

Effect of Some Inhibitors on *Clostridium Chauvoei* (Jakari Strain) Neuraminidase

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Abstract: The effect of some inhibitors [silver nitrate (AgNO_3), paranitrophenyl oxamic acid (PNPO), Salicyl Oxamic Acid (SOA) and Ethylene Diamine Tetra Acetic acid (EDTA)] on dialysed *Clostridium chauvoei* (jakari strain) neuraminidase was investigated *in vitro* and it was found that these compounds inhibited neuraminidase activity. The concentrations of the inhibitors used were $1.5 \times 10^3 \mu\text{Mol}$, $2.9 \times 10^3 \mu\text{Mol}$, $4.4 \times 10^3 \mu\text{Mol}$, $5.9 \times 10^3 \mu\text{Mol}$ and $7.4 \mu\text{Mol}$ for all the inhibitors. Neuraminidase activity was measured by assaying the amount of sialic acid cleaved from fetuin by the enzyme on incubation with the substrate. In all studies, the inhibition was not proportional to the concentration of inhibitor used. It is suggested, based on the present results, that proper clinical trials should be conducted to ascertain the clinical application of these neuraminidase inhibitors in the management of blackleg in cattle.

Key words: Inhibitors, *clostridium chauvoei* (Jakari strain), neuraminidase

INTRODUCTION

Blackleg is a disease of cattle, sheep and other ruminants caused by *Clostridium chauvoei* that was first reported in 1870^[1]. In Nigeria, the disease was first reported in 1929 by Anon^[2] and has remained a major problem of cattle in the country. Although vaccination against it has been carried out since 1930, sporadic outbreaks are recorded annually.

Clostridium chauvoei (jakari Strain), which cause blackleg in indigenous Nigerian cattle, has been found to produce neuraminidase^[3]. Neuraminidases (sialidases, EC 3.2.1.18) are involved in the pathogenesis of infectious diseases, whose aetiologic agents produce the enzyme^[4,5]. The role of neuraminidase in the pathogenesis of blackleg is that of spreading the disease in host tissues^[3]. Neuraminidases are of great importance in medicine and the pharmaceutical industry for the analysis of oligosaccharides and development of neuraminidase inhibitors^[6]. The use of neuraminidase inhibitors to manage diseases clinically, especially human influenza virus infections has been advocated^[7,8]. As a prerequisite to developing an alternative management protocol against blackleg in cattle, the effect of dialysed *Clostridium chauvoei* (jakari strain) neuraminidase on fetuin was investigated *in vitro* (both in the presence and absence of neuraminidase inhibitors) and we report here for the first time, the inhibition of *Clostridium chauvoei* (jakari strain)

neuraminidase catalysed hydrolysis of fetuin by silver nitrate (AgNO_3), paranitrophenyl oxamic acid (PNPO), Salicyl Oxamic Acid (SOA) and Ethylene Diamine Tetra Acetic acid (EDTA).

MATERIALS AND METHODS

Bacterial strain: *Clostridium chauvoei* (jakari strain) isolated from clinically infected Zebu cattle was obtained in its lyophilized form from the National Veterinary Research Institute (NVRI), Vom, Plateau State, Nigeria for this experiment. The pathogenicity indices of this bacterium have been fully determined^[9].

Bacterial cultivation: Three media were used for cultivating *Clostridium chauvoei* (jakari strain) to isolate neuraminidase for this study namely: Reinforced Clostridial Medium (RCM), blood agar and Cooked Meat Medium (CMM). Media preparation and bacterial cultivation were carried out as described previously^[10,3] with all the procedures for microbiological asepsis strictly maintained.

Processing of cultivated bacteria: CMM containing the cultivated bacteria was placed in a refrigerated centrifuge (Mistral 4L, MSE) and spun at 9000 g for 40 min at 4°C. The supernatant containing extracellular (crude) neuraminidase released by the bacteria during growth^[3]

was decanted immediately into two sterile plastic containers of one (1) litre capacity each. The plastics containing the culture supernatant were kept at -20°C until when required.

