

# Asian Journal of Animal and Veterinary Advances



www.academicjournals.com

Asian Journal of Animal and Veterinary Advances 10 (12): 911-917, 2015 ISSN 1683-9919 / DOI: 10.3923/ajava.2015.911.917 © 2015 Academic Journals Inc.



# Giardia duodenalis Infection in Dairy Cattle of Assam, India

<sup>1</sup>M. Das, <sup>2</sup>D.K. Deka, <sup>2</sup>P.C. Sarmah, <sup>2</sup>S. Islam and <sup>1</sup>R. Laha

<sup>1</sup>Division of Animal Health, ICAR-Research Complex for NEH Region, Umroi Road, Umiam, 793103, Meghalaya State, India

<sup>2</sup>Department of Parasitology, College of Veterinary Science, Khanapara, 781022, Assam State, India

Corresponding Author: M. Das, Division of Animal Health, ICAR-Research Complex for NEH Region, Umroi Road, Umiam, 793103, Meghalaya State, India Tel: 8486356433

# ABSTRACT

To determine the prevalence of *Giardia duodenalis* infection in dairy cattle present in Assam, India, a total of 1176 numbers of faecal samples of cattle comprising 535 numbers of faecal samples of calves and 641 numbers of faecal samples of heifers were screened for one year using Formol-Ether technique and zinc sulphate (33%) floatation method. The overall prevalence of infection in cattle was 17.94%. Age-wise infection rate was 13.45 and 21.68% in calves and heifers, respectively. Season-wise highest infection was recorded during monsoon (30.90%) followed by pre-monsoon (21.97%), post-monsoon (5.50%) and winter (4.45%). Infection was higher among the diarrhoeic (83.41%) than non-diarrhoeic (16.58%) animals. The prevalence of G. duodenalis infection in cattle of Assam has significance because it infects a wide range of hosts, including humans worldwide and responsible for causing diarrhoea.

Key words: Giardia duodenalis, prevalence, dairy cattle, giardiasis, gastrointestinal protozoan

# INTRODUCTION

*Giardia* is a ubiquitous enteric protozoan of class-Mastigophora and family-Hexamitidae. Amongst the six currently accepted species of Giardia, G. duodenalis (syn. intestinalis/lamblia) has the broadest host range and is the species with the greatest public and animal health significance in terms of gastrointestinal diseases. This parasite has a wide host range, including mammals, birds and amphibians (Minetti et al., 2014). Giardia duodenalis is detected frequently in many mammals (Feng and Xiao, 2011) and is one of the most common intestinal parasites in pets like dogs (Batchelor et al., 2008) and in livestock (Thompson et al., 2008). Because of the impact on socio-economic development, especially in developing countries, it was included in the "Neglected Disease Initiative" of the World Health Organization (Savioli *et al.*, 2006). The parasite occurs in two morphologically distinct forms one being vegetative trophozoite and the other is thin walled cyst. Giardiosis in cattle is of particular concern because of the reported high prevalence of infection combined with the large output of faeces, potentially leading to contamination of water. They may act as a reservoir of infection with the potential to cause disease in humans either through direct contact or by contamination of food and water supplies. Transmission may occur through either direct contact in the case of farmers, veterinarians and petting zoo, or through indirect routes such as contaminated surface water or foods (Dixon, 2009). The infections are usually self-limiting in people with normal immune systems but can be severe in immuno-compromised individuals (Hunter and Thompson, 2005). The clinical signs associated with G. duodenalis infections in cattle may vary. Subclinical infections often reported; however, infection can result in the onset of diarrhoea, ill thrift and decreased weight gain in young calves (O'Handley et al., 1999). Morphological alterations in the intestinal epithelium of scouring calves with *Giardias* is has been

observed (Barigye *et al.*, 2008) and a significant improvement in weight gain was noticed in calves that were treated for *Giardia*sis with fenbendazole compared with untreated infected animals (Geurden *et al.*, 2010).

In India, Deshpande and Shastri (1981) from Maharastra reported the first incidence of *Giardia* infection in calves. Thereafter, Singh *et al.* (2008), Khan *et al.* (2011) and Sabu *et al.* (2011) reported *G. duodenalis* infections in cattle from three states of India viz. Punjab, West Bengal and Kerala, respectively. The state Assam situated in the North Eastern region of India and is the gateway of other six north-eastern states of India. It also shares international borders with Bhutan and Bangladesh, thus having importance in transmission of transboundary parasitic diseases. However, from Assam and also from other six North Eastern states of India, there is no report on prevalence of *G. duodenalis* infection in dairy cattle. Thus, the present study was undertaken to determine the prevalence of *G. duodenalis* infection in dairy cattle present in Assam, India.

