Toxicity of *Anagallis arvensis* Plant

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**Abstract:** Sixty adult rats of both sexes were used for determination of LD\(_{50}\) of *Anagallis arvensis*. Other eighteen rats were used for repeated successive dose for determination of blood picture, kidney function and histopathological changes associated with *A. arvensis* toxicity (1/5 and 1/10 LD\(_{50}\)) IP for 15 days. The LD\(_{50}\) was 10.718 mg/kg,b.wt. of alcoholic extract of *A. arvensis*. The clinical signs included anorexia, restlessness, diarrhea, thirst, difficult breathing, tremors and ended by coma and death. Hematologically, there were a significant reduction in PCV\(\%\), Hb concentration and RBCs count of the intoxicated rats. Concerning kidney function tests, there were a significant increase in urea and creatinine level of the intoxicated rats. Pathologically, the lesions were primarily confined to the urinary system.

**Key word:** *Anagallis arvensis*, rat, alcoholic extracts, Saudi Arabia

**Introduction**

Poisonous plants are widely distributed all over the world. These plants are considered those which, when eaten, can give rise to a departure from the normal health of man and domestic animals. The toxicity of plants has been known since the beginning of recorded history. Records of harmful effects by many of these plants are based solely on poisoning of livestock rather than on actual cases of human ingestion.

Many weed infest Saadian field: one of these is *Anagallis arvensis*, which is decidedly poisonous to small animals and sheep (Miller, 1935; Wahby, 1942), and that it may adversely affect large animals specially horses fed in great quantities of green fodder heavily infested with this weed (Georgia, 1923; Lander, 1944). Also, *Anagallis arvensis* has been caused poisoning in sheep in Australia on various occasions (Hurst, 1942) and its toxicity has been confirmed by experiment(Pullar, 1939). However, the plant is reported to produce gastrointestinal symptom in the dog and horse, to be toxic to poultry and rabbit and (the seed) to birds (Walt and Breyer-Brandwijk, 1962). More recently in South Africa (Schneider, 1978) poisoning of sheep, all of which died, was attributed to *Anagallis arvensis* after other sheep had been fed experimentally with the plant and developed the same clinical signs. These included difficult breathing, depression, stiffness of gait, leg weakness, recumbency and in the terminal stages, coma and rapid drop in body temperature. Typical post-mortem lesions are hemorrhage of the kidneys, heart and intestines and congestion of the lungs and liver. Therefore, (Kotb, 1985) reported that the active principle of *Anagallis arvensis* were acied volatile oil, enzymes, saponins, tannins, bitter principle and a compound known as primin. Roots contain cyclamen, a crystalizable glucosidal saponin. The oil produce headache lasts for 24 hrs. The plant was toxic to dog, rabbits and sheep. Signs of toxicity were general depression, thirst and diarrhea. Also, Chevallier (1996), recorded that *Anagallis arvensis* contains saponins (including anagallin), tannins and cytotoxic. However, Riet-Correa et al. (1998) reported that four cases of *Anagallis arvensis* poisoning were diagnosed in the Department of Psysandu, Uruguay during December, 1994 and January, 1995, in barely and stubble fields. Cattle of different ages were affected. Morbidity was 7-30% and case fatality was 50-66%. In two cases the animals had been introduced in the stubble field 7-15 days before the observation of clinical case. In to others the animals were in the fields 30-45 days before developing clinical signs. Eight of 289 ewes died after grazing in the same field that had affected cattle. Another case was observed in sheep and cattle with calves with no clinical signs in the calves. Clinical signs included anorexia, restlessness, weight loss, haemorrhagic diarrhea, muscular tremors and convulsion. Serum urea, creatinine and magnesium were increased. Clinical manifestation periods was 2-15 days. Gross lesions were characterized by petechial haemorrhage and edema of the mesentery, presence of clear yellowish fluid in the cavities, erosive and ulcerative lesions in the esophagus, haemorrhagic abomasitis and entritis, perirenal edema and yellowish discoloration of the kidneys. The main histologic
lesion was a severe tubular nephrosis. Sadekar et al., 1996 have been observed dullness, anorexia and constipation of feeding *Anagallis arvensis* also pathological changes in kidneys and livers.

