

aJava

Asian Journal of Animal and Veterinary Advances



Academic
Journals Inc.

www.academicjournals.com

Maternal Blood and Milk Lead Concentrations Following Exposure During Pregnancy with Emphasis to its Residues in Tissues of Aborted Foeti of Goats

¹A.R. Abd El-Hameed, ¹Samy I.A. Shalaby and ²Amira Hassan Mohamed

¹Department of Animal Reproduction and A.I.,
National Research Center, Cairo, Egypt

²Department of Clinical Pathology,
Faculty of Veterinary Medicine, Cairo University, Cairo, Egypt

Abstract: The study was carried out on two equal (7) groups of baladi goats, the first was kept as control and the second received daily oral dose of 6.0 mg lead acetate 1 kg body weight from early stage of pregnancy till abortion occurs around 14 week of pregnancy in all exposed animals. Blood samples were collected every two weeks from pregnant animals for plasma progesterone assay. Also blood and milk samples were obtained monthly for lead residue levels. Tissues from aborted foeti including bone, muscles, kidney, liver and brain were analyzed for lead residues. Results revealed a significant decline in progesterone levels from the beginning of the 6th week of gestation until abortion occurs around the fourteen week of pregnancy. Reproductive efficiency of animals after abortion revealed low pregnancy rate percentage and increase service interval period. In addition, maternal post-abortion lead level in both blood and milk showed a significant elevation. An increase in lead concentration in different organs noticed especially for femur bone, thigh muscle, kidney and liver of aborted foeti.

Key words: Goat, blood, milk, lead, pregnancy, aborted foeti tissues

INTRODUCTION

Lead is one of the most hazardous and cumulative environmental pollutants that introduced into the environment by humans. It affects embryonic development and functioning of reproductive system (Milnes *et al.*, 2006).

Lead exposure appears to reduce fertility and increase the risks of spontaneous abortion (Bellinger, 2005). Females subjected to lead for a long period suffered from disorders deals with hormonal function (Gorbel *et al.*, 2002).

High blood lead levels in animals have been reported in different localities in the world such as in India (Swarup *et al.*, 2005) and Egypt (Khalaf Alla and Abd El Aal, 1999), particularly in urban localities.

Lead is a well-known reproductive toxin affecting reproduction in female animals. In rats, it was associated with delayed sexual maturity, irregular estrus and reduced numbers of corpora lutea (Iavicoli *et al.*, 2006). In farm animals, it causes endometritis in ewes (Stoev *et al.*, 1997) impaired fertility in cows (Buhatel *et al.*, 1985; McEvoy and McCoy, 1993) as well as poor conception rate, reduced detection of heat and increased service interval in buffalo-cows (El-Tohamy *et al.*, 1997).

This study aimed to clarify a more reproductive details concerning the effect of chronic exposure of goats to lead throughout gestation as well as studying some reproductive indices during post-

abortion period. In addition, measuring of both maternal blood and milk lead concentration proving its crossing through placenta with emphasis to its residues and distribution through tissues of aborted foeti.

MATERIALS AND METHODS

Lead used was in the form of lead acetate crystalline powder having a molecular weight of 379.34 g mol⁻¹ purchased from Merck (Germany) and it dissolved in distilled water for oral dosing of pregnant baladi goats reared at National Research Center, experimental farm from October until March.

Fourteen pregnant baladi goats were divided into two equal groups, the first one kept as a control and the other drenched daily with 6.0 mg lead acetate/kg body weight until abortion take place.

Blood samples were collected every two weeks for plasma progesterone radioimmunoassay. After occurrence of abortion, both milk and whole blood samples were collected monthly for three successive times for determination of lead levels according to Dwivedi *et al.* (1995) and Yee *et al.* (1994). Lead level was determined by using graphite furnace atomic absorption spectrophotometry at a wavelength of 283.7 nm.

Tissues from aborted foeti (bone, muscle, liver, kidney and brain) were analyzed for lead residues according to O'Hara *et al.* (1995).

