Effects of Vitamin E on Growth Performance, Survival Rate, Hematological Parameters Response to Heat Stress in Rainbow Trout (Oncorhynchus mykiss) at Two Stocking Densities

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Abstract: The effects of vitamin E were studied in fingerlings rainbow trout (*Oncorhynchus mykiss*) kept at two different densities. Fingerlings (initial weight of $2.1\pm0.1~g$) were reared in triplicate groups in fiberglass tanks at densities of 400 and 800 m³. The fish were fed with 4 diets containing different levels of vitamin E (0, 100, 200 and 1000 mg vitamin E kg $^{-1}$ diet) for 9 weeks. The at end of feeding experiment fish were exposed to different temperatures (24, 26 and 28°C) and evaluate survival. The growth performance in fish fed diets containing different levels of vitamin E (100, 200 and 1000 mg kg $^{-1}$ diet) was significantly (p<0.05) higher than those fed diet without vitamin E. Levels of vitamin E significantly (p<0.05) affected the hematocrit, hemoglobin, Red Blood Cells (RBC) count, Mean Corpuscular Hemoglobin (MCH) and eosinophil in rainbow trout. However, it did not change Mean Corpuscular Hemoglobin Concentration (MCHC), lymphocyte and monocyte values. Stocking density had significant effect on eosinophil count only. Survival of the treated fish was significantly (p<0.05) higher than the control in response to heat stress. The highest effects of vitamin E was at level of 200 mg kg $^{-1}$ and a density of 400 pieces m $^{-3}$.

Key words: Vitamin E, growth, Oncorhynchus mykiss, hematological parameters, temperature stress

INTRODUCTION

Aquaculture industry is one of the most important industries in the developing world increasing annual aquaculture production. Fishes are normally reared in closed environments, such as pond aquaculture in earthen, cement and cages tanks. Attempts on rising fish stocking density account for one of the main causes of the rise and progress of this industry and also, as one of the key factors in the efficiency and economic viability of aquaculture. Undoubtedly, regardless of the control environment and stressful factors, raising density will adversely affect the health status of fish. This situation creates difficult and stressful environmental conditions, as well as increased susceptibility to fish pathogens. Therefore, beside the control of stressful environmental factors by adopting methods, the survival and growth of fish kept, as well as the immune system in such conditions can be boosted using immune stimulants in diet against infectious diseases (Huang and Huang, 2004; Irwin et al., 1999; Montero et al., 1999a; Soltani, 2008). Vitamins are important nutrients that stimulate

the immune system of fish and vitamin E, as an essential nutrient in aquaculture is proposed to maintain physiological processes, such as normal growth and immune function reproductive processes (De Menezes *et al.*, 2006; Andrade *et al.*, 2007). Vitamin E is a fat-soluble vitamin and diets with high levels of vitamin E, as opposed to normal levels lead to storage of the vitamin in the body fat tissues. Furthermore, excessive vitamin E in the diet results in hypervitaminosis which reduces growth and increases mortality in response to liver toxicity. Appropriate amounts of this vitamin, therefore should be added in the diet.

The vitamins and antioxidants, as well as macro-molecules such as DNA, lipids, proteins and other molecules form oxidation by free radicals during normal metabolism or under such conditions as pollution, corruption stress. Antioxidant properties of vitamin E may be effective in increasing the tolerance of fishes to environmental stress (Chen *et al.*, 2004). Vitamin E is needed to protect fish's immunity, meat quality, resistance of red blood cells against hemolysis, capillary permeability heart muscle (Blaxhall and Daisley, 1973). It also increases

the amount of Thiobarbituric Acid (TBARs) and reduces RBC fragility (Paul et al., 2004). Research has also shown that vitamin E is effective on blood parameters of fish (Randall et al., 1992; Chen et al., 2004; Trenzado et al., 2006; Garcia et al., 2007). Although, numerous studies have been conducted on the effects of vitamins and the resultant fish immune response, as well as the relationship between dietary vitamin and fish response to stress, there are few studies mostly focusing on stress in fish driven by vitamin C, a small number of studies were conducted on the effect of vitamin E and stress (Montero et al., 1999a, 2001).

