

Microbiological Assessment of Gram Negative Bacterial Isolates from Ear, Eye, Nose and Throat among Patients Attending Aresho Advanced Medical Laboratory, Addis Ababa, Ethiopia

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Abstract: ENT samples from patients who referred to Arsho Advanced Medical Laboratory were isolated for further microbiological assessment. About 91 from both male and female patients comprising 42 ear, 21 nasal, 1 eye and 27 throat swab, respectively were screened between April-2017-July-2017 and the samples were analyzed using culture technique, direct microscopy and identification of bacteria (Vtek 2 compact). Finding from the study identified 12 species of bacteria, i.e., *Pseudomonas aeruginosa*, *Proteus mirabilis*, *Klebsiella oxytoca*, *Enterobacter cloacae*, *Citrobacter koseri*, *Citrobacter pauculus*, *Pseudomonas fulorescens*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *E. coli*, *Leclerciaa decarboxylate* and *Proteus vulgaris*. *Proteus mirabilis* recorded the highest rates (30%) followed by *Klebsiella pneumoniae* (20%) which was isolated in ear and nose swabs while the least were recorded on *Citrobacter koseri*, *Citrobacter pauculus*, *Pseudomonas fulorescens*, *Proteus vulgaris*, *Acinetobacter baumannii*, *E. coli*, *Leclerciaa decarboxylate*, *Klebsiella pneumonia* (3.3%).

Key words: Bacteria, species, swab, nasal, twelve species, culture technique

INTRODUCTION

Ear, eye and nose infection are among the most widespread and serious infections that compel an individual to seek medical attention. It represents some of the most common bacterial disease encountered affecting people of all ages. These infections are one of the leading causes of morbidity and mortality in critically ill patients (Witsell *et al.*, 2001). The ear, nose and throat are the frequent sites of infection because they come in direct contact with the physical environment and are exposed to air borne microorganisms.

Disease of ear and nose affect the functioning of adults as well as children, often with significant impairment of the daily life of affected patients (Witsell *et al.*, 2001). It has been predicted that with increase in global population, infection remain the most important causes of disease with upper respiratory infections causing hearing loss and learning disability particularly in children (Alberti, 1999; Ibe *et al.*, 2013; Obiajuru and Anaelechi, 2013). Ear infection such as chronic otitis (tinnitus) have serious consequences in developing countries such as retarded language development and progress in school among children (Newton *et al.*, 2001). Tinnitus which is now known to be the most common childhood infections, lead annually to the death of over 50,000 children under 5 year (Rovers *et al.*, 2006).

In addition, antimicrobial resistance profile of bacteria varies among population because of the difference in geography, local antimicrobial prescribing practices and prevalence of resistant bacterial strains in a given area (Noh and Kim, 1985). So, there should be up to date information on microbial resistance pattern at national and local levels to guide the rational use of the existing antimicrobial drugs.

The human Ear, Nose and Throat (ENT) are closely related and inter connected parts of the body. Infections, diseases and health problems related to the ENT are therefore jointly studied and managed like the most other part of the human body. The ear, nose and throat were found to be colonized by a wide range of microorganisms some of which are more or less harmless under normal condition (Ibe *et al.*, 2013).

In Ethiopia particularly in the study area there is no such type of recent data that shows the magnitude of the problem. Therefore, the aim of this study was to determine the gram negative bacterial isolates and their drug susceptibility patterns from patients who gave ear and nose swab samples at Arsho Advanced Medical Laboratory.

Objectives

General objectives: To determine the frequency of gram negative bacterial outline and antibiotic susceptibility pattern isolated from ENT at Arsho Advanced Medical Laboratory.

Specific objectives:

- To assess the gram negative pathogens responsible for the ENT infection
- To verify the antimicrobial resistance and sensitivity pattern of commonly isolated gram negative bacteria from ENT patients

MATERIALS AND METHODS

Study area and period: A study was conducted from, April-2017-July-2017 at Arsho Advanced Medical Laboratory, Addis Ababa, Ethiopia.

