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Anti-Oxidant and Anti Tyrosinase Effect of *Ziziphus spina-christi* Seed Extract

¹Ibrahim Hassan Maina, ²Keneth Idowu Onifade, ¹Bashir Sa'idu, ³Adamu Abdul Abubakar, ¹Sadiq Habibullah, ¹Abdullahi Ishaq Ja'afar, ¹Yahaya Abubakar Yabo and ¹Abdurahman Jibreen Hassan

¹Department of Physiology and Biochemistry,

²Department of Pharmacology and Toxicology,

³Department of Surgery and Radiology, Faculty of Veterinary Medicine, Usmanu Danfodiyo University, P.M.B. 2346, Sokoto, Nigeria

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Corresponding Author:

Hassan Ibrahim Maina

Department of Physiology and Biochemistry,

Faculty of Veterinary Medicine,

Usmanu Danfodiyo University, P.M.B. 2346, Sokoto, Nigeria

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Abstract: Medicinal plant such as *Ziziphus Spina-Christi* (ZSC) has been used for the treatment and prevention of several ailments in human and animals. It is recommended for the management of diseases in which free radical species are produced as a result of oxidative stress. However, there is lack of systematic study on the antioxidant and antityrosinase capacities of ZSCF from Nigeria. The present study quantifies the anti-oxidant and anti tyrosinaseability of the ZSCF grown in the Gwaski, Southern Borno state Nigeria. Antioxidant activity was assessed by using 2, 2-Diphenyl-1-Picrylhydrazyl (DPPH), Superoxide-Radical Scavenging Activity (SRSA), Ferric Reducing/Antioxidant Power (FRAP assay). Overall the ZSCF showed strong antioxidant ability and high anti tyrosinase effect. There is variation in the oxidation power (DPPH) with 15.5% at the concentration of $7.8 \mu\text{g mL}^{-1}$ -88.4% at $1000 \mu\text{g mL}^{-1}$ and 22.5% at $7.8 \mu\text{g mL}^{-1}$ as well as 63.5% at $1000 \mu\text{g mL}^{-1}$ for FRAP assay. The percentage antityrosinase activities varied from 4.1% at $7.8 \mu\text{g mL}^{-1}$ -70.5% at $1000 \mu\text{g mL}^{-1}$ for plant crude extract. The observed antioxidant and anti tyrosinase ability of ZSCF may be due to abundant presence of phenolic contents and high electron donating ability to neutralize free radicals.

INTRODUCTION

Ziziphus spina-christi ordinarily known as Christ's Thorn Jujube is a persistence tree commonly found in warm temperate zone, subtropical region including North and West Africa, South Europe, Mediterranean, Australia, tropical America, South and East of Asia and middle East^[1,2]. It is member of Rhamnaceae family in the order

of Rosales that contains up to 60 genera and >850 species^[3]. There are about 100 species of deciduous or evergreen trees and shrubs under the genus *Ziziphus* all over the world^[4,5]. The tree and its parts have been used in Pharaonic industry (carpentry) as well as dietary supplement, antioxidant and in folk medicine as a demulcent, depurative, anodyne, emollient, stomach-ache for toothaches, astringents and as a mouth wash^[6,7].

Scientific finding reveal that some plants contained peptides, unsaturated long chain aldehydes, alkaloidal constituents, some essential oils, phenols and water, ethanol, chloroform, methanol and butanol soluble compounds^[8-10]. Some works also, shows that the plant contain betulinic and ceanothic acid, three cyclopeptide alkaloids as well as four saponin glycosides^[11, 12] and several flavonoids have been isolated from the leaves of *Z. spina-christi*^[13, 14]. The present study aimed to evaluate the antioxidant and antityrosinase effect of 80% methanolic extracts of *Z. spina-christi* seed.

MATERIALS AND METHODS

Plant

Plant collection and identification: *Ziziphus spina-christi* seeds were collected from Gwaski, Hawaul LGA Borno state and identified at Department of Botany, Faculty of science, University of Maiduguri. Seed were washed with clean water to separate it from the white membranes and allowed to dry room temperature for four weeks and ground to semi-powder form.

