

Influence of Longdan Xiegan Decoction on Body Weights and Immune Organ Indexes in Ducklings Intoxicated with Aflatoxin B₁

¹S.N. Guo, ³S.Q. Liao, ¹R.S. Su, ¹R.Q. Lin, ¹Y.Z. Chen, ¹Z.X. Tang, ²H. Wu and ¹D.Y. Shi
¹College of Veterinary Medicine, ²College of Life Science, South China Agricultural University,
³Institute of Veterinary Medicine, Guangdong Academy of Agricultural Sciences,
510640 Guangzhou, Guangdong Province, China

Abstract: To explore the influence of Longdan Xiegan Decoction (LXD) on body weights and the immune organ indexes in ducklings administrated with Aflatoxin B₁ (AFB₁), ninety, 7 days old ducklings were randomly divided into three groups (groups I-III). Group I was used as a blank control. Group II was administered with AFB₁ (0.1 mg kg⁻¹ body weight). Group III was administered with AFB₁ (0.1 mg kg⁻¹ body weight) plus LXD (as 2% of feed). All treatments were given once daily for 21 days. It showed that the ducklings' bursa of fabricius and thymus indexes, body weights in group II significantly decreased when compared with group I (p<0.01). Furthermore, the spleen indexes significantly decreased (p<0.01). However, the ducklings' bursa of fabricius and thymus indexes, body weights in group III ducklings significantly increased when compared with group II (p<0.01). In addition, the spleen indexes significantly decreased (p<0.01). These results revealed that AFB₁ significantly affect ducklings' growth and immune organs development. However, LXD significantly ameliorated the negative effects induced by AFB₁.

Key words: Longdan Xiegan Decoction, body weights, immune organ indexes, aflatoxin B₁, growth, thymus

INTRODUCTION

Aflatoxins, a group of well-known toxic metabolites are produced primarily by certain strains of fungi *Aspergillus flavus* and *Aspergillus parasiticus* under favorable temperature and humidity. In subtropical and tropical areas, the contamination of aflatoxins in food crops is common especially for the crops containing high starch and lipid content (Ostrowski-Meissner *et al.*, 1995; Bokhari, 2002; Aba-Alkhalil *et al.*, 2004). It was reported that aflatoxins could be detected in 96.1% of the 334 tested commercial feeds and raw materials collected from Asia (Chen and Rawlings, 2008). Among them Aflatoxin B₁ (AFB₁) is the most prevalent and toxic for human, land animals and aquatic organisms, mostly by its strong carcinogenic, mutagenic and teratogenic effects (Santacroce *et al.*, 2008; Han *et al.*, 2008). Aflatoxicosis in birds is highly relevant as a problem affecting the broiler chicken industry due to its socioeconomic impact on mortality, feed conversion index, production costs and the negative effects on public health related to the human consumption of exposed animals.

Pretreatment of animals with substances tested before hepatotoxin administration, e.g., carbon tetrachloride, aflatoxin B₁, is a common experimental model for investigation potential hepatoprotective activity

(Rafatullah *et al.*, 2008; Chavda *et al.*, 2010; Bigoniya *et al.*, 2010; Bitiren *et al.*, 2010). In the studies, researchers have used this model to test whether Longdan Xiegan Decoction (one of the most famous Traditional Chinese Medicine prescriptions) could mitigate toxicity of aflatoxin B₁ with respect to growth performance and the relative weights of the immune organs (spleen, thymus and bursa of fabricius) in ducklings intoxicated with aflatoxin B₁.

MATERIALS AND METHODS

Longdan xiegan decoction preparation: In this research, Longdan Xiegan Decoction (LXD), a famous traditional Chinese medicine prescriptions was comprised of 10 medicinal materials including Radix Gentianae, Radix Scutellariae, Fructus gardenia, Rhizoma alismatis, Caulis Aristolochiae Manshuriensis, Semen plantaginis, Radix Angelica Sinensis, Radix rehmanniae, Radix bupleuri and Radix Glycyrrhizae. In a certain ratio, the ten ingredient herbs were crushed fully and used to feed ducklings (as 2% of feed).

Animals and treatments: All experimental protocols were approved by the Ethics Committee of South China Agricultural University. This study was carried out on 90

Guangdong white ducklings weighing 180-200 g body weight. About 7 days old ducklings were obtained from Research Center of Experimental Animals at South China Agricultural University. The animals were randomly divided into three equal groups containing 30 ducklings with an equal number of male and female ducklings. Group I was used as control and intragastrically administered with Dimethyl Sulfoxide (DMSO). Group II was intragastrically administered with AFB₁ (0.1 mg kg⁻¹ body weight). Group III was intragastrically administered with AFB₁ (0.1 mg kg⁻¹ body weight) plus LXD (as 2% of feed). These treatments were administrated once daily for a period of 21 days under the same condition. AFB₁ was diluted with DMSO for experimental ducklings. All ducklings had free access to water and food at room temperature during the study.

