

Prevalence of Gastrointestinal Parasites of Stray Dogs with *in vivo* Assessment using the Garlic (*Allium sativum*)

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Page No.: 161-168 Volume: 20, Issue 6, 2021 ISSN: 1680-5593 Journal of Animal and Veterinary Advances Copy Right: Medwell Publications Abstract: A surveillance of zoonotic and other gastrointestinal parasites of stray dogs along with assessing the efficacy of garlic-based treatment was conducted in Alexandria, Egypt. Therefore, forty stray dogs were examined and divided into two groups; group 1: Dogs were given a high dose of five garlic cloves/dog twice daily, group 2: Dogs received a proposed strategy of gradual lower doses based on weight; small weight dogs (13-14 kg) received 1/4 clove twice/day, medium weight (16-18 kg) dogs received 1/2 cloves twice/day, larger dogs (19-20 kg) received 3/4 clove twice/day, heavy weight (>20 kg) dogs received one clove twice/day. The overall prevalence of gastrointestinal parasitic infections was 90.0%. Identified parasites were Isospora (100%), Toxocara canis (27.78%) and both Taenia spp. and Dipylidium caninum (5.56% each). Single infection with at least one parasite was revealed in 20 dogs and co-infections with more than one parasitic species was found in 16 dogs. Sex, age and weight of dogs were non-significant. All female dogs were infected. Upon the treatment with garlic, the coprological examination revealed a significant reduction in helminth eggs among dogs of the group 1 (79.9%; t = -3.121, $p = 0.006^*$) rather than those of group 2 while the number of protozoaloocysts was significantly reduced in both groups (t = -4.211, p = 0.001^* and t = -6.872, p = 0.000^* , respectively). The mean values of most of blood parameters measured were significantly positive like

HCT, MCV, MCH, MCHC, blood platelets and neutrophils. Kidney function tests revealed that uric acid significantly increased post treatment in both groups (t = 5.257, p = 0.000^{*} and t = 6.945, p = 0.000^{*} , respectively) while creatinine level remained within the normal values. Moreover, liver enzymes, particularly

INTRODUCTION

Stray dogs are abundant carnivores distributed worldwide. Egypt has a great community of stray dogs wandering freely among cities and villages and keeping in touch with humans and residential domesticated animals^[1]. Moreover, they are known to be the definitive hosts of various parasitic helminths and protozoa; some of them have a potential zoonotic importance, like *Toxocara canis, Dipylidium caninum* and *Taenia multiceps*^[2, 3]. As they communicate with the wildlife via. the direct contact or via. the sharing of ingested food, water or even soil infected with parasitic stages, stray dogs may acquire parasitic infections as well as they infect newly introduced dogs^[4-6].

In developing countries, infectious parasitic diseases are growing with the occurrence of unregulated populations of stray dogs in particular with the lack of veterinary care to improve the risk of the disease transmission^[7,8]. Meanwhile, the persistent indiscriminate use of anthelmintic drugs has led to the problem of resistance of parasites to conventional treatments^[9].

Garlic (*Allium sativum*) has been reported to be a parasiticide, amoebicide, larvicide as well as immunostimulant^[10]. Garlic oil has a broad antimicrobial spectrum as it possesses antibacterial and anti-parasitic effects. Moreover, it influences the growth of at least 12 different human and nonhuman parasites with a potent immunomodulatory activity^[11]. However, it contains 17 amino acids, like arginine, at least 33 organosulphate compounds, as alliinandallicin, eight minerals (germanium, calcium, copper, iron, potassium, magnesium, selenium and zinc), enzymes as allinase and vitamins A, B1 and C. The efficacy of garlic against parasitic infections has been reported also in rabbits^[12].