Ammonium sulphate [(NH₄)₂SO₄] fractionation of crude neuraminidase from clostridium chauvoei (jakari strain): This was carried out using the method of Useh^[3] to determine the fraction with the highest enzyme activity. The highest activity [55-70% (NH₄)₂SO₄ fraction] was pooled and dialysed by modifying the method of Heuermann^[11].

Dialysis of pooled neuraminidase: Thirty (30) mL of pooled neuraminidase [55-70% (NH₄)₂SO₄ fractionated enzyme] was pipetted into a 20 cm long and 6 cm diameter dialysis bag and placed in a 2 litre capacity dialysis beaker containing 1.5 litres of 50 mM acetate buffer pH 4.5. The dialysis beaker and its content was placed on a speed plate magnetic stirrer at 4°C for 28 h.

Preparation of 50 mM acetate buffer pH 4.5: This was carried out using the methods described by Useh^[3].

Preparation of Acid Citrate Dextrose (ACD): This was prepared using the method previously described^[12].

Preparation of fetuin: Fetuin in the various concentrations used in the study was prepared using the procedures described elsewhere^[3].

Preparation of inhibitors: Specified concentrations (1.5x10³ μMol, 2.9x10³ μMol, 4.4x10³ μMol, 5.9x10³ μMol and 7.4x10³ μMol) of AgNO₃, PNPO, SOA and EDTA, respectively were prepared as previously described^[3].

Hydrolysis of fetuin by dialysed Clostridium chauvoei (jakari strain) neuraminidase: The hydrolysis (inhibition studies) was conducted as described by Useh^[13]. The neuraminidase activity and the degree of enzyme inhibition were quantified by assaying the sialic acid cleaved from fetuin using the thiobarbituric acid assay method^[14].

RESULTS

Hydrolysis of fetuin by dialysed neuraminidase: The activity of dialysed Clostridium chauvoei (jakari strain) neuraminidase was inhibited *in vitro* by all the compounds tested i.e AgNO₃, PNPO, SOA and EDTA. For all the inhibitors used, the rate of enzyme inhibition was not proportional to the concentration of inhibitors used.

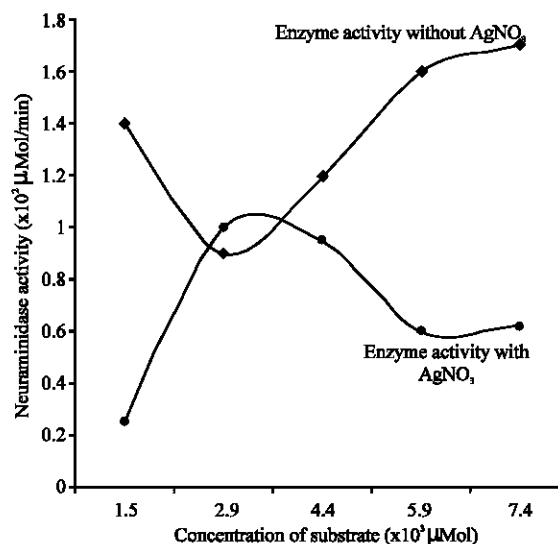


Fig. 1: Effect of silver nitrate (AgNO₃) on clostridium chauvoei (jakari strain) neuraminidase

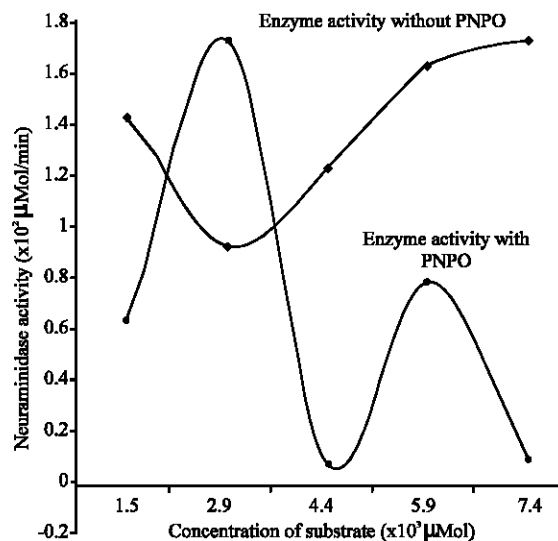


Fig. 2: Effect of paranitrophenyl oxamic acid (PNPO) on clostridium chauvoei (jakari strain) neuraminidase