## MATERIALS AND METHODS

**Study area:** The present study was carried out in Assam, India located South of the Eastern Himalayas. It lies within the latitude of 24°8 to 28°2' N and longitude of 89°42' to 96°E. Assam comprises of the Brahmaputra valley and the Barak river valleys along with the Karbi Anglong and the North Cachar Hills with an area of 30,285 square miles (78,438 km<sup>2</sup>). With the 'Tropical Monsoon Rainforest Climate', Assam is a temperate region and experiences heavy rainfall and humidity (https://en.wikipedia.org/wiki/Assam). The maximum and minimum temperature ranges from 35-38 and 6-8°C, respectively.

**Study period:** The study was conducted for one calendar year from August 2012 to July 2013 and divided into four seasons viz. Pre-monsoon (March, April, May), monsoon (June, July, August, September), post-monsoon (October, November) and winter (December, January, February).

**Faecal sample examination:** A total of 1176 numbers of faecal samples were collected per rectum from 535 calves (<1 year) and 641 heifers (1-3 years) randomly. The faecal samples were screened by using Formol-Ether technique (Ruprah, 1985) and zinc sulphate (33%, sp. gr. 1.18) flotation technique (Georgi and Georgi, 1990) for detection of *Giardia* cyst. Direct wet mount method was used to observe motile trophozoites in diarrhoeic samples as per standard method (Rajurkar *et al.*, 2012). Faecal samples were collected directly from the rectum of individual animal and kept in marked plastic pouch/vials. Precautions were taken to avoid contamination from one specimen to the other. Samples not being examined on the same day were preserved and stored at refrigerated temperature (4°C) for next day examination.

**Statistical analysis:** Data was statistically analyzed by Chi-square tests for significance using SPSS 15 version.

## RESULTS

The results of the faecal examination are summarized in Table 1. The overall prevalence of *G. duodenalis* infection in cattle was 17.94%. However, age-wise the infection rate in calves and heifer was 13.45 and 21.68%, respectively. In zinc sulphate floatation, *G. duodenalis* appeared as oblique cyst having ventral concavity (Fig. 1). However, when stained with iodine solution, two prominent nuclei occupying most of the anterior portion of the cyst was observed (Fig. 2).

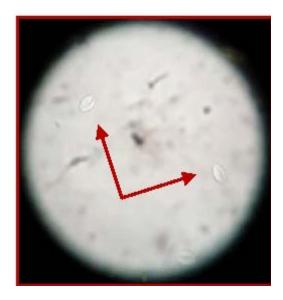


Fig. 1: Unstained G. duodenalis cyst in 33% zinc sulphate solution (400X)



Fig. 2: Stained G. duodenalis cyst (400X) showing two nuclei

Season-wise highest infection was recorded during monsoon (30.90%) followed by pre-monsoon (21.97%), post-monsoon (5.50%) and winter (4.45%) (Table 1). In calves, highest infection was recorded during monsoon (20.39%) followed by pre-monsoon (18.01%), winter (5.64%) and post-monsoon (4.04%). However, infection pattern in heifer was comparatively higher during monsoon (40.95%) followed by pre-monsoon (24.69%), post-monsoon (6.93%) and winter (3.57%). Though, there were apparent differences in infection rate according to season yet, no statistical significance could be drawn upon applying Chi-quare test. However, significant difference (p<0.05)

Seasons	Sample screened	Number positive	Sample consistency	
			Diarrhoeic	Non-diarrhoeid
Pre-monsoon	Calves (111)	20 (18.01)	17	3
	Heifer (162)	40 (24.69)	28	12
	273	60 (21.97)	45 (75.00)	15 (25.00)
Monsoon	Calves (201)	41 (20.39)	39	2
	Heifer (210)	86 (40.95)	73	13
	411	127 (30.90)	112 (88.18)	15 (11.81)
Post-monsoon	Calves (99)	4 (4.04)	3	1
	Heifer (101)	7 (6.93)	7	-
	200	11 (5.50)	10 (90.90)	1 (9.09)
Winter	Calves (124)	7 (5.64)	3	4
	Heifer (168)	6 (3.57)	6	-
	292	13 (4.45)	9 (69.23)	4 (30.76)
Overall	Calves (535)	72 (13.45)	62	10
	Heifer (641)	139 (21.68)	114	25
	1176	211 (17.94)	176 (83.41)	35 (16.58)
$\chi^2$ value, df 6		11.36 <sup>NS</sup>	113.86**	

Table 1: Seasonal prevalence of giardiosis in cattle of Assam

\*\*p<0.01, NS: Non-significant, -: Negative, Value in parentheses are percent positivity

existed according to the nature of sample consistency i.e., infection was higher among the diarrhoeic (83.41%) than non-diarrhoeic (16.58%) animals.