To the author's knowledge, there is a paucity data regarding the estimation of the prevalence of gastrointestinal parasites among stray dogs in Egypt, particularly zoonotic parasites. Therefore, the current work was conducted to provide recent data on the distribution of zoonotic and other gastrointestinal parasites of stray dogs in Alexandria province as well as the promising use of the garlic as a natural remedy against parasitic infections. AST and AP were significantly increased post treatment. There was a high risk of human zoonotic parasites transmission in the study area and the garlic is strongly recommended as an anthelmintic and a potential alternative to overcome rising resistance to conventional anthelmintics.

MATERIALS AND METHODS

Animals and husbandry: Forty stray dogs of different sexes (22 males and 18 females) and ages were collected randomly from the center and west Alexandria province, Egypt and trapped inregistered animal house with registration number 584813328 under control of Ministry of Supply and Internal Trade, Alexandria. Clinical examination of dogs including body weight, temperature and pulse was done. Infected dogs were divided into two groups; group 1 (n = 18) included dogs aged 9-36 (average 22.5) months and weighed 12-24 (average 18) kg and group 2 (n = 18) included dogs aged 12-36 (average 22) months and weighed 13-24.5 (average 17.5) kg.

Collection of fecal/blood samples: From the rectum of stray dogs, each fresh fecal sample was collected in a sterile glass bottle labeled with serial number, age and sex. Visual examinationwas done to identify adult nematodes and cestode proglottids. Then, the direct microscopy was applied using concentration and flotation techniques^[13]. Identification of parasitic stages was based on morphological characteristics using $\times 40$ and $\times 100$ magnifications^[14]. Moreover, blood samples were collected for Complete Blood Count (CBC) analysis, kidney and liver function tests. The determination of the intensity of infection also estimated through counting protozoan cysts and helminth eggs in 0.1 mL of sediment then multiplying by 10 to obtain the number of cysts or eggs per one gram stool^[15].

Protocol of treatment: Coprologically, enteroparasitesinfected dogs (protozoan cysts/helminth eggs) were allocated into two groups receiving the dose of garlic for ten days. Group 1 received a high dose of garlic (five cloves per dog twice daily) while group 2 received a proposed strategy of gradual lower doses based on weight; small weight dogs (13-14 kg) received ¼ clove twice a day, medium weight (16-18 kg) dogs received ¼ cloves twice a day, larger dogs (19-20 kg) received ¾ clove twice a day, heavy weight (>20 kg) dogs received one clove twice a day. Garlic clove can weigh up to 5 g administered as minced raw garlic mixed the drinking water. Assessment of treatment: The treatment assesses by comparing findings of fecal examination both macroscopically and microscopically prior to and post treatment. Blood samples were collected on the third day after the end of the administration of garlic for all treated dogs to perform the CBC and biochemical analyses comparing results prior to and post treatment.

Statistical analysis: Data were statistically analyzed using a Microsoft Excel spreadsheet and descriptive statistics. The prevalence of infection was calculated for all data as the number of infected divided by the number of individuals and multiplied by 100. Chi-square was used to assess the association of risk factors on the prevalence of parasites. T-Independent Samples Test (Levene's Test) was used to compare the normal distribution quantitative data and z-Mann-Whitney U for non-normal distribution quantitative data between two groups and t-Paired Samples Test for normal distribution quantitative data, z-wilcoxon for non-parametric quantitative data comparing two related samples. SPSS was used to analyze the data statistically. The significance level was considered at p<0.05.

RESULTS

The present study revealed that 36 (90.0%) out of 40 surveyed stray dogs exhibited endoparasitism by the use of coprological examination. Among infected dogs, 20 (55.56%) animals had a single infection and 16 (44.44%) dogs were co-infected with helminth

eggs/protozoaloocysts. The identified parasites species were *Isospora* sp. (100%), *Toxocara canis* (27.78%), *Taenia* spp. and *Dipylidium caninum* (5.56% each) (Table 1).

Clinically, it has been found that there was nonsignificant differences among infected and uninfected dogs relative sex, age and weight. Dogs aged >18 months were highly infected with a considerable risk factor of 95% Cl 1.2(1.003-1.435). All female dogs were infected (Table 2).