AgNO₃ inhibited neuraminidase activity from 1.4x10² μMol min⁻¹ to 2.5x10¹ μMol min⁻¹, 1.2x10² μMol min⁻¹ to 9.5x10¹ μMol min⁻¹ and 1.7x10² μMol min⁻¹ to 6.2x10¹ μMol min⁻¹ at concentrations of 1.5x10³ μMol, 4.4x10³ μMol, 5.9x10³ μMol and 7.4x10³ μMol AgNO₃, respectively. There was no inhibition of enzyme activity when 2.9x10³ μMol AgNO₃ was used Fig. 1.

For PNPO, neuraminidase activity was inhibited from 1.4x10² μMol min⁻¹ to 6.0x10¹ μMol min⁻¹, 1.2x10² μMol min⁻¹ to 5.0 μMol min⁻¹, 1.6x10² μMol min⁻¹ to 7.6x10¹ μMol min⁻¹ and 1.7x10² μMol min⁻¹ to 7.0 μMol min⁻¹

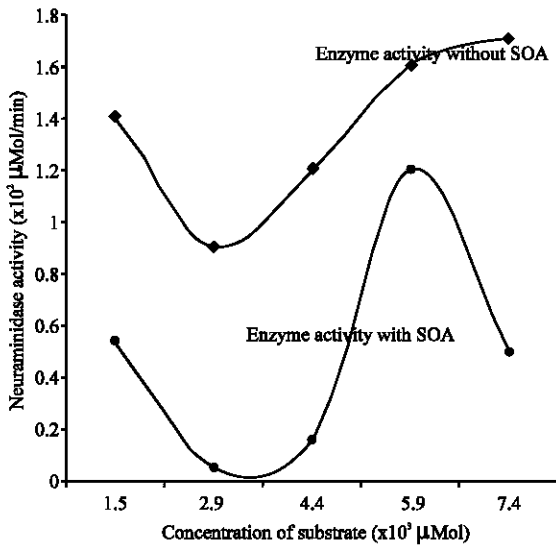


Fig. 3: Effect of salicyl oxamic acid (SOA) on clostridium chauvoei (jakari strain) neuraminidase

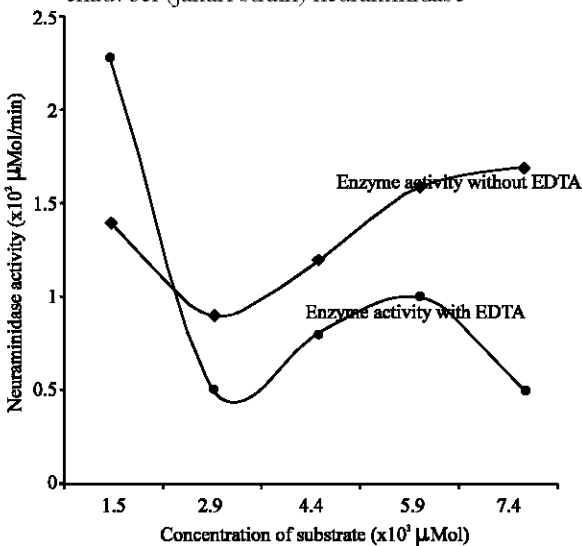


Fig. 4: Effect of ethylene diamine tetraacetic acid (EDTA) on clostridium chauvoei (jakari strain) neuraminidase

SOA inhibited neuraminidase activity from $1.4 \times 10^2 \mu\text{Mol min}^{-1}$ to $5.4 \times 10^1 \mu\text{Mol min}^{-1}$, $9.0 \times 10^1 \mu\text{Mol min}^{-1}$ to $5.0 \mu\text{Mol min}^{-1}$, $1.2 \times 10^2 \mu\text{Mol min}^{-1}$ to $1.6 \times 10^1 \mu\text{Mol min}^{-1}$, $1.6 \times 10^2 \mu\text{Mol min}^{-1}$ to $1.2 \times 10^2 \mu\text{Mol min}^{-1}$ and $1.7 \times 10^2 \mu\text{Mol min}^{-1}$ to $5.0 \times 10^1 \mu\text{Mol min}^{-1}$ when $1.5 \times 10^3 \mu\text{Mol}$, $2.9 \times 10^3 \mu\text{Mol}$, $4.4 \times 10^3 \mu\text{Mol}$, $5.9 \times 10^3 \mu\text{Mol}$ and $7.4 \times 10^3 \mu\text{Mol}$ SOA, respectively were used Fig. 3.