Determinations of body weights and of the immune organ indexes: On 7th, 14th and 21st day after treatment, five ducklings from every experimental group were randomly taken out to be weighed. Afterwards, the ducklings' immune organ (spleen, thymus and bursa of fabricius) were taken out and weighed. The duckling's body weights are expressed in grams. The immune organ indexes are expressed in the ducklings' immune organs (spleen, thymus and bursa of fabricius) relative weight (g kg⁻¹). The immune organ relative weight equal to the immune organ weight/the duckling's body weight.

Statistical analysis: The statistical significance of differences between groups in these studies was determined using a one-way analysis of variance and the results were presented as the mean±SE. The significance level was p<0.05.

RESULTS AND DISCUSSION

As shown in Table 1, the body weight was significantly affected after ducklings were intragastrically administered with AFB₁ for 7th, 14th and 21st days. The body weight in group I ducklings decreased by 35.38, 52.38 and 51.92%, respectively compared with group II (p<0.01). However, the body weight increased in group III ducklings by 11.38, 17.89 and 24.41%, respectively compared with group II (p<0.01).

As shown in Table 2, the spleen index was significantly affected after ducklings were intragastrically administered with AFB₁ for 7th, 14th and 21st days. The spleen index in group I ducklings increased by 30.67, 41.85 and 49.33%, respectively compared with group II (p<0.01). However, the spleen indexes in group III ducklings decreased by 10.54, 17.62 and 18.30%, respectively compared with group II (p<0.01).

Table 1: Body weights for 7th, 14th and 21st days after the ducklings exposed to AFB₁

On 7th, 14th and 21st day after the administration of AFB ₁			
Groups	7	14	21
I	475.83±37.00	929.17±75.87	1460.00±128.73
II	307.50±26.42 ^{aa}	442.50±33.16 ^{aa}	702.00±58.280 ^{aa}
III	342.50±24.50 ^b	521.67±46.87 ^b	873.33±61.670 ^{bb}

Table 2: The spleen indexes for 7th, 14th and 21st days after the ducklings exposed to AFB₁

On 7th, 14th and 21st day after the administration of AFB ₁			
Groups	7	14	21
I	2.25±0.20	1.84±0.14	1.50±0.12
II	2.94±0.27 ^{aa}	2.61±0.19 ^{aa}	2.24±0.22 ^{aa}
III	2.63±0.25 ^b	2.15±0.18 ^{bb}	1.83±0.21 ^{bb}

Table 3: The thymus indexes for 7th, 14th and 21st days after the ducklings exposed to AFB₁

On 7th, 14th and 21st day after the administration of AFB ₁			
Groups	7	14	21
I	8.22±0.70	10.22±0.88	9.79±0.76
II	5.08±0.41 ^{aa}	7.17±0.68 ^{aa}	6.09±0.62 ^{aa}
III	6.26±0.54 ^{bb}	8.63±0.59 ^{bb}	7.85±0.68 ^{bb}

Table 4: The bursa of fabricius indexes for 7th, 14th and 21st days after the ducklings exposed to AFB₁

On 7th, 14th and 21st day after the administration of AFB ₁			
Groups	7	14	21
I	2.55±0.25	2.24±0.16	1.78±0.14
II	1.74±0.10 ^{aa}	1.51±0.13 ^{aa}	1.24±0.16 ^{aa}
III	2.03±0.24 ^b	1.85±0.15 ^b	1.54±0.18 ^b

^ap<0.05, ^{aa}p<0.01, significantly different, group II compared with group I. ^bp<0.05, ^{bb}p<0.01, significantly different, group III compared with group II. (the same following)

As shown in Table 3, the thymus index was significantly affected after ducklings were intragastrically administered with AFB₁ for 7th, 14th and 21st days. The thymus index in group I ducklings decreased by 38.20, 29.84 and 37.80%, respectively compared with group II (p<0.01). However, the thymus index in group III ducklings increased by 23.23, 20.36 and 28.89%, respectively compared with group II (p<0.01).

As shown in Table 4, the bursa of fabricius index was significantly affected after ducklings were intragastrically administered with AFB₁ for 7th, 14th and 21st days. The bursa of fabricius index in group I ducklings decreased by 31.76, 32.59 and 30.34%, respectively compared with group II (p<0.01). However, the bursa of fabricius index in group III ducklings increased by 16.67, 22.52 and 24.19%, respectively compared with group II (p<0.01).

