

Ruminal and Post-Ruminal Protein Disappearance of Chemically Treated Alfalfa Silage and the Effect of the Silage Containing Diets on Performance of Holstien Lactating Dairy Cows

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Abstract: Alfalfa hay (30% dry matter) was ensiled as untreated (AS) or treated with formic acid (ASF, 24 mL kg⁻¹ DM) or formic acid+urea (ASFU, 24 mL and 4 g kg⁻¹ DM, respectively). Disappearance of protein and dry matter from the samples was determined using the *in situ* mobile bag procedure. In lactation trail, 12 Holstein cows received diets containing 28.5% AS or ASF or ASFU, 30% corn silage, 7% alfalfa hay and 34.5% concentrate for 8 weeks. The chemically processed applied caused a significant decrease in ruminal and an increase in post-ruminal dry matter disappearances of the silages evaluated (p<0.05). Dry matter intake and yields of milk did not influenced by the experimental diets. Milk urea nitrogen concentration of the cows received the diet containing ASFU were significantly lower than the others (p<0.05). Blood plasma concentration of glucose was increased when the diet containing of ASF was fed to the animals (p<0.05) compared with the others.

Key words: Alfalfa silage, formic acid, urea, degradability, ASFU, glucose

INTRODUCTION

Alfalfa protein is subject to extensive degradation during ensiling as much as 75-87% of the total nitrogen present in alfalfa silage may be non-protein nitrogen (Muck, 1987). This resulted in inefficient N use especially in diets in which fermentable energy is limiting. Formic acid commonly is used as a preservative for direct-cut silage in northern Europe. Formic acid-treated alfalfa hay silage had lower pH and NH₃ concentrations than untreated controls and higher water-insoluble N compared with the intact silage (Barry *et al.*, 1978; Lancaster and Brunswick, 1977).

Formic acid was more consistent than bacterial inoculants in reducing protein degradation and deamination in clover silage (Woolford and Sawczyk, 1984). In contrast to enzymes and inoculants that stimulate silage fermentation, formic acid restricts fermentation and decreases silage pH by direct acidification (Muck and Kung, 1997; Nagel and Broderick, 1992; Waldo *et al.*, 1971). Therefore, formic acid is commonly used in crops with low DM and sugar concentrations. Under these conditions, it is especially important to decrease pH rapidly (<4.2) to prevent clostridial growth (McDonald *et al.*, 1991; Muck and Kung, 1997). Well fermented, highly digestible silages

containing high concentration of lactic acid and low acetic acid, ammonia-N and cell wall concentrations are associated with higher intake and improved animal performance (Wilkins *et al.*, 1971). Increased dry matter intake and N retention have been reported in sheep (Barry *et al.*, 1978) and dairy heifers (Waldo *et al.*, 1971) fed treated alfalfa silage. Little information is available on milk production when formic acid-treated alfalfa silage is fed to dairy cattle. Glenn *et al.* (1986) reported a trend for higher milk yields when cows were fed alfalfa silage treated with formic acid plus formaldehyde however, alfalfa comprised only 30% of the diet DM. The objective of this study was to test the effect of treating alfalfa silage with formic acid and urea on *in situ* mobile bag DM and protein disappearances and the effect of diets containing the silages on dry matter intake, milk and milk component yields and blood plasma metabolite concentrations of Holstein lactating dairy cattle.

MATERIALS AND METHODS

Ensiling and chemical analysis: Freshly chopped alfalfa hay at second cutting that was prepared with a commercial forage harvester and contained 30% dry matter was used in this experiment. Alfalfa hay was ensiled as untreated (AS) or chemically treated using

formic acid (ASF, 24 mL kg⁻¹ DM) or formic acid+urea (ASFU, 24 mL and 4 g kg⁻¹ DM, respectively) for 40 days. Formic acid was carried and used under the safety protocol of Ferdowsi University of Mashad using special instruments.

The acid (95%) was diluted with water (acid:water 1:4, vol/vol) and stored in an artificial plastic container until it was mixed with the forage. Dry matter, organic matter, silage pH and Chemical composition including Crude Protein (CP), Non-Protein Nitrogen (NPN) and ammonia-N concentrations were determined using standard methods (AOAC, 1990).

Mobile nylon bag technique: Ruminal and post-ruminal disappearance of protein and DM from the silage samples were determined using the *in situ* mobile bag procedure as described by Danesh and Stern (2005). Two Holstein steers (450±20 kg Body weight) fitted with ruminal fistulae and T-shaped intestinal cannulae were used. Animals were fed 5.1 kg of DM of alfalfa hay, 3.2 kg of DM corn silage and 2.5 kg of DM concentrate (170 g CP kg⁻¹ of DM) per head per day, two times per day at 8.00 and 18.00 h.

Dried silage samples were ground to pass a 2 mm screen. Approximately 5 g DM of each sample were placed into a polyester bag (17×12 cm with pore size of 48 µm) and incubated in the rumen for 12 h (8 bags per each sample). After removal from the rumen, bags were hand washed with tap water and subsequently dried using an aired oven (60°C, 48 h).

