Quantitative Trait Gene Responsible for Intramuscular Fat Content in the Rat

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Abstract: Marbling defined by the amount and distribution of intramuscular fat, so-called Shimofuri is an economically important trait of beef cattle in Japan. A rat Quantitative Trait Locus (QTL) responsible for intramuscular fat content has been mapped to the \( \sim 10 \)-cM genomic region on chromosome 1 by utilizing the Otsuka Long-Evans Tokushima Fatty (OLETF) rat and OLETF-derived congenic strain. In this study, we have attempted to identify the Quantitative Trait Gene (QTG) for the intramuscular fat content QTL by using the OLETF rat. For this purpose, we performed refinement of the intramuscular fat content QTL genomic region to a \( \sim 2.3 \)-cM region by use of informative recombinants selected from the OLETF-derived congenic strain, functional retrieval of positional candidate gene, mRNA level analysis and polymorphism search of physiologically relevant positional candidate gene, Pnilp and analysis of cosegregation of the intramuscular fat content QTL with Variable Number of Tandem Repeat (VNTR) in Pnilp by use of informative recombinants. This study suggests that Pnilp encoding pancreatic lipase is the QTG for the intramuscular fat content QTL.

Key words: Candidate gene, intramuscular fat, marbling, QTL, QTL, rat

INTRODUCTION

Generally, marbling means the amount of intramuscular fat (Cameron et al., 1994). In Japan, marbling is characterized as the amount and distribution of intramuscular fat in a cross section of Musculus longissimus muscle and is called Shimofuri. High levels of such marbling improve the palatability and acceptability of beef by affecting the taste and tenderness of the meat (Busboom et al., 1993; Boylston et al., 1995). Because of the importance of marbling on the economics of beef production, there is great interest in gaining a better understanding of the molecular architecture of marbling that is intramuscular fat content.

The Otsuka Long-Evans Tokushima Fatty (OLETF) rat, an animal model for obese type 2 diabetes, exhibits higher levels of intramuscular fat content in Musculus longissimus than the F344 rat (Tanomura et al., 2002). We previously reported that a genomic region between D1Wox8 and D1Rat90 on chromosome 1 influences intramuscular fat content with the OLETF allele acting on an increase in fat content by Quantitative Trait Locus (QTL) analysis using \( F_2 \) progenies derived from the OLETF and F344 rats (Tanomura et al., 2002). Further, we demonstrated that the QTL on chromosome 1 responsible for intramuscular fat content is located in the \( \sim 10 \)-cM genomic region between D1Rat166 and D1Rat90 using a congenic strain trapping the OLETF allele of the \( \sim 2.3 \)-cM region on the F344 genetic background in a monogenic context (Tanomura et al., 2003).

The present study has attempted to identify the Quantitative Trait Gene (QTG) for the intramuscular fat content QTL by utilizing the OLETF rat. For this purpose, we performed refinement of the intramuscular fat content QTL genomic region to a \( \sim 2.3 \)-cM region by use of informative recombinants selected from the congenic strain, functional retrieval of positional candidate gene, mRNA level analysis and polymorphism search of physiologically relevant positional candidate gene, Pnilp and analysis of cosegregation of the intramuscular fat content QTL with Variable Number of Tandem Repeat (VNTR) in Pnilp by use of informative recombinants. This study suggests that Pnilp encoding pancreatic lipase is the QTG for the intramuscular fat content QTL.

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MATERIALS AND METHODS

Animals: The researchers used the inbred strains of OLETF (OLETF/Ole) (Kawano et al., 1992), normal F344 (F344/Cj) and normal BN rats (BN/F1). We also used the congenic strain constructed by introgressing the OLETF allele on the ~10-cM intramuscular fat content QTL region between D1Rat66 and D1Rat90 into the normal F344 rat strain (Tanomura et al., 2003). Furthermore, researchers constructed informative recombinants in which a part of the intramuscular fat content QTL region between D1Rat66 and D1Rat90 in the congenic strain was replaced by the F344 background genome, to refine the intramuscular fat content QTL region. For breeding of informative recombinants, the F2 rats were generated by female congenic rats bred with male F344 rats and male backcross progenies were obtained by mating the female F1 with male congenic rats.

While the male OLETF rat possesses ~2.4-fold higher levels of intramuscular fat content in Musculus longissimus than male F344 rat, no difference in the levels is detected between OLETF and F344 females (Tanomura et al., 2002). Thus, using males was assumed to facilitate exclusively the detection of the quantitative difference of intramuscular fat content. All rats were kept under specific pathogen-free conditions. The temperature (21±2°C), humidity (55±5%) and air conditioning were all controlled.