Samples collections: The throat swab, nasal (nose) swab, eye swab and aural (ear) swab were collected by using a sterile cotton swab and then the swabs were transported with amies transport media to the Microbiology Laboratory.

Culture technique: Throat swab were inoculated on blood agar plates were as nasal (nose), eye and ear swab were inoculated on chocolate, blood and MacConkey agar plates. All the plates were incubated for 48 h 5% CO₂ an aerobically with the exception of MacConkey agar plate that was inculcated in aerobic. The plates were examined for the growth of gram negative bacteria and the *pathogenic colonies* were identified by VITEK 2 compact.

Bacteria identification test: Pure colonies of gram negative colonies which were grown on macConkey agar, chocolate agar and blood agar were used for identification in VITEK 2 compact.

Suspension preparations for ID and AST card

Suspension preparation for ID card: First transfer 3 mL of 0.45% saline into a test tube. Select an isolated colony and dissolve it. Mix well and check the density within dens check. Inoculum density for gram positive and gram negative colonies should be between 0.5-0.63 MCF. Then place the Identification (ID) card into the test tube and then transfer the test tube into the cassette.

Suspension preparation for AST card: First transfer 3 mL of 0.45% saline into a test tube. Transfer 145 mL for gram negative of the ID suspension into the saline test tube. Then place the AST card into the test tube and then transfer the test tube into the cassette.

Filling and loading the card into VITEK 2 system: Put all the test tubes containing cards and suspension a cassette. From computer work station print cassette worksheet and record job ID and bar code for each card. When instrument status is OK then press start fill button. Remove the cassette from the loading station when the machine indicates.

Antibiotics used: The antibiotics used in this investigation were Ampicillin, Amoxicillin/Clavulanic Acid, piperacillin/Tazobactam, Cefalotin, Cefazolin, cefuroxime axil, Cefoxitin, Cefpodoxime, Ceftazidime, Ceftriaxone, Cefepime, Gentamicin, Toberamycin, Ciprofloxacin, Levofloxacin, Teteracyclin, Nitrofurantoin and Trimethoprim/sulfamethoxazole for gram negative. The control strains were run simultaneously with the test organisms.

Data analysis: Data were checked for completeness, manually, entered and analyzed using SPSS Version 20 Statistical Software and Excel.

RESULTS AND DISCUSSION

A total number of 42 ear swabs, 21 nasal swabs, 1 eye and 27 throat swabs were collected from the patients attending at Aresho Advanced Medical Laboratory. Out of these, ear swabs were positive in 20 (22%), 10 (11%) in nose swabs.

The ear swabs was positive in 9 (9.9%) male and 11(12.1%) in female, nose swabs was positive 4 (4.4%) was positive in male and 5(5.5%) in female (Table 1). The findings in this study show that 12 genera of gram negative bacteria which are *P. mirabilis*, *K. pneumoniae*, *E. coli*, *P. vulgaris*, *P. aeruginosa*, *P. fulorescens*, *K. oxytoca*, *E. cloacea*, *C. koseri*, *C. pauculus*. Leclerciaa decarboxylata and *A. baumannii* were involved in various ear and nose amongst patients examined at the study area. Out of all the organisms isolated 20 (66.67%) occurred in ear swabs followed by nasal 10 (33.3%) while there is no growth in eye and throat.

