

ajava

Asian Journal of Animal and Veterinary Advances



Academic
Journals Inc.

www.academicjournals.com

Genetic Polymorphisms of α -lactalbumin and β -lactoglobulin in South Anatolian and East Anatolian Red Cattle

¹H. Yardibi, ¹G. Turkey, ¹A. Mengi, ²F. Kaygısız and ¹K. Oztabak

¹Department of Biochemistry,

²Department of Animal Breeding and Husbandry, Faculty of Veterinary Medicine, University of Istanbul, Istanbul, Turkey

Abstract: The objective of the present study was to determine the genotype and allele frequencies for alpha-lactalbumin (α -LA) ve β -lactoglobulin (β -LG) that are claimed to be associated with milk production traits in cattle in South Anatolian Red (SAR) and East Anatolian Red (EAR) cattle. In this study, 40 cattle for each of SAR and EAR were used. Genomic DNA samples were isolated by using standard salt-out method. After Polymerase Chain Reaction (PCR), α -LA and β -LG genes were digested with *MspI* and *RsaI* (R5), *AvaI* (R3), *MspI* (R1), *Sau3A* (R2) restriction enzymes, respectively. As a result, SAR and EAR cattle breeds have the lower allele frequencies for α -LA and β -LG gene than high-yielding European dairy cattle breeds. Because of that reason we may claim that applying the selection programs for developing the alleles belonging to both genes may contribute to the trials to improve the production parameters in SAR and EAR breed bovines.

Key words: SAR, EAR, genetic polymorphism, α -LA, β -LG

INTRODUCTION

A number of studies have been performed on milk proteins since Aschaffenburg and Drewy (1955) demonstrated the A and B variants of β -LG in cow milk in 1955. Each of milk proteins that have been determined basing on homology of their structures (Farrell *et al.*, 2004). Milk proteins follow condominant Mendel's law of inheritance. Many of these genes have been mapped and sequenced. Nowadays, polymorphisms of milk proteins can be determined at protein level and DNA level. Some of these polymorphisms in milk proteins are known to affect milk yield, milk composition, micelle organization, coagulation characteristics and cheese yield. Scientific investigations on polymorphisms of genes related to milk proteins usually focused on cow milk. According to Mendel's laws, 6 large milk protein fractions which are controlled by non-dominant autosomal genes exist at different allele forms in cattle; α_{s1} casein, α_{s2} casein, β casein, κ casein, α -LA and β -LG. Four casein variants are encoded by a cluster of genes located on the 4th chromosome whereas those encoding α -LA and β -LG are located on the 5th and 11th chromosomes, respectively (Erhardt *et al.*, 1997).

α -LA which is the least studied milk protein is required for lactose biosynthesis in mammary gland and therefore it plays an important role in milk production. α -LA has two genetic variants as A and B. Variant A differs form variant B because glutamine amino acid at the 10th position in variant B replaces with arginine. Such change at the 10th codone results in a Single Nucleotide Polymorphism (SNP) which can be recognized by *MspI* restriction in the presence of α -LA-B or in the lack of α -LA-A (Mitra *et al.*, 1998). A lots of investigations shows that SNP in α -LA change the gene expression and deal with differences in milk yield and quality (Ramesha *et al.*, 2002).

The gene encoding β -LG has been sequenced in sheep, cow and goat. It has been mapped on the 3rd chromosomes in sheep and on the 11th chromosome in goat and cattle. Although, biological function of β -LG which is the major milk serum protein in swine, horse, cat, whale, dolphins and ruminants is not entirely known yet, it has been thought that it may play a role in transport of retinol and fatty acids. β -LG is composed of 162 amino acids and is one of the major milk components. Its function has still not been known except for playing a role in transport of hydrophobic molecules such as retinol and small fatty acids (Godovac *et al.*, 1985). It shows structural homologies with human retinol-binding protein and it has been speculated that it functions in vitamin A transport by binding retinol (Erhardt, 1989). In addition, β -LG has been reported to have a positive effect on digestion of milk proteins (Perez *et al.*, 1992).