Plant extraction: Plant seed was grounded into semi powder (40-60 mesh) with a blender. A total of 70 g of samples was immersed for 3 days subsequently in 1000 mL of 80% methanol in 2.5 L flat bottom flask. The sample was shaking ceaselessly for 3 days at 25°C to acquire 80% methanolic extract. The extract obtained was then filtered with what-man filter paper after which was concentrated to semi-solid form with a rotary evaporator at the temperature of 42°C. Crude extracts obtained were then weighed, transferred to the sample bottle and stored at 4°C.

Plant sample dilution and dose preparation: A stock solution (100 mg mL⁻¹) was prepared by dissolving (100 mg) in 1 mL of 100% DMSO. Sub-stocks were prepared in a microliter (µg mL⁻¹) using the formulae $M_1V_1 = M_2V_2$ and diluted to the concentration of interest by twofold serial dilution using 100% methanol at eight concentrations ranging from 7.81-1000 µg mL⁻¹ prepared in a 96-well microplate. DMSO (vehicle) was maintained at 0.1% in all concentration of samples.

Antioxidant assay

DPPH radical-scavenging activity assay: The free radical scavenging activity of sample extracts was evaluated by 1, 1-Diphenyl-2-Picryl-Hydrazyl (DPPH) according to the method previously reported by Shen etc. with minor modifications. Briefly, 0.1 mM solution of DPPH in methanol was prepared and 1 mL of this solution was added to 3 mL of methanol and then exposed to different concentration plant extract (7.81-1000 µg mL⁻¹). DPPH reagent (270 µL) and sample solutions (30 µL) were mixed in 96-well plates shake vigorously and allowed to stand at room temperature for

30 min. Then the absorbance was measured at 517 nm using a UV-VIs spectrophotometer. Trolox and quercetin were used as the reference. Lower absorbance values of reaction mixture indicate higher free radicalscavenging activity. The capability of scavenging the DPPH radical was calculated by using the following equation:

$$\text{DPPH scavenging effect (inhibition\%)} = \frac{A_0 - A_1}{A_0} \times 100$$

Where:

A₀ : The absorbance of the control reaction

A₁ : The absorbance in the presence of all of the extract samples and reference

All the tests were performed in triplicates and the results were averaged.

Ferric Reducing/Antioxidant Power (FRAP) assay:

The FRAP assay was carried out according to the previously reported procedure by Wojdylo with slight modifications. Sample extracts were dissolved in 0.1% DMSO-methanol (v/v) to prepare the stock solution (1 mg mL⁻¹). Briefly, the working FRAP reagent was prepared by mixing 300 mM acetate buffer (pH 3.6), a solution of 10 mM 2, 4, 6-Tripyridyl-s-Triazine (TPTZ) in 40 mM hydrochloric acid and 20 mM ferric chloride at 10:1:1 (v/v/v). The working FRAP reagent (270 µL) and sample solutions (30 µL) were mixed in 96-well plates and warmed at 37°C in constant temperature oven for 5 min. The absorbance was taken at 593 nm. Trolox and quercetin were used as a standard. All determinations were performed in triplicate:

$$\text{FRAP scavenging activity (\%)} = \frac{(\text{Abs}_{\text{control sample}}) - \text{Abs}_{\text{control}}}{\text{Abs}_{\text{control}}} \times 100$$

Where:

Abs_{control} : The absorbance of FRAP+methanol

Abs_{sample} : Absorbance FRAP+Sample extract/standard

L-DOPA and mushroom tyrosinase were purchased from Sigma Chemical. The 20 µL of mushroom tyrosinase (1000 U mL⁻¹), 20 µL of 0.1 M phosphate buffer (pH 6.8) and 100 µL of the test sample solution (20%) containing 20 µL of plant extracts were mixed (called sample solution with enzyme). Sample solutions without enzyme were also, prepared by repeating all previous steps but with no plant extracts added. Blank solutions with and without enzyme were also prepared with no test sample solution added. We also prepared positive controls of 0.5 mg mL⁻¹ kojic acid solutions (with water) with and without enzyme. The 20 µL of 0.85 mM L-DOPA solution as the substrate was added into every sample and blank. These assay mixtures were incubated at 25°C for 10 min. The amount of dopachrome

produced in the reaction mixture was measured at 475 nm ($\epsilon_{475} = 3600/\text{m}/\text{cm}$) using microplate reader (Zenyth 2000, Anthos Labtech Instrument). Percent inhibition of tyrosinase activity was calculated as the following: % tyrosinase:

$$\text{Inhibition} = \frac{(A-B)-(C-D) \times 100}{(A-B)}$$

Where:

- A : Absorbance of blank solution with enzyme
- B : Absorbance of blank solution without enzyme
- C : Absorbance of sample solution with enzyme
- D : Absorbance of sample solution without enzyme

RESULTS AND DISCUSSION

Present of phenolics in the fruits and vegetables have been extensively studied due to their potential biological activities. Phenolic compounds such as flavonoids, phenolics acid and tannins possess diverse biological activities including anti-inflammatory, anti-carcinogenic and antiatherosclerotic activities. These activities might be related to their antioxidant activity.

Yield of 35.2 g was obtained following extraction of 100 g seed samples with 80% methanol and concentrated to semisolid form with a rotary evaporator at a temperature of 42°C (Table 1). Percentage scavenging of *Zizyphus spina-christi* (seed) on DPPH with different concentration 7.8-10000 $\mu\text{g mL}^{-1}$ of plant crude extract as compared with Quercetin and Trolox (reference). Percentage of inhibition (mean \pm SD) (n = 3) is shown versus concentration of the tested sample. The antioxidant activity of *Zizyphus spina-christi* seed extract showed variation in the oxidation power with 15.5% at the concentration of 7.8 $\mu\text{g mL}^{-1}$ -88.4% at 1000 $\mu\text{g mL}^{-1}$ for *Zizyphus spina-christi* seed extract. While the percentage scavenging power for trolox varied from 56.4% at the concentration of 7.8 $\mu\text{g mL}^{-1}$ -96.5% at 1000 $\mu\text{g mL}^{-1}$. For quercetin the percentage scavenging power varied from 89.1% at 7.8 $\mu\text{g mL}^{-1}$ -100% at 1000 $\mu\text{g mL}^{-1}$. The largest capacity to neutralize DPPH radicals was found at 1000 $\mu\text{g mL}^{-1}$ for both *Zizyphus spina-christi* seed extract, Trolox and quercetin (Fig. 1).

Percentage inhibition of *Zizyphus spina-christi* (seed) on FRAP revealed differences in activities at concentration range 7.8-1000 $\mu\text{g mL}^{-1}$ of plant crude extract compared with Trolox and Quercetin (reference). Percentage of inhibition (mean \pm SD) (n = 3) is shown versus concentration of the tested sample. The antioxidant activities values varied from 22.5% at the concentration 7.8 $\mu\text{g mL}^{-1}$ -63.5% at 1000 $\mu\text{g mL}^{-1}$ for *Zizyphus spina-christi*. For Trolox the percentage inhibition activities

vary from 46.3% at 7.8 $\mu\text{g mL}^{-1}$ -100% at 91.5 $\mu\text{g mL}^{-1}$. Percentage antioxidant power of quercetin differed from 79.1% at 7.8 $\mu\text{g mL}^{-1}$ -100% at 1000 $\mu\text{g mL}^{-1}$. The largest capacity to inhibit FRAP was found at 1000 $\mu\text{g mL}^{-1}$ (Fig. 2).

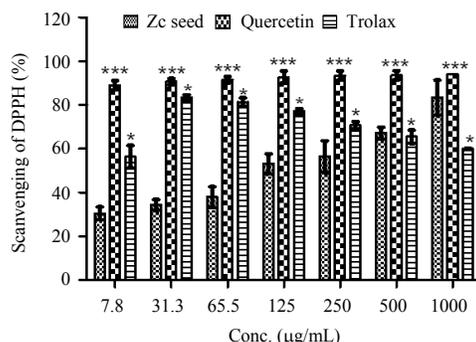


Fig. 1: Antioxidant activities of *Zizyphus spina-christi* crude extracts (2 mg mL⁻¹), quercetin and trolox (500 $\mu\text{g mL}^{-1}$). Free radical scavenging inhibitory activities was measured using DPPH assay. Absorbance was taken at 517 nm with quercetin and trolox as standards. ***p<0.0001 represented significantly different values from standards and tested plants. The values represent mean \pm SD from three independent experiments

