)</td>
</tr>
<tr>
<td>FICT-3 score</td>
<td>≥1.0</td>
<td>0.001</td>
<td>2.17 (1.55-4.04)</td>
<td>&lt;0.001</td>
<td>1.77 (1.12-3.47)</td>
</tr>
</tbody>
</table>

95%CI: 95% confidence interval, P: Probability value and significance at p<0.05, BMI: Body mass index, hsCRP: High sensitive C-reactive protein, GGT: Gamma glutamyl transferase, HOMA-IR: Homeostasis model assessment for insulin resistance, CK18-Fs: Cytokeratin 18-fragments, FIB-4: Fibrosis-4, FICT-3: score: Sum of FIB-4, HOMA-IR and Cytokeratin 18-Fs

Table 5: Spearman’s rho correlation between FICT-3 score and histological features and fibrosis grade

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Correlation coefficient (r)</th>
<th>Sig. (2-tailed) (p)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Steatosis</td>
<td>0.990</td>
<td>0.0001</td>
</tr>
<tr>
<td>Lobular inflam.</td>
<td>0.855</td>
<td>0.0001</td>
</tr>
<tr>
<td>Ballooning</td>
<td>0.666</td>
<td>0.0001</td>
</tr>
<tr>
<td>Fibrosis (F1-2)</td>
<td>0.867</td>
<td>0.0001</td>
</tr>
<tr>
<td>Fibrosis (F3-4)</td>
<td>0.683</td>
<td>0.0001</td>
</tr>
</tbody>
</table>

0.803 (0.650-0.956), 0.835 (0.561-0.903), respectively; p = 0.05]. Moreover, their diagnostic accuracies in detecting the advanced-stages of liver fibrosis (F3-4) increased significantly especially in FICT-3 score (AUC 95%CI: 0.907 (0.822-0.991, p = 0.001). Contrasted with MACK-3 and FICT-22 scores, the FICT-3 score provided the highest significant diagnostic performance in discriminating cases with fibrotic NASH as defined by NAS ≥4 and F ≥2 (AUC 95%CI: 0.954 (0.917-0.992, p = 0.001).

Multivariate analysis were performed using the significant covariates in univariate analysis and displayed in Table 4. At specific cutoff values obtained from the ROC curve analyses, FIB-4 index (≥1.46), HOMA-IR (≥2.11), CK18-Fs (≥307 U L⁻¹) and the combined FICT-3 score (≥1 point) demonstrated high statistical significant predictions for NASH and liver fibrosis in patients with NAFLD (p<0.001). The correlations between FICT-3 score and histological characteristics were elucidated in Table 5. The FICT-3 score showed highly significant positive correlations with the histological features and fibrosis grade (p = 0.0001).

The receiver operating characteristics (ROC) curves for FIB-4 index, HOMA-IR, CK18-Fs and FICT-3 score as predictors of NASH in patients with NAFLD were demonstrated in Fig. 1. The FICT-3 score had the highest significant diagnostic performance for the prediction of NASH (AUC 0.817, p<0.001). In Fig. 2, the combined FICT-3 score significantly predicted early-stage liver fibrosis (F1-2) compared with individual variables (AUC 0.835, p<0.001). As shown in Fig. 3, the FICT-3 score also demonstrated highly significant diagnostic value for detection of advanced-stage liver fibrosis (F3-4) compared with other parameters (AUC 0.907, p<0.001). While in Fig. 4, FICT-3 score had the best
Fig. 1: Receiver operating characteristics (ROC) Curves for fibrosis-4 index (FIB-4), HOMA-IR, cytokeratin 18-fragments (CK18-Fs) and FICK-3 score as predictors of non alcoholic steatohepatitis (NASH) in patients with NAFLD (NASH without liver fibrosis).

Fig. 2: Receiver operating characteristics (ROC) curves for fibrosis-4 index (FIB-4), HOMA-IR, cytokeratin 18-fragments (CK18-Fs) and FICK-3 score in the detection of NASH with early-stage liver fibrosis (F1-2) in patients with NAFLD (Early-stage fibrosis, F1-2).