Concerning the counting of helminth eggs, there was a significant reduction in number of eggs by -79.9% (t =-3.121, p = 0.006*) among dogs of group 1. On the other hand, group 2 showed no significant reduction (t = -1.031, p = 0.317). Regarding the counting of protozoaloocysts, both groups exhibited a significant reduction (t = -4.211, p = 0.001* and t = -6.872, p = 0.000*, respectively). The percentage of reduction among dogs of group 1 was higher (-62.8%) than that of group 2 (-62.8%) (Table 3).

Furthermore, findings of the blood analysis showed that in both groups, there was a significant decrease in the RBCs count post treatment (t = -14.875, p = 0.000* and t = -9.386, p = 0.000*, respectively). On the other hand, blood platelets significantly increased post treatment (t = 3.629, p = 0.002* and t = 3.056, p = 0.007*, respectively). Hemoglobin percent was significantly higher in group 2 (1.81%). The mean hematocrit (HCT) values were significantly decreased in both groups (-26.67%; -30.08%, respectively). The mean values of MCV, MCH, MCHC, platelets, RDW-SD and RDW-CV were significantly increased in dogs of group 1 compared to those of group 2 with the exception of the mean corpuscular hemoglobin

Table 1: The prevalence of parasitic gastrointestinal infections among examined dogs

	Number		Percentage		
Dog fecal samples	Single infection	Mixed infection	Single infection	Mixed infection	
Infected	20	16	55.56	44.44	
Isospora spp.	36		100		
Taenia spp.	2		5.56		
Toxocara canis	10		27.78		
Dipylidium caninum	2		5.56		
Uninfected	4		10.0		
Total	40		100		

Table 2: The clinical examination of examined dogs with determining associated risk factors

Risk factors	Infected dogs $(n = 36)$		Uninfected dogs					
	No.	Percentage	No.	Percentage	x2/t	p-values	95% Cl	
Sex								
Males	18	50	4 100		0.114^		$0.818(0.672-0.996)^{P}$	
Females	18	50	0	0				
Age								
<18 months	16	44.4	0 0 0.136^		$1.2(1.003-1.435)^{R}$			
>18 months	20	55.6	4	100				
Mean age	22.33±10.696		24±0.0		-0.935	0.356 ^t	1.068(0.922-1.237) ^R	
Mean body weight	16.639±3.1884		16.875±2.7195		-0.142	0.888^{t}	0.941(0.590-1.501) ^p	
Mean temperature	39.233 ± 0.7657		40±1.1547		-1.811	0.078^{t}	$2.95(0.746-11.671)^{R}$	
Mean pulse	82.22±8.656		87.5±8.66		-1.302	0.193 ^z	1.079(0.928-1.255) ^R	

No = Number of examined dogs; % = Percentage of infected/uninfected dogs; $^$ = Fisher's Exact Test; R = Risk factors; P = Protective factors; Z = Mann-Whitney U; t = Independent Samples Test (Levene's Test)

(MCHC) which was higher in dogs of group 2. The platelet distribution width (PDW) was significantly decreased (-54.17%; -49.315) in both groups.

Non-significantly, the Mean Platelet Volume (MPV) values were 1.92 and 0.93% in both groups. A significant reduction in the mean value of WBCs in dogs of group 1 (-6.27%) compared to that of group 2 (29.45%).

Neutrophils count was significantly higher (11.53%) in dogs of group 1 than those of group 2 (-4.58%).

Significantly, lymphocytes percent increased in group 1 (260.71%) compared to group 2 (114.20%). The number of eosinophils significantly increased in group 2 (137.5%) than in group 1 (48.38%). Finally, the number of monocytes was significantly decreased in group 1 (-65.33%) and group 2 (-51.60%) (Table 4). Concerning the safety margin of the garlic, kidney function tests revealed that among both groups, uric acid significantly increased post treatment (t = 5.257, p = 0.000° and t =

Table 3: The efficacy of the administration of various doses of garlic against parasitic infections