**Materials and Methods**

**Plant materials:** *Anagallis arvensis* were freshly collected from various farms in Al-ahsa. Approximately 1 kg of dried plant materials was ground and extracted with 80% ethanol by shaking and percolation for 24 hours at a room temperature. The extract was pooled and centrifuged at 1000 rpm for 10 minutes. The solvent of supernatant liquid was removed completely by evaporation under vacuum. Prior to drug administration the residue was dissolved or suspended in distilled water. White albino rats of both sexes were used in this investigation. These animals were clinical healthy and weighing 150-200 g. Rats were housed in hygienic fiber glass cages. They were obtained from college of Veterinary Medicine and Animals Resources, King Faisal University. Animals were fed on commercial pellets, obtained from Grain Silos and Flour Mills Organization, Riyadh. These rats were used for assay of different toxic dose levels in experimental toxicity tests. Through the study all animals were observed once daily for clinical and signs of toxicity.

**The determination of the LD₅₀:** According to the method of Weil (1952) for determination of the dose of LD₅₀, exploratory trials were performed in five groups each of two rats, alcoholic extract was administrated intraperitoneally (IP) at doses of 5, 10, 20 and 30 mg/kg b.wt. Correspondingly in the five groups to find the smallest toxic dose to start with. The dose 5mg/kg b.wt. which was the first dose to cause signs of toxicity multiplied by constant factor (2) for each succeeding groups of rats. Five groups of rats were used (10 of each), four groups were 5, 10, 20 and 40mg/kg b.wt. alcoholic extract respectively. Groups five was used kept as a control. Mortality rats was record after 24 hr. Studies of repeated administration of *Anagallis arvensis* extract in the rats, eighteen adult rats of both sexes were divided into 3 equal groups (6 rats each). The 1st and 2nd groups were daily IP injected with (2.359 and 1.072) about 1/5 and 1/10 LD₅₀ aqueaus solution of *Anagallis arvensis* alcoholic extract respectively. The 3rd group was injected by sterile saline solution and kept as a control. All rats were kept under observation for 15 days. At the end of experiment all rats were sacrificed. Hemoglobin concentration was determined according to Sahl’s metods (Schalm,1975) red and white cell count were done using the double improved Neubauer chamber (Wintrobe,1967) and packed cell volume (PCV) was calculated by microhematocrite tube as described by (Schalm,1975).

Serum samples, following sacrifice were used to determine urea according to Fawcett and Scott (1960) and serum creatinine according to Seeling and Wust (1969) methods.

The post mortem examination was applied for all the sacrificed rats of the three groups and specimens were collected from various organs. The latter specimens were fixed 10% neutral buffer formaline and then routinely processed in paraffin technique, for the microscopic examination after staining by haematoxylin and eosin stain, Culling (1974).

**Results**

**LD₅₀ Determination:**

<table>
<thead>
<tr>
<th>No of animals/group</th>
<th>Dose of <em>A. arvensis</em> extract mg/kg b.wt</th>
<th>No of animals died</th>
<th>% of animals died</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>5</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>10</td>
<td>10</td>
<td>4</td>
<td>40</td>
</tr>
<tr>
<td>10</td>
<td>20</td>
<td>10</td>
<td>100</td>
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<tr>
<td>10</td>
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<tr>
<td>10</td>
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</tr>
</tbody>
</table>

Log m= Da+d (f+1), Where:  
Da = The log of the lowest of the four dosage level used.  
d = The logarithm of the constant ratio between dosage levels. f = A constant factor from Weil tables  
Log m = log 5 + log 2 (0.1+1)  
= 0.69867+0.30103(1-1)  
= 0.69867+0.331133  
= 1.03013  
Antilog = 10.718  
LD₅₀ = 10.718 mg/kg, b.wt.