Fig. 3: Receiver operating characteristics (ROC) curves for fibrosis-4 index (FIB-4), HOMA-IR, cytokeratin 18-fragments (CK18-Fs) and FICK-3 score in the detection of NASH with advanced-stage liver fibrosis (F3-4) in patients with NAFLD (Advanced-stage fibrosis, F3-4).

Fig. 4: Receiver operating characteristics (ROC) curves comparing the FICK-3 score, MACK-3 score and FIC-22 score in diagnosis of fibrotic NASH in patients with NAFLD (NAS≥4, F≥2).

**DISCUSSION**

As specific markers discriminating NASH in NAFLD patients are lacking, the authors in this study investigated the predictive value of other noninvasive markers and scores.
like FIB-4 index, HOMA-IR, CK18-Fs (either individually or combined in new scoring system; FICK-3) and correlated them with the histopathological characteristics. Also, there is no available data sufficiently assessing liver fibrosis noninvasively in NAFLD patients. Therefore, in this study, the reliabilities of FIB-4 index, HOMA-IR, CK-18Fs and FICK-3 were investigated not only in prediction of NASH but also in the determination of the absence or presence of liver fibrosis (F0, F1-4) in NAFLD patients.

In the current study, it was found that NASH was present in (66%) of Egyptian patients with NAFLD. In Egypt, the prevalence of NAFLD was reported to range from 10-18% with no reports for prevalence of NASH among Egyptian population. This relatively high percentage may be explained by increasing the epidemic of obesity and westernization of the life style including junk food and sedentary behavior.

The relations between the clinical, biochemical parameters and NAFLD were extensively analyzed in several studies. Some authors found that MetS was an independent predictor for NASH and its individual histological features especially ballooning and fibrosis. Hepatic enzymes (ALT, AST), triglycerides were reported to be increased significantly in NAFLD. However, NASH can’t be excluded in those with normal aminotransferases and triglycerides. DM had a higher prevalence among patients with NASH. A previous study developed a simple model with significant accuracy for predicting NASH incorporating AST and diabetes. Another study demonstrated an association of GGT with increased mortality in NASH. Moreover, the majority of previous studies demonstrated that serum tests of aminotransferases, GGT, AST/ALT ratio, components of extracellular matrix (hyaluronic acid; HA) are not helpful or may have little or modest diagnostic/predictive ability in identifying the presence of liver fibrosis. In the current study, no significant predictive values were demonstrated for serum aminotransferases, GGT, AST/ALT ratio in discriminating liver fibrosis in NASH patients.

Of the extensively investigated tests and models with consistent results were Nice model incorporating (CK18-Fs, ALT and MetS), Apoptosis Panel incorporating (CK18-Fs, soluble Fas and Fas ligand). The Fibrosis-4 index and the AST-to-Platelet Ratio Index (APRI) score are validated noninvasive serologic measure for liver fibrosis based on routine laboratory data (platelets, ALT, AST and age). These scores are promising and were able to reliably determine mild to advanced fibrosis (F0-F4). This study demonstrated a great significant predictive and diagnostic ability of FIB-4 index (accuracy > 92%, AUC > 0.739, p < 0.01) for NASH as well as for liver fibrosis regardless its stage.