## DISCUSSION

The prevalence of G. duodenalis infection in cattle ranges from 3 to 64% world-wide (Geurden et al., 2008). In the present study, the prevalence of G. duodenalis infection in cattle was comparable to that of Wade et al. (2000) and Singh et al. (2008), who reported 21.68 and 34.78% infection in heifer and young calves, respectively. In a recent study from Al-Qadisiya, Iraq, Ahmed and A'aiz (2015) reported 85.71% infection in calves ( $\geq$  1year) with a significant difference (p<0.05) with calves (< 6 months) and calves (6 months to 1 year). Higher prevalence in younger animals is also in accordance with the findings of Ehsan et al. (2015) and Wang et al. (2014) from Bangladesh and China, respectively who reported 22 and 22.7% infections in calves, respectively. Due to decreased level of acquired immunity in younger animals they become more susceptible to infections (Reinemeyer, 1995). Ralston et al. (2003) observed that the infection rate (15%) and the number of *Giardia* cysts in the cows faeces (38.49 cysts/g) numerically increased at 1 week post calving compared to levels at calving and thus they act as a carrier of infections for calves. Higher infection among the diarrhoeic (83.41%) animals in the present findings was in accordance with the findings of Singh et al. (2008). Giardiosis in cattle with variation have been reported from different geographical areas viz. 5.0% from Thailand (Inpankaew et al., 2015), 7.2% from Germany (Gillhuber et al., 2014), 14% from Canada (Budu-Amoako et al., 2012) and 34.5% from Argentina (Tiranti et al., 2011). The variation in different geographical localities may be due to differences in the levels of management practices employed at farm level, housing-related factors and nature of the study (Swai and Schoonman, 2010). As compared to other countries and other studies of India, a moderate percentage of animals (17.94%) were found to be infected with G. duodenalis in Assam. The climate of Assam is characterized by heavy monsoon downpours; the cycle/flow of water may be responsible for transmission of infections from place to place.

In the present study, there is seasonal variation in the prevalence of infection and highest infection was recorded during monsoon (30.90%) season. Ayaz *et al.* (2012) also observed seasonal variation of infection in Pakistan. They observed highest infection (36.66%) in autumn (August,

September, October) followed by 34.58% infections during summer (May, June, July), spring (30.83%) and winter (25%). The lowest percentage of infections during winter as observed in the present study was similar as observed in neighbouring country Pakistan, although the rate of infections varied. A lowest percentage of infection during winter might be due to less availability of water thereby less organisms in contaminated water. Similarly, Castro-Hermida *et al.* (2009) in Spain detected more *G. duodenalis* cysts in faecal samples of cows during spring (16.0%) and summer (6.9%). High temperature and humidity along with frequent rains in the monsoon season enabled the transmission of cysts faster by contaminating feed and water of the animals. Thus, the warm and humid monsoon season was observed to be the most amicable season for the propagation of the disease than other seasons.

The present finding has significance because G. duodenalis is zoonotic and has public health importance in terms of gastrointestinal protozoan disease. Moreover, the infectious dose for susceptible humans may be as low as 10 cysts of G. duodenalis (Rendtorff, 1954). The prevalence of *Giardia*sis was significantly higher among children who had close contact with cattle (18.7%) compared to children who had no contact with cattle (9.6%) (Wegayehu *et al.*, 2013). In Assam, Devi *et al.* (2011) reported 1.5% *Giardia* cysts in indigenous people of Dibrugarh district during a community-based study. Thus, it is important to minimize contact between faecal materials containing infectious cysts and water supplies. So far our knowledge is concerned, there is no report of *G. duodenalis* infection in cattle of North East region of India and this may be considered as the first report of *G. duodenalis* in cattle from North East region of India, particularly from the state Assam.

#### CONCLUSION

The present study revealed that there is the prevalence of *G. duodenalis* in dairy cattle of Guwahati, Assam and infection rate was higher in heifers than calves. Season-wise highest infection was recorded during monsoon and recorded higher among the diarrhoeic animals.