Obtained data was computed and statistically analyzed by student t-test (Snedecor and Cochran, 1980).

RESULTS

Oral administration of lead acetate to pregnant baladi goats at a level of 6.0 mg kg⁻¹ body weight abrupt pregnancy to full term causing abortion in all exposed animals around 14 weeks of pregnancy that is accompanied by placental retention in some cases. Progesterone hormone showed a significant decline in its level from the beginning of the 6th week of pregnancy until abortion occurred around the 14 week of gestation (Table 1).

Post abortion and during post partum period, pregnancy rate percentage in exposed group was 42.9%; service interval period was 100.57±10.72 days compared to 100% and 36.42±1.01 days in control group animals.

Maternal post-abortion lead level in both blood and milk showed a significant elevation (Table 2 and 3). Analyzed tissues for lead from foeti of lead exposed animals showed a highly significant (p<0.01) increase in aborted foeti tissues except for brain tissues (Table 4).

Table 1: Effect of oral administration of lead (6 mg kg⁻¹ b.wt.) during pregnancy until abortion on progesterone level (Mean±SE, ng mL⁻¹)

Weeks	Control	Exposed group
2	4.77±0.58	5.60±1.32
4	4.55±0.44	3.57±0.58
6	15.40±5.10	1.35±0.06**
8	6.63±1.50	1.22±0.30**
10	6.35±1.48	1.23±0.04**
12	7.10±1.30	1.31±0.04**
14	10.30±2.10	0.70±0.02**

*: p<0.05; **: p<0.01

Table 2: Blood lead level in goats subjected to 6.0 mg lead kg⁻¹ b.wt. after abortion (µg dL⁻¹)

Weeks	Control	Exposed group
Just after abortion	22.50±1.84 ^a	39.80±0.19** ^{aa}
After 1 month	21.80±1.54 ^a	38.00±0.16** ^{ab}
After 2 month	22.00±1.48 ^a	34.50±0.14** ^{ac}
After 3 month	21.40±1.92 ^a	31.30±0.16** ^{ad}
F-value	0.36	15.80**

** : p<0.01; Different letter(s) means significance within the group

Table 3: Milk lead level in goats subjected to 6.0 mg lead kg⁻¹ b.wt. after abortion ($\mu\text{g dL}^{-1}$)

Weeks	Control	Exposed group
Just after abortion	2.27 \pm 0.07 ^a	3.98 \pm 0.19 ^{*#a}
After 1 month	2.13 \pm 0.14 ^a	3.80 \pm 0.16 ^{*#b}
After 2 month	2.33 \pm 0.11 ^a	3.45 \pm 0.14 ^{*#c}
After 3 month	2.37 \pm 0.11 ^a	3.13 \pm 0.15 ^{*#cd}
F-value	0.83	5.40 ^{**}

*: p<0.05; **: p<0.01; Different letter(s) means significance within the group

Table 4: Concentration of lead in organ samples from aborted foeti ($\mu\text{g g}^{-1}$) Mean \pm SE

Organ	Femur	Muscle	Kidney	Liver	Brain
Control	0.525 \pm 0.10 ^B	0.123 \pm 0.01 ^A	0.260 \pm 0.001 ^A	0.232 \pm 0.05 ^A	0.167 \pm 0.04 ^A
Exposed group	2.620 \pm 0.37 ^{B**}	0.724 \pm 0.06 ^{E**}	0.398 \pm 0.02 ^{A**}	0.425 \pm 0.04 ^{A**}	0.192 \pm 0.02 ^A

Difference between mean values of group bearing different capital letter(s) means significant within the group (between different organs); **: p<0.01

DISCUSSION

Lead is a reproductive toxicant. Exposure to high concentration of lead affects fertility and reproductive success causing early embryonic mortality which increased as much as 2-3 times (Butkauskas and Sruoga, 2004).