Rats had free access to tap water and standard laboratory chow (CRF-1; Oriental Yeast, Tokyo, Japan) and were maintained on a 12 h light and dark cycle (6 a.m./6 p.m.). This study conformed to the guidelines for animal experimentation of the Graduate School of Science and Technology, Niigata University (Niigata, Japan).

Genotyping: Male rat tails were cut and frozen at -80°C for subsequent isolation of high-molecular-weight DNA. Genotyping of D1Rat166 and D1Rat90 with tail biopsy DNA of male backcross progenies was performed as described previously (Tanomura et al., 2003), to select male backcross progenies which harbored heterozygote at either one of D1Rat166 or D1Rat90 and homozygote at the other as informative recombinants (Table 1). Subsequently, genotyping of D1Rat225 located on a genomic interval between D1Rat66 and D1Rat90 (Wei et al., 1998) was performed for the informative recombinants by PCR with primers purchased from Research Genetics (Huntsville, AL) according to a previous report (Wei et al., 1998) to determine a refined region of the intramuscular fat content QTL (Table 1).

Phenotype evaluation: Male rats were sacrificed under deep anesthesia at 35 weeks of age. The body weight was measured before the sacrifice of rats. Immediately after exsanguination, the abdominal fat pads consisting of mesenteric, retroperitoneal and epididymal fat pads were removed and weighed. Furthermore, the first to third lumbar region of Musculus longissimus was removed from the right side of the carcass and rapidly frozen in dry ice-cooled isopentane. Five serial coronal sections (6 μm thick) were obtained at intervals of 180 μm from frozen Musculus longissimus and the sections were stained with Oil Red O to detect the presence of lipid droplets in adipocyte cytoplasm. The section area, occupied by cells positive for Oil Red O staining were measured in a given area (1 cm²) using the MacSCOPE computer program. The fat-staining-positive area expressed as an absolute value was used as the phenotypic value of the level of intramuscular fat content (Tanomura et al., 2003).

Pulip mRNA level analysis: The pancreases were removed after overnight starvation from male rats at 15 and 35 weeks of age and frozen at -80°C. Total RNAs were prepared from the pancreases using ISOGEN (Wako Pure Chemical Industries, Osaka, Japan). First-strand cDNA was reverse-transcribed from total RNA sample (1 μg) using RNA PCR Kit ver. 2.1 (Takara Bio, Shiga, Japan). Competitive PCR was performed for Pulip mRNA level analysis using reverse-transcribed reaction mixture as described previously (Shikamoto and Morita, 1999). PCR primers used in competitive PCR were designed from the rat cDNA sequences of Pulip (GenBank accession number NM_013161) and glyceraldehyde-3-phosphate dehydrogenase gene (Gapdh) (GenBank accession number NM_017008). Primer sequences for Pulip were 5'-ATACACCCAGGCTACCCA GAAC-3' (nucleotide positions 419-440 of NM_013161) and 5'-CCAATAGCTCGAATGTGCTCT-3' (nucleotide positions 589-568 of NM_013161). Primer sequences for Gapdh were 5'-TCCTCGACCAACCAAG TGCTTA-3' (nucleotide positions 1294-1314 of NM_017008) and 5'-ACCACCCGTGTGGCTAGCCA-3' (nucleotide positions 880-858 of NM_017008).
positions 1817-1797 of NM_017008). DNA competitors used in competitive PCR were generated by PCR with PCR composite primers which were engineered to contain the competitor template-specific sequences with the target gene-specific primer sequences, using Competitive DNA Construction Kit (Takara Bio). Sequences of PCR composite primers for Pnlp polymorphism were 5'-ATACACCGCGACTCCACACGCAGATGCAGACGACAC-3' (nucleotide positions 419-440 of NM_017008 plus 1-20 of competitor template) and 5'-CCAATAGCTCCGAAATGTCCCTCTTCAATTACGCCATCGCTATTAC-3' (nucleotide positions 589-568 of NM_017008 plus 200-181 of competitor template). Sequences of PCR composite primers for Gapdh competitor were 5'-TCCTGCACCAACCTGCTTATGTGCTGCTGACAC-3' (nucleotide positions 1294-1314 of NM_017008 plus 1-20 of competitor template) and 5'-ACCACCTCTGCGTGCTGACCCAGGCTGAGATTGAAAGTG-3' (nucleotide positions 1817-1797 of NM_017008 plus 400-381 of competitor template). Concentration of DNA competitors was measured by spectrophotometry (A260) and subsequently adjusted to 1 x 10^4 copies/µL. A series of competitive PCR was performed with 0.5 µM of each PCR primer, DNA competitor (10 fold serial dilution from 10^-1 to 10^-5 copies/reaction) and 0.1 µL reverse-transcribed reaction mixture, using RNA PCR Kit ver. 2.1 (Takara Bio).