Some of the investigators indicated that synthesis of milk protein is not affected by environmental factors but affected by differences between A and B alleles which are produced by SNP in gene regions (Gelderman *et al.*, 1996; Folch *et al.*, 1999; Ford *et al.*, 1993; Prosser *et al.*, 2000)

The aim of this study is to determine the frequencies of alleles and genotypes of β -LG and α -LA genes which effect milk protein contents in SAR and EAR cattle.

MATERIALS AND METHODS

In the present study, unrelated 40 cattle for each of SAR and EAR breeds were used. SAR breed cattle were selected from the herds in South Anatolian region of Turkey (Diyarbakir, Hatay) and EAR cattle were selected from those located in Eastern Anatolian region of Turkey (Kars). Blood samples were collected in sterile 2 mL tubes containing EDTA. Genomic DNAs were isolated using a standard salt-out method (Miller *et al.*, 1988). The PCR for α -LA and β -LG was carried out in a final volume of 25 μ L containing 1 U Taq DNA polymerase (Fermentas Life Sciences, Canada), 2-2.5 μ L 10 \times PCR buffer (750 mM Tris-HCl (pH 8.0), 200 mM (NH₄)₂SO₄, 0.1% Tween 20, 1.5 mM MgCl₂, 50-100 ng genomic DNA, 100 μ M dNTP (Takara, Biotechnology Co, Ltd, Japan) and 10 pmol of each primer. The primer sequence used for the β -LG; primer 1: AGCAACACACCCAGCACCAG and primer 2: CAAGCAGGAGGCACTTCATG (Wagner *et al.*, 1994). Amplification program were 94°C for 3 min; 32 cycles of 94°C for 1.30 min, 58°C for 1.30 min and 72°C for 3 min and a final extension at 72°C for 5 min. Those samples that were positive at the end of PCR, the target region of β -LG gene which is a 854 bp sequence containing 795 bp of the promoter region and 59 bp of exon I is subjected to digestion by *MspI* (Fermentas Life Sciences, Canada), *Sau3AI*, *AvaI* and *RsaI* restriction enzymes. *MspI*: allele A 730, 119, 5 and allele B 730, 89, 30 and 5 bp; *Sau3AI*: allele A 468, 386 and allele B 433, 386, 35; *AvaI*: allele A 734, 83, 37 and allele B 734, 120; *RsaI*: allele A 588, 266 and allele B 854 bp. The digestion products were run through 2% agarose gel. The primer sequence used for the α -LA; primer 1: TTGGTTTACTGGCCTCTCTTGTCATC and primer 2: TGAATTATGGGACAAAGCAAAATAGCAG (Mitra *et al.*, 1998). Amplification program were 94°C for 5 min; 30 cycles of 95°C for 15 sec, 60°C for 30 sec, 72°C for 30 sec and a final extension at 72°C for 10 min. The amplified 309 bp is subjected to cleavage by *MspI* enzyme and then the resulting products were separated by 2% agarose gel electrophoresis to differentiate the alleles B (220 and 89 bp) and AB (309, 220 and 89 bp).

Direct counting was used to estimate genotype and allele frequencies of α -LA, β -LG variants. The Chi-square test (χ^2) was used to determine whether the populations were in Hardy-Weinberg equilibrium using PopGene32 software (Yeh *et al.*, 2000).

RESULTS

In both breeds, frequency of A alleles of R₂, R₃ and R₅ were found higher than that of B alleles. In both breeds, frequency of B alleles of R₁ was found higher than that of A alleles. BB genotypes of

R1 were found higher than the AA genotypes (Table 1). AA genotypes were found higher than the BB genotypes for R3. The PCR products of α -LA is shown in Fig. 1 and agarose gel separation of *Msp*I PCR amplified products from α -LA AA, BB and AB is shown in Fig. 2. Genotype and allele frequencies determined in EAR and SAR cattle for *Msp*I polymorphism of the α -LA are shown in Table 2. BB genotypes were found higher than the AA genotypes significantly in both breeds. In both breeds, frequency of B alleles were found higher than that of A alleles.