Stocking density has been shown to affect behavioral interactions in several fish species (Irwin et al., 1999; Chen et al., 2004; Trenzado et al., 2006) which may ultimately affect growth rates. The effect of stocking density on growth was reported for a range of cultured fish species, such as gilthead sea bream, Sparus aurata (Montero et al., 1999a), juvenile turbot, Scophthalmus maximus (Irwin et al., 1999), Arctic charr (Jorgensen et al., 1993), Japanese flounder, Paralichthys olivaceus (Bolasina et al., 2006) rainbow trout Oncorhynchus mykiss (North et al., 2006; Trenzado et al., 2008), silver perch Bidyanus bidyanus (Rowland et al., 2006). Both positive and negative relationships between stocking density and growth have been reported the pattern of this interaction appears to be species specific. Therefore, the aim of this study was to investigate the effects of different levels of vitamin E on growth, blood parameters resistance against heat stress in rainbow trout fingerlings at two stocking densities.

MATERIALS AND METHODS

Experimental design, procedures and diets: This research was performed for 9 weeks in Tehran Ghezel Ala Co. located in Firoozkooh City, Northern Iran. The 4 diets with similar energy and protein levels were tested (Table 1). The diets contained different levels of vitamin E (0.0, 100, 200 and 1000 mg kg⁻¹ diet). The fish were stocked in densities of 400 pieces (typical density in the workshop) and 800 pieces m⁻² (high density) in a factorial (2×4) experimental design, considering 3 replicates per treatment. Rainbow trout fingerlings with average initial weight of 2.1±0.15 g stored in 24 experimental tanks (0.9×0.3×0.4 m). During the test, the experimental conditions were as: Water flow rate = 0.5 L/S, pH = 7.8, the average dissolved oxygen = 8.5 ppm, water temperature 10-13°C a natural photoperiod (10 L, 14 D). In order to feed the fingerlings, fish meal and meat meal were milled separately twice using tiny-meshed screens to obtain a homogeneous raw material. To mix food items to better

Table 1: Diet formulations and proximate composition (protein, lipid, NFE, moisture and energy)

Ingredients	Amount
Fish meal	58
Wheat flour	14
Meat flour	12
Dextrin	5
Fish oil	6
Vegetable oil	2.2
Filler (sawdust)	0.8
Mineral mixturec	1
Vitamin mixture (vitamin E free)	1
Proximate composition (analyzed) moistur	10±0.8
Crude protein	49±1
Crude lipid	15.1±0.8
NFE	16.5±.92
Energy (Kcal g ⁻¹)	380

¹1 kg mineral premix contained 130.6 g calcium phosphate dibasic, 327 g calcium lactate, 29.7 g ferric citrate, 137 g magnesium sulfate, 239.8 g potassium phosphate dibasic, 87.2 g sodium phosphate dibasic, 43.5 g sodium chloride, 0.15 g, aluminum chloride hexahydrate, 0.15 g potassium iodine, 0.1 g cupric chloride, 0.8 g manganese sulfate monohydrate, 1 g cobalt chloride hexahydrate and 3 g zinc sulfate heptahydrate; ²Vitamin E free premix containing 6600 IU vitamin A (retinol palmitate), 2400 IU vitamin D3, 28 mg vitamin K (menadione sodium bisulfate), 47 mg thiamin, 53 mg biotin, 12.7 mg folic acid, 0.06 mg vitamin B12 and 300 mg inositol per kg feed; Vitamin E used in the diets was DI-all-rac-α-tocopherol (Sigma chemical CO., Germany) which had purity ≥96% and with diet were mixed by vegetable and fish oil

work being done in the 1st dry ingredients that the greater the amount of rations were allocated to mixed together and then dry food items that were smaller (supplements, vitamins, minerals and salts), both blended after it's all dry ingredients together mixed well for 5 min with the ratios specified and then vitamin E with oil diet (tofu and soy oil) mixed and later received a higher volume was added to the diet. Then, the resulting mixture with specific humidity was ground using a meat grinder (mesh size from 5.1-2.2 mm) to yield string feeds. The food was dried (10% moisture) under a fan and open air for 24 h. It was then broken manually into small sizes (the size of the fish's mouth) and was stored in separate packages for each treatment at a cool and dry place away from sunlight. It was fed to the juveniles at 3-5% of body weight 6 times a day (8, 11, 13, 15 and 18 h).