A thermocycling protocol was 30 cycles of denaturation for 45 sec at 94°C, annealing for 45 sec at 60°C and elongation for 45 sec at 72°C. PCR products were separated by electrophoresis on a 3.0% Metaphor agarose gel (Takara Bio). Agarose gels were stained with ethidium bromide and photographed under an ultraviolet light. Digital images of photographs were imported into the Scion Image computerized densitometry program (Scion, Frederick, MD).

cDNA copy numbers of Pnlp and Gapdh were calculated according to the method previously reported (Shikamoto and Morita, 1999). Briefly, intensity ratio of target DNA bands to corresponding competitor DNA bands was plotted on a logarithmic scale against copy number of competitors and then an equivalence point (log ratio = 0) was calculated from a linear regression standard curve. Results of competitive PCR for Gapdh were used for normalization of cDNA amounts subjected to competitive PCR for Pnlp.

Pnlp polymorphism search: Pnlp polymorphisms between the F344 and OLETF rats were searched by the direct sequencing. About 95 pairs of PCR primers were designed to cover -25.6 kb Pnlp genomic sequences (nucleotide positions 1788220-1814849 of GenBank accession number NW_047570) which included the 5'-flanking region (-9.8 kb), the exon/intron region (-13.1 kb) and the 3'-flanking region (-3.6 kb). PCR was carried out with 0.5 µM of each PCR primer and genomic DNA (100 ng) isolated from tails of the F344 and OLETF rats, using TaKaRa Ex Taq (Takara Bio). A thermocycling protocol was the same as described in the mRNA level analysis section. DNA sequencing of PCR products was performed by the direct sequencing with capillary DNA sequencer RISA-384 system in Shimadzu Corporation (Kyoto, Japan). GENETYX-WIN3.1 program (Software Development, Tokyo, Japan) was used for analysis of DNA sequence.

Genotyping of Pnlp VNTR: Pnlp VNTR was genotyped with tail biopsy DNA by PCR amplification of a ~2.8 kb genomic segment which contains tandem repeats of ~38-bp core sequence between the bases 5517 and 2698 upstream from the transcription start site of Pnlp (nucleotide positions 1792587-1795406 of NW_047570). Primer sequences were 5'-TGTCAGAGCAGCAGAGAGGCTGAC-3' (nucleotide positions 1792587-1792608 of NW_047570) and 5'-CACAGTGCTGCTTGTTGGA-3' (nucleotide positions 1795406-1795587 of NW_047570). PCR was carried out as described in the polymorphism search section.

A thermocycling protocol was 30 cycles of denaturation for 60 sec at 94°C, annealing for 60 sec at 60°C and elongation for 180 sec at 72°C. PCR products were electrophoresed on a 1.0% agarose gel (Takara Bio).

Statistical analysis: Comparisons of phenotypes and Pnlp mRNA levels among different genotypic groups were performed by an Analysis of Variance (ANOVA) with a post hoc test using Scheffe’s F test (StatView; SAS Institute, Cary, NC).

RESULTS AND DISCUSSION

Phenotype evaluation in congenic and (congenic x F344) F1 rats: The level of intramuscular fat content was increased and decreased in the congenic rat as compared with the F344 and OLETF rats, respectively and both the differences in the changes were statistically significant (p<0.05) (Fig. 1). This result of an increase in the level of intramuscular fat content of the congenic strain over the F344 rat was consistent with the previous finding (Tanomura et al., 2003). Additionally, the level in (congenic female x F344 male) F1 rat was intermediate between those of the F344 and the congenic rats and all the differences between the F1 and the F344 or congenic were statistically significant (p<0.05) (Fig. 1). This result indicates that the OLETF allele at the intramuscular fat
Functional retrieval of positional candidate gene: About 46 genes were detected within the ~2.3-cM intramuscular fat content QTL region on the rat genome (rat genome resources). Functional retrieval of the 46 genes revealed that Pulpip which encodes pancreatic lipase involved in energy income and fat regulation was regarded as the most prominent and physiologically relevant positional candidate gene for the intramuscular fat content QTL. Pulpip is composed of 13 exons spanning >13-kb and encodes a 56-kD protein of 465 amino acid residues (Sims et al., 1993).

The hydrolysis of dietary triglycerides to fatty acids by pancreatic lipase encoded by Pulpip is essential for the intestinal absorption of long-chain triglyceride fatty acids (Lowe, 2002), indicating an involvement of pancreatic lipase in energy income. Further, the treatment with Orlistat that blocks predominantly pancreatic lipase activity in the small intestine promotes significant fat loss and prevents fat regain in obese patients (Sjoström et al., 1998; Hill et al., 1999) indicating an involvement of pancreatic lipase in fat regulation. Thus, an increase in pancreatic lipase amounts is likely to result in excess energy income and then increase of intramuscular fat content.

