

ajava

Asian Journal of Animal and Veterinary Advances



Academic
Journals Inc.

www.academicjournals.com



Review Article

Phytobiotics: Could the Greens Inflate the Poultry Production

¹Govinthasamy Prabakar, ²Marappan Gopi, ³Kumarakurubaran Karthik, ¹Subramaniyan Shanmuganathan, ³Arumugam Kirubakaran and ⁴Selvaraj Pavulraj

¹ICAR-Indian Veterinary Research Institute, Izatnagar, India

²ICAR-Central Avian Research Institute, Izatnagar, India

³TANUVAS, Chennai, India

⁴National Research Centre on Equines, Hisar, Haryana, India

Abstract

The ever increasing consumer's awareness and their concern over the presence of antibiotic residues in poultry products necessitate looking for an alternative to antibiotic growth promoters. Among the numerous alternatives like probiotics, prebiotics, acidifiers, the plant origin compound attracts more interest than else. The phytochemicals consists of various natural products that may be nutritional, non-nutritional or anti-nutritional in nature. These phyto-chemicals also act as an antibacterial, antiviral, antifungal, antioxidant, digestive stimulant, immune-modulator, hypo-lipidemic agent and also heat stress alleviator. A compound of multi-functional can be considered as additive in animal production system. Their anti-oxidant activity and hypo-lipidemic property of these plant derived compounds will be attributed for the improvement of shelf-life of various animal or poultry products. Their hypolipidemic properties are used in production of lean meat production. These phytobiotics also impart readily acceptable flavour to the products especially meat and eggs. They aid in digestive process by stimulating the digestive secretions throughout the gastro-intestinal tract thereby increasing the overall digestibility of the nutrients and reduce the environmental pollution. Moreover, these botanical products are becoming more prominent in insect and pest control strategies due to their availability and cost. With their wide range of activities, these phytobiotics will go to be a new group of feed additives for better growth rather than simply as another alternate to in-feed antibiotics in food production industry.

Key words: Phytobiotics, poultry, antimicrobial, alternate, digestive stimulant, flavouring agent

Received: April 26, 2016

Accepted: May 26, 2016

Published: June 15, 2016

Citation: Govinthasamy Prabakar, Marappan Gopi, Kumarakurubaran Karthik, Subramaniyan Shanmuganathan, Arumugam Kirubakaran and Selvaraj Pavulraj, 2016. Phytobiotics: Could the greens inflate the poultry production. Asian J. Anim. Vet. Adv., 11: 383-392.

Corresponding Author: Govinthasamy Prabakar, ICAR-Indian Veterinary Research Institute, Izatnagar, India

Copyright: © 2016 Govinthasamy Prabakar *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Phytogenic feed additives (phytobiotics or botanicals) are commonly defined as plant-derived compounds incorporated in to diets to improve the livestock productivity through amelioration of feed properties, improvement of nutrient digestibility, absorption and elimination of pathogens in the gut^{1,2}. Pphytogenic compounds included or incorporated in diet to improve the animals' production performance and improving the quality of food derived from those animals³. According to their origin and treatment variety of plant derivatives used such as herbs (flowering, non woody and non-persistent plants), spices (herbs with an intensive smell or taste commonly added to human food) like cinnamom, coriandor, pepper, chilli, rosemary, oregano, anise, thyme and garlic. Essential oils (volatile lipophilic compounds derived by cold expression or by steam or alcohol distillation), or oleoresins (extracts derived by non-aqueous solvents)⁴. Some of the other compounds extracted from fruits such as flavonoids, are water soluble in nature, as this type of fruit extraction compound also used in poultry feed as additive⁵. Within phytogenic feed additives, the content of active principles in products may vary widely, depending on the plant part used (e.g., seeds, leaf, root or bark), harvesting season and geographical origin. The technique for processing (e.g., cold expression, steam distillation, extraction with non-aqueous solvents etc.) modifies the active substances and associated compounds within the final product.