Analyses of growth indices: The fish were biometried in breeding period once every 10 days. To do this, 30 juveniles from each tank were randomly picked and anesthetized using clove powder. The fish weight and length, respectively were measured on a digital scale (accuracy 0.01 g) and on a biometry board (accuracy 1 mm) and transferred to rearing tanks afterwards. At the end of week 9, all the fish in each tank were analyzed for growth indices (weight gain, survival, Specific Growth Rate (SGR), Feed Conversion Rate (FCR) and Condition Factor (CF)).

Weight gain (g) = Final weight – Initial weight

FCR = Dry feed intake/Wet weight gain

$$SGR = \frac{Ln final weight - Ln initial weight}{63 days} \times 100$$

$$Survival(\%) = \frac{Initial\ fish\ number - Dead\ fish\ number}{Initial\ fish\ number} \times 100$$

$$CF = (W/L^3) \times 100$$

Hematological measurements: At the completion of the experiment, changes in the blood factors (hemoglobin, hematocrit, Mean Corpuscular Hemoglobin (MCH), Mean Corpuscular Hemoglobin Concentration (MCHC), White Blood Cells (WBC) Red Blood Cells (RBC)) of 15 fish per treatment were determined after they had anesthetized with clove powder. Blood samples were taken through the caudal fin (Haghighi, 2009) and stored in EDTA tubes until use. WBC and RBC were counted under a Neubaur hemocutometer using diluents Gaur and torque (Blaxhall and Daisley, 1973). Hemoglobin was measured by a test kit (Pars Azmun Co., Tehran, Iran) with colorimetric method on a spectrophotometer (in 540 nm). Hematocrit was estimated through centrifuge (Haghighi, 2009). MCH and MCHC were calculated using the following equations (Haghighi, 2009):

$$MCH = (Hb/RBC) \times 10$$

$$MCHC = (Hb/Ht) \times 100$$

Experimental procedures for thermal stress: After the end of the rearing period to evaluate resistance of rainbow, trout fingerlings against thermal stress, thermal points of 24, 26 and 28°C were employed. To perform this

evaluation, 5 fish from low-density treatments and 10 fish from those with high density were randomly selected and stored separately in 12 L tanks. The water temperature in each tank was set using an aquarium heater and survival of juveniles was assessed after 35 min.

Statistical analyses: For statistical evaluation of the data, normality of the data was analyzed by one-sample Kolmogorov-Smirnov test. In the case of normal data, two-way ANOVA analyzed the data by the use of SPSS statistical software. The data were compared using Duncan's test at a significance level of 5%.

RESULTS

The effect of vitamin E on growth: After 9 weeks of the experiment, fish fed diets containing different levels of vitamin E significantly showed increased body weight, SGR and lower FCR than fish fed diets without supplemental vitamin E. As shown in Table 2, most of the weight gain (average 49.16 g) was associated with 200 mg of vitamin E supplementation and the lowest value (average 6.13 g) was obtained by fish in the control. The highest level (average 43.3 g) of SGR was also resulted from a dietary vitamin E of 200 mg and the lowest estimation (average 1.3 g) obtained by fish fed control diets. The best FCR from 200 mg of vitamin E was not significantly different from the diets containing 100 and 1000 mg of vitamin E but had significant differences with diets without vitamin E. Supplementation of 200 mg vitamin E led to significant differences (p<0.05) in the CF values with the non-complemented diets with average values of 1.16 and 0.9, respectively. The examined levels of vitamin E did not differ significantly (p>0.05) in respect of survival rates (Table 2). In addition, the fish not fed with vitamin E showed symptoms such as sclerosis, lordosis muscle and skin atrophy.

