Seroprevalence of Bovine Paratuberculosis Specific Antibodies in Khartoum and Al-Jazeera States, Sudan

K.B. Mohammed, A.M.S. El-Eragi and A.M. Zakia
Central Veterinary Research Laboratories, Department of Pathology and Diagnosis, Khartoum, Sudan

Abstract: This study was conducted to estimate the bovine paratuberculosis seroprevalence in Khartoum and Al-Jazeera states, Sudan. The generated results showed that in Khartoum state the seroprevalence of paratuberculosis was 66.7% at the herd level and 10.2% at the individual animal level. The lowest seroprevalence (8%) was found at Kuku and the highest (18.8%) was found at El-Sealeot localities. All sera collected from El-Kadaro, Wad-Medani and El-Salama localities were found negative for Mycobacterium avium Subspecies Paratuberculosis (MAP) antibodies. Khartoum North showed the highest rate of seropositivity whereas Omdurman showed the lowest. Relationship between seroprevalence and clinical manifestation were described. The results of this study reported a widespread of bovine paratuberculosis in the Khartoum state.

Key words: Seroprevalence, bovine, paratuberculosis, ELISA, antibodies, Sudan

INTRODUCTION

Paratuberculosis (Johne’s Disease, JD) is a chronic progressive, granulomatous enteritis caused by MAP (Harris and Barletta, 2001). Paratuberculosis has been recognized as a major disease of ruminants for more than a century and has significant economic and welfare effects on livestock in all continents (Chiodini et al., 1984). Several animal species are known to be susceptible to infection but the primary hosts are ruminants including cattle, sheep, goats, llamas, alpacas and non domesticated hoofed stock such as bison and deer (Harris and Barletta, 2001). Paratuberculosis has also been reported in a variety of free range animal species including antelope (Dukes et al., 1992), bighorn sheep (Williams et al., 1983), white-tailed deer (Chiodini and van Kruiningen, 1983), tule elk (Cook et al., 1996), moose (Soltys et al., 1967), red deer, roe-deer, fallow deer and moufflon (Pavlak et al., 2000), rabbits (Angus, 1990) and earthworms (Fischer et al., 2003). Some of these free range animals have been implicated as potential carriers of MAP for domestic ruminants (Dukes et al., 1992; Chiodini and van Kruiningen, 1983; Daniels et al., 2001).

Infections with MAP occur in the first 6 months of life and can persist for several years with mild clinical manifestations (Merkal et al., 1975; Kennedy and Benedictus, 2001; Manning and Collins, 2001; Storset et al., 2001). Recently, MAP has received an increasingly wide interest because of a rapidly growing body of scientific evidence which suggests that human infection with MAP may cause some and possibly all cases of Crohn’s disease (Naser et al., 2004; Uzoigwe et al., 2007).

ELISA was the most widely used test for screening herds. Detection of infection by ELISA appears to be dependent upon the stage of disease (Yayo-Ayele, 2001). ELISA sensitivity for clinical cases has been reported to be 87 and 15% in subclinical cases (Sweeney et al., 1995). Most paratuberculosis experts recommend that ELISA positive reactors should be confirmed by faecal cultures (Yayo-Ayele et al., 2001).

In general, it is accepted that the sensitivity of the antibody ELISA is 50% in all infected adults and this ratio rises up to 90% in clinical cases (Sweeney et al., 1995). A series of reports from North America and Australia have indicated that the specificity of the test may reach 97% up to 99% or above (Cox et al., 1991; Reichel et al., 1999). ELISA tests to detect MAP antibodies in milk do not correlate well with the ELISA results on serum from the same animals. Similarly, the bulk-milk ELISA test does not accurately predict the percentage of cattle in herds that were positive by ELISA on serum samples (Hardin, 1995).