There was also inhibition of enzyme activity when EDTA was used. Neuraminidase activity was inhibited from $9.0 \times 10^1 \mu\text{Mol min}^{-1}$ to $5.0 \times 10^1 \mu\text{Mol min}^{-1}$, $1.2 \times 10^2 \mu\text{Mol min}^{-1}$ to $8.0 \times 10^1 \mu\text{Mol min}^{-1}$, $1.6 \times 10^2 \mu\text{Mol min}^{-1}$ to $1.0 \times 10^2 \mu\text{Mol min}^{-1}$ and $1.7 \times 10^2 \mu\text{Mol min}^{-1}$ to 5.0×10^1

$\mu\text{Mol min}^{-1}$ when $2.9 \times 10^3 \mu\text{Mol}$, $4.4 \times 10^3 \mu\text{Mol}$, $5.9 \times 10^3 \mu\text{Mol}$ and $7.4 \times 10^3 \mu\text{Mol}$ of EDTA, respectively were used. There was no inhibition of enzyme activity when $1.5 \times 10^3 \mu\text{Mol}$ of EDTA was used Fig. 4.

DISCUSSION

The terminal sialic acid from sugar residues and glycoproteins is cleaved by the enzyme neuraminidase and the detection of the neuraminidase rests on the assay of free N-acetylneuraminic (sialic) acid split from the substrate^[15]. The activity of Clostridium chauvoei (jakari strain) neuraminidase is therefore a measure of the amount of N-acetylneuraminic (sialic) acid cleaved from fetuin.

The result of this study suggests that the compounds used inhibited the activity of dialysed Clostridium chauvoei (jakari strain) neuraminidase *in vitro*. This agrees with the findings of Oladele^[16] who also reported a similar finding when these inhibitors were tested on Newcastle disease Kudu 113 virus neuraminidase *in vitro* at different concentrations from those reported in the present study. The inhibition of neuraminidase activity in this study was not proportional to the concentrations of inhibitors used. This disagrees with the former report^[16] that the inhibition of Newcastle disease Kudu 113 virus neuraminidase was proportional to the concentrations of inhibitors used. In the present study, there was no inhibition of neuraminidase activity when concentrations of $2.9 \times 10^3 \mu\text{Mol AgNO}_3$, $2.9 \times 10^3 \mu\text{Mol PNPO}$ and $1.5 \times 10^3 \mu\text{Mol EDTA}$, respectively were used. With this finding, it is tempting to speculate that these concentrations of the compounds that failed to inhibit neuraminidase activity may not be of significance in clinical situations. In the present study, the type of inhibition, whether competitive, non-competitive or uncompetitive was not investigated.

In another study, natural products (Tamarindus indicus and Combretum fragrans) were reported to inhibit neuraminidase activity in a dose dependent fashion and the pattern of inhibition was non competitive^[17]. Blackleg is a devastating disease of cattle in Nigeria and annual losses of cattle to the disease in the country have been estimated at six hundred (600) million naira^[8]. This is the first time the effect of the compounds used in the present study on Clostridium chauvoei (jakari strain) neuraminidase activity is investigated, in an attempt to circumvent the use of conventional penicillin in the chemotherapy of blackleg. The use of neuraminidase inhibitors to manage diseases clinically has been advocated^[18,19]. These inhibitors (AgNO_3 , PNPO, SOA and EDTA) showed good therapeutic prospect against blackleg *in vitro* and should therefore, be properly investigated *in vivo* to ascertain their efficacy clinically to manage the disease.