Generally, the most prevalent organisms were *K. oxytoca* were isolated in both ear and nose swabs, *E. coli*, *P. aeruginosa*, *P. fuloresce*, *E. cloacea*, *C. koseri*, *C. pauculus* and *A. baumannii* were isolated in ear only while *L. decarboxylata* and *P. vulgaris* were isolated from only nasal swabs (Table 2)

Table 1: Sex wise distribution and culture result of ENT patients diagnosed from April-2017-July-2017 in Arsho Advanced Medical Laboratory

Sample/Sex	Negative	Positive	Total examined
Ear			
M	10	9	19
F	12	11	23
Nose			
M	3	4	7
F	7	7	14
Throat			
M	13	0	13
F	14	0	14
Eye			
M	1	0	1
F	0	0	0

The age related prevalence of the ENT infection in the study area showed that the highest prevalence of the bacterial infection (40%) was amongst those aged 15-24 years, (26.67%) 25-44 years, (23.33%) 1-14 year and (10%) 45-64 while no growth observed amongst <1 and >65 years aged as shown in Table 3.

During the study period 18 different antimicrobial agents were used to test the antibiotic susceptibility patterns of the pathogenic gram negative bacteria isolated from ENT patients. The general susceptibility profiles of bacterial isolates are shown in Table 4. Out of the total antibiotics examined during the study period, Ampicillin had the highest overall resistance rate (80.8%) followed by Cephalothin (58.66%), Cefazolin (57.14%), Cefuroxime and Cefuroxime Axetil (53.6%) and Cefpodoxime (48.27%). on the other hand, majority of bacterial isolates were susceptible to Levofloxacin, Tobramycin, Ciprofloxacin and Gentamycin with overall resistance rates of 0, 0, 6.6 and 11.11%, respectively.

The 86.67% of the isolated bacteria were found to be resistant to one and more of the commonly used antibiotics Table 5. Among the total *Klebsiella* spp.

Isolated all (100%) of the isolates have developed resistant to one or more antibiotics used, among the total *Proteus* spp. isolated, the majority (77.8 %) of the isolates have developed resistance to one and more antibiotics used. Similarly, about three-fourth of (75%) *Pseudomonas* spp. isolates were able to resist one and more antibiotics commonly used to treat them. All other bacterial isolates have also showed overall antibiotic resistance to one and more antibiotics used except *L. decarboxylata* and *C. koseri* which shows 100% sensitivity (Fig. 1).

ENT infection is a more frequent treat able health care problem worldwide, yet if left untreated it can cause a serious complication such as a speech disorder, pain in patients and their family quality of life and economic burden on the health care system (Edwin *et al.*, 2014). The burden and prevalence of ENT infection are more intense in developing countries due to the poor living standard and hygienic conditions along with lack of proper nutrition (Argaw-Denboba *et al.*, 2016). Kumar *et al.*

Table 2: Gram negative bacteria associated with ENT infection in Arsho Advanced Medical Laboratory

Isolates	Ear	Nose	Throat	Eye	Total (%)
<i>Pseudomonas aeruginosa</i>	3	-	-	-	3(10)
<i>Proteus mirabilis</i>	7	2	-	-	9(30)
<i>Kelebsiella oxytoca</i>	2	1	-	-	3(10)
<i>Entrobacter cloacae</i>	2	-	-	-	2(6.67)
<i>Citrobacter koseri</i>	1	-	-	-	1(3.33)
<i>Citrobacter pauculus</i>	1	-	-	-	1(3.33)
<i>Pseudomonas fulorescens</i>	1	-	-	-	1(3.33)
<i>Klebsiella pneumoniae</i>	1	5	-	-	6(20)
<i>Acinetobacter baumannii</i>	1	-	-	-	1(3.33)
<i>E. coli</i>	1	-	-	-	1(3.33)
<i>Leclteciaa decarboxylate</i>	-	1	-	-	1(3.33)
<i>Proteus vulgaris</i>	-	1	-	-	1(3.33)

Table 3: Age related prevalence of gram negative bacterial isolates in ENT infection in Arsho Advanced Medical Laboratory