The effects of vitamin E on hematological indices: Vitamin E had significant effects on hematological

Table 2: Growth indicators of rainbow trout fed different levels of vitamin E at two stocking densities

Vitamin E						
(mg kg ⁻¹ diet)	Density (m)	Weight gain (g)	SGR	FCR	CF	Survival (%)
0	400	13.73 ± 1.4^{ab}	3.15 ± 0.08^a	1.39±0.05 ^b	0.99±0.5°	87±9.8ª
	800	13.6 ± 0.72^{a}	3.14 ± 0.07^{a}	1.39 ± 0.04^{b}	1.01 ± 0.07^{ab}	89±6.7ª
100	400	15.96±1.8ac	3.5 ± 0.09^{bc}	1.22 ± 0.08^a	1.06 ± 0.017^{abc}	92±6.4°
	800	15.29±0.74 ^{abc}	3.3 ± 0.05^{abc}	1.27 ± 0.01^a	1.07 ± 0.2^{abc}	89±5.8°
200	400	16.48±0.57°	3.4 ± 0.08^{bc}	1.21 ± 0.01^a	1.14 ± 0.1^{bc}	92±6.2°
	800	16.06±0.62°	$3.43\pm0.03^{\circ}$	1.21 ± 0.05^a	1.16 ± 0.09^{c}	88±8.8°
1000	400	15.7 ± 1.2^{abc}	3.13 ± 0.09^{abc}	1.26 ± 0.01^a	1.1 ± 0.11^{abc}	90±4.5°
	800	14.23 ± 1.6^{abc}	3.22 ± 0.18^{ab}	1.29 ± 0.03^a	1.07 ± 0.05^{abc}	86±9.7°
The effect of each	h variable and their	interactions				
Vitamin		0.00 <i>6</i> *	0.002^*	0.00^{*}	0.02^{*}	0.2
Density		0.25	0.39	0.26	0.86	0.9
Interactions		0.69	0.74	0.77	0.92	0.11

^{*}Means significance at a level of 0.05

Table 3: Hematological indicators of rainbow trout fed different levels of vitamin E at two different densities

Vitamin E	Density			RBC	Hb	MCHC			
		MOTT ()	TT-4 (0/)				E00 (0/)	MONT (0/)	T 373 ((0/)
(mg kg ⁻¹ diet)	(m ²)	MCH (pg)	Hct (%)	$(10^6 L^{-1})$	$(g L^{-1})$	$(g L^{-1})$	EOS (%)	MON (%)	LYM (%)
0	400	119.06±4.9 ^a	29.73±4.83°	0.51 ± 0.06^a	6 ± 0.36^{a}	20.76 ± 2.7^a	5.9 ± 0.6^{cd}	5.06 ± 1.5 ab	76±4.3°
	800	107.5±5.9 ^a	31.5 ± 4.73 ab	0.56 ± 0.05^a	6 ± 0.27^a	19.3±2.08 ^a	6.2 ± 0.6^{d}	5.4±1.2 ^b	78.6 ± 4.04 ab
100	400	97.1 ± 2.1^{ba}	34.6 ± 5.2 ab	0.65 ± 0.11^{b}	6.23 ± 0.3^{ab}	18.1 ± 1.78^a	5.6 ± 0.4^{bcd}	4.7 ± 1.3^{ab}	76.3 ± 2.3^{a}
	800	92.42±1.6ª	32.3 ± 6.86 ab	0.65 ± 0.06^{b}	6.0±0.45ª	18.9±2.6ª	$6\pm0.7^{\rm cd}$	3.5 ± 1.4^{ab}	80 ± 3.6^{ab}
200	400	100.1±8.1a	42.06±2.95 ^b	$0.74\pm0.13^{\circ}$	7.40±.05°	17.52 ± 0.27^a	4.3±0.5a	2.8 ± 1.2^{a}	85.3 ± 4.1^{b}
	800	98.1±6.1ª	38.3 ± 6.02 ab	0.71 ± 0.09^{bc}	6.93 ± 0.4^{bc}	18.25 ± 1.7^a	5.3±0.6 ⁶	3.9 ± 1.6^{ab}	82 ± 4.3^{ab}
1000	400	100.7±8.1a	37.96 ± 5.7 ab	0.96 ± 0.1^{bc}	6.9 ± 0.4^{bc}	18.3 ± 1.6^a	5.1±0.6 [∞]	3.8 ± 1.3^{ab}	80.3 ± 6.6^{ab}
	800	100.9±9.9°	41.23 ± 6.02^{b}	0.72 ± 0.1^{bc}	7.2±0.3°	17.6±1.89 ^a	6.1 ± 0.3^{d}	4.9 ± 1.5 ab	80.3 ± 2.9^{ab}
The effect of each variable and their interactions									
Vitamin		0.03^*	0.018^{*}	0.009^*	0.00^{*}	0.78	0.63	0.48	0.15
Density		0.29	0.92	0.77	0.54	0.86	0.014^{*}	0.55	0.67
Interactions		0.78	0.64	0.85	0.39	0.78	0.63	0.48	0.51