It was well known the AFB₁ and its metabolites can accumulate in animal tissues after AFB₁ exposure. Aflatoxin contamination has been a potential threat to the health of humans and animals (Agag, 2004; Meissonnier *et al.*, 2007). In addition, the greatest economic impacts include reduced productivity,

suppressed immune function, pathological effects on organs and tissues (Anonymous, 1989). It was reported that the inclusion of AFB₁ 200 ppb in the diet significantly reduced the growth of mule ducklings, significantly affect immune organ development (Cheng *et al.*, 2001). Compared to the controls, the relative weights of the livers and kidneys increased significantly in the broilers exposed to aflatoxins (Raju and Devegowda, 2000).

In this study for 7th, 14th and 21st days after ducklings administrated with AFB₁, respectively it showed that the ducklings' bursa of fabricius indexes, thymus indexes and body weights in group II significantly decreased, compared with group I ($p < 0.01$). Furthermore, the spleen indexes significantly increased ($p < 0.01$). In all, AFB₁ reduced the growth of the ducklings caused tymus and bursa of fabricius atrophy and induced swelling of the spleen.

Traditional Chinese Medicines (TCMs) have been practiced in China and other eastern countries over long period of time. Usually, TCMs are administrated in the form of formulated preparations consisting of several even >10 ingredient herbs in a certain ratio which may provide polyvalent biological actions. Longdan Xiegan Decoction (LXD), a traditional Chinese formulation has been widely practiced to treat jaundice, conjunctival congestion, earache, scrotum and extremitas inferior eczema, etc. in Chinese medicine for thousands of years (Chinese Pharmacopoeia Commission, 2005).

It has been suggested that for some herbs it is the natural antioxidants they contain conferred their biological activities (Loliger, 1991; Zhu *et al.*, 2004). Most of the antioxidant potentials in herbs and spices are due to the redox properties of phenolic compounds that allow them to act as reducing agents, hydrogen donors and free radicals quenchers (Shahidi *et al.*, 1992).

Longdan Xiegan Decoction is one of the most famous traditional Chinese medicine prescriptions which is a combinatorial formula including ten drugs. It has been used to treat damp-heat in the liver, gall bladder, congested eyes or as an important drug for acute/chronic cholecystitis (Meng, 2006; Leng, 2006). The prescription consists of 10 medicinal materials including Radix Gentianae, Radix Scutellariae, Fructus gardenia, Rhizoma alismatis, Caulis Aristolochiae Manshuriensis, Semen plantaginis, Radix Angelica Sinensis, Radix rehmanniae, Radix bupleuri and Radix Glycyrrhizae.

Although, LXD preparation as a whole has not been elucidated for its bioactive constituents, some of the compounds contained in the ingredient herbs have been tested and have shown pharmacological properties. For instance, gentiopicroside isolated from Radix Gentianae

root indicated antibacterial and free radical scavenging activities (Kumarasamy *et al.*, 2003). The flavonoids baicalin and baicalein from Radix Scutellariae root showed multi functional efficacies including anti-bacterial, antiviral, anti-inflammation and liver protective activities (Pan and Feng, 1994). In the study, researchers assessed the effect of LXD on body weight and immune organs indexes in ducklings administrated with AFB₁. It showed that the bursa of fabricius indexes, thymus indexes and body weights in group III ducklings (administrated with AFB₁ plus LXD), significantly increased when compared with group II ($p < 0.01$). In addition, the spleen indexes in group III ducklings significantly decreased, compared with group II ($p < 0.01$).

CONCLUSION

In this research, these results showed that the ducklings' bursa of fabricius and thymus indexes, body weights in group II significantly decreased, compared with group I. In addition, the spleen indexes significantly decreased. However, the ducklings' bursa of fabricius and thymus indexes, body weights in group III ducklings significantly increased, compared with group II. Besides, the spleen indexes significantly decreased. So it was concluded that Longdan Xiegan Decoction could significantly ameliorate AFB₁ negative effect on ducklings' growth performance and the immune organs' development.

ACKNOWLEDGEMENTS

This research was supported by the Guangdong Province Science Foundation of China (Grant No.: 9151008002000003), the National Natural Science Foundation of China (Grant No. 38071900), the National Twelve-Five Technological Supported Plan of China (No. 2011BAD34B01), the Guangdong province and Ministry of education Cooperation project by Production, Study and research of China (Grant No. 2008B090500250). The S.N. Guo, S.Q. Liao and R.S. Su contributed equally to this research.

REFERENCES

- Aba-Alkhalil, A.R.A., G.H. Ibrahim, S.M. Al-Rehiayani, A.I. Askar and K.A. Osman, 2004. Susceptibility of some varieties of date fruits to support the production of aflatoxins: Analysis by high performance liquid chromatography. *Pak. J. Biol. Sci.*, 7: 1937-1941.