Isolates	Age					
	<1	1-14	15-24	25-44	45-64	>65
<i>Pseudomonas aeruginosa</i>	-	-	2	1	-	-
<i>Proteus mirabilis</i>	-	4	3	1	1	-
<i>Kelebsiella oxytoca</i>	-	-	2	1	-	-
<i>Entrobacter cloacae</i>	-	-	-	2	-	-
<i>Citrobacter koseri</i>	-	-	-	-	1	-
<i>Citrobacter pauculus</i>	-	-	1	-	-	-
<i>Pseudomonas fulorescens</i>	-	-	1	-	-	-
<i>Klebsiella pneumoniae</i>	-	-	2	3	1	-
<i>Acinetobacter baumannii</i>	-	1	-	-	-	-
<i>E. coli</i>	-	1	-	-	-	-
<i>Leclteciaa decarboxylate</i>	-	1	-	-	-	-
<i>Proteus vulgaris</i>	-	-	1	-	-	-
Total	0	7	12	8	3	0
Percentage	0	23.33	40	26.67	10	0

Table 4: Overall antibiotic susceptibility profiles of isolated bacteria from ear discharge

Type of antibiotics used	Frequency of each antibiotic tested	Susceptibility patterns		
		Resistant (%)	Sensitive (%)	Intermediate (%)
Ampicillin	26	80.8(21)	15.4(4)	3.8(1)
Augumentin	28	25(7)	75(21)	-
Cefazolin	28	57.14(16)	39.3(11)	3.6(1)
Cefepime	30	26.7(8)	73.3(22)	-
Cefoxitin	29	44.83(13)	48.27(14)	6.9(2)
Cefpodoxime	29	48.27(14)	51.73(15)	-
Ceftazidime	30	30(9)	70(21)	-
Ceftriaxone	30	36.67(11)	63.33(19)	-
Cefuroxime	28	53.6(15)	39.3(11)	7.14(2)
Cefuroxime Axetil	28	53.6(15)	46.4(13)	7.14(2)
Cephalothin	29	58.6(17)	37.9(11)	3.4(1)
Ciprofloxacin	30	6.67(2)	90(27)	0.33(1)
Co-trimoxazole	27	29.63(8)	70.37(19)	-
Gentamycin	27	11.11(3)	23(85.2)	3.7(1)
Levofloxacin	29	-	93.1(27)	6.9(2)
Piperacillin/Tazobactam	28	28.6(8)	71.4(20)	-
Tetracycline	28	42.9(12)	57.1(16)	-
Tobramycin	28	-	96.4(27)	3.6(1)

Table 5: Antibiotic susceptibility test of pathogenic bacteria isolated from ENT patients

Organisms	AMP	AMC	ZP	CFA	CFZ	CFU	CFXA	FOX	CPD	CAZ	CRO	CFP	GM	TBM	CIP	LEV	TEC	SXT
<i>P. mirabilis</i>	-	S	S	-	-	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>P. mirabilis</i>	R	S	R	R	R	R	R	R	R	R	R	R	R	S	R	S	R	R
<i>P. mirabilis</i>	R	S	R	R	R	R	R	R	R	R	R	R	R	S	R	S	R	R
<i>P. mirabilis</i>	R	S	R	R	R	R	R	R	R	R	R	R	I	S	S	S	R	R
<i>P. mirabilis</i>	-	-	-	-	-	S	-	S	S	S	S	-	-	-	S	-	-	-
<i>P. mirabilis</i>	S	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S	R	S
<i>P. mirabilis</i>	R	S	R	R	R	R	R	S	R	R	R	R	R	S	S	S	R	R
<i>P. mirabilis</i>	R	S	R	R	R	R	R	I	R	R	R	R	S	S	S	S	R	R
<i>P. vulgaris</i>	R	S	R	R	R	R	R	R	R	R	R	R	S	S	S	S	R	S
<i>P. aeruginosa</i>	R	R	R	R	R	R	R	R	R	R	R	S	S	S	S	S	R	R
<i>P. aeruginosa</i>	R	R	S	R	R	R	R	R	R	S	R	S	S	S	I	I	R	R
<i>P. aeruginosa</i>	R	R	S	R	R	R	R	R	R	S	R	S	S	S	S	S	R	R
<i>P. fluorescens</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. oxytoca</i>	R	S	S	I	I	R	R	S	S	S	S	S	S	S	S	S	S	S
<i>K. oxytoca</i>	R	R	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. oxytoca</i>	R	S	S	S	S	R	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	R	S	R	S	R	R	S	S	S	S	S	S	S	S	S
<i>K. pneumonae</i>	R	S	S	R	R	S	I	R	S	S	S	S	S	S	S	S	S	S
<i>E. cloacae</i>	R	R	S	R	R	I	R	R	R	S	S	S	S	S	S	S	S	S
<i>E. cloacae</i>		R	S	R	R	I	R	R	R	S	S	S	S	S	S	S	R	S
<i>C. koseri</i>		S	S	S	S	S	I	S	S	S	S	S	S	S	S	S	S	S
<i>C. pauchus</i>	S	S		R	R	S	S	S	S	S	S	S			S	S		
<i>A. baumannii</i>	I	S	S	S	S	R	R	R	S	S	S	S	S	S	S	S	S	S
<i>E. coli</i>	R	R	S	R	R	R	R	I	R	R	R	R	R	I	R	I	S	S
<i>L. adecarboxylata</i>	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S	S

