Distribution and Effect of Plant-Parasitic Nematodes Associated with Cashew in North Central-Nigeria

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Abstract: Roots and soil were sampled for the presence of plant-parasitic nematodes from seven selected locations in North Central Nigeria noted for cashew production. Ten genera of plant-parasitic nematodes were found associated with cashew in North Central Nigeria. Meloidogyne sp., Helicotylenchus coffeeae and Radopholus sp., were widespread in all the locations while Rotylenchulus reniformis was recorded only in Ochaja, Ayingba and Ejule. There were significant reduction in the height of cashew seedlings inoculated with root-knot nematodes, Meloidogyne incognita, in the nursery. Therefore, pre-planting sampling strategies should be taken to predict the impact of the nematode populations and enable preventive measures to be taken, most especially at the seedlings stage.

Key words: Cashew, plant-parasitic nematodes, Meloidogyne incognita, Radopholus sp., Nigeria

INTRODUCTION

Cashew, Anacardium occidentale L., is an important commodity cash crop which grows in most agroecological zone of Nigeria, right from the coast in the South to the sub-desert areas of the North (Topper et al., 2001). It has tremendous potential as a cash crop to generate foreign exchange, create employment to curb desertification in the Northern states while ornamental and alley trees are used to prevent soil erosion by protecting the watersheds and dams in the South (Adejumo, 2010).

Limited information on nematodes attacking cashew exists. High populations of Criconemoides sp., Xiphinema sp. and Scutellonema sp. have been found around unthriftly trees in Brazil and da Ponte recognized Xixinematose caused by X. index as one of the more common diseases of cashew in North-East Brazil, although data on its economic impact are lacking. A review of cashew diseases in Brazil concluded that nematodes supported by the plant cause no evident damage (Freire et al., 2002). Rotylenchulus reniformis, apparently in its migratory form, was reported from around cashew trees in Costa Rica but again, evidence of damage is unclear (Lopez and Salazar, 1987). Hemicycliophora attapadi was described from the cashew rhizosphere in India. Therefore, the study was undertaken to survey and consider the effects of plant-parasitic nematodes on cashew in selected locations of Nigeria.

MATERIALS AND METHODS

Nematode survey: Nematological surveys were carried out in selected locations in North Central-Nigeria noted for cashew production. These locations include: Ochaja, Ayingba, Ejule, Kabba, Okene (Kogi state), Ibadan (Oyo state) and Oro (Kwara state). Soil and root samples were collected from the rhizosphere region about 50-70 cm from the base of the plants and at a depth of 20 cm.

Processing of soil and root samples: Aliquots of 250 cm³ sub-sample soil from 500 cm³ each composite sample were assayed for nematodes by sieving and decanting (Cobb, 1918). After decanting, the sediment was assayed for nematodes using the Whitehead and Hemming (1965) tray modification of Baermann’s technique as described by Coyne et al. (2007). The root samples were washed, pooled, chopped into approximately 1 cm pieces and thoroughly mixed. A 5 g sub-sample was put in 100 mL water in a kitchen blender. The root was macerated 3 times for 10 sec, separated by 5 sec intervals and the nematodes were extracted from the resulting homogenate using Sieve Method (Speijer et al., 1997). The nematode suspension was diluted with water in a graduated cylinder to 10 mL.

Prior to counting, solution containing nematodes were agitated thoroughly and nematode populations were determined in 1 mL distilled water suspension in a counting dish (Doncaster, 1962) under a stereomicroscope and expressed per 250 cm³ soil or 5 g roots. A mean of 3 counts was taken in each case. Nematodes were transferred with an eye lash picker to a slide with a drop of water, covered (with a cover slip) and examined under a compound microscope with a 40, 60 and 100 X objective for identification using taxonomic keys (Hunt et al., 2005).
and counted. The identification and counting was repeated 3 times and mean population of nematodes/sample calculated.

**Pathogenicity tests:** Cashew seedlings collected from the locations were allowed to stabilize in the greenhouse for 2 weeks and thereafter subjected to pathogenicity tests. The pots with the cashew seedlings were inoculated with 5,000 *Meloidogyne incognita* eggs obtained from the pure culture on the roots of *Celesa argentea* using Hussey and Barker (1973)'s Sodium Hypochlorite (NaOCl) Method. Uninoculated units served as control. Normal watering of seedlings as obtains in coffee nurseries was carried out. Fortnightly, growth parameters such as plant height, stem girth and numbers of leaves were recorded. The experiment was terminated 24 weeks after inoculation.

To assess infection the roots were carefully freed of soil, washed under a gentle stream of tap water, mopped and galls counted using a hand lens at 3-5 X magnification. Root galling was assessed using the 0-5 gall index (Sasser *et al.*, 1984). Nematode eggs were collected from each root system using Sodium Hypochlorite Method (NaOCl) of Hussey and Barker (1973) and counted. Aliquots of 250 cm² soil samples from each pot were assayed for juveniles of *M. incognita* using the modified Baermann Technique (Coyne *et al.*, 2007).