*Anagallis arvensis* produce gastrointestinal symptoms. Rats suffered from anorexia, restlessness, diarrhea, thirst (the animal goes toward drinking water), difficult breathing, tremors and ended by coma and death of some individuals. There was haematological changes RBCs, Hb and PCV were significantly decreased P< 0.05 as compared with the normal in both dose levels. WBCs count was showed non-significant change, Table 2. Biochemical changes were urea and creatinine level was increased significantly in both dose levels, Table 3.

**P. M. Lesions:** There were hemorrhage of the kidneys, heart and intestine and congestion of the lung and liver. Prostate gland was enlarged as compared with normal, Fig. 2.

**Histopathological changes**

**High Dose**

**Lung:** Areas of chronic interstitial pneumonitis and
Table 2: Duncan multiple range test for hematological studies of rats administered *Anagallis arvensis* alcoholic extracts

<table>
<thead>
<tr>
<th>Treatment</th>
<th>PVC% Mean ± S.E.</th>
<th>Hb gm/100 ml Mean ± S.E.</th>
<th>RBCs 10mm Mean ± S.E.</th>
<th>WBCs 10 mm Mean ± S.E.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>41.25±0.50</td>
<td>14.4±0.18</td>
<td>6.20±0.14</td>
<td>8.58±0.10</td>
</tr>
<tr>
<td>A</td>
<td>34.75±0.96</td>
<td>10.98±0.17</td>
<td>6.00±0.08</td>
<td>8.15±0.31</td>
</tr>
<tr>
<td>B</td>
<td>37.85±2.04</td>
<td>12.20±0.20</td>
<td>7.33±0.19</td>
<td>8.02±0.16</td>
</tr>
</tbody>
</table>

All means within the same column and having the same superscript are not significantly different from each other. Where Control = mean of control group, A = mean of group 1/5 of LD<sub>50</sub>, B = mean of group 1/10 of LD<sub>50</sub>, P< 0.05.

Table 3: Duncan multiple range test for some of kidney function tests of administrated *Anagallis arvensis* alcoholic extracts

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Urea mg/dl Mean ± S.E.</th>
<th>Creatinine mg/dl Mean ± S.E.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>31.80±2.50</td>
<td>0.50±0.04</td>
</tr>
<tr>
<td>A</td>
<td>50.75±5.74</td>
<td>0.74±0.12</td>
</tr>
<tr>
<td>B</td>
<td>41.67±2.16</td>
<td>0.62±0.03</td>
</tr>
<tr>
<td>C</td>
<td>50.75±5.74</td>
<td>0.74±0.12</td>
</tr>
</tbody>
</table>

All means within the same column and having the same superscript are not significantly different from each other. Where A = mean of control group. B = mean of group 1/10 of LD<sub>50</sub>, C = mean of group 1/5 of LD<sub>50</sub>.

alveolar emphysema were seen. Bronchitis and bronchiolitis associated with peri-bronchial lymphocytic hyperplasia were also detected. The epithelium cells lining, the bronchial mucosa showed marked goblet cell hyperplasia, Fig. 3, 4 and 5.

Liver: In one case marked venous congestion has been observed. Hepatocyte appeared swollen and fine vacuolated. Occasional foci and single cell necrosis associated with inflammatory cell infiltration and kupper cell hyperplasia were seen in four animals, Fig. 6.

Kidney: Thickening of the basement membrane of the glomeruli and degeneration of the renal tubular epithelium characterized by dilatation of renal tubules and vacuolization of the aggregation were seen. Occasional granuli appeared atrophy with dilatation of Bowman's capsule and accumulation of serous fluid.

Prostate Gland: In one case acute prostatitis characterized by massive accumulation of neutrophils in the lumen of the prostatic acini as well as the interstitial tissue in addition to congestion and edema of interstitial tissue in addition to congestion and edema of interstitial tissue have been observed, Fig. 7.

Heart: Mild peri-vascular edema

Testis: Congestion with small areas of intertubular hemorrhage were seen, Fig. 8.

Fig. 1: Varieties of *Anagallis arvensis* plant

Brain: Congestion of meningeal vessels was detected.