Coprological examination	Infected dogs							
	Group 1 (n = 18)			Group 2 (n = 18)				
	Pre-treatment	Post-treatment	Percentage	Pre-treatment	Post-treatment	Percentage		
Helminth eggs								
Mean±SD	27.78±35.572	5.56 ± 7.048	-79.9	7.78±19.268	4.44 ± 8.556	-42.93		
t	-3.121			-1.031				
р	0.006*			0.317				
Protozoalo ocysts								
Mean±SD	68.89±37.083	25.56±17.564	-62.8	68.89±24.944	35.56±15.424	-48.38		
t	-4.211			-6.872				
р	0.001*			0.000*				

*Significance at the level $p \le 0.05$

Table 4: The Complete Blood Count (CBC) of infected dogs prior to and post treatment with garlic

	Infected dogs							
	Group 1 (n = 18)		Group 2 (n = 18)					
CBC findings	Pre-treatment	Post-treatment	Percentage	Pre-treatment	Post-treatment	Percentage		
RBCs								
Mean±SD t p	7.027±0.6225 -14.875 0.000*	4.51±0.4126	-35.81	7.05±0.9596 -9.386 0.000*	4.822±0.5247	-31.60		
HB	0.000			0.000				
Mean±SD t	15.367±1.2127 0.058	15.389±1.3394	0.14	15.311±1.9944 0.533	15.589±1.410	1.81		
р	0.954			0.601				
HCT			A 6 67			20.00		
Mean±SD	47.400±12.407	34.756±2.8126	-26.67	50.856±5.7996	35.556±3.962	-30.08		
t	-2.376a 0.018*			-10.104 0.000*				
p MCV	0.018			0.000				
Mean±SD	72.867±1.8337	77.111±2.3492	5.82	64.178±20.976	73.767±2.209	14.94		
t	8.811	77.111±2.5472	5.62	-2.684 ^z	13.101±2.20)	14.74		
p	0.000*			0.007*				
MCH	0.000			01007				
Mean±SD	22.022±0.9188	34.144±1.6614	55.04	29.144±16.01	33.156±2.311	13.76		
t	38.929			-2.071 ^z				
р	0.000*			0.038*				
MCHC								
Mean±SD	31.122±2.9775	43.911±2.8419	41.09	30.633±2.1321	43.956±1.544	43.49		
Z	22.745			33.391				
p	0.000*			0.000*				
Platelets	224.00.100.04	400 - 112 2421	42.22	202.00.100.26	476 000 . 50 0	04.07		
Mean±SD	334.89±128.84	480±113.2421	43.33	382.89±108.26	476.222±50.2	24.37		
t	3.629 0.002*			3.056 0.007*				
p RDW_SD	0.002			0.007				
Mean±SD	13.9±0.63	38.6±3.07	177.69	16.01±4.7	37.3±1.8	132.97		
t	40.608	50.0±5.07	177.09	-3.728a	57.5±1.0	152.97		
p	0.000*			0.000*				
RDW CV	0.000							
Mean±SD	1.8±0.2	11.9±0.6	561.11	3.8±4.5	12.05±0.2	217.10		
t	73.557			-3.597a				
р	0000*			0.000*				