#### ACKNOWLEDGMENTS

The authors are grateful to the Dean, Faculty of Veterinary Science and Director Post Graduate Studies, Assam Agricultural University, Khanapara, Guwahati, Assam for providing financial assistance and necessary facilities to conduct the research study.

#### REFERENCES

- Ahmed, H.S. and N.N. A'aiz, 2015. Detection of *Giardia*sis in apparently healthy cattle by using direct ELISA technique. Al-Qadisiya J. Vet. Med. Sci., 14: 1-3.
- Ayaz, S., A. Maqbool, S. Khan, S.N. Khan and M. Hussian et al., 2012. Prevalence of *Giardiasis* in selected dairy cattle farms in Lahore, Pakistan. Afr. J. Microbiol. Res., 6: 5383-5386.
- Barigye, R., N.W. Dyer, T.K. Newell, M.L. Khaitsa, J.M. Trout, M. Santin and R. Fayer, 2008. Molecular and immunohistochemical detection of assemblage E, *Giardia duodenalis* in scouring North Dakota calves. Vet. Parasitol., 157: 196-202.
- Batchelor, D.J., S. Tzannes, P.A. Graham, J.M. Wastling, G.L. Pinchbeck and A.J. German, 2008. Detection of endoparasites with zoonotic potential in dogs with gastrointestinal disease in the UK. Transboundary Emerg. Dis., 55: 99-104.
- Budu-Amoako, E., S.J. Greenwood, B.R. Dixon, H.W. Barkema and J.T. McClure, 2012. *Giardia* and *Cryptosporidium* on dairy farms and the role these farms may play in contaminating water sources in Prince Edward Island, Canada. J. Vet. Internal Med., 26: 668-673.

- Castro-Hermida, J.A., I. Garcia-Presedo, A. Almeida, M. Gonzalez-Warleta, J.M.C. Da Costa and M. Mezoa, 2009. Detection of *Cryptosporidium* spp. and *Giardia duodenalis* in surface water: A health risk for humans and animals. Water Res., 43: 4133-4142.
- Deshpande, P.D. and U.V. Shastri, 1981. Incidence of *Giardia* infection in calves in Maharashtra state, India. Trop. Anim. Health Prod., 13: 34-34.
- Devi, U., B. Borkakoty and J. Mahanta, 2011. Strongyloidiasis in Assam, India: A community-based study. Trop. Parasitol., 1: 30-32.
- Dixon, B.R., 2009. The Role of Livestock in the Food Borne Transmission of *Giardia duodenalis* and *Cryptosporidium* spp. to Humans. In: *Giardia* and *Cryptosporidium*: From Molecules to Disease, Ortega-Pierres, M.G., S.M. Caccio, R. Fayer and H. Smith (Eds.). CAB International, Wallingford, UK., pp: 107-122.
- Ehsan, A.M., T. Geurden, S. Casaert, S.M. Parvin and T.M. Islam et al., 2015. Assessment of zoonotic transmission of *Giardia* and *Cryptosporidium* between cattle and humans in rural villages in Bangladesh. PloS ONE, Vol. 10. 10.1371/journal.pone.0118239
- Feng, Y. and L. Xiao, 2011. Zoonotic potential and molecular epidemiology of *Giardia* species and *Giardia*sis. Clin. Microbiol. Rev., 24: 110-140.
- Georgi, J.R. and M.E. Georgi, 1990. Parasitology for Veterinarians. 5th Edn., WB Saunders Co., Philadelphia, pp: 297-301.
- Geurden, T., P. Geldhof, B. Levecke, C. Martens and D. Berkvens *et al.*, 2008. Mixed *Giardia duodenalis* assemblage A and E infections in calves. Int. J. Parasitol., 38: 259-264.
- Geurden, T., E. Vandenhoute, H. Pohle, S. Casaert, N. De Wilde, J. Vercruysse and E. Claerebout, 2010. The effect of a fenbendazole treatment on cyst excretion and weight gain in calves experimentally infected with *Giardia duodenalis*. Vet. Parasitol., 169: 18-23.
- Gillhuber, J., D. Rugamer, K. Pfister and M.C. Scheuerle, 2014. Giardiosis and other enteropathogenic infections: A study on diarrhoeic calves in Southern Germany. BMC Res. Notes, Vol. 7. 10.1186/1756-0500-7-112
- Hunter, P.R. and R.C.A. Thompson, 2005. The zoonotic transmission of *Giardia* and Cryptosporidium. Int. J. Parasitol., 35: 1181-1190.
- Inpankaew, T., T. Jiyipong, N. Thadtapong, C. Kengradomkij, N. Pinyopanuwat, W. Chimnoi and S. Jittapalapong, 2015. Prevalence and genotype of *Giardia duodenalis* in dairy cattle from Northern and Northeastern part of Thailand. Acta Parasitologica, 60: 459-461.
- Khan, S.M., C. Debnath, A.K. Pramanik, L. Xiao, T. Nozaki and S. Ganguly, 2011. Molecular evidence for zoonotic transmission of *Giardia duodenalis* among dairy farm workers in West Bengal, India. Vet. Parasitol., 178: 342-345.
- Minetti, C., W. Taweenan, R. Hogg, C. Featherstone, N. Randle, S.M. Latham and J.M. Wastling, 2014. Occurrence and diversity of *Giardia duodenalis* assemblages in livestock in the UK. Transboundary Emerg. Dis., 61: e60-e67.
- O'Handley, R.M., C. Cockwill, T.A. McAllister, M. Jelinski, D.W. Morck and M.E. Olson, 1999. Duration of naturally acquired giardiosis and cryptosporidiosis in dairy calves and their association with diarrhea. J. Am. Vet. Med. Assoc., 214: 391-396.
- Rajurkar, M.N., N. Lall, S. Basak and S.K. Mallick, 2012. A simple method for demonstrating the *Giardia* lamblia trophozoite. J. Clin. Diagn. Res., 6: 1492-1494.
- Ralston, B.J., T.A. McAllister and M.E. Olson, 2003. Prevalence and infection pattern of naturally acquired *Giardia*sis and cryptosporidiosis in range beef calves and their dams. Vet. Parasitol., 114: 113-122.
- Reinemeyer, C.R., 1995. Should you deworm your clients' dairy cattle? Vet. Med., 90: 496-498.