- Agag, B.I., 2004. Mycotoxins in foods and feeds: 1-aflatoxins. Ass. Univ. Bull. Environ. Res., 7: 173-206.
- Anonymous, 1989. Mycotoxins Economic and Health Risks. Council for Agricultural Science and Technology, Ames, pp: 91.
- Bigoniya, P., A. Shukla and C.S. Singh, 2010. Evaluation of hepatic microsomal enzyme functional integrity on picroliv pretreatment against CCl₄ induced hepatotoxicity. Int. J. Pharmacol., 6: 200-207.
- Bitiren, M., D. Musa, A. Ozgonul, M. Ozaslan and A. Kocyigit *et al.*, 2010. Protective effects of green tea (*Camelia sinensis*), *Hypericum perforatum* and *Urtica dioica* on hepatic injury and lymphocyte DNA damage induced by carbon tetrachloride in wistar rats. Int. J. Pharmacol., 6: 241-248.
- Bokhari, F.M., 2002. Aflatoxins production by *Aspergillus flavus*, isolated from different foodstuffs commonly used in Jeddah region, Saudi Arabia. Pak. J. Biol. Sci., 5: 69-74.
- Chavda, R., K.R. Vadalia and R. Gokani, 2010. Hepatoprotective and antioxidant activity of root bark of *Calotropis procera* R. Br (Asclepediaceae). Int. J. Pharmacol., 6: 937-943.
- Chen, H.Y. and R. Rawlings, 2008. The truth of mycotoxin contamination of feed in Asia region. China Poult., 30: 33-35.
- Cheng, Y.H., T.F. Shen, V.F. Pang and B.J. Chen, 2001. Effects of aflatoxin and carotenoids on growth performance and immune response in mule ducklings. Comp. Biochem. Physiol. C. Toxicol. Pharmacol., 128: 19-26.
- Chinese Pharmacopoeia Commission, 2005. Pharmacopoeia of the People's Republic of China. Vol. 1, People's Medical Publishing House, Beijing, China, pp: 64-524.
- Han, X.Y., Q.C. Huang, W.F. Li and Z.R. Xu, 2008. Changes in growth performance, digestive enzyme activities and nutrient digestibility of cherry valley ducks in response to aflatoxin B₁ levels. Livest. Sci., 119: 216-220.
- Kumarasamy, Y., L. Nahar and S.D. Sarker, 2003. Bioactivity of gentiopicoside from the aerial parts of *Centaurium erythraea*. Fitoterapia, 74: 151-154.
- Leng, F.D., 2006. Modified longdan xiegan decoction in treating 30 cases of acute cholecystitis. J. Emerg. Trad. Chin. Med., 15: 425-425.
- Loliger, J., 1991. The Use of Antioxidants in Foods. In: Free Radicals and Food Additives, Aruoma, I. and B. Halliwell (Eds.). Taylor Francis, London, pp: 121-150.
- Meissonnier, G.M., J. Laffitte, N. Loiseau, E. Benoit and I. Raymond *et al.*, 2007. Selective impairment of drug-metabolizing enzymes in pig liver during subchronic dietary exposure to aflatoxin B₁. Food. Chem. Toxicol., 45: 2145-2154.
- Meng, Q.C., 2006. The experience of treating ache of Shingles and the residual ache with modified longdanxiegan decoction. Gansu. J. Trad. Chin. Med., 19: 22-22.
- Ostrowski-Meissner, H.T., B.R. LeaMaster, E.O. Duerr and W.A. Walsh, 1995. Sensitivity of the Pacific whiteshrimp, *Penaeus vannamei*, to aflatoxin B₁. Aquaculture, 131: 155-164.
- Pan, F. and Y.X. Feng, 1994. Research progress in Radix Scutellariae. Nat. Prod. Res. Dev., 6: 61-65.
- Rafatullah, S., A. Al-Sheikh, S. Alqsoumi, M. Al-Yahya, K. El-Tahir and A. Galal, 2008. Protective effect of fresh radish juice (*Raphanus sativus* L.) against carbon tetrachloride-induced hepatotoxicity. Int. J. Pharmacol., 4: 130-134.
- Raju, M.V. and G. Devegowda, 2000. Influence of esterified glucomannan on performance and organ morphology, serum biochemistry and haematology in broilers exposed to individual and combined mycotoxicosis. Br. Poult. Sci., 41: 640-650.
- Santacroce, M.P., M.C. Conversano, E. Casalino, O. Lai, C. Zizzadoro, G. Centoducati and G. Crescenzo, 2008. Aflatoxins in aquatic species: Metabolism, toxicity and perspectives. Rev. Fish Biol. Fish., 18: 99-130.
- Shahidi, F., P.K. Janitha and P.D. Wanasundara, 1992. Phenolic antioxidants. Crit. Rev. Food Sci. Nutr., 32: 67-103.
- Zhu, Y.Z., S.H. Huang, B.K. Tan, J. Sun, M. Whiteman and Y.C. Zhu, 2004. Antioxidants in Chinese herbal medicines: A biochemical perspective. Nat. Prod. Rep., 21: 478-489.