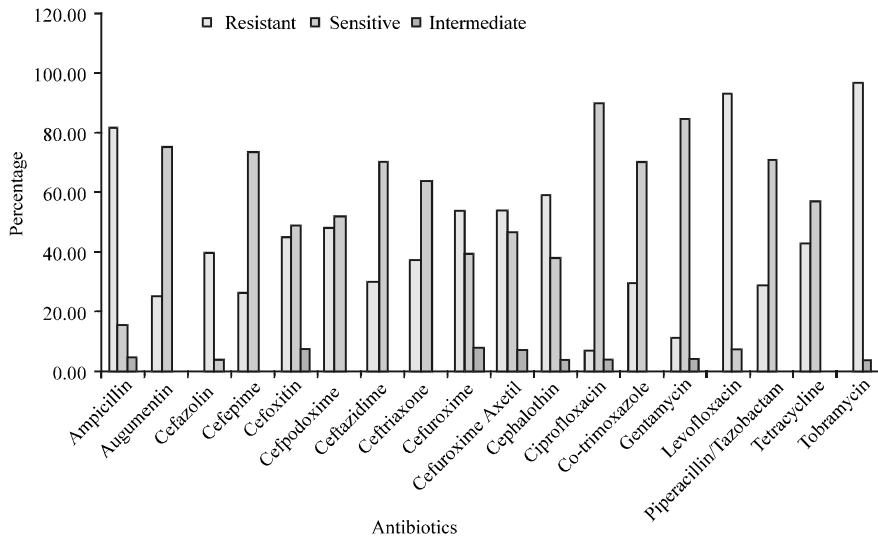


Fig. 1: Antibiotic susceptibility profiles of isolated bacteria from ENT

(2013) reported that from gram negative bacteria spp., *Proteus* spp. were responsible for most cases of ENT infections Whereas (Edwin *et al.*, 2014) find out that *Pseudomonas* spp., *Proteus* spp. and *Klebsiella* spp. are the common bacteria that cause ENT infection in Japura India, similarly (Argaw-Denboba *et al.*, 2016) find out that *P. aeruginosa*, *Klebsiella* spp. *E. coli* are the

bacteria associated with ENT infection and *Pseudomonas aeruginosa* was the most prevalent aetiological agent of ENT infection in Benin city.