	Infected dogs								
	Group 1 (n = 18)	Group 1 (n = 18)			Group 2 (n = 18)				
CBC findings	Pre-treatment	Post-treatment	Percentage	Pre-treatment	Post-treatment	Percentage			
PDW									
Mean±SD	49.1±5.1	22.5±1.5	-54.17	45.4±12.4	23.01±1.5	-49.31			
t	-21.391			-3.597b					
р	0.000*			0.000*					
MPV									
Mean±SD	10.4±0.3	10.6 ± 0.44	1.92	10.7±0.97	10.6±0.46	-0.93			
t	1.759			-0.291					
р	0.097			0.774					
WBCs									
Mean±SD	16.076 ± 3.3	15.067 ± 3.7	-6.27	12.9 ± 2.8	16.7±3.0	29.45			
t	-1.417 ^z			4.831					
р	0.157			0.000*					
Neutrophils (%)									
Mean±SD	31.778±18.4	35.444±8.2	11.53	32.7±15.0	31.2±5.7	-4.58			
t	-1.025 ^z			-0.719 ^z					
p	0.306			0.472					
Lymphocytes (%)									
Mean±SD	11.2 ± 4.4	40.4 ± 5.1	260.71	18.3±19.1	39.2 ± 5.9	114.20			
t	15.991			-2.725 ^z					
p	0.000*			0.006*					
Monocytes (%)	0.000			01000					
Mean±SD	42.7±20.2	14.8 ± 5.02	-65.33	37.4±23.4	18.1 ± 4.7	-51.60			
t	-6.294	11.0±5.02	05.55	-3.397	10.121.7	51.00			
p	0.000*			0.003*					
Eosinophils (%)	0.000			0.005					
Mean±SD	6.2±3.4	9.2±3.3	48.38	4.8±3.6	$11.4{\pm}1.8$	137.5			
t	2.442	<i></i>	10.50	6.597	11.1±1.0	107.0			
p	0.026*			0.000*					
Basophils (%)	5.020			0.000					
Mean±SD	$0.089 \pm .0583$	0.0 ± 0.0	-100	0.244±0.3	0.0 ± 0.0	-100			

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Table 4: Continue

Table 5: Findings of kidney and liver function tests among infected dogs prior to and post treatment with garlic Infected dogs

T7'1 (1'	Group 1 (n = 18)			Group 2 (n = 18)					
Kidney/liver function tests	Pre-treatment	Post-treatment	Percentage	Pre-treatment	Post-treatment	Percentage			
Uric acid									
Mean±SD	1.7±0.5	2.9±1.1	70.58	1.7±0.5	3.49±0.93	105.29			
t	5.257			6.945					
р	0.000*			0.000*					
Creatinine									
Mean±SD	0.86±0.13	1.19±0.17	38.37	1.02 ± 0.24	1.1±0.12	7.8400			
t	5.907			2.826					
р	0.000*			0.012*					
AST (SGOT)									
Mean±SD	55.33±14.6	150.93±28.02	172.78	65.11±23.7	151.07±39.6	132.02			
t	24.372			6.904					
р	0.000*			0.000*					
ALT (SGPT)									
Mean±SD	65.22±63.7	45.28±31.128	-30.57	53.22±34.2	50.6±23.4	-49.22			
t	-1.243w			-0.283b					
р	0.214		0.777						
ALP									
Mean±SD	52.44±13.9	69.52±33.2	32.57	44.7±31.7	77.52±40.6035	73.420			
t	2.75			2.297					
р	0.014*			0.035*					

Significance at the level p≤0.05

6.945, p = 0.000^* , respectively) but the creatinine level remained within the normal values. Meanwhile, liver enzymes, particularly Aspartate Aminotransferase (AST) and Alkaline Phosphatase (AP) were significantly increased post treatment with the garlic (Table 5).

DISCUSSION

Canine gastrointestinal parasitism is common with a worldwide distribution affecting various species of dogs, particularly stray dogs^[16-18]. The condition is frequent in Egypt, since, it causes harmful impacts on the health and productivity of dogs^[19-22]. The results of the current study revealed a high infection rate (90.0%) of gastrointestinal parasites of zoonotic potential and a public health hazard, like *Taenia* spp., *Toxocara canis*, *Dipylidium caninum* and *Isospora* spp. The current finding was almost similar to that reported in Tunisia (98·89%)^[23] but it was higher than thatgiven in Ethiopia (78.8%)^[24], Tanzania (67.2%)^[25], Nigeria (72.5%)^[26], Ethiopia (89.3%)^[27], Turkey (34.68%)^[28], Pakistan (26.8%)^[2] and Ghana (62.6%)^[29]. The widespread of gastrointestinal parasites revealed the lack of preventive control measures of those examined dogs and highlighted the existence of risk of zoonotic potentials from dogs in the study areas.