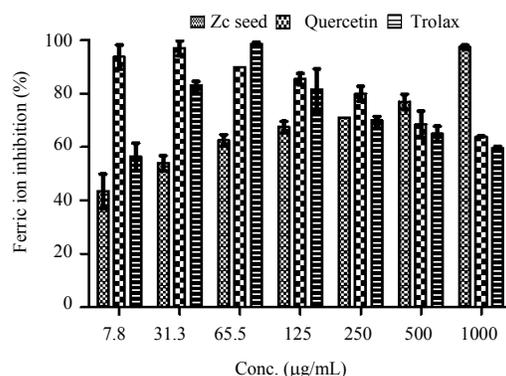


Fig. 2: Antioxidant activities of *Zizyphus spina-christi* crude extracts (2 mg mL⁻¹), quercetin and trolox (500 $\mu\text{g mL}^{-1}$). Free radical scavenging inhibitory activities was measured using DPPH assay. Absorbance was taken at 517 nm with quercetin and trolox as standards. There is no significant different value from standards and tested plants. The values represent mean \pm SD from three independent experiments

Table 1: Result of seed samples

Plant's	Part	Weight of samples in powder form (g)	Weight of samples crude extract (g)	Percentage yield/100 g (DW)
<i>Zizyphus spina-christi</i>	Seed	100	67.1	39.74

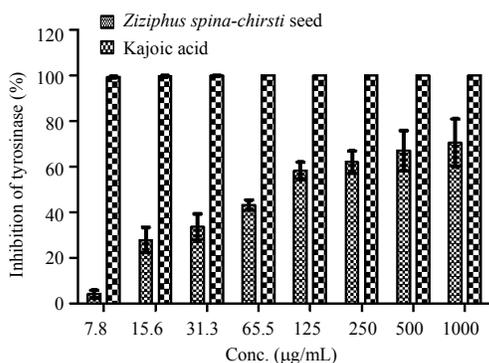


Fig. 3: Antityrosinase activities of *Ziziphus spina-christi* crude extracts (1000 µg mL⁻¹), kajoic acid (500 µg mL⁻¹). A tyrosinase inhibitory activity was measured using and the absorbance was taken at 517 nm with kajoic acid as standards

Percentage inhibition of *Zizyphus spina-christi* (seed) on tyrosinase shows variation in activities with increased concentration 7.8-1000 µg mL⁻¹ of plant crude extract as compared to Kajoic acid (reference). Percentage of inhibition (mean±SD) (n = 3) is shown versus concentration of the tested sample. The percentage antityrosinase activities varied from 4.1% at 7.8 µg mL⁻¹ -70.5% at 1000 µg mL⁻¹ for plant crude extract. Percentage kajoic acid activities was observed to be 99.3% at 7.8 µg mL⁻¹ to 100% at 1000 µg mL⁻¹. The largest capacity to inhibit tyrosinase was found at 1000 µg mL⁻¹ for both plant extracts and Kajoic acid (Fig. 3).

Phytochemicals including phenolic compounds, present in many herbs have received much attention in recent years due to their many health benefits including antioxidant and anti-inflammatory activities^[15,16]. For this reason there are interests in using some herbs not only as culinary products but also as a supplement to functional products^[17-19]. Therefore, the study on the stability of some bioactive compounds in the plant material during some processing (drying, freezing, etc.) or preparation of plant extracts would be useful in the selection of the bioactive compounds extraction procedure^[20-22]. For extracting phenolic compounds typically from plant material previously used solvents are methanol, ethanol, acetone and ethyl acetate^[23-25]. In the present study for extracting phenolic compounds from the seed 80% methanol was used.

Antioxidants are exceptionally leading substances which influence the proficiency to defense the body system from detrimental effect ROS including superoxide anion, free radical, hydroxyl radical, hydrogen peroxide, nitrogen dioxide, nitric oxide radical induced by oxidative stress^[26-28]. ROS can stimulate the production of melanin pigment cells and promote the generation of

hyper pigmentation. The DPPH radical scavenging measure is a valuable technique for antioxidant evaluation^[29,30]. Neutralizing free radicals can repress the production of melanin in skin and other tissue which result in skin whitening.