In this study, the main pathogenic bacterium associated with ENT infection was *Proteus mirabilis*. Followed by *Klebsiella pneumonia*, *Kelebsiella oxytoca* and *Pseudomonas aeruginosa*., respectively. Similarly,

previously published articles from our study area and other parts of Ethiopia also reported *Proteus* spp. were the foremost bacteria associated with middle ear infection (Edwin *et al.*, 2014; Argaw-Denboba *et al.*, 2016). Although, it needs a further nationwide study, taking into account our study finding and others it seems that *Proteus mirabilis* are the leading bacterial isolates associated with ENT infection in Ethiopia. Conversely, several other published data pieces from Africa and elsewhere in the world reported *Pseudomonas* spp., mainly *P. aeruginosa* is the primary pathogenic bacteria associated with middle ear infection (Afolabi *et al.*, 2012). One possible explanation for this difference might be due to climate, poor living standard, hygienic condition and geographical variations between Ethiopia and other countries.

In general, Ampicillin, Cephalothin, Cefazolin, Cefuroxime and Cefuroxime had shown the highest antibiotic resistance rates to all bacterial pathogens isolated from ENT discharge, respectively. Correspondingly, Tetracycline were the most clinically used antibiotics that showed a higher resistance rate to *Proteus* spp. which is in line with earlier reports from Ethiopia, Nigeria and Egypt (Seid *et al.*, 2013). Among the total *Klebsiella* spp. and *Proteus* spp. isolated during the study period about 100 and 77.8% of the isolated bacteria have developed resistance to one and more antibiotics that were once in clinical use, respectively. Overall, more than 60% of the bacterial isolates of this study were characterized as multi-antibiotic-resistant pathogenic bacteria. The reason for this high degree of multi-antibiotic resistance might be connected to inappropriate use of antibiotics, including incomplete dose, self-medication and poor infection prevention and control practices as indicated by the recent WHO antimicrobial resistance report. An interestingly our study revealed almost all the isolated pathogenic bacteria were considerably susceptible to Levofloxacin, Tobramycin, Ciprofloxacin and Gentamicin. Particularly, Levofloxacin, and Tobramycin were shown to be highly effective for pathogenic bacteria associated with ENT infection in this study. Likewise with a slight variation several other researchers have shown a similar high efficiency of ciprofloxacin against these bacterial species (Afolabi *et al.*, 2012). Thus we propose that Levofloxacin and Tobramycin can be taken as a first-line optional treatment for ENT infection attending in Arsho Advanced Medical Laboratory, Addis Ababa, Ethiopia. This suggestion holds true, since, the current first-line treatment for both acute and chronic otitis media in Ethiopia is Ampicillin and Augmentin

which already showed a high resistance rate (80.8 and 25%, respectively) for the majority of the bacterial isolates in this study.

CONCLUSION

This study showed that the risk of acquiring particularly ear infection is strongly associated in *Proteus mirabilis*, *Pseudomonas aeruginosa* and *Klebsiella oxytoca* were the three predominant bacteria isolates from patient ear discharges suspected of otitis media whereas in nose infection *Klebsiella pneumonia* was the most predominant isolated organism. Almost all the isolated bacteria showed a considerable level of resistance to one antibiotic that are commonly used in health institutes; particularly, majority of isolated bacteria were found to be highly resistant to Ampicillin Cephalothin, Cefazolin, Cefuroxime and Cefuroxime Axetil and Cefpodoxime treatments. Among the total *Klebsiella* spp. Isolated all (100%) of the isolates have developed resistant to one or more antibiotics used, among the total *Proteus* spp. isolated, the majority (77.8%) of the isolates have developed resistance to one and more antibiotics used. This study also indicated Levofloxacin and Tobramycin are effective against all the bacterial isolates and most was highly sensitive to Tobramycin. In general, the result of this study revealed that antibiotic resistant bacteria are alarmingly increasing in Arsho Advanced Medical Laboratory study area, Addis Ababa, Ethiopia and becoming a major public health problem in patients with ENT infection.

RECOMMENDATIONS

Therefore we recommend nationwide antimicrobial inspection to make the right recommendation of alternative antibiotics along with strict devotion to antibiotic policy to reduce the spread of drug resistant microbes in the country.

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