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REFERENCES

1. Armstrong, H.L. and J.K. MacNamee, 1950. Blackleg in deer. *J. Am. Vet. Med. Assoc.*, 117: 212-214.
2. Osiyemi, T.I.O., 1975. The aetiology and data on seasonal incidence of clinical blackleg in Nigerian cattle. *Bull. Anim. Hlth. Prod. Afri.*, 23: 367-370.
3. Useh, N.M., 2002. The production and characterization of neuraminidase (sialidase) from *Clostridium chauvoei* (jakari strain). M.Sc. Thesis, Ahmadu Bello University, Zaria, Nigeria, pp: 224.
4. Muller, H.E., 1976. Neuraminidase als pathogenitätsfaktor bei mikro biellen infektionen. *Zbl. Bakt. Hyg.*, 235: 106-110.
5. Esievo, K.A.N., D.I. Saror, A.A. Ilemobade and M.H. Hallaway, 1982. Variation in erythrocyte surface and free serum sialic acid concentrations during experimental *Trypanosoma vivax* infection in cattle. *Res. Vet. Sci.*, 32: 1-5.
6. Von Itzstein, M., W.Y. Wu, G.B. Kok, M.S. Pegg, J.C. Dyason and B. Jin, 1993. Rational design of potent sialidase-based inhibitors of influenza virus replication. *Nature*, 363: 418-423.
7. Hayden, F.G., A.D. Osterhalls, J.J. Treanor, D.M. Fleming, F.Y. Aoki and K.G. Nicholson, 1997. Efficacy and safety of the neuraminidase inhibitor Zanamivir in the treatment of influenza virus infections. *New Engl. J. Med.*, 337: 874-880.
8. Useh, N.M., 2006. The possible role of clostridial neuraminidase (sialidase) in the pathogenesis of blackleg. PhD Dissertation, Ahmadu Bello University, pp: 172.
9. Princewill, T.J.T., 1965. Effect of calcium chloride on germination and pathogenicity of spores of *Clostridium chauvoei*. *J. Comp. Pathol.*, 75: 343-351.
10. Dowell, V.R. and T.M. Hawkins, 1981. Laboratory methods in anaerobic bacteriology. Centre for Disease Control (CDC) Laboratory Manual, HHS Publication, Atlanta, Georgia, pp: 1-96.
11. Heuermann, D., P. Roggentin, R.G. Kleinneidam and R. Schauer, 1991. Purification and characterization of a sialidase from *Clostridium chauvoei* NC08596. *Glycoconjugate J.*, 8: 95-101.
12. Mollison, P.L., 1979. Preparation of acid citrate dextrose. In: blood transfusion in clinical medicine, sixth edition. Blackwell Scientific Publications. London, pp: 65-69; 724-725.
13. Useh, N.M., O.J. Ajanusi, K.A.N. Esievo and A.J. Nok, 2006. Characterization of a sialidase (neuraminidase) isolated from *Clostridium chauvoei* (jakari strain). *Cell Biochem. Func.*, 24: 347-352.
14. Aminoff, D., 1961. Methods for the estimation of N-acetylneuraminic acid and their application to hydrolysis of sialomucoids. *Biochem. J.*, 81: 384-392.
15. Webster, R.G. and C.H. Campbell, 1972. An inhibition tests for identifying the neuraminidase antigen of influenza virus. *Av. Dis.*, 16: 1057-1066.
16. Oladele, S.B., P. Abdu, A.J. Nok, K.A.N. Esievo and N.M.N.M. Useh, 2002. Effect of some inhibitors on neuraminidase of newcastle disease virus Kudu 113 strain. *Vet. Arh.*, 72: 185-194.
17. Useh, N.M., A.J. Nok, S.F. Ambali and K.A.N. Esievo, 2004. The inhibition of neuraminidase activity by methanolic extracts of the stem barks of *tamarindus indicus* and *cssombretum fragrans*. *J. enzy. Inhib. Med. Chem.*, 19: 339-342.
18. Allen, U., 2000. The battle against influenza: The role of neuraminidase inhibitors in children. *Paediatr. Child Health*, 8: 457-460.
19. Thobhani, S., B. Ember, A. Siriwardena and G. Boons, 2003. Multivalency and the mode of action of bacterial sialidases. *J. Am. Chem. Soc.*, 125: 7154-7155.