Tyrosinase inhibition is desired as tyrosinase catalyzes the oxidation of phenolic compounds present in fruits and vegetables into quinone which gives an undesirable taste and color and also decreases the availability of certain essential amino acids as well as the digestibility of the products^[31-33]. As such highly effective tyrosinase inhibitors are also needed by the body system^[34-36]. The antioxidant and tyrosinase inhibition potential of *Z. spina-christi* 80% methanol extracts was investigated in the search for new bioactive compounds from natural resources. It has clearly shown that *Z. spina-christi* seed extract present high antioxidant and tyrosinase inhibition activities compared with reference quercetin and trolox for DPPH and FRAP assay as well as kajoic acid for anti tyrosinase assay. Previous studies reported that DPPH radical scavenging activity of fruit and seed extract showed good free radical scavenging activity when compared with ascorbic acid^[37-39]. Presences of poly phenols such as flavonoid might be responsible for the antioxidant activities of the extract^[40-42]. Scavenging of the free radical might be the major mode of action of this plant extract when use for the treatment of diseases^[43]. Component with antioxidants activities are of interest to biologists and clinicians because they help in protecting human body against damages induced by reactive free radicals generated in atherosclerosis, ischemic heart disease, cancer, Alzheimer's disease, Parkinson's disease and even in aging process^[44]. Scientific finding has reveal that most of the natural products and their derivatives have efficient anti-oxidative characteristics, consequently linked to anti-cancer, hypolipidemic, anti-aging and anti-inflammatory activities^[45,46].

In human, skin it is the largest organ of the integument system. It covers the whole surface of the human body, representing around 15% of the body weight^[47]. It connect the body with the outer environment, protect the basic structures against scraped spot and lack of hydration and assumes an essential part in invulnerability and other body defense mechanisms^[48]. Intensity and skin pigmentation varies with environmental, state of origin, climate and gender. Individual skin color may varies with time or when exposed to climatic or environmental changes^[49]. Thickness and melanin contained of the skin may alter hemoglobin and melanocytes resulting in minor changes in pigmentations such as carotenoids affect the perceived color and determination of skin color. Therefore, melanin synthesis is very vital in determining the skin functions and color formation^[50]. The key enzyme that catalysis the synthesis of melanin and intervenes in

several intermediate stages of pigment formation is tyrosinase. Report shows that present of bioactive compounds such as (collagen gels in *Aesculus hippocastanum*) (Aloesin [2-acetonyl-8-beta-dglucopyranosyl-7-hydroxy-5-methylchromone], Tyrosine hydroxylase and 3, 4-dihydroxyphenylalanine oxidase in *Aloe vera*) (polysaccharides, flavonoids, hyaluronan synthase-3 and hyaluronan synthase-2 in *Astragalus membranaceus*), (polyphenols catechin, epigallocatechin, epigallocatechin-3-gallate in *Camellia sinensis*) (triterpenoids, saponins, madecassoside, asiaticoside, centelloside and asiatic acid in *Centella asiatica*) (phenolic compounds such as anthocyanins, flavanones, hydroxycinnamic acids and ascorbic acid in *Citrus sinensis*) competitively inhibit tyrosinase enzyme^[51].

Microscopic differences for photo aged skin in contrast can be associated with either increased epidermal thickness or pronounced epidermal atrophy with histological changes like accumulation of elastin-containing material just below the derma lepidermal junction or disorganization of collagen may be responsible for microscopic differences for photoaged skin. For example, one of the main building blocks of human skin is collagen, it is synthesized from its precursor molecules called procollagen which is derived from dermal fibroblasts regulated by Transforming Growth Factor- β (TGF- β), a cytokine that promotes collagen production and Activator Protein-1 (AP-1), a transcription factor promotes collagen breakdown by up regulating enzymes called matrix metalloproteinases^[51].

CONCLUSION

High antioxidant and tyrosinase inhibitory ability of 80% methanolic extract of *Ziziphus spina-christi* seed has been recorded in this study. Other scientific evaluation on the use of *Ziziphus spina-christi* for traditional medicinal used should be explored further based on different models. Fractions from the all part of the plant need to be evaluated for antioxidant and tyrosinase inhibition potential. Bioactive constituent from the all part of the plant should also be identified to find effective leads from natural resources useful in the treatment of skin wrinkling.