In the Sudan, paratuberculosis was first diagnosed in goats by Fawi and Obied (1964). Thereafter, 9 clinical cases at El Gurashi dairy farm were diagnosed by El Derdiri. Later it was diagnosed in kuku area and Belgravia dairy farm by Abu Buiker and Elsanousi (1975). Abbas et al. (1986) recovered the organism from clinical

Corresponding Author: K.B. Mohammed, Central Veterinary Research Laboratories, Department of Pathology and Diagnosis, Khartoum, Sudan

2098
cases of goats. About 11 isolates of MAP were isolated from one thousand of cows. The investigation of Mongash in the Sudan, revealed 23 cows were positive by Ager Gel Immunodiffusion (AGID) test where as 13 out of 830 were positive by ELISA.

In the same farm 13 out of 120 were found to have acid fast bacteria in the rectal scrapings (Mongash, 1989). The aim of this study was to determine the seroprevalence rate of bovine paratuberculosis in Khartoum and AlJazeera states, Sudan.

MATERIALS AND METHODS

Sera: About 225 serum samples were collected from nine dairy herds of cross bred between Friesian and local Butana breeds. Eight herds were from Khartoum state and one from Al Jazeera state. Collected sera were labelled and stored at -20°C till being used.

ELISA procedure: ELISA kits specific for detection of MAP antibodies were purchased from Institut Pourquier, Montpellier, France (Cat. No P07110). The kit procedure was similar to that described by OIE. The test was performed according to the manufacturer’s instructions. Briefly, the microplate wells were coated with a protoplasmic extract of MAP.

In order to minimize cross-reactions, samples along with positive and negative controls were diluted 1:20 in a buffer containing an extract of Mycobacterium phlei and placed on a shaker for 15 min at 24°C. All samples and reagents were added at 100 µL well-1. Diluted samples and controls were dispensed into the antigen coated wells and incubated for 1 h at 24°C. The wells were then washed manually. After washing, a peroxidase labelled monoclonal anti-ramoniun IgG conjugate was added to each well and incubated for 30 min. The plates were then washed and Tetra-Methyl-Benzidine (TMB) buffer was added in each well and incubated for 10 min.

The reaction was stopped by using 0.5 M sulphuric acid and Optical Densities (OD) were read at 450 nm using a microplate reader (Digital and analog systems s.r.l, Pabonera Sabina, Roma, Italy). Each test serum and both controls were evaluated in two adjacent wells. The obtained results were interpreted according to the formula given by the manufacturer:

\[ S = \frac{\text{Corrected OD 450 value of the sample}}{\text{Mean corrected OD 450 value of the positive control}} \times 100 \]

- Any sample with an S/P is between 60 and 70% is considered to be doubtful. A second test will be necessary to confirm its status.
- Any sample with an S/P equivalent or >70% is considered coming from an animal which has been infected by MAP.

RESULTS AND DISCUSSION

The results of ELISA test showed that 23 out of 225 (10.2%) serum samples were positive (Table 1). This indicates that the overall prevalence of seropositive cattle was 10.2%. Paratuberculosis seropositive animals were detected in 6 herds (66.7%). Regarding localities, the lowest value of seroprevalence (8%) was found at Hilat kuku and the highest (18.8%) was found at El-Sealeet areas (Table 1). All sera collected from El-Kadaro, Wad-Medani and El-Salama were found negative for MAP antibodies. Sero-positive animals were found in all locations with different prevalence rates except for Wad-Madani. Khartoum North showed the highest rate of seropositivity while Khartoum showed the lowest (Table 1). The seroprevalence among clinical and subclinical cases were 100 and 5.6%, respectively (Table 2). In El-Salma herd although, there was a history of paratuberculosis no specific antibodies for MAP were detected.