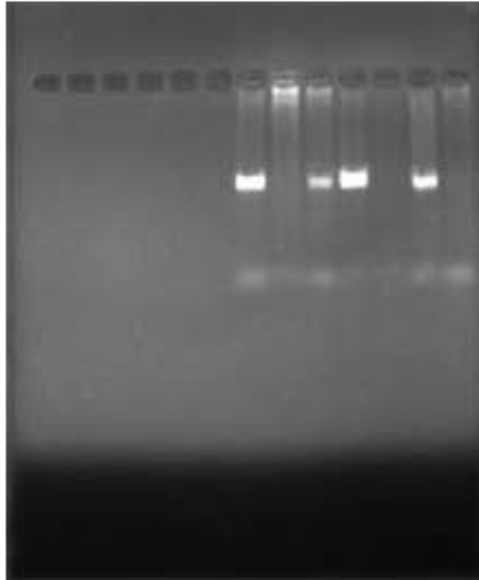


Fig. 1: PCR products for alpha-lactalbumin



Fig. 2: Agarose gel separation of *Msp*I-digested PCR amplified products from alpha-lactalbumin

Table 1: Distribution of R₅, R₃, R₁, R₂ genotypes and allele frequencies in SAR and EAR cattle

Locus	Breed	n	Allele frequency (%)		Genotype			$(\chi^2)^1$
			A	B	AA	AB	BB	
R ₅	SAR ²	40	0.68	0.32	20	14	6	1.8617 ^{ns}
	EAR ³	40	0.75	0.25	20	20	0	4.1864*
R ₃	SAR	40	0.74	0.26	27	5	8	19.3294***
	EAR	40	0.53	0.47	20	2	18	33.3091***
R ₁	SAR	40	0.40	0.60	16	0	24	41.1146***
	EAR	40	0.04	0.96	0	3	37	0.0399 ^{ns}
R ₂	SAR	40	0.66	0.34	19	15	6	1.2198 ^{ns}
	EAR	40	0.48	0.52	7	24	9	1.4497 ^{ns}

¹Test of Hardy-Weinberg equilibrium; ^{ns}: Not significant, ²South anatolian red cattle, ³East anatolian red cattle, *, **Indicate significant values

Table 2: Distribution of α -LA *MspI* genotypes and allele frequencies in SAR and EAR

Locus	Breed	n	Allele frequency (%)		Genotype			$(\chi^2)^1$
			A	B	AA	AB	BB	
β -LG (<i>MspI</i>)	SAR ²	40	0.14	0.86	0	11	29	0.9143 ^{ns}
	EAR ³	40	0.46	0.54	12	13	15	5.1570*

¹Test of Hardy-Weinberg equilibrium; ^{ns}: Not significant, ²South anatolian red cattle, ³East anatolian red cattle, *, **Indicate significant values

DISCUSSION

Ehrmann *et al.* (1997) reported that the frequencies of variants A in the β -LG encoding gene were between 0.38-1.0 in Brown Swiss, Simmental, German Friesian and Jersey cattles. In a study by Gustafson and Lunden (2003), the frequency of allele A was higher compared to the others in Swedish Red and White cattle which had higher milk fat concentration. In this study, the A allele frequencies associated with high milk fat concentration in SAR and EAR cattle were found between 0.52- 0.75. The frequency of allele A is known to be as high as 0.80-0.91 in high-yielding European dairy cattle breeds (Udina *et al.*, 2001).

Some of the investigators reported that the β -LG genotype AA was found to be associated with high whey protein and milk-protein quantities (Lin *et al.*, 1989; Mc Lean *et al.*, 1984; Ng-Kwai-Hang *et al.*, 1986, 1990). In present study, SAR and EAR those cattle with AA genotype relate to the high production traits for R₅ polymorphism was higher than the other genotypes.