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REFERENCES

01. Renner, S.S., J.S. Strijk, D. Strasberg and C. Thebaud, 2010. Biogeography of the Monimiaceae (Laurales): A role for East Gondwana and long-distance dispersal but not West Gondwana. *J. Biogeogr.*, 37: 1227-1238.
02. Sakaguchi, S., Y.X. Qiu, Y.H. Liu, X.S. Qi and S.H. Kim *et al.*, 2012. Climate oscillation during the quaternary associated with landscape heterogeneity promoted allopatric lineage divergence of a temperate tree *Kalopanax septemlobus* (Araliaceae) in East Asia. *Mol. Ecol.*, 21: 3823-3838.
03. Karar, M.G.E., L. Quiet, A. Rezk, R. Jaiswal and M. Rehders *et al.*, 2016. Phenolic profile and in vitro assessment of cytotoxicity and antibacterial activity of *Ziziphus spina-christi* leaf extracts. *Med. Chem.*, 6: 143-156.
04. Pringle, E.G., R.I. Adams, E. Broadbent, P.E. Busby and C.I. Donatti *et al.*, 2011. Distinct leaf-trait syndromes of evergreen and deciduous trees in a seasonally dry tropical forest. *Biotropica*, 43: 299-308.
05. Dolatkahi, M., A. Dolatkahi and J.B. Nejad, 2014. Ethnobotanical study of medicinal plants used in Arjan-Parishan protected area in Fars Province of Iran. *Avicenna J. Phytomedicine*, 4: 402-412.
06. Dinda, B., A.M. Kyriakopoulos, S. Dinda, V. Zoumpourlis and N.S. Thomaidis *et al.*, 2016. *Cornus mas* L. (Cornelian cherry), an important European and Asian traditional food and medicine: Ethnomedicine, phytochemistry and pharmacology for its commercial utilization in drug industry. *J. Ethnopharmacol.*, 193: 670-690.
07. Chen, H.C., L.W. Peng, M.J. Sheu, L.Y. Lin and H.M. Chiang *et al.*, 2013. Effects of hot water treatment on the essential oils of calamondin. *J. Food Drug Anal.*, 21: 363-368.
08. Isman, M.B., S. Miresmailli and C. Machial, 2011. Commercial opportunities for pesticides based on plant essential oils in agriculture, industry and consumer products. *Phytochem. Rev.*, 10: 197-204.
09. Jakopic, J., R. Veberic and F. Stampar, 2009. Extraction of phenolic compounds from green walnut fruits in different solvents. *Acta Agriculturae Slovenica*, 93: 11-15.
10. Jayanegara, A., E. Wina and J. Takahashi, 2014. Meta-analysis on methane mitigating properties of saponin-rich sources in the rumen: Influence of addition levels and plant sources. *Asian-Australasian J. Anim. Sci.*, 27: 1426-1435.
11. Zohra, S.F., B. Meriem, S. Samira and M.A. Muneer, 2012. Phytochemical screening and identification of some compounds from Mallow. *J. Nat. Prod. Plant Resour.*, 2: 512-516.