In the present investigation, baladi goats subjected to 6.0 mg lead /kg b.wt. suffered from abortion around 14 weeks of pregnancy as well as placental retention in few cases. Exposure to high concentration of lead has been associated with a variety of adverse reproductive outcomes such as spontaneous abortion, impaired fecundity and sterility (Foster *et al.*, 1996; Pace *et al.*, 2005). Dearth *et al.* (2002) mentioned that gestational exposure appeared more sensitive to the effects of lead. Milnes *et al.* (2006) reported that high lead exposure causes endocrine-disruptive effects that exhibit an organizational effect on the developing embryo causing adversing effect on embryonic development. Progesterone hormone assay in exposed pregnant goats showed a steady decline in its level. Similar results were recorded by Sierra and Castiglioni (1992) in guinea pigs and by Abd El-Hameed *et al.* (1998) in goats.

Abortion may occur either as a results of crossing of lead through placenta (Neathery and Miller, 1975) reaching fetus itself or as a result of placentitis (O'Hara *et al.*, 1995) causing its death.

Poor pregnancy rate and increase of service period were the more pronounced reproductive features noted on exposed animals during postpartum periods. Females subjected to chronic high dose of lead suffered from reduced fertility as chronic lead exposure causes a double sexual disorder: first disorder deals with the hormonal function, which is affected at the early stages of poisoning but rapidly corrected; second disorder deals with the genital tract affecting the ovary resulting in a reduction in fertility of females in spite of presence of normal estrus (Gorbel *et al.*, 2002).

Lead acts both at the level of hypothalamus and directly at gonadal sites to disrupt reproductive physiology (Ronis *et al.*, 1996). At the level of pituitary, it is thought that lead may interfere with pituitary hormone release via interaction with calcium-dependant secondary messengers systems, which mediate hormone release from secretory granules storage (Klein *et al.*, 1994). At the gonads, lead accumulates in the ovary resulting in dysfunction of folliculogenesis (Taupeauet *et al.*, 2003). It has been reported that exposure to lead during pregnancy resulted in reduction in hypothalamic GnRH levels in both mothers and newborn (Camoratto *et al.*, 1993). Lead has a direct effect on gonadal germinal epithelium causing adverse effects on reproduction (Stoev *et al.*, 1997). Moreover, lead causes a decrease in gonadal weight or even act synergistically to reduce DNA gonadal content (Corpas and Antonia, 1998).

In the present study, a significant increase in both maternal blood and milk lead throughout three months post-abortion. This result is in agreement with Koyama *et al.* (1984) in goats and with Khalaf-Allah and A bd El-Aal (1999) in sheep grazing in heavy industrized area polluted with lead. It has been reported that lead retention is greater in lactating than in non-lactating dams (Hallen *et al.*, 1996).

In the present study concentration of lead showed significant increase in tissues of aborted foeti from exposed animals. This increase is mainly related to transplacental transfer (Franklin *et al.*, 1997; Synder *et al.*, 2000). It was reported that lead accumulates in areas of active bone formation suggesting the deposition of lead salt and subsequent binding to organic matrix (Milton *et al.*, 1982).

Increased lead concentration in kidney and liver was observed in this study. It is known that kidney secretes lead by glomerular filtration. An increased urinary lead level has been correlated with elevated lead nephropathy, which is seen as intranuclear inclusion bodies in the proximal tubules of lead poisoned animals (Milton *et al.*, 1982). High lead concentration in liver of aborted foeti attributed to erythropoiesis taking place in liver during gestation (Danielsson *et al.*, 1983).

Lead concentration in brain tissue was lower than in other organs, which may be attributed to uptake of lead by the liver firstly and reduction of blood flow to brain of fetus (O'Hara *et al.*, 1995). In addition, restriction of lead uptake by the brain may take place due to blood brain barrier (Crowe and Morgan, 1996).

We can conclude that there is a relationship between exposure of pregnant goats to lead and abortion, high blood and milk lead concentration and adverse effects on reproduction post-abortion periods. It is recommended to build animal farms far away from industrialized areas and high ways.