Otherwise, the present study showed that both single and mixed infections with helminth eggs/protozoanoocysts were revealed in 55.56 and 44.4% of surveyed dogs. Such finding coincided with that reported by Trasvina-Munoz et al. [30] in Mexico, whereas, in Ethiopia, Abere *et al.*^[24] detected that the mixed infection with two and more parasite species was common rather than the single infection. The most frequently occurred parasites in the infected dogs was *Isospora* spp. (100%), although, absence of zoonotic importance, it might be the causative agent of evidential intestinal epithelium damage. Moreover, the common canid arrow-headed worm, Toxocara canis was recovered in an infection rate of 27.78%. The percentage was higher than that recordedin Turkey (12.9%)^[28] and in Pakistan (10.5%)^[2]. In Ethiopia, Abere et al.^[24] showed a higher percentage (53.8%) than revealed in this study. Regarding Dipylidium caninum and Taenia spp., the current study revealed a prevalence of 5.56% for each. Concerning Dipylidium, the finding of the present work was higher than that recorded in Turkey $(3.22\%)^{[28]}$ and lower than that determined in Pakistan (11.8%)^[2] and in Mexico (16.50%)^[30]. Dipylidiasis is an important helminthic infection for the public health that occasionally occursin young children causing various gastrointestinal troubles^[31]. Children are noted more likely to be infected with Dipylidium caninum rather than adults^[32]. Conversely, the infection rate of Taenia spp. was higher than that reported in Turkey (4.84%)^[28] and lower than the finding recorded in Pakistan (6.57%)^[2] and in Mexico $(6.79\%)^{[30]}$.

Concerning sexes and ages of stray dogs, the prevalence of canine GIT parasitic infection wasnon-significant (p>0.05). Meanwhile, the elderly stray dogs (>18 months) were more infected rather than young dogs with a non-significant difference (p>0.05). Such finding agreed with that reported in Ghana by Johnson *et al.*^[33] who reported that 61.0% of dogs aged >12 months more infected.

The effect of curative efficacy of garlic with various doses on the parasitic intensity showed that administration of garlic reduced the intensity of protozoan cysts and helminth eggs with highly significant differences $(p \le 0.05)$. Based on the complete blood count analyses, not all dogs had anemia and this suggests that the routine clinical examination might be insufficient to diagnose anemia. Currently, the count of white blood cells increased. There was a non-significant difference in Hb estimation while neutrophils had no significant difference $(p>0.05)^{[34]}$. Regarding the prevalence of kidney and liver function tests, it has been found the concentration of uric acid and creatinine significantly increased post administration with garlic, moreover, the levels of the liver AP and AST were significantly increased ($p \le 0.05$). Thus, confirmatory, the current study revealed a clear therapeutic efficacy against the gastrointestinal parasites including helminths and protozoa, of stray dogs. Furthermore, Shenaway *et al.*^[29] reported an increased level of natural killer cells activity that promotes the immune system function and streng then the body's defense mechanism during the duration of treatment by garlic.

CONCLUSION

The present study showed a higher prevalence of the gastrointestinal parasites in stray dogs in Alexandria, Egypt with various species of intestinal parasites, somewitha potential zoonotic importance. The administration with garlic could reduce the density of protozoan cysts and helminth eggs. There was a high risk of human zoonotic parasites transmission in the study area and the garlic is strongly recommended as an anthelmintic and a potential alternative to overcome rising resistance to conventional anthelmintics

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Author's contribution: All authors organized the research, conducted the experiments and created the manuscript.

Compliance with ethical standards: Dogs were collected randomly from the center and west Alexandria province, Egypt and trapped in registered animal house with registration number 584813328 under control of Ministry of Supply and Internal Trade, Alexandria, Egypt.

REFERENCES

 Kutdang, E.T. and U. Onyejiaka, 2010. Prevalence of gastrointestinal parasites of dogs (*Canis familaris*) in Maiduguri, Northeastern Nigeria. Rep. Opin., 2: 1-4.