As for the influence of α -LA *MspI* on the production traits of cows Mitra *et al.* (1998) reported that Zebu cattles (Sahiwal, Hariana, Tharparkar) with BB genotypes had higher milk yields compared to the other genotypes. Aschaffenburg (1968) reported that the frequency of α -LA A was higher in Indian Zebu (0.22-0.44) than in the most African Zebu (0.03-0.15). In this investigation the frequency of α -LA A was 0.14 in SAR cattles as like in the Zebu cattles. Anatolia is accepted as a primary centre of domestication for *Bos taurus* cattle and it is also widely accepted that considerable levels of *Bos indicus* introgression have occurred at this centre (Loftus *et al.*, 1999; Kumar *et al.*, 2003). The results found by Edwards (2007) which observed Zebu admixture in SAR and EAR cattle, also support this findings. In this study the frequencies for allele A for, which is characteristic for taurine breeds were estimated lower in SAR cattle than EAR cattle. It is possible that SAR cattle are genetically closer to Zebu cattle. This possibility is also supported by the phenotypic characteristics of SAR cattle, because some of the SAR cattle have a hump on their cidago region like Zebu cattle. Although, both of SAR and EAR cattles belong to the *Bos taurus*, they are similar to the *Bos indicus* in genetic position. So that the frequency of α -LA A alleles were found lower like Zebu cattles.

As a result, in both SAR and EAR cattles, the frequencies of A alleles of the β -LG gene which associated with milk protein quantities were lower than similiar to the high yielding European cattles. The frequencies of A alleles of the α -LA were found like Zebu cattles (*Bos indicus*). Because of that

reason we may claim that applying the selection programs for developing the alleles belonging to both genes may contribute to the trials to improve the production parameters in SAR and EAR breed bovines.

ACKNOWLEDGMENT

This study was supported by Research Fund of the University of Istanbul. Project No. 346.

REFERENCES

- Aschaffenburg, R. and J. Drewry, 1955. Occurrence of different β -lactoglobulins in cow's milk. *Nature*, 176: 218-219.
- Aschaffenburg, R., 1968. Genetic variants of milk proteins: Their breed distribution. *J. Dairy Res.*, 35: 447-447.
- Edwards, C.J., J.F. Baird and D.E. MacHugh, 2007. Taurine and zebu admixture in near eastern cattle: A comparison of mitochondrial, autosomal and y-chromosomal data. *Anim. Genet.*, 38: 520-524.
- Ehrmann, S., H. Bartenschlager and H. Geldermann, 1997. Polymorphism in the 5 flanking region of the bovine lactoglobulin-encoding gene and its association with beta lactoglobulin in the milk. *J. Anim. Breed. Genet.*, 114: 49-53.
- Erhardt, G., 1989. Evidence for a third allele at the β -lactoglobulin (β -Lg) locus of sheep and its occurrence in different breeds. *Anim. Genet.*, 20: 197-204.
- Erhardt, G., J. Juszcak, L. Panicke and H. Krick-Saleck, 1997. Genetic polymorphism of milk proteins in polish red cattle: A new genetic variant of beta lactoglobulin. *J. Anim. Breed. Genet.*, 115: 63-71.
- Farrell, H.M., R. Jimenez-Flores, R. Bleck, G.T. Brown and E.M. Butler *et al.*, 2004. Nomenclature of the proteins of cows milk-sixth revision. *J. Dairy Sci.*, 87: 1614-1674.
- Folch, J.M., V. Dova and J.F. Medrano, 1999. Differential expression of bovine beta lactoglobulin A and B promoter variants in transiently transfected HC 11 cells. *J. Dairy Res.*, 66: 537-544.
- Ford, C.A., M.B. Connett and R.J. Wilkins, 1993. Beta lactoglobulin expression in bovine mammary tissue. *Proceedings of the New Zealand, 1993, Society of Animal Production*, pp: 167-169.
- Geldermann, H., J. Gogol, M. Kock and G. Tacea, 1996. NA variants within the 5 flanking region of bovine milk protein encoding genes. *J. Animal Breeding Genetics*, 113: 261-267.
- Godavac, Z., A. Conti, J. Liberatori and G. Braunitzer, 1985. The amino acids sequence of beta lactoglobulin II from horse colostrum: Beta lactoglobulins are retinol binding proteins. *Biol. Chem. Hoppe Seyler*, 366: 601-608.
- Gustafsson, V. and A. Lunden, 2003. Strong linkage disequilibrium between polymorphisms in the 5 flanking region and the coding part of the bovine β -lactoglobulin gene. *J. Anim. Breed. Genet.*, 120: 68-72.
- Kumar, P., A.R. Freeman, R.T. Loftus, C. Gaillard, D.Q. Fuller and D.G. Bradley, 2003. Admixture analysis of South Asian cattle. *Heredity*, 91: 43-50.
- Lin, C.Y., A.J. McAllister, K.F. Ng-Kwai-Hang, J.F. Hayes and T.R. Batra *et al.*, 1989. Relationships of milk protein types to lifetime performance. *J. Dairy Sci.*, 72: 3085-3090.
- Loftus, R.T., O. Ertugrul, A.H. Harba, M.A.A. El-Barody, D.E. MacHugh and D.G. Bradley, 1999. A microsatellite survey of cattle from a centre of origin: The near east. *Mol. Ecol.*, 8: 2015-2022.
- McLean, D.M., E.R.B. Graham and R.W. Ponzoni, 1984. Effects of milk protein genetic variants on milk yield and composition. *J. Dairy Res.*, 51: 531-546.
- Miller, S.A., D.D. Dykes and H.F. Polesky, 1988. A simple salting out procedure for extracting DNA from human nucleate cells. *Nucl. Acids Res.*, 16: 1215-1215.