^{*}Means significance at a level of 0.05

parameters, such as Hb, Hct, RBC, eosinophils MCH but not on MCHC, monocytes lymphocytes, however there was not significant changes among the treatments (p<0.05). Values of Hb, Hct, erythrocytes and lymphocytes in fish fed diets containing vitamin E (100, 200 and 1000 mg kg⁻¹) were significantly higher than the control diet. Levels of monocytes, eosinophils, MCH MCHC in the fish fed diets containing vitamin E were less than those in fish fed the control diet. The fish fed 1000 and 200 mg of vitamin E were not significantly different whereas groups fed levels of 1000 and 200 mg differed significantly from both 100 mg and control groups (Table 3).

The effects of stocking density on growth and hematological parameters: Although, the effect of crowding on growth was not significant, the fish stored in a density of 400 m⁻² had relatively better growth performance (Table 2). The storage density had a significant effect on eosinophil (p<0.05), other blood parameters were not significantly different (p>0.05) (Table 3).

The interaction of vitamin E and stocking density on growth and hematological parameter: The highest weight gain, CF, SGR, FCR survival rate were found in fish fed 200 mg kg⁻¹ of vitamin E stocked at low density (400 m⁻²). These factors were lowest in fish without vitamin E feeding at high density (800 m⁻²). The interaction of vitamin E and stocking density on hematological indices did not give rise to significant effects (p>0.05).

The effect of vitamin E on thermal stress: Thermal stress showed that regardless of the density, survival rates in fish fed all levels of vitamin E (1000, 200 100 mg kg⁻¹) in 24 and 26°C yielded no significant differences. However, survival rates in the control group was significantly lower than those in the supplemented levels (p<0.05). Fish fed diets containing 200 mg kg⁻¹ of vitamin E at both

Table 4: Percentage survival of rainbow trout under heat stress at different levels of vitamin E and stocking density

	veis of vital	nin e and stocking	density	
Vitamin E				
$(mg kg^{-1}$	Density			
diet)	(m^2)	24°C	26°C	28°C
0	400	33.30±6.7ª	33.30±6.7a	19.86±6.8 ^{ab}
	800	47.7 ± 5.09 ab	44.04±1.09b	17.73±1.96 ^a
100	400	53.26±11.54bc	46.63±6.65bc	26.63 ± 6.65^{ab}
	800	63.30±3.30°	56.63±6.65 ^{cd}	29.93±5.7 [∞]
200	400	59.93±11.54bc	55.53 ± 7.73^{bcd}	42.16 ± 7.67^{d}
	800	53.30±3.30 ^{bc}	63.30 ± 3.30^{d}	31.06±3.8 ^{bc}
1000	400	46.6 ± 6.65 ab	59.96±6.65d	39.69±6.65°d
	800	55.5±1017 ^{bc}	53.30 ± 6.7^{bcd}	28.83±6.9abc
The effect	of each var	iable and their in	teractions	
Vitamin		0.006^*	0.00^{*}	0.00^*
Density		0.057	0.04^{*}	0.049*
Interactions	;	0.155	0.076	0.145

^{*}Means significance at a level of 0.05

densities (400 and $800\,\mathrm{m}^{-2}$) differed statistically from fish fed diets containing $100\,\mathrm{mg}\,\mathrm{kg}^{-1}$ of vitamin E and also the control group but not from fish fed $1000\,\mathrm{mg}\,\mathrm{kg}^{-1}$ of vitamin E. Density affected significantly in 24, 26 and 28°C with the highest survival rate in 28°C and a density of 400 pieces m^{-2} (Table 4).