The present study was designed to study the prevalence of MAP specific antibodies among dairy cattle in both Khartoum and Al-Jazeera states. The overall seroprevalence detected by ELISA in Khartoum (10.2) was found to be higher than that reported by Mongash (1989). This may indicate that the infection is progressing in the area of the study. In this study, the positive reactors were

### Table 1: Seroprevalence of bovine paratuberculosis in Khartoum and Al-Jazeera states

<table>
<thead>
<tr>
<th>State</th>
<th>Province</th>
<th>Locality</th>
<th>No. of test animals</th>
<th>No. of reactive reactors</th>
<th>Prevalence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Khartoum</td>
<td>Khartoum North</td>
<td>El-Sealeet</td>
<td>16</td>
<td>3</td>
<td>18.8</td>
</tr>
<tr>
<td></td>
<td></td>
<td>El-Bagair</td>
<td>50</td>
<td>9</td>
<td>18.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Hilat Kuku</td>
<td>25</td>
<td>2</td>
<td>8.0</td>
</tr>
<tr>
<td>Khartoum</td>
<td></td>
<td>El-Kadaro</td>
<td>19</td>
<td>0</td>
<td>0.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>El-Salama</td>
<td>33</td>
<td>0</td>
<td>0.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Soba</td>
<td>37</td>
<td>5</td>
<td>13.5</td>
</tr>
<tr>
<td>Omdurman</td>
<td>Omdurman</td>
<td></td>
<td>32</td>
<td>4</td>
<td>12.5</td>
</tr>
<tr>
<td>Al-Jazeera</td>
<td>Wad-Medani</td>
<td>Wad-Medani</td>
<td>13</td>
<td>0</td>
<td>0.0</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>225</td>
<td>23</td>
<td>10.2</td>
</tr>
</tbody>
</table>

### Table 2: Seroprevalence of bovine paratuberculosis in relation to clinical signs

<table>
<thead>
<tr>
<th>Clinical signs</th>
<th>Animals tested (No)</th>
<th>ELISA positive (No)</th>
<th>Prevalence (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Clinical cases</td>
<td>8</td>
<td>8</td>
<td>100.0</td>
</tr>
<tr>
<td>Subclinical cases</td>
<td>217</td>
<td>15</td>
<td>6.9</td>
</tr>
<tr>
<td>Total</td>
<td>225</td>
<td>23</td>
<td>10.2%</td>
</tr>
</tbody>
</table>
found at Hilat kuku, Soba and El-Bagair areas, a finding similar to that reported by Mongash (1989). These findings were direct evidence that these areas were endemic with bovine paratuberculosis and they may constitute a continuous source of shedding the infection. Although, there is a history of the disease in El-Salma area, no MAP antibodies were detected. This result might be attributed to the good husbandry management, culling and disposability of the clinical and heavy fecal shredders by the animal owners in the area as was explained previously by Whitlock et al. (2000).

The source of bovine paratuberculosis in Sudan was suggested to be through the importation of foreign dairy cattle from European countries (Mongash, 1989). In the study, it is reported the prevalence of MAP specific antibodies in crossbred cattle between Friesian and Butana breeds, since all the sera were collected from these cattle.

Seroprevalence in clinical cases was higher (100%) than in subclinical cases (6%), a finding similar to that reported by Sweeney and colleagues in 1995.

CONCLUSION

The results of this survey indicated that MAP antibodies are widely distributed in cattle in the Khartoum North. This may be attributed to the crowding of the animals in this area without clear partitions between the farms which might increase the potential for exposure to contaminated faeces. This area requires special attention of veterinarians and producers in order to establish an efficient control programme with regular revaluation.

It needs to apply seroprevalence of bovine paratuberculosis as nationwide to provide information used in control programme and eradication. Due to its low cost, accuracy and ease of sample collection and shipment, it suggest ELISA can be used for this purpose as a screening test on herd or individual levels followed by other methods to confirm animals which were positive for antibodies specific for MAP by ELISA.

ACKNOWLEDGEMENTS

We gratefully acknowledge Pro. Osman, A.Y for critical reading of manuscript. This research was funded by Central Veterinary Research Laboratories, Khartoum, Sudan.

REFERENCES


Mongash, B.M., 1989. Diagnosis of Johne’s disease in cattle. M.Sc. Thesis, Department of Medicine, Pharmacology and Toxicology, Faculty of Veterinary Medicine, University of Khartoum, Sudan.