Aproximately 1 g of rumen-incubated residual of each bag was transfered into a polyester mobile bag (3×6 cm, pore size of 48 µm). Then the mobile bags were inserted into the small intestine via the intestinal cannulae at the rate of one bag every 30 min. Removal bags from the voided faces were collected and rinsed in tap water. The bags were dried using an aired oven (60°C, 48 h) and weighed to determine DM disappearance. Nitrogen concentration of un-incubated, rumen and post-rumen incubated samples was determined by the kjeldahl method (Kjeltec 2300 Auto analyzer, Foss Tecator AB, Hoganas, Sweden).

Lactation trail: Twelve early lactating Holstein dairy cows (50±16 DIM, 33±4.5 kg/milk yield per day) were used in three groups (4 head per group) in a complete random design for 8 weeks.

The animals were fed with same ration in the 1st week and the milk production data were used as covariate for the experimental data. The cows were fed the experimental diets containing 28.5% AS OR ASF ASFU, 30% corn silage, 7% alfalfa hay and 34.5% concentrate (13.5% corn,

42% barley, 12.9% soybean meal, 10.9% sugar beet pulp, 12.9% wheat bran, 5.8% cottonseed meal, 0.55% CaCO₃, 1% mineral and vitamin premix, 0.45% salt), two times per day. Feed intake and milk production were recorded daily. The samples of milk were prepared at the end of each week of the experimental period.

Blood samples were taken at the end of the last week of the experiment at 4 h after the morning feeding. Milk protein and milk urea-N concentrations were determined using the standard procedures (AOAC, 1990). Plasma from the blood samples were isolated using centrifuge at 4000 RPM per min for 10 min. Plasma glucose and urea-N concentrations were determined as described by Nasri *et al.* (2007).

Statistical analysis: Data of silage chemical composition, silage pH, ruminal and post-ruminal disappearance of DM and CP of the silages and plasma metabolites of the cows were analyzed according to a statistical model of $y = \text{overall mean} + \text{treatment effect} + \text{residual}$.

Data of milk yield and milk composition and dry matter intake obtained during lactation trial were analyzed as repeated in time using PROMIX of SAS (1999) according to a model of $y = \text{overall mean} + \text{treatment} + \text{time} + \text{treatment} \times \text{time} + \text{residual}$. Duncan's multiple comparison test was used to determine the the significant different among the means at $p < 0.05$.

RESULTS AND DISCUSSION

Chemical compositions and pH (g kg⁻¹ of DM) of the alfalfa silages are shown in Table 1. Formic acid and urea caused to reduce pH, NH₃-N and NPN and increased CP and DM ($p < 0.05$). Formic acid may reduce proteolysis during ensiling by either reduction of pH or by providing additional substrate to enhance the reduction of pH. In addition, acid treatment of AS caused to a decrease in soluble N and resulted in lower ruminal degradation of the silage CP.

Lower NPN and NH₃-N in alfalfa silage treated with urea and formic acid AS indicated that the pH drop was sufficiently more rapid, resulting in less protein being degraded in the AS treated with urea and formic acid (Behgar *et al.*, 2008; Agbossamey *et al.*, 1998).

Reducing silage NPN will improve utilization of CP in lactating dairy cows (Nagel and Broderick, 1992; Makonia *et al.*, 1997). Data obtained by Nadeau *et al.* (1996) has shown no effects of formic acid on cell-wall concentration. Alfalfa insoluble N was greater for the treated silages compared with the untreated silage which indicates less proteolysis occurred during

Table 1: Chemical composition (g kg⁻¹) of alfalfa silage treated with urea and formic acid

| Chemical factor ¹ | Alfalfa silages ^{2,3,4e} | | | SEM ⁵ | p-value |
|------------------------------|-----------------------------------|---------|---------|------------------|---------|
| | AS | ASF | ASFU | | |
| pH | 4.780 | 3.750 | 3.930 | 0.385 | 0.212 |
| DM | 292.410 | 349.720 | 395.330 | 34.180 | 0.038 |
| OM | 925.510 | 908.320 | 899.940 | 9.030 | 0.785 |
| CP | 177.000 | 168.550 | 196.200 | 11.700 | 0.177 |
| NPN | 17.950 | 15.950 | 14.750 | 1.040 | 0.353 |
| NPN/total N | 0.630 | 0.590 | 0.470 | 0.058 | 0.635 |
| N-NH ₃ | 0.131 | 0.998 | 0.937 | 0.170 | 0.194 |

¹DM = Dry Matter, OM = Organic Matter, CP = Crude Protein, NPN = Non Protein Nitrogen, ²AS = Untreated Alfalfa Silage, ASF = Alfalfa Silage Treated with 24 mL formic acid kg⁻¹ DM, ASFU = Alfalfa Silage treated with 24 mL Formic acid and 4 g Urea kg⁻¹ DM, ³when the difference between means is greater than two times the SEM, it is considered as significant (p<0.05), ⁴values were reported as the mean of 6 sampling, ⁵SEM = Standard Error of Mean