DISCUSSION

The present study indicates that dietary vitamin E is essential for normal growth of rainbow trout which showed significant effects on growth, blood parameters thermal stress mitigation. Vitamin E requirements were evaluated in several species and the results of the present study are in agreement with the reports of the earlier works, such as common carp requirements of 100, 80 and 100 mg kg⁻¹ (Halver, 1995), Atlantic salmon demand of 120 mg kg⁻¹ (Hamre *et al.*, 1994), rohu up to 131.19 mg (Sau *et al.*, 2004), grouper 100 mg (Lin and Shiau, 2005; North *et al.*, 2006) rainbow trout 120 mg (Cowey *et al.*, 1981).

Because fish growth is related to food quality and food intake, feeding is done on the basis of growth rates and percent body weight. Thus, weight gain and growth rate of rainbow trout in this study was found to be related to the food quality that is changes in the supplemented levels of vitamin E in diet. Also, it was shown that with increasing vitamin E levels up to levels needed for fish, fish weight increased, so that with supplementing from 200-1000 mg kg⁻¹ no weight gain was observed. This is in agreement with those of Sau et al. (2004), Paul et al. (2004), Lin and Shiau (2005) and Chen et al. (2004). However, studies of Cowey et al. (1981, 1983) on salmon showed that with increasing vitamin E, weight gain was not observed. Blazer (1982) demonstrated that rainbow trout reared without vitamin E for 4 months showed no differences in growth compared with fish fed 400 mg of dietary alpha-tocopherol. Similar results were obtained for other species, such as Atlantic salmon (Raynard et al., 1991; Hardie et al., 1990), gilthead seabream (Montero et al., 2001), rainbow trout (Blazer and Wolke, 1984; Furones et al., 1992). Nevertheless, some researchers reported effects of deficiency in dietary vitamin E on the growth and survival of fish. For instance, consequences observed in Atlantic salmon (Hamre et al., 1994) rainbow trout (Cowey et al., 1983) chinook salmon (Thorarinsson et al., 1994) agree with the results of present study.

The vitamin E requirement in this study was higher than that in other reports (Lin and Shiau, 2005; Sau *et al.*, 2004; North *et al.*, 2006) in that the best performance was obtained with 200 mg of vitamin E. The differences in vitamin E requirements may be dependent on ration formulation such as the type and quality of used oils, species, fish size, form of vitamin E and/or the experimental conditions (Montero *et al.*, 2001). Besides, fluctuations and/or high FCR estimations can reflect difficulties in the diet or feeding methods (Montero *et al.*, 1999b). In this investigation, as feeding method was similar for all treatments, therefore different FCR values depended on the type of feed rations, i.e., differences in vitamin E levels.

Some symptoms of vitamin E deficiency have been reported, such as reduced growth, damage to collagen, darkened skin, lordosis, sclerosis mortality (Fracalossi et al., 1998; Montero et al., 1999a; Wang et al., 2003; Chen et al., 2004). The present study revealed some symptoms of vitamin E deficiency, such as reduced growth, atrophy of muscle darkened skin. Many deficient fish showed darkened skin which might be caused by impaired production and/or malfunction of melanocyte-stimulating hormones or broken tyrosine phosphorylation affecting the regulation of melanosome distribution in melanocytes (Chen et al., 2004). Melanin concentration was not measured in this study and further work is needed to verify the cause of the darkened skin

associated with vitamin E deficiency in the rainbow trout. Feeding the fish with ration without vitamin E rendered thinner (atrophied muscle) than other groups of fish. Muscle atrophy in fish and other animals may be caused by lack of antioxidants, particularly vitamin E in the diet because dietary vitamin E deficiency can cause severe oxidative stress (Puangkaew *et al.*, 2004) which plays a role in animals having muscle atrophy.

After 9 weeks, despite signs of vitamin E deficiency in the control fish, there was no significant differences in survival rates since the lowest survival was observed in the control; this is likely due to the short duration of the test period leading to non-significant fish death.