- Rendtorff, R.C., 1954. The experimental transmission of human intestinal protozoan parasites. II. *Giardia lamblia* cysts gwen in capsules. Am. J. Epidemiol., 59: 209-222.
- Ruprah, N.S., 1985. Textbook of Clinical Protozoology. Oxonian Press, New Delhi, India, pp: 156-157.
- Sabu, L., H. Subramanian and R. Sreekrishnan, 2011. Acute Giardiosis in bovine calves and its treatment with fenbendazole. J. Vet. Parasitol., 25: 96-96.
- Savioli, L., H. Smith and A. Thompson, 2006. *Giardia* and *Cryptosporidium* join the neglected diseases initiative. Trends Parasitol., 22: 203-208.
- Singh, B.B., R. Sharma, J.K. Sharma, H.S. Banga, H. Kumar, R.S. Aulakh and J.P.S. Gill, 2008. Prevalence of *Giardia intestinalis* infection in cattle. J. Vet. Parasitol., 22: 91-92.
- Swai, E.S. and L. Schoonman, 2010. Investigation into the prevalence of *Cryptosporidium* infection in calves among small-holder dairy and traditional herds in Tanzania. Vet. Med. Int. 10.4061/2010/676451
- Thompson, R.A.A., C.S. Palmer and R. O'Handley, 2008. The public health and clinical significance of *Giardia* and *Cryptosporidium* in domestic animals. Vet. J., 177: 18-25.
- Tiranti, K., A. Larriestra, C. Vissio, N. Picco, F. Alustiza, A. Degioanni and A. Vivas, 2011. Prevalence of *Cryptosporidium* spp. and *Giardia* spp., spatial clustering and patterns of shedding in dairy calves from Cordoba, Argentina. Revista Brasileira Parasitologia Veterinaria, 20: 140-147.
- Wade, S.E., H.O. Mohammed and S.L. Schaaf, 2000. Prevalence of *Giardia* sp., *Cryptosporidium* parvum and *Cryptosporidium muris* (C. andersoni) in 109 dairy herds in five counties of Southeastern New York. Vet. Parasitol., 93: 1-11.
- Wang, H., G. Zhao, G. Chen, F. Jian and S. Zhang *et al.*, 2014. Multilocus genotyping of *Giardia duodenalis* in dairy cattle in Henan, China. PLoS ONE, Vol. 9. 10.1371/journal.pone.0100453
- Wegayehu, T., H. Adamu and B. Petros, 2013. Prevalence of *Giardia duodenalis* and *Cryptosporidium* species infections among children and cattle in North Shewa Zone, Ethiopia. BMC Infect. Dis., Vol. 13.