Table 2: Ruminal and post-ruminal dry matter and protein disappearance (g kg⁻¹) of Alfalfa silage treated with urea and formic acid using *in situ* mobile bag technique

| Items | Factors | Alfalfa silages ¹ | | | SEM ² | p-value |
|----------------------------|---------|------------------------------|------------------|------------------|------------------|---------|
| | | AS | ASF | ASFU | | |
| Ruminal disappearance | DM | 536 ^a | 512 ^b | 524 ^a | 9.30 | 0.01 |
| | CP | 767 | 767 | 763 | 23.20 | 0.93 |
| Post-ruminal disappearance | DM | 432 ^a | 482 ^b | 472 ^b | 6.20 | 0.01 |
| | CP | 868 | 881 | 875 | 6.50 | 0.80 |

^{a,b}:In each row, difference between means with different letter were significant (p<0.05), ¹AS = Untreated Alfalfa Silage, ASF = Alfalfa Silage treated with 24 mL Formic acid kg⁻¹ DM, ASFU = Alfalfa Silage treated with 24 mL Formic acid and 4 g Urea kg⁻¹ DM, ²SEM = Standard Error of Mean

Table 3: Dry matter intake, milk yield, milk composition and blood metabolites of lactating cows fed diets containing LS treated with urea and formic acid

| Items | Experimental diets containing alfalfa silage ¹ | | | SEM ² | p-value |
|---|---|--------------------|--------------------|------------------|---------|
| | AS | ASF | ASFU | | |
| DMI (kg day ⁻¹) | 23.2 | 23.1 | 22.8 | 0.242 | 0.6937 |
| Milk (kg day ⁻¹) | 32.8 | 31.6 | 30.9 | 0.930 | 0.8383 |
| Milk protein (g kg ⁻¹) | 33.1 | 32.1 | 33.6 | 0.240 | 0.2419 |
| Milk urea Nitrogen (g kg ⁻¹) | 18.8 ^a | 16.5 ^a | 14.7 ^b | 2.070 | 0.0021 |
| Milk dry matter (g kg ⁻¹) | 109.2 ^a | 101.2 ^b | 114.6 ^a | 2.420 | 0.0174 |
| Plasma glucose (mg dL ⁻¹) | 61.9 | 56.6 | 61.3 | 2.920 | 0.4430 |
| Plasma urea nitrogen (mg dL ⁻¹) | 19.6 | 17.6 | 19.7 | 1.160 | 0.1985 |

^{a,b}: In each row, difference between means with different letter were significant (p<0.05), ¹AS = untreated Alfalfa Silage, ASF = Alfalfa Silage treated with 24 mL Formic acid kg⁻¹ DM, ASFU = Alfalfa Silage treated with 24 mL Formic acid and 4 g Urea kg⁻¹ DM, ²SEM = Standard Error of Mean

ensiling (Ohshima and McDonald, 1977). The formic acid treated silages had lower energy, cellulose, lignin and protein but higher sugar concentrations than the untreated silages. In addition, the silages that were treated with formic acid had lower pH (p<0.001), lactic acid, acetic acid, butyric acid, total acids and ammonia nitrogen than the untreated silages (Derbyshire *et al.*, 1976).

Ruminal and post-ruminal dry matter and protein disappearance of the alfalfa silages are shown in Table 2. The ruminal dry matter disappearance was lower in the silages treated with formic acid and urea than the untreated silage but the post-ruminal dry matter disappearance in the control silage was lower than the other silages. Ruminal and post-ruminal protein disappearance in all silages were similar. Nadeau *et al.* (1996) reported greater total DM and NDF disappearances during early ruminal fermentation *in situ* in cellulase plus formic acid treated orchard grass and alfalfa silages compared with the control silage but the differences between treatments became smaller as the fermentation proceeded up to 96 h of incubation. Results of dry matter intake, milk yield and milk composition and plasma blood metabolites are shown in Table 3. Milk yield and milk protein were not significantly affected by the treatments

(p>0.05). Experimental diets had a significant effect on milk urea nitrogen concentration and milk dry matter (p<0.05). Lower concentrations of milk urea nitrogen in cows fed the diet containing ASF might be reflecting of a difference in CP component intake (Broderick and Clayton, 1997). Blood glucose and urea nitrogen were all similar among the diets at each sampling time (p>0.05). However, blood glucose was numerically higher in animals fed the treated silage compared with those were fed the control silage.

CONCLUSION

In conclusion, results of the of present study demonstrate that formic acid treatment of the alfalfa silage is effective procedure in reducing N degradation in the silo and in the rumen and has not negative effect on milk production. In addition, the formic acid treated silage resulted in a significant shift in the disappearance site of dry matter and CP as post-ruminal digestion was greater in the chemically treated AS compared with the untreated. However, additional research must be done to develop an accurate on-farm test to predict when formic acid application is appropriate and what application rate is optimal.

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