**Data analysis:** Prior to statistical analyses, data were checked for normality and homogeneity of variances and transformed where necessary. A log transformation \[\log_{10}(x+1)\] was applied to the data on nematodes (densities per 250 cm² soil, densities per gram root and gall index). The determination of disease incidence was based on the nematode population per root system and it was expressed by the number of egg masses and gall index. The numbers of egg masses were counted and Gall Index (GI) and Egg Mass Index (EMI) were determined on the following scale: 0-10; 1-10; 2-3; 3-10; 4-11-30; 5-31-100 and 6-101-1000 galls or egg masses per root system (Taylor and Sasser, 1978, Sasser *et al.*, 1984).

Reproduction factor of the nematode (RF) = Final nematode population (pf) × Initial nematode population (pi)⁻¹.

All data collected were subjected to analysis of variance and significant differences between means were evaluated using Least Significant Difference Method at p<0.05. All analyses were performed using GenStat (Version 7.1, VSN International Ltd., Lawes Agricultural Trust, Hampstead, UK).

## RESULTS AND DISCUSSION

Ten genera of plant-parasitic nematodes were found associated with cashew in North Central-Nigeria. *Meloidogyne* sp., *Helicotylenchus* *coffeanus* and *Radopholus* sp., were widespread in all the locations while *Rotylenchulus reniformis* was recorded only in Ochaja, Ayinbba and Ejule (Table 1). An earlier survey of plant-parasitic nematodes associated with cashew in South Eastern-Nigeria revealed *Xiphinema*, *Scutellonema* and *Criconemella* sp., as the most important based on frequency of occurrence (Agu, 2007). The prominence and importance values of *Scutellonema* and *Criconemella* sp., were highest and differed significantly from others due to the presence of sweet potato and *Cynodon dactylon* in the plantations sampled which, respectively are good hosts to *Scutellonema* and *Criconemella* sp., (Jatala and Bridge, 1990). *Rotylenchulus reniformis* was reported from around cashew trees in Costa Rica, though evidence of damage is unclear (Lopez and Salazar, 1987).

There were significant reductions in the height of cashew seedlings inoculated with root-knot nematodes, *Meloidogyne incognita* (Table 2). Although, there were no significant differences in the number of leaves and stem girth of the seedlings between the inoculated and the control; yellow discoloration of the leaves was noticed in the inoculated plants. The nematode successfully reproduced on the cashew seedlings with tiny galls seen on the roots of the inoculated. It is important to emphasize that mature cashew trees has

### Table 1: Distribution of plant-parasitic nematodes associated with cashew in selected locations of Nigeria

<table>
<thead>
<tr>
<th>Nematode species</th>
<th>Ochaja</th>
<th>Ayinbba</th>
<th>Ejule</th>
<th>Kabba</th>
<th>Okene</th>
<th>Badaun</th>
<th>Oro</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Meloidogyne</em> <em>sp.</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td><em>Helicotylenchus</em> <em>coffeanus</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td><em>Xiphinema</em> <em>sp.</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td><em>Criconemoides</em> <em>senex</em></td>
<td>o</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Pratylenchus</em> <em>coffeanus</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td><em>Scutellonema</em> <em>brachyurus</em></td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Hemicyclophora</em> <em>sp.</em></td>
<td>o</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td><em>Radopholus</em> <em>sp.</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td><em>Rotylenchulus</em> <em>reniformis</em></td>
<td>+++</td>
<td>+++</td>
<td>+++</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td><em>Dichodus</em> <em>sp.</em></td>
<td>o</td>
<td>++</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
</tbody>
</table>

*o* = Not recorded; + = Present in survey; ++ = Common; +++ = Widespread
Table 2: Effect of root-knot nematode, Meloidogyne incognita, on some growth parameters of cashew seedlings at 24 weeks after inoculation

<table>
<thead>
<tr>
<th>Cashew seedlings</th>
<th>Plant height (cm)</th>
<th>No. of leaves (count)</th>
<th>Stem girth (cm)</th>
<th>GI/EMI (count)</th>
<th>Reproduction factor (RD) p/f/p</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oro series</td>
<td>32.70</td>
<td>11.13</td>
<td>0.81</td>
<td>3/3</td>
<td>8.1</td>
</tr>
<tr>
<td>Control</td>
<td>38.54</td>
<td>10.14</td>
<td>0.80</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>CRIN series</td>
<td>35.65</td>
<td>10.54</td>
<td>0.82</td>
<td>3/3</td>
<td>7.6</td>
</tr>
<tr>
<td>Control</td>
<td>39.12</td>
<td>11.14</td>
<td>0.82</td>
<td>-</td>
<td>-</td>
</tr>
</tbody>
</table>

*Means followed by the same letter in the same column are not significantly different (p<0.05); p/f = Final nematode population; p = Initial nematode population; GI = Gall Index; EMI = Egg Mass Index.

been shown clearly to be highly resistant to different populations of the root-knot nematodes in West Africa and in Brazil but the seedlings are susceptible to the nematode.

CONCLUSION

From the study, pathogenicity tests showed that Xiphinema, Criconemella and Scutellenema sp., also caused significant reductions in cashew seedling growth at all levels of inoculation (Agu, 2007). Therefore, pre-planting sampling strategies should be taken to predict the impact of the nematode populations and enable preventive measures to be taken, most especially, at the seedlings stage.

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REFERENCES