REFERENCES

- Abd El-Hameed, A.R., S.I. Shalaby and H.M. Amira, 1998. Effect of oral administration of lead on fertility, plasma progesterone and cortisol levels and tissues residue in baladi goats. *Egypt. J. Applied Sci.*, 13: 1-12.
- Bellinger, D.C., 2005. Teratogen update: Lead and pregnancy birth defects. *Res. A. Clin. Mol. Teratol.*, 73: 409.
- Buhatel, T., S. Tamas and S. Vesa, 1985. Dynamics of lead poisoning in relation to blood values of cows in different physiological states in an industrial area. *Zootech. Sci. Med. Vet.*, 39: 55-61.
- Butkauskas, D. and A. Sruoga, 2004. Effect of lead and chromium on reproductive success of Japanese quail. *Environ. Toxicol.*, 19: 412-415.
- Camoratto, A.M., L.M. White, Y.S. Lau, G.O. Ware, W.D. Berry and C.M. Moriarty, 1993. Effect of exposure to low level lead on growth and growth hormone release in rats. *Toxicology*, 83: 101-114.
- Corpas, I. and M.T. Antonia, 1998. Study of alterations produced by cadmium/lead administration during gestational and early lactation periods in the reproductive organs of the rat. *Ecotoxicol. Environ. Saf.*, 41: 180-188.
- Crowe, A. and E.H. Morgan, 1996. Interactions between tissue uptake of lead and iron in normal and iron-deficient rats during development. *Biol. Trace. Elem. Res.*, 52: 249-261.
- Danielsson, B.R., L. Dencker and A. Lindgren, 1983. Transplacental movement of inorganic lead in early and late gestation in the mouse. *Arch. Toxicol.*, 54: 97-107.
- Dearth, R.K., J.K. Hiney, V. Srivastava, S.B. Burdick, G.R. Bratton and W.L. Dees, 2002. Effects of lead exposure during gestation and lactation on female pubertal development in the rat. *Reprod. Toxicol.*, 16: 343-352.
- Dwivedi, S.K., S. Dey and D. Swarup, 1995. Lead in blood and milk from urban cattle and buffalo. *Vet. Hum. Toxicol.*, 37: 471-472.
- El-Tohamy, M.M., A.M. Hamam and U.A. Ali, 1997. Reproductive efficiency of buffalo-cows and its relationship with some heavy metals in the soil. *Egypt. J. Applied Sci.*, 12: 75-88.
- Foster, W.G., A. McMahon and D.C. Rice, 1996. Sperm chromatin structure is altered in cynomolgus monkeys with environmentally relevant blood lead levels. *Toxicol. Health*, 12: 723-735.
- Franklin, C.A., M.J. Inskip, C.L. Baccanale, C.M. Edwards, W.I. Manton, E. Edwards and E.J. O'Flaherty, 1997. Administration of stable lead isotopes to investigate changes in blood lead during pregnancy in a non human primate. *Fundam. Applied Toxicol.*, 39: 109-119.

