

## Pharmacokinetics of Tylosin in Desert Sheep after Intramuscular Injection

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**Abstract:** The pharmacokinetics of tylosin after intramuscular administration at a dose of 1g kg<sup>-1</sup> body weight was studied in eleven healthy desert sheep. A peak level of tylosin C<sub>max</sub> of 198±0.95 µg mL<sup>-1</sup> was achieved after t<sub>max</sub> of 0.9±0.13 h and the mean distribution half-life (t<sub>1/2β</sub>) was 2±0.3 h and the apparent elimination half-life (t<sub>1/2α</sub>) was 72.2±9.3 h. The volume of distribution (V<sub>d</sub>) was found to be 53.299±3.907 L. The area under the curve (AUC<sub>0-240</sub>) of tylosin was calculated as being 1947.8±177.8 µg mL<sup>-1</sup> and the Area Under Maximum Concentration Curve (AUMC) was 74285.4±5420.75 µg/mL/h<sup>2</sup>. The total clearance (cl<sub>T</sub>) was 511.7±207 µl h<sup>-1</sup> and the mean resident time was 30±9.83 h.

**Key words:** Pharmacokinetic, intramuscular, tylosin, sheep, AUMC

### INTRODUCTION

Tylosin, an antibiotic of the macrolides group, is widely used in veterinary medicine either for prophylaxis or for treatment of a wide range of anaerobic bacteria. Gram-positive bacteria and mycoplasma (Duthu 1985; Prescott and Baggot, 1988). Tylosin could be administered in feed, drinking water, or injected intramuscularly mainly for prevention and treatment of Chronic Respiratory Disease (CRD). The pharmacokinetic characteristics and tissues disposition profiles for macrolide antibiotics are influenced by structural and physicochemical properties (Baggot and Gingerich, 1976; Carlier *et al.*, 1987; Ziv *et al.*, 1995). The pharmacokinetic aspects of tylosin have been evaluated in different species (Ziv *et al.*, 1995; Kowalski *et al.*, 2002; Parts *et al.*, 2002; Saurit *et al.*, 2002), goats (Atef *et al.*, 1991) and camels (Ziv *et al.*, 1995). The purpose of the present research is to study the pharmacokinetic parameters of tylosin after intramuscular administration in healthy sheep.

### MATERIALS AND METHODS

**Animals:** Twelve male's desert sheep, aged 9-12 month, weighing between 30-35 kg were used. Sheep were put in

pens (3×3 m) and fed with balanced ratio of concentrate and forage and water was available *ad libitum*. After clinical examination and liver and kidney tests, all animals were apparently healthy and allowed to acclimatize for two weeks before the start of the experiment.

**Drug administration and sample collection:** A single dose of 1g kg<sup>-1</sup> body weight of tylosin ((Macrolan-200, Interchemie, Holland) was injected intramuscularly in the cervical muscle. Blood samples were collected from the jugular vein, in heparinized tubes from each sheep at 0 (pre-treatment), 0.25, 0.05, 0.75, 1, 6, 24, 96, 168 and 240 h after drug administration. Plasma samples were separated by centrifugation (1200×g for 5 min) and stored at -20°C until analysis.

**Plasma assay:** Tylosin plasma concentration (µg mL<sup>-1</sup>) were estimated microbiologically by the modification one-plate test method described elsewhere (Koenen *et al.*, 1995; Koenen and DeBeer, 1998).

**Pharmacokinetic analysis:** Following Intramuscular (IM) administration, the tylosin plasma concentration versus time data were best fitted by a biexponential equation, C<sub>p</sub>=A<sub>e</sub><sup>-αt</sup> + B<sub>e</sub><sup>-βt</sup>. Consequently a two-compartment open

model was used to describe the plasma disposition of tylosin. The relevant pharmacokinetic parameters were calculated following I.M. injection of a single dose of 1 mg kg<sup>-1</sup> where C<sub>max</sub> is the maximum attained drug concentration in the plasma and was featured from the drug plasma-time plot, t<sub>max</sub> is the time at which C<sub>max</sub> was attained and also featured from the plot. A and B are intercepts of slopes of the initial distribution phase and the terminal elimination phase, respectively. α and β are the rate constants associated with distribution and elimination phases and obtained from the slopes of the two lines, respectively; t<sub>1/2α</sub> and t<sub>1/2β</sub> are the distribution and total body elimination half-lives of the drug and were obtained as 0.693/α and 0.693/β, respectively; k<sub>d</sub> is the rate constant for the drug elimination from the central compartment and calculated by αβ(A+B)/(αA+βB); k<sub>12</sub> is the rate constant for drug distribution from central to peripheral compartment and calculated by AB(β-α)<sup>2</sup>/(A+B)/(Aβ+Bα); k<sub>21</sub> is the rate constant for drug distribution from peripheral to central compartment and calculated by (Aβ+Bα)/(A+B). V<sub>d(are)</sub> is the apparent volume of drug distribution calculated by dose/(AUC<sup>0</sup>α×β). AUC<sup>0</sup><sub>240</sub> is the area under log-transformed drug plasma concentration-time curve from time curve from time zero to infinity and calculated as AUC<sup>0</sup><sub>240</sub> + (detected drug plasma concentration at 240h/k<sub>d</sub>). Cl<sub>β</sub> is the total body clearance of the drug based on the total elimination phase β and was calculated using the formula V<sub>d(are)</sub>×β. AUC<sup>0</sup><sub>α</sub> is the area under momentum of drug plasma-time curve from time zero to infinity and calculated by trapezoidal rule where it equals AUC<sup>0</sup><sub>240</sub> + (concentration of the drug at time 240h×240)/k<sub>d</sub> + (concentration of the drug at time 240h/ke<sub>12</sub>). MRT is the mean resident time for the drug in the body and was given by AUMC<sup>0</sup><sub>α</sub>/AUC<sup>0</sup><sub>α</sub>.