Properties of phytobiotics: Plant extracts have antimicrobial action, immune enhancement, anti-stress property⁶, antioxidant and gut microflora manipulation⁷, nutrigenomics effects⁸, digestibility enhancer⁹, stress lowering effect¹⁰, cholesterol-lowering effect¹¹. The plant herbs have these kinds of properties to increase the animal's health and performance (Fig. 1).

Phytogenic effects have been proven in poultry for feed palatability and quality (sensory aspects), growth promotion (improved weight gain and feed conversion ratio, reduced mortality), gut function and nutrient digestibility (improved growth), gut microflora (less diseases of the GIT, improved growth, reduced mortality), immune function (improved health) and carcass meat safety and quality (reduced microbial load, improved sensory¹²).

ANTIBACTERIAL AND ANTIVIRAL ACTIVITY

Phytobiotics especially spices and herbs exert well known antimicrobial activity *in vitro* including some major pathogens and fungi¹³. A common growth enhancing and digestion modifiers present in hawthorn fruit, this is having more than seventy organic chemicals, bioactive, active components and some unidentified growth factors. In phytobiotics, having some complex active molecules that exhibits synergistic effect to increase the enhancement of growth as well as contain antimicrobial property^{14,15}. Tannic acid phytochemical compound it have the antimicrobial

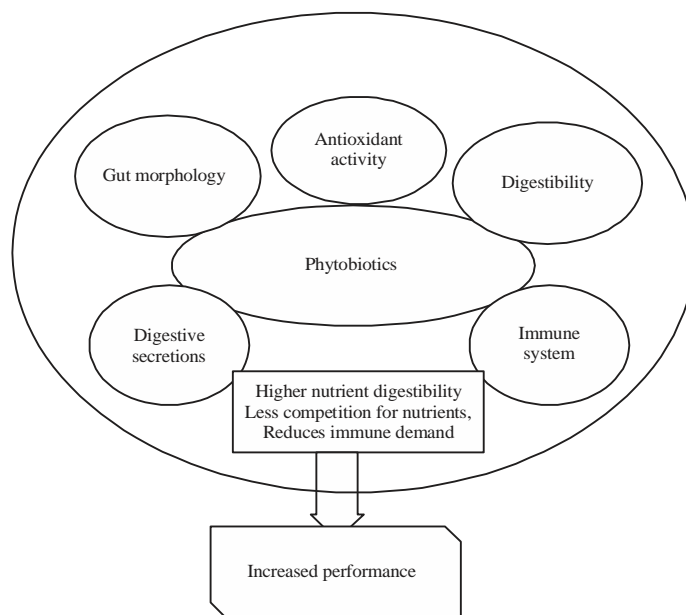


Fig. 1: Activities of phytobiotics in poultry system

property to inhibit the intestinal bacteria such as *Clostridium perferingens*, *E. coli*, *Enterobactor cloaceae* and *Bacteriodes fragilis*¹⁶. The DNA synthesis was inhibited by phytochemical compound alkaloid which is act as a intercalator or DNA inhibitor¹⁷. Saponin is another phytochemical compound, have antimicrobial property through its ability to form complexes with sterols present in the microorganisms membrane, it cause membrane damage and followed by collapse of cells¹⁸. Essential oils much their potential as alternative to antibiotics in broiler chicken and potent it has antimicrobial effect¹⁹. Some of the essential oils have potent anti-microbial effect against *Clostridium perfringens* and *E. coli*^{20,21}. The essential oils act as antimicrobial action through their lipolytic property²² and chemical structure²³. In this mechanism of essential oils like terpenoids and phenylpropenoids have the lipolytic property it penetrate the bacterial membrane and reach the cells inner part²⁴ and aromaticity²⁵ also responsible for essential oils anti-microbial property. Using *Achillea millefolium*, *Hypericum perforatum* and *Levisticum officinale* was efficiently used as substitute for antibiotics and also this diet favourably affected meat sensory characteristics²⁶.