- Gorbel, F., M. Boujelbene, A.F. Makni, C.F. Guermazif, J.P. Soleihavoup and A. El Feki, 2002. Cytotoxic effects of lead on the endocrine and exocrine sexual function of pubescent male and female rats. Demonstration of apoptotic activity. Laboratoire d'eco-physiologic animale, Faculte des sciences de Sfax, 3018 Sfax Tunisie.
- Hallen, I.P., S. Jonsson, M.D. Karlsson and A. Oskarsson, 1996. Kinetic observation in neonatal mice exposed to lead via milk. *Toxicol. Applied Pharmacol.*, 140: 13-18.
- Iavicoli, I., G. Garelli, E.J. Stanek, N. Castellino, Z. Li and E.J. Calaabrese, 2006. Low Doses of Dietary Lead are Associated with a Profound Reduction in the Time to the Onset of Puberty in Female Mice. *Reproductive Toxicology*. Elsevier Inc. www.sciencedirect.com.
- Khalaf-Allah, S.A. and S.A. Abd El-Aal, 1999 Effect of lead emissions on sheep grazing in heavy industrized area in Helwan, Egytp. *Assiut. Vet. Med. J.*, 40: 147-155.
- Klein, D.W., S.O. Kamyab and R.Z. Sokol, 1994. Effects of toxic levels of lead on gene regulation in the male axis: Increase in messenger ribonucleic acids and intracellular stores of gonadotrophins within the central nervous systems. *Biol. Reprod.*, 50: 802-811.
- Koyama, T., S. Myamoto and S. Maeda, 1984. Concentration of lead in milk. *Bull. Natl. Inst. Anim. Indus Japan*, 42: 15.
- McEvoy, J.D. and M. McCoy, 1993. Acute lead poisoning in a beef herd associated with contaminated silage. *Vet. Rec.*, 132: 89-90.
- Milnes, M.R., D.S. Bermudez, T.A. Bryan, T.M. Edwards, M.P. Gunderson, T.L. Larkin, B.C. Moore and L.J. Guillette, 2006. Contaminant-induced feminization and demasculinization of non mammalian vertebrate males in aquatic environments. *Environ. Res.*, 100: 3-17.
- Milton, R.H., B. Earl, B.B. Dawson and J. Williams, 1982. Tissue distribution of lead in rat pups nourished by lead poisoned mothers. *J. Toxicol. Environ. Health*, 9: 77-86.
- Neathery, M.W. and W.J. Miller, 1975. Metabolism and toxicity of cadmium, mercury and lead in animals. *Review. J. Dairy Sci.*, 58: 1767-1780.
- O'Hara, T.M., L. Bennett, C.P. McCoy, S.W. Jack and S. Fleming, 1995. Lead poisoning and toxicokinetics in a heifer and fetus treated with CaNa_2EDTA and thiamine. *J. Vet. Diagn Invest.*, 7: 531-537.
- Pace, B.M., D.A. Lawrence, M.J. Behr, P.J. Parsons and J.A. Dias, 2005. Neonatal lead exposure changes quality of sperm and number of macrophages in testes of BALB/C mice. *Toxicology*, 210: 247-256.
- Ronis, M.J., M.B. Thomas, J.S. Sarah, K.R. Paula and S. Fatima, 1996. Reproductive toxicity and growth effects in rats exposed to lead at different periods during development. *Toxicol. Applied Pharmacol.*, 136: 361-371.
- Sierra, M.E. and E.T. Castiglioni, 1992. Effect of low level lead exposure on hypothalamic hormones and serum progesterone levels in pregnant guinea pigs. *Toxicology*, 72: 89-97.
- Snedecor, G.W. and W.G. Cochran, 1980. *Statistical Method*. 1st Edn., Iowa State University Press, Iowa, USA.
- Stoev, S.D., V. Manov and N. Vassilev, 1997. Morphological investigation in experimental cases of chronic lead poisoning in pregnant sheep. *Bulgarian. J. Agric. Sci.*, 3: 795-801.
- Swarup, D., R.C. Patra, R. Naresh, P. Kumar and P. Shekhar, 2005. Blood lead levels in lactating cows reared around polluted localities; transfer of lead into milk. *Sci. Total Environ.*, 347: 106-110.
- Synder, J.E., N.M. Filipov, P.J. Parsons and D.A. Lawrence, 2000. The efficiency of maternal transfer of lead and its influence on plasma IgE and Splenic cellularity of mice. *Toxicol. Sci.*, 57: 87-94.
- Taupeauet, C., J. Poupon, D. Treton, A. Brosse, P. Richard and V. Machelon, 2003. Lead reduces messenger RNA and protein levels of cytochrome P450 aromatase and estrogen receptor beta in human ovarian granulosa cells. *Biol Reprod.*, 68: 1982-1988.
- Yee, H.Y., D.N. John and B. Jackson, 1994. Measurement of lead in blood by graphite furnace Atomic absorption spectrometry. *J. Anal. Toxicol.*, 18: 415-418.