- 02. Khan, W., N.N. Nisa, S. Ullah, S. Ahmad and S.A. Mehmood *et al.*, 2019. Gastrointestinal helminths in dog feces surrounding suburban areas of Lower Dir district, Pakistan: A public health threat. Braz. J. Biol., 80: 511-517.
- Trasvina-Munoz, E., G. Lopez-Valencia, F.J. Monge-Navarro, J.C. Herrera-Ramirez and P. Haro *et al.*, 2020. Detection of intestinal parasites in stray dogs from a farming and cattle region of Northwestern Mexico. Pathogens, Vol. 9, No. 7. 10.3390/pathogens9070516
- Savilla, T.M., J.E. Joy, J.D. May and C.C. Somerville, 2011. Prevalence of dog intestinal nematode parasites in South central West Virginia, USA. Vet. Parasitol., 178: 115-120.
- 05. Gompper, M.E., 2014. Free-Ranging Dogs and Wildlife Conservation. Oxford University Press, Oxford, England, UK.,.
- Hemy, M., J. Rand, J. Morton and M. Paterson, 2017. Characteristics and outcomes of dogs admitted into Queensland RSPCA shelters. Animals, Vol. 7, No. 9. 10.3390/ani7090067
- 07. GDVS., 2011. Annual Report of the General Direction of Veterinary Services (DGSV). General Directorate of Veterinary Services, Ministry of Agriculture, Tunisia.
- Smith, L.M., S. Hartmann, A.M. Munteanu, D.P. Villa, R.J. Quinnell and L.M. Collins, 2019. The effectiveness of dog population management: A systematic review. Animals, Vol. 9, No. 12. 10.3390/ani9121020
- Mortensen, L.L., L.H. Williamson, T.H. Terrill, R.A. Kircher, M. Larsen and R.M. Kaplan, 2003. Evaluation of prevalence and clinical implications of anthelmintic resistance in gastrointestinal nematodes in goats. J. Am. Vet. Med. Assoc., 223: 495-500.
- Duke, J.A., 2002. Handbook of Medicinal Herbs. 2nd Edn. CRC Press, Boca Raton, FL, USA., ISBN: 0849312795, pp: 896.
- Shalaby, H.A. and T.K. Farag, 2014. Body surface changes in gastrointestinal helminthes following *in vitro* treatment with *Allium sativum* oil. J. Vet. Sci. Technol., Vol. 5, No. 1. 10.4172/2157-7579.1000153
- Toulah, F.H. and M.M. Al-Rawi, 2007. Efficacy of garlic extract on hepatic coccidiosis in infected rabbits (*Oryctolagus cuniculus*): Histological and biochemical studies. J. Egypt. Soc. Parasitol., 37: 957-968.
- Cringoli, G., L. Rinaldi, M.P. Maurelli and J. Utzinger, 2010. FLOTAC: New multivalent techniques for qualitative and quantitative copromicroscopic diagnosis of parasites in animals and humans. Nat. Protocols, 5: 503-515.
- Urquhart, G.M., J. Armour, J.L. Duncan, A.M. Dunn and F.W. Jennings, 1996. Veterinary Parasitology. 2nd Edn., Blackwell Science Ltd., Oxford, England, pp: 224-234.