The initial studies of CK-18Fs in the diagnosing NASH were promising yet the subsequent studies were disappointing. Cusi et al. demonstrated that CK-18 when used alone had a limited diagnostic accuracy in distinguishing NASH from NAFLD or in determining the disease severity. They suggested combining CK-18 with other biological markers to achieve an accurate discrimination of NASH and liver fibrosis. Polyzos et al. demonstrated that the CHA index with the following variables (CK-18, HOMA and AST) was significantly higher in NASH compared to NAFL or control groups and was significantly correlated with the severity of the histological lesions. Boursier et al. proposed a new score MACK-3 combining the same variables in CHA index (HOMA-IR, AST and CK-18Fs) as a promising biomarker for diagnosing fibrotic NASH. The MACK-3 score was shown to be superior to other scores such as BARD score, NFS, FIB-4. They proposed MACK3 score, the algorithm of which is available as an online calculator, as a screening test for the fibrotic NASH with excellent diagnostic accuracy. Nevertheless, Chuah et al. found that MACK-3 was superior to NFS and FIB-4 only in diagnosing the active component of fibrotic NASH. In advanced fibrosis stage ≥F2, MACK-3 was similar to NFS and FIB-4 in diagnosing fibrotic NASH. The CHA index and MACK-3 score included the same parameters which are considered circulating reflectors of insulin resistance, hepatocellular dysfunction and apoptosis contributing to the pathogenesis of NAFLD. Higher ALT rather than AST is typical for NAFLD yet higher AST is more indicative of advanced liver disease. In this study, the replacement of AST by FIB-4 index in the current FICK-3 score significantly improved its diagnostic accuracy in predicting both early and advanced stages of fibrosis in NASH.

The results in this study showed not only significantly increased CK18-Fs levels in NASH, but also close correlations with the histological features like steatosis, ballooning, lobular inflammation and fibrosis stage in patients with NAFLD. The results in the present study evidenced that the CK18-Fs could serve as a reliable surrogate biomarker predicting not only NASH development, severity and progression but also the presence or absence of liver fibrosis in NAFLD patients. It is the only validated biomarker with the most consistent results differentiating steatosis from NASH.

Interestingly, the newly proposed FICK-3 score combining FIB-4, HOMA-IR and CK-18Fs in this study showed better diagnostic accuracy with higher prediction of NASH and liver fibrosis. Also, FICK-3 showed significant close correlations with the histological features like steatosis, ballooning, lobular inflammation and fibrosis stage in patients with NAFLD.
We compared the FICK-3 score proposed in this study with the previously proposed scores (such as FIC-22 and MACK-3 score) in the determination of fibrotic NASH. The FICK-3 score had the highest AUC95%CI, sensitivity and specificity denoting the best diagnostic performance in discrimination of advanced fibrosis in patients with NASH. The FICK-3 score had higher diagnostic ability for detecting early-stage liver fibrosis (F1-2) contrasted with FIC-22 score. Moreover, this study incorporated larger numbers of NAFLD patients with higher percentage of histopathologically evidenced NASH (66%). The results of FICK-3 score obtained in this work could ascertain the presence of NASH and/or liver fibrosis in NAFLD patients.

CONCLUSION AND RECOMMENDATIONS

In Egyptian patients with NAFLD, FIB-4 index, HOMA-IR and CK18-Fs were the most significant independent noninvasive predictors for NASH development and the presence or absence of liver fibrosis. The proposed FICK-3 score had reliable and acceptable diagnostic performance in discriminating NASH and liver fibrosis in NAFLD Egyptian patients. These observations could minimize the use of liver biopsy in diagnosis of NASH. Being a new score combining three important variables with greater diagnostic accuracy reflecting hepatic dysfunction, insulin resistance and apoptosis, it would offer adequate nucleus for a further validation study aiming to construct a screening test for fibrotic NASH.

Further large scale, multicenteric and validation cohort studies are advised to validate this score as a screening test for NASH and liver fibrosis in patients with NAFLD.

SIGNIFICANCE STATEMENT

This study elucidated that FIB-4 index, HOMA-IR and CK18-Fs were reliable significant independent predictors for NASH development and progression in NAFLD Egyptian patients. Moreover, the new combined FICK3 score (the sum of HOMA-IR ≥2.11, CK18-Fs ≥307 U L⁻¹ and FIB-4 index ≥1.46; one point for each variable) had significantly higher diagnostic accuracy for the prediction of early-as well as late-stage fibrotic NASH.

REFERENCES