- Mitra, A., Sashikanth and B.R. Yadav, 1998. Alpha lactalbumin polymorphism in three breeds of Indian zebu cattle. *J. Anim. Breed Genet.*, 115: 403-405.
- Ng-Kwai-Hang, K.F., J.F. Hayes, F.E. Moxley and H.G. Monardes, 1986. Relationships between milk protein polymorphisms and major milk constituents in Holstein-Friesian cows. *J. Dairy Sci.*, 69: 22-26.
- Ng-Kwai-Hang, K.F., H.G. Monardes and J.F. Hayes, 1990. Association between genetic polymorphism of milk proteins and production traits during three lactations. *J. Dairy Sci.*, 73: 3414-3420.
- Perez, M.D., L. Sanchez, P. Aranda, J.M. Ena, R. Oria and M. Calvo, 1992. Effect of beta lactoglobulin on the activity of pregastric lipase. A possible role for this protein in ruminant. *Biochim. Biophys. Acta*, 1123: 151-155.
- Prosser, C.G., S.A. Turner, R.D. McLaren, B. Langley, P.J. LHuillier, P. Molan and M.J. Auld, 2000. Milk whey protein concentration and mRNA associated with beta lactoglobulin phenotype. *J. Dairy Res.*, 67: 287-293.
- Ramesha, K.P., T. Saravanan, M.K. Rao, M.M. Appannavar and A. Obi Reddy, 2002. Genetic distance among South Indian breeds of zebu cattle using Random Amplified DNA markers. *Asian Aust. J. Anim. Sci.*, 15: 309-314.
- Udina, I., S. Turkova, M. Kostuchenko, L. Lebedeva and G. Sulimova, 2001. Polymorphism of bovine prolactin gene, microsatellites, PCR-RFLP. *Russian J. Genet.*, 4: 407-411.
- Wagner, V.A., T.A. Schild and H. Gelderman, 1994. DNA variants within 5 flanking region of milk-protein-encoding genes. II. The b-Lactoglobulin-encoding gene. *Theor. Applied Genet.*, 89: 121-126.
- Yeh, F., R.C. Yang and T. Boyle, 2000. Popgene (v.1.32), microsoft windows-based freeware for population genetic analysis. <http://www.ualberta.ca/~fyeh/Pop32.exe>.