12. Mohammed, G.T., F.I. Abdulrahman, I.Z. Khan, M.I. Hussaini and J. Muazu *et al.*, 2013. Antimicrobial efficacies of ethanolic extract and active column fractions of the stem bark of *Zizyphus spina-christi* L. Intl. J. Pharm. Pharm. Sci., 5: 455-460.
13. Su, Z., 2012. Anthocyanins and flavonoids of *Vaccinium* L. Pharm. Crops, 3: 7-37.
14. Gani, A., S.M. Wani and F.A. Masoodi, 2012. Whole-grain cereal bioactive compounds and their health benefits: A review. J. Food Process. Technol., Vol. 3. 10.4172/2157-7110.1000146
15. Bellik, Y., L. Boukraa, H.A. Alzahrani, B.A. Bakhotmah, F. Abdellah, S.M. Hammoudi and M. Iguer-Ouada, 2012. Molecular mechanism underlying anti-inflammatory and anti-allergic activities of phytochemicals: An update. Molecules, 18: 322-353.
16. Dzoyem, J.P., V. Kuete, L.J. McGaw and J.N. Eloff, 2014. The 15-lipoxygenase inhibitory, antioxidant, antimycobacterial activity and cytotoxicity of fourteen ethnomedicinally used African spices and culinary herbs. J. Ethnopharmacol., 156: 1-8.
17. Singh, D., R. Gupta and S.A. Saraf, 2012. Herbs-are they safe enough? An overview. Crit. Rev. Food Sci. Nutr., 52: 876-898.
18. Ottenbacher, M.C. and R.J. Harrington, 2013. A case study of a culinary tourism campaign in Germany: Implications for strategy making and successful implementation. J. Hospitality Tourism Res., 37: 3-28.
19. Wijesinghe, W.A.J.P. and Y.J. Jeon, 2012. Enzyme-Assistant Extraction (EAE) of bioactive components: A useful approach for recovery of industrially important metabolites from seaweeds; A review. Fitoterapia, 83: 6-12.
20. Luis, A., F. Domingues and A.P. Duarte, 2011. Bioactive compounds, RP-HPLC analysis of phenolics and antioxidant activity of some Portuguese shrub species extracts. Nat. Prod. Commun., 6: 1863-1872.
21. Gupta, V.K., M.G. Tuohy, A. O'Donovan and M. Lohani, 2015. Biotechnology of Bioactive Compounds: Sources and Applications. John Wiley and Sons, Hoboken, New Jersey, USA., ISBN:9781118733493.
22. Deng Y., G. Yang, J. Yue, B. Qian and Z. Liu *et al.*, 2014. Influences of ripening stages and extracting solvents on the polyphenolic compounds, antimicrobial and antioxidant activities of blueberry leaf extracts. Food Control, 38: 184-191.
23. Fasakin, C.F., C.C. Udenigwe and R.E. Aluko, 2014. Antioxidant properties of chlorophyll-enriched and chlorophyll-depleted polyphenolic fractions from leaves of *Vernonia amygdalina* and *Gongronema latifolium*. Food Res. Intl., 44: 2435-2441.
24. Shalmashi, A. and F. Golmohammad, 2010. Solubility of caffeine in water, Ethyl acetate, Ethanol, Carbon Tetrachloride, Methanol, Chloroform, Dichloromethane and Acetone between 298 and 323 K. Lat. Am. Appl. Res., 40: 283-285.
25. Gill, S.S. and N. Tuteja, 2010. Reactive oxygen species and antioxidant machinery in abiotic stress tolerance in crop plants. Plant Physiol. Biochem., 48: 909-930.
26. Wu, D. and P. Yotnda, 2011. Production and detection of Reactive Oxygen Species (ROS) in Cancers. J. Visualized Exp., 21: 1-5.
27. GroB, F., J. Durner and F. Gaupels, 2013. Nitric Oxide, antioxidants and prooxidants in plant defence responses. Front. plant Sci., 4: 1-15.
28. Garel, M., L. Caeymaex, F. Goffinet, M. Cuttini and M. Kaminski, 2011. Ethically complex decisions in the neonatal intensive care unit: Impact of the new French legislation on attitudes and practices of physicians and nurses. J. Med. Ethics, 37: 240-243.
29. Barrett, T., K. Clark, R. Gevorgyan, V. Gorelenkov and E. Gribov *et al.*, 2011. BioProject and BioSample databases at NCBI: Facilitating capture and organization of metadata. Nucleic Acids Res., 40: D57-D63.
30. Solimine, J., E. Garo, J. Wedler, K. Rusanov and O. Fertig *et al.*, 2016. Tyrosinase inhibitory constituents from a polyphenol enriched fraction of rose oil distillation wastewater. Fitoterapia, 108: 13-19.
31. Komori, Y., M. Imai, T. Yamauchi, K. Higashiyama and N. Takahashi, 2014. Effect of p-aminophenols on tyrosinase activity. Bioorg. Med. Chem., 22: 3994-4000.
32. Da Silva, L.M.R., E.A.T. de Figueiredo, N.M.P.S. Ricardo, I.G.P. Vieira, R.W. de Figueiredo, I.M. Brasil and C.L. Gomes, 2014. Quantification of bioactive compounds in pulps and by-products of tropical fruits from Brazil. Food Chem., 143: 398-404.
33. Lee, S.Y., N. Baek and T.G. Nam, 2016. Natural, semisynthetic and synthetic tyrosinase inhibitors. J. Enzyme Inhib. Med. Chem., 31: 1-13.
34. Mendes, E., M.D.J. Perry and A.P. Francisco, 2014. Design and discovery of mushroom tyrosinase inhibitors and their therapeutic applications. Expert Opin. Drug Discovery, 9: 533-554.
35. Lee, Y.C., N.W. Hsiao, T.S. Tseng, W.C. Chen and H.H. Lin *et al.*, 2015. Phage display-mediated discovery of novel tyrosinase-targeting tetrapeptide inhibitors reveals the significance of N-terminal preference of cysteine residues and their functional sulfur atom. Mol. Pharmacol., 87: 218-230.