- 15. Sapero, J.J. and D.K. Lawless, 1953. The MIF stainpreservation technic for the identification of intestinal protozoa. Am. J. Trop. Med. Hyg., 2: 613-619.
- 16. Morey, D.F., 1994. The early evolution of the domestic dog. Am. Sci., 82: 336-347.
- Kantere, M., L.V. Athanasiou, D.C. Chatzopoulos, V. Spyrou, G. Valiakos, V. Kontos and C. Billinis, 2014. Enteric pathogens of dogs and cats with public health implications. Am. J. Anim. Vet. Sci., 9: 84-94.
- Kohansal, M.H., A. Fazaeli, A. Nourian, A. Haniloo and K. Kamali, 2017. Dogs gastrointestinal parasites and their association with public health in Iran. J. Vet. Res., 61: 189-195.
- Oliveira-Sequeira, T.C.G., A.F.T. Amarante, T.B. Ferrari and L.C. Nunes, 2002. Prevalence of intestinal parasites in dogs from Sao Paulo State, Brazil. Vet. Parasit., 103: 19-27.
- Macpherson, C.N., 2005. Human behaviour and the epidemiology of parasitic zoonoses. Int. J. Parasitol., 35: 1319-1331.
- Youssef, A.I. and S. Uga, 2013. Review of parasitic zoonoses in Egypt. Trop. Med. Health, 42: 3-14.
- Awadallah, M.A. and L.M. Salem, 2015. Zoonotic enteric parasites transmitted from dogs in Egypt with special concern to *Toxocara canis* infection. Vet. World, 8: 946-957.
- Lahmar, S., I. Arfa, S.B. Othmen, W. Jguirim, Y. Said, A. Dhibi and B. Boufana, 2017. Intestinal helminths of stray dogs from Tunisia with special reference to zoonotic infections. Parasitology Open, Vol. 3, 10.1017/pao.2017.21
- Abere, T., B. Bogale and A. Melaku, 2013. Gastrointestinal helminth parasites of pet and stray dogs as a potential risk for human health in Bahir Dar town, North-Western Ethiopia. Vet. World, 6: 388-392.
- Muhairwa, A.P., H.E. Nonga and L.J. Kusiluka, 2008. A retrospective study of intestinal helminthosis as a cause of clinical disease in dogs. Tanzania Vet. J., 25: 24-30.
- Mahmuda, A., A.A. Magaji, M.D. Salihu, M.D. Lawal, M. Usman and A. Danmaigoro, 2012. Prevalence of intestinal parasites of dogs slaughtered at Mami market area, Sokoto, Nigeria. Sci. J. Anim. Sci., 1: 126-130.
- Mekbib, B., A. Regassa and D. Sheferaw, 2013. Gastrointestinal helminthes of dogs and owners perception of dogs parasitic zoonoses in Hawassa, Southern Ethiopia. J. Vet. Med. Anim. Health, 5: 20-26.
- Karakus, A. and V. Denizhan, 2019. Prevalence of gastrointestinal Helminths in stray dogs in Van Province. Turk. J. Vet. Res., 3: 27-32.

- Shenawy, E.L., S. Nahla, M.F. Soliman and S.I. Reyad, 2008. The effect of antioxidant properties of aqueous garlic extract and Nigella sativa as antischistosomiasis agents in mice. J. Inst. Trop. Med. Sao Paulo, 50: 29-36.
- Trasvina-Munoz, E., G. Lopez-Valencia, P.A. Centeno, S.A. Cueto-Gonzalez and F.J. Monge-Navarro *et al.*, 2017. Prevalence and distribution of intestinal parasites in stray dogs in the Northwest area of Mexico. Aust. J. Vet. Sci., 49: 105-111.
- Neafie, R.C. and A.M. Marty, 1993. Unusual infections in humans. Clin. Microbiol. Rev., 6: 34-56.

- 32. Molina, C.P., J. Ogburn and P. Adegboyega, 2003. Infection by *Dipylidium caninum* in an infant. Arch. Pathol. Lab. Med., 127: e157-e159.
- 33. Johnson, S.A.M., D.W. Gakuya, P.G. Mbuthia, J.D. Mande and N. Maingi, 2015. Prevalence of gastrointestinal helminths and management practices for dogs in the Greater Accra region of Ghana. H e l i y o n , V o l . 1 , N o . 1 . 10.1016/j.heliyon.2015.e00023
- 34. Kumar, M., B. Sharma, A. Kumar, H.P. Lal, V. Kumar and M.K. Tripathi, 2014. Prevalence and Haemato-Biochemical Studies of *Toxocara canis* Infestation in Dogs and Risk Perception of Zooneses by Dog Owners in Mathura, India Asian J. Anim. Vet. Adv., 9: 653-663.