The antimicrobial mode of action is considered to arise mainly from phytobiotics through the potential of the hydrophobic essential oils to have the ability to forcefully enter into the bacterial cell membrane; this will cause structural membrane disintegration hence its leads to leakage of ions. The properties of anti-microbial action have shown the ability to improve the carcass keeping quality through maintaining the microbial hygiene of carcass. Phytobiotic compound like oregano have antimicrobial properties it reduces the pathogens specifically salmonella and total viable bacteria on broiler carcass²⁷. Some of the non-phenolic compounds extracted from limonene and extract of *Sanguinaria canadensis* have anti-bacterial activity^{28,29}. Phytochemical substances show a clear antimicrobial activity *in vivo*³⁰. Peppermint oil also showed strong anti-microbial activity³¹. In broilers fed with essential oils of oregano and garlic through the diet and observed that the oregano and garlic oil reduced the small intestinal *Clostridium* counts³². Marinating chicken breast filletes treated with the extract of lemon green tea and turmeric showed effective against *Salmonella enteritidis* and *Campylobacter jejuni*. The combination of these three essential oils was found to be most effective against both the microbes³³.

PHYTOCHEMICAL ACTS AS ANTIFUNGAL

Chemical control of fungi and mycotoxins also result in environmental pollution, health hazard and affects the natural

ecological balance³⁴. Use of plant products informs of plant extracts and essential oils provide an opportunity to avoid synthetic chemical preservatives and fungicide risks³⁵. Essential oil oregano has antifungal activity³⁶.

Some phytochemicals are injurious to fungi and could be used to protect crops, animals, humans, food and feeds against toxigenic fungi and mycotoxin³⁷. Phyto-fungicides could be prepared or formulated from the leaves, seeds, stem bark or roots of plants of pesticidal significance and could be applied in form of extract, powders and cakes or as plant exudates^{38,39}. Phytochemicals vary in plants depending on their growing conditions, varietal differences, age at harvest, extraction methods, storage conditions and age of sample. *Allium sativum* has a wide antifungal spectrum, reached about 60-82% inhibition in the growth of seed borne *Aspergillus* and *Penicillium* fungi⁴⁰. This was attributed to phytochemical properties of garlic plant, allicin which could decompose into several effective antimicrobial compounds such as diallyl sulphide, diallyl disulphide, diallyl trisulphide, allyl methyl trisulphide, dithiols and ajoene^{41,42}. Aflatoxins refer to a group of four mycotoxins (B1, B2, G1 and G2) produced primarily by two closely related fungi, *A. flavus* and *A. parasiticus*. Aflatoxin contamination of crops is a worldwide food safety concern. Neem extract have an inhibitory effect on biosynthesis of aflatoxins (groups B and G) in fungal mycelia⁴³. More than 280 plant species have been investigated for their inhibitory effect on toxigenic *Aspergillus* and nearly 100 of these plants had some activity on their growth or toxin production. The inhibitory effect of crude extracts from mint, sage, bay, anise and ground red pepper on the growth of *A. parasiticus*. Plants like *Ocimum gratissimum*, *Cymbopogon citratus*, *Xylopiia aethiopicia*, *Monodora myristica*, *Syzygium aromaticum*, *Cinnamomum verum* and *Piper nigrum* are effective in inhibiting formation of non sorbic acid, a precursor in aflatoxin synthesis pathway⁴⁴. In addition lemon and orange oils (at concentrations of 0.05-2.0%) affected more than a 90% reduction in aflatoxin formation by *A. flavus*⁴⁵. Marjoram essential oil culture broth reduced the growth of fungal species *Aspergillus flavus*, *A. niger*, *A. ochraceus* and *A. parasiticus* by up to 89%⁴⁶.

ANTIOXIDATIVE ACTION

Herbes and spices have well known for its anti-oxidative property⁴⁷. Among variety of plant constituents possessing