**Statistical analysis:** Statistical analysis was performed using software Miercal Origin 8, 2002 (Miercal Inc., USA). Pharmacokinetic parameters were expressed as mean±SEM.

## RESULTS

The plasma concentration of intramuscular injection of tylosin at a dose of 1g to eleven sheep at different times were represented in Table 1.

Tylosin pharmacokinetic parameters were recorded in Table 2. Intramuscular injection of tylosin induced a peak concentration C<sub>max</sub> 198±0.95 mg mL<sup>-1</sup> in plasma of sheep after t<sub>max</sub> 0.93±0.1 while (t<sub>1/2α</sub>) was 2±0.3h and the biological half-life (t<sub>1/2β</sub>) was 72.2±9.3h. The volume of distribution was found to be 53.299±3.907L.

Table 1: Mean values±SEM of tylosin plasma concentrations following intramuscular of 1g kg<sup>-1</sup> body weight to healthy sheep. (n = each 11)

Time (h)	Tylosin concentration (μg mL <sup>-1</sup> )
0	0
0.25	88.3±18
0.5	153.5±25
0.75	180.5±29
1	198±34
6	43±13
24	8±1.8
96	4±0.6
168	2±0.5
240	1±0.07

Table 2: Pharmacokinetic parameters (mean±SEM) of tylosin after single intramuscular administration of 1g kg<sup>-1</sup> body weight to healthy sheep. (n = each 11)

Pharmacokinetic value	Pharmacokinetic parameters
C <sub>max</sub> (μg mL <sup>-1</sup> ).	198±95
T <sub>max</sub> (h)	0.93±0.1
A (μg mL <sup>-1</sup> )	265.5±110
B (μg mL <sup>-1</sup> )	9.98±2.1
A (h <sup>-1</sup> )	0.3448±0.07
B (h <sup>-1</sup> )	0.0096±0.001
t <sub>1/2α</sub> (h)	2.00±0.3
t <sub>1/2β</sub> (h)	72.2±9.3
K <sub>d</sub> (h <sup>-1</sup> )	0.1522±0.083
K <sub>12</sub> (h <sup>-1</sup> )	0.1804±0.052
K <sub>21</sub> (h <sup>-1</sup> )	0.0217±0.0038
V <sub>d(are)</sub> (L)	53.299±3.907
AUC <sup>0</sup> <sub>240</sub> (μg/ml/h)	1947.8±177.8
AUC <sup>0</sup> <sub>α</sub> (μg/ml/h)	1954.4±205.4
AUMC <sup>0</sup> <sub>α</sub> (μg/ml/h)	74285.4±5420.75
Cl <sub>β</sub> (ml/h)	511.7±20.7
MRT (h)	38.01±9.83

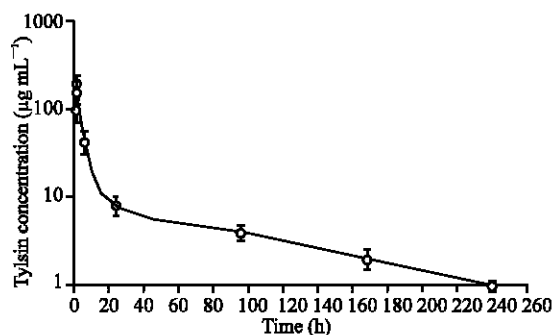


Fig. 1: Mean semi-log serum concentrations of tylosin versus time following intramuscular administration of a single dose 1g kg<sup>-1</sup> body weights to healthy sheep (n = 11 each)

The actual rate constant k<sub>12</sub>, k<sub>21</sub> and k<sub>d</sub> were calculated as being 0.1511±0.083, 0.184±0.050 and 0.0217±0.0038h<sup>-1</sup>, respectively. The area under concentration-time curve was determined as being 74285.4±5420.75 mg/ml/h<sup>2</sup> while the area under the curve was 194.8±177.8 mg/ml/h (Table 2).

The concentrations reached a level of 198±34 μg mL<sup>-1</sup> after 1 h while the minimum concentrations were evaluated as 1±0.07 μg mL<sup>-1</sup> after 240 h post dose. The

drug plasma concentrations revealed a biexponential decline which could be described by the open two-compartment model (Fig. 1).