36. Alhakmani, F., S.A. Khan and A. Ahmad, 2014. Determination of total phenol, *in-vitro* antioxidant and anti-inflammatory activity of seeds and fruits of *Zizyphus spina-christi* grown in Oman. Asian Pac. J. Trop. Biomed., 4: S656-S660.
37. Shekhar, T.C. and G. Anju, 2014. Antioxidant activity by DPPH radical scavenging method of *Ageratum conyzoides* Linn, leaves. Am. J. Ethnomed., 1: 244-249.
38. Marinova, G. and V. Batchvarov, 2011. Evaluation of the methods for determination of the free radical scavenging activity by DPPH. Bulg. J. Agric. Sci., 17: 11-24.
39. Khatua, S., T. Roy and K. Acharya, 2013. Antioxidant and free radical scavenging capacity of phenolic extract from *Russula laurocerasi*. Asian J. Pharm. Clin. Res., 6: 156-160.
40. Ravipati, A.S., L. Zang, S.R. Koyyalamudi, N. Reddy and J. Bartlett *et al.*, 2012. Antioxidant and anti-inflammatory activities of selected Chinese medicinal plants and their relation with antioxidant content. BMC Complementary Altern. Med., 12: 173-178.
41. Hidalgo, M., C. Sanchez-Moreno and S. De Pascual-Teresa, 2010. Flavonoid-flavonoid interaction and its effect on their antioxidant activity. Food Chem., 121: 691-696.
42. Trivellini, A., M. Lucchesini, R. Maggini, H. Mosadegh and T. Villamarin *et al.*, 2016. Lamiaceae phenols as multifaceted compounds: Bioactivity, industrial prospects and role of positive-stress. Ind. Crops Prod., 83: 241-254.
43. Lobo, V., A. Patil, A. Phatak and N. Chandra, 2010. Free radicals, antioxidants and functional foods: Impact on human health. Pharmacogn. Rev., 4: 118-126.
44. Asgarpanah, J. and E. Haghghat, 2012. Phytochemistry and pharmacologic properties of *Zizyphus spina christi* (L.) Willd. Afr. J. Pharm. Pharmacol., 6: 2332-2339.
45. Cho, J.Y., S.C. Park, T.W. Kim, K.S. Kim and J.C. Song *et al.*, 2006. Radical scavenging and anti-inflammatory activity of extracts from *Opuntia humifusa* Raf. J. Pharm. Pharmacol., 58: 113-119.
46. Haslam, I.S., E.W. Roubos, M.L. Mangoni, K. Yoshizato and H. Vaudry *et al.*, 2014. From frog integument to human skin: Dermatological perspectives from frog skin biology. Biol. Rev., 89: 618-655.
47. Lapsley, D.K. and P.L. Hill, 2010. Subjective invulnerability, optimism bias and adjustment in emerging adulthood. J. Youth Adolescence, 39: 847-857.
48. Blach-Overgaard, A., J.C. Svenning, J. Dransfield, M. Greve and H. Balslev, 2010. Determinants of palm species distributions across Africa: The relative roles of climate, non-climatic environmental factors and spatial constraints. Ecography, 33: 380-391.
49. Yoshida-Amano Y., A. Hachiya, A. Ohuchi, G.P. Kobinger and T. Kitahara *et al.*, 2012. Essential role of RAB27A in determining constitutive human skin color. PLoS One, 7: 1-11.
50. Mukherjee, P.K., N. Maity, N.K. Nema and B.K. Sarkar, 2011. Bioactive compounds from natural resources against skin aging. Phytomedicine, 19: 64-73.
51. Pittayapruerk, P., J. Meephansan, O. Prapapan, M. Komine and M. Ohtsuki, 2016. Role of matrix metalloproteinases in photoaging and photocarcinogenesis. Intl. J. Mol. Sci., 17: 1-20.