Pulpip mRNA level analysis: Pulpip mRNA levels in the pancreas of the congenic rat (2.81±0.21 for 15 weeks of age and 3.03±0.43 for 35 weeks of age) were similar to those of the OLETF rat (2.98±0.40 for 15 weeks of age and 3.13±0.31 for 35 weeks of age) at both 15 and 35 weeks of age (p<0.05). The (congenic x F344) F1 rat (1.92±0.20 for 15 weeks of age and 1.85±0.19 for 35 weeks of age) exhibited an intermediate value between the F344 and congenic rats in the pancreatic Pulpip mRNA levels at both 15 and 35 weeks of age and all the differences between the F1 and the F344 or congenic were statistically significant (p<0.05). Further, the normal BN rat showed the similar pancreatic Pulpip mRNA levels to the F344 but their levels were ~3-fold lower as compared to those of the congenic rat (data not shown). These results indicate that Pulpip possesses an OLETF allele-specific increase of mRNA levels in the pancreas in an additive manner, consistent with the result of phenotype evaluation in the F344, congenic and F1 rats. These suggest that the increased mRNA levels of Pulpip may cause the increase of intramuscular fat content in the OLETF rat.

Pulpip polymorphism search: We could not find any polymorphisms between the F344 and OLETF rats in the exon/intron and 3'-flanking regions of Pulpip by the direct
sequencing. Additionally, no polymorphisms were detected between the F344 and OLETF rats in the Pnlip 5′-flanking region, except for a ~2.8-kb genomic segment which contains tandem repeats of ~38-bp core sequence, between the bases 5517 and 2698 upstream from the transcription start site of Pnlip (nucleotide positions 1792587-1795406 of NW_047570). PCR amplification of the ~2.8-kb genomic segment identified approximately 2.8-kb PCR product in the F344 rat but about 3.0-kb PCR product in the OLETF and congenic rats. It is likely that the PCR-product length polymorphism is attributable to a variation in the number of repeats of core sequence. Further, the normal BN rat exhibited ~2.8-kb PCR amplification product similar to the F344 rat. These results indicate that the Pnlip OLETF allele is longer in VNTR within the 5′-flanking region than the normal alleles.

A computer scan of a ~2.8 kb genomic segment (nucleotide positions 1792587-1795406 of NW_047570) containing tandem repeats of core sequence with MOTIF program disclosed dozens of putative binding sites for known transcription factors in the tandem repeat region. Of the notables were binding sites for API and USF1 which are highly expressed and seem to exert a function in the pancreas.

It has been reported that the longer variants of VNTR in the regulatory region of serotonin transporter gene were associated with enhanced mRNA levels of serotonin transporter gene (Lesch et al., 1996). Additionally, the class III longer alleles (140-200 repeats) of VNTR in the regulatory region of insulin gene (INS) were associated with 2-3 fold higher INS mRNA levels as compared with the class I shorter alleles (23-63 repeats) of which transmission predisposes to type 1 diabetes in human (Vafadis et al., 1997; Pagliese et al., 1997). The tandem repeat region of INS has been reported to also contain binding sites for the transcription factor PURA and to regulate INS transcription (Kennedy et al., 1995). These suggest that the longer OLETF allele in Pnlip VNTR may be associated with the increased Pnlip mRNA levels in the OLETF rat.

**Cosegregation of the intramuscular fat content QTL with Pnlip VNTR:** We performed genotyping of Pnlip VNTR for the 12 informative recombinants selected from the congenic strain. The genotyping allowed the 12 informative recombinants to be classified into two types, Pnlip VNTR OLETF homozygote (O/O, n = 7) and Pnlip VNTR heterozygote (O/F, n = 5). Allele segregation pattern of the intramuscular fat content QTL in the 12 informative recombinants was completely coincident with that of Pnlip VNTR indicating complete cosegregation of the intramuscular fat content QTL with Pnlip VNTR. This suggests that Pnlip is the QTL for the intramuscular fat content QTL and that the longer allele of Pnlip VNTR may be a causal mutation for phenotype of increase of intramuscular fat content in the OLETF rat. Taken together, the present study might support a hypothesis that the longer allele of Pnlip VNTR causes phenotype of increase of intramuscular fat content through increasing Pnlip mRNA levels in the OLETF rat. There is an evidence for a linkage of marbling trait to genomic region containing PNLIP on bovine chromosome 26 (Takasuga et al., 2007). Based on the present results, it will be important to use PNLIP as candidate gene for marbling trait in bovine association study.

**CONCLUSION**

In this study, we suggests that Pnlip encoding pancreatic lipase is the QTL responsible for the intramuscular fat content in the rat. This study will provide an useful information for the understanding of the molecular architecture of marbling that is an economically important trait of beef cattle.

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