The total body clearance of the drug was found to be  $511.7 \pm 20.7 \text{ mL h}^{-1}$  and the mean resident time was  $30.01 \pm 9.83 \text{ h}$ .

## DISCUSSION

As shown in Fig. 1, the decline in plasma tylosin concentration following single intramuscular injection of  $1 \text{ g kg}^{-1}$  in sheep could be described by the two compartment open model (Baggot 1977; Burrow 1980). The observed  $C_{\text{max}}$  ( $198 \text{ } \mu\text{g mL}^{-1}$ ) was much higher than those obtained after I. M. injection in goat  $C_{\text{max}}$  ( $2.38 \text{ } \mu\text{g mL}^{-1}$ ) (Atef *et al.*, 1991) and in camel  $C_{\text{max}}$  ( $1.16 \text{ } \mu\text{g mL}^{-1}$ ) following I. V. administration (Ziv *et al.*, 1995). The difference in  $C_{\text{max}}$  values could be attributed to difference in species and dosage level. The time to reach this concentration  $t_{\text{max}}$  (0.93h) was shorter than that reported in goat (4.19h) following I.M. injection (Atef *et al.*, 1991).

As shown in Table-2, the  $t_{1/2\alpha}$  (2 h) value was higher than that achieved in goat (0.2 h), after I.V. injection (Atef *et al.*, 1991) but it was nearly similar to that of tylosin in camel (3 h) (Ziv *et al.*, 1995). The observe  $t_{1/2\beta}$  (72.2 h) value was higher than that recorded following I.V. injection in goat (3.04 h) (Atef *et al.*, 1991) and camel (54.97 h) (Ziv *et al.*, 1995). The prolong elimination half-life following I.M. injection indicated continual release of the drug either from the injection site or from tissues which serve as reservoirs for the drug, a phenomenon characteristic of macrolide antibiotics (Burrows *et al.* 1983; 1986).

The value of  $k_{\text{dis}}$  ( $0.1522 \text{ h}^{-1}$ ),  $k_{12}$  ( $0.1804 \text{ h}^{-1}$ ) and  $k_{21}$  ( $0.0217 \text{ h}^{-1}$ ) were less than those reported in goat ( $k_{\text{dis}}$ , 0.02,  $k_{12}$  2.5,  $k_{21}$  2.5  $\text{min}^{-1}$ ) (Atef *et al.*, 1991) and in camel ( $k_{\text{dis}}$ , 4.58,  $k_{12}$  3.66,  $k_{21}$  4.91  $\text{min}^{-1}$ ) (Ziv *et al.*, 1995).

The low values indicate the slow distribution of tylosin between the body compartments. The  $V_d$  (53.299L) value was much higher than those reported after IV administration in goat (1.7L) (Atef *et al.*, 1991) and in pig (14.6L) (Parts *et al.*, 2002) but it was lower than that in cow (307L) (Duthu 1985) and another macrolide antibiotic tilmicosin; which in sheep (165L) (Modric *et al.*, 1998) and this indicated a wide distribution of the drug.

As shown in Table 2, the high values of  $\text{AUC}^{\circ}_{240}$  ( $1947.8 \text{ } \mu\text{g mL}^{-1}/\text{h}$ ),  $\text{AUC}^{\circ}_{\alpha}$  ( $1954.4 \text{ } \mu\text{g mL}^{-1}/\text{h}$ ) and  $\text{AUMC}^{\circ}_{\alpha}$  ( $74285.4 \text{ } \mu\text{g mL}^{-1}/\text{h}^2$ ) following I.M. injection were different from those obtained in goat ( $\text{AUC}^{\circ}_{\text{IM}}$   $47.4 \text{ } \mu\text{g mL}^{-1}/\text{h}$ ,  $\text{AUC}^{\circ}_{\text{IV}}$   $65.3 \text{ } \mu\text{g mL}^{-1}/\text{h}$ ) (Atef *et al.*, 1991) which indicated more absorption of the drug, while the difference in values may be attributed to dosages. The observed  $\text{Cl}_p$  ( $511.7 \text{ mL}^{-1}$ ) was much higher than that in

goat ( $6.8 \text{ mL min}^{-1}$ ) (Atef *et al.*, 1991) and in cow ( $4602 \text{ mL/min}^{-1}$ ) (Duthu 1985) but less than those achieved in normal and water deprived camel ( $97156 \text{ mL/min}^{-1}$ ) and ( $13468 \text{ mL/min}^{-1}$ ), respectively, which indicates large clearance of the drug. MRT value (38.01 h) was within the range achieved in cattle (36.9 h) and sheep (40.6 h) following tilmiosin administration (Modric *et al.*, 1998). But lower than that obtained in normal camel (207.58 min) (Ziv *et al.*, 1995) which indicate that the extent of absorption was high. The different in pharmacokinetics in this study may be attributed to difference in species or dosage of drug.

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