Low Dose

Kidney: Some of the cortical tubules were lined with vacuolated epithelial cells with hyaline casts in their lumen. Few basophilic cortical tubules were seen, Fig. 9 and 10.

Spleen: No abnormality detected.

Lung: Areas of chronic interstitial pneumonitis and endarteritis obliterans have been seen in the four cases, Fig. 11.

Liver: Hyperplasia of kuffer cell and small foci of parenchymal necroses infiltrated by mononuclear cells were seen Fig. 12 and 13.
Heart: Mild myodegeneration.

Testis: No abnormalities detected.

Brain: Congestion of the meningeal vessels.

Prostate Gland: Acute suppurative prostatitis characterized by multiple abscesses in the interstitial tissue and intensive neutrophilic cell aggregation in the lumen of the prostatic acini, Fig. 14.

Control

Kidney: Occasional cortical tubules were lined with vacuolated epithelium in addition to atrophy of the glomeruli associated with dilated Bowman's capsule and accumulation of serous fluid in the lumen.

Testis: Degeneration of the germinal epithelial cells of some seminiferous tubules with formation of spermatid giant cells were seen, Fig. 15.

Lung: Perivascular accumulation of round cell areas and areas of chronic interstitial pneumonitis were seen.

Discussion

Studies on the toxicity of *Anagallis arvensis* in rats gave LD₅₀ of 10.718 mg/kg.b.wt. i.p. Acute toxic manifestation characterized by producing gastrointestinal symptom. Rats suffered from anorexia, restlessness, diarrhea, thirst (the animals go towards drinking water), difficult breathing, tremors and ended by coma and death of some individuals by Watt and Breyer - Brandwijk (1962); Forsyth (1968); Kob (1985).

Hematological results revealed severe anemia as remarked by decrease hemoglobin concentration, lowering of packed cell volume and marked fall in erythrocytic count. These results could be attributed to the harmful effect of saponin, an active principle of *Anagallis arvensis*, concerning to total WBCs count, there was no changes.

The elevated urea could be attributed to acute renal failure rather than to inadequate fluid intake, fluid diversion, or excessive external output (Oliver and
Platonow, 1960) and tubular damage seen at the time of killing (Klein et al., 1972; Tryphonas and Neilsen, 1973). The other opinion concerning normal level of urea are attributed to the plant, dose and period of treatment by the active principles.

The level of creatinine of the intoxication rats was increased significantly (p < 0.05). This increment is an indicator for renal function impairment due to toxicosis by Anagallis arvensis. The results are impairment due to toxicosis by Anagallis arvensis. The results are in agreement with that obtained by Coles (1974), Metwalli (1987); El. Garieb (1990) who reported that a significant increase in creatinine level was indicative of kidney impairment and dysfunction. The significant increase in the level of creatinine may be regarded to glomerular
damage and excessive muscular catabolism. Such as conclusion is confirmed by the present histopathological investigation which indicated glomerular damage (membranous glomerulopathy) and corticotubular degeneration mainly the proximal convoluted tubule (nephrosis). These changes appeared dose related as by moderate and seen pathological change in the kidney at low and high dose groups respectively. Such conclusion is confirmed with those reported by Kelly (1984) who recorded that there was an increase in blood creatinine during severe renal damage, where the degree of rise can be more accurately correlated to be extent of glomerular damage in chronic nephritis than in acute renal impairment in which excessive muscular catabolism will artificially elevated the blood creatinin value. The relation of the inflammatory changes observe in the prostatic gland of one animals of groups 2 and 3 are low significance as there is no indication of an edema toxic effect on the genital organ have been detected. The change observed in liver and kidneys of treaties groups are more or less similar to that of control animal and have no pathological significance. It could be concluded that Anagallis arvensis was highly toxic to rats and induces several changes in the animals tissues. It is potentially nephrotoxic.

References


Weill, C. 1952. Tables for convenient calculation of median effective dose (LD50 or ED50) and instructions in their use. Biometrics, 8: 249.