Huang and Huang (2004), suggested that decreased survival at high stocking density is related to low quality of the water physicochemical and also to competition for food and space which will increase the stress on fish. The results obtained in this study are in accordance with research on stocking densities of rainbow and brown trout in which fish growth and survival were not affected (North et al., 2006). When the effects of stocking density on the growth of fish are examined, different terms of expression are normally used and density is expressed in terms of either biomass or numbers of fish per unit area or volume. Sometimes, numbers of fish are reduced as the fish grow, so as to maintain constant biomass stocking densities (Irwin et al., 1999). In this research, numbers of rainbow trout were held constant for the duration of the experiment and therefore, biomass density was allowed to increase with time due to the growth of the fish. Accordingly, maximum biomass was observed in treatment with a density of 400 piece m⁻² rationing with 200 mg of vitamin E on the other hand, the lowest biomass was noticed in fish fed without vitamin E with a density of 800 pieces m⁻².

researchers have suggested increased density has negative effects on growth factors (Irwin et al., 1999; Montero et al., 1999a; Farhad et al., 2006; Trenzado et al., 2006, 2007; Andrade et al., 2007; Trenzado et al., 2007, 2008). Overcrowding is a common chronic stressor in aquaculture (Cristana et al., 2006; Chen et al., 2004), hence suppressed growth were reported as a result of high stocking density (Ross and Watten, 1998; Irwin et al., 1999; Rowland et al., 2006). This effect has been attributed to several factors including decreased food consumption. High stocking density also imposes increased energy demands that require fish to cope with metabolic adjustments, such as changes of gluconeogenic and glucolytic enzyme activities. This is because under such conditions, food consumption is reduced and the extra expenditure of energy has to be met by the body reserves resulting in reduced growth (Trenzado *et al.*, 2007, 2008; Garcia *et al.*, 2007; Trenzado *et al.*, 2006, 2007, 2008).

Agradi *et al.* (1993), evaluated positive effect of dietary vitamin E on sturgeon growth and expressed that vitamin E indirectly increased growth by controlling or reducing metabolic costs resulting in reduced tissue damage during stress. In the study, juveniles reared at low density had higher growth rates than those reared at high density, nonetheless density impact on fish growth was not significant. Yet, weight gain and SGR of fish reared at low density (400 m⁻², 16.48 g) was greater than the fish raised at high density (800 m⁻², 13.6 g) this may partially confirm the hypothesis that increased fish density will reduce growth. However, there were no significant differences in weight gain between treatments after 9 weeks which probably needs longer testing times to be verified.

It has also been reported that high stocking density affects some hematological parameters. Though, the results are widely variable and not decisive all the time, many researchers reported haemoconcentration, as a common effect of fish crowding evidenced as increases in both Hct and Hb values. This response may be a strategy for increasing the oxygen carrying capacity of blood under high energy demand situations such as chronic stress (Trenzado et al., 2006; Montero et al., 2001; Trenzado et al., 2007). In this study, Het and Hb concentrations were not significantly affected which may be related to regulative role of vitamin E for metabolic activities in fish and also to reduced oxygen consumption under conditions of oxygen deficiency (Randall et al., 1992; McKenzie et al., 1995). The fish fed without vitamin E under conditions of high density probably increased oxygen consumption rising amount of Hb, RBC, MCHC and MCH, as opposed to the fish kept at low density indicating likely elevated oxygen demand at high density (Srivastava and Sahai, 1987). Significantly lower Hct, Hb RBC was determined in the control fish than fish fed with different levels of vitamin E. The small amount of blood parameters in the control refers to anemia in fish, as was demonstrated by Taveekijakarn et al. (1996) and Chen et al. (2004) as well.

Chen et al. (2004), suggested that antioxidant properties of vitamin E might also influence temperature tolerance in fish. Vitamin E is a component of cell membranes that prevents long-chain, highly unsaturated fatty acids in phospholipids from oxidative damage these fatty acids maintain cell membrane fluidity. Vitamin E deficient fish might have had impaired membrane structure and function due to lipid oxidation because of heat stress, hence those fish received less vitamin E showed greater mortality rates.

CONCLUSION

This study reveals that different levels of vitamin E can affect growth and hematology of rainbow trout fed 200 mg kg⁻¹ dietary vitamin E and stocked in a density of 400 pieces m⁻². The results of thermal stress show that fish fed different levels of vitamin E had better survival rate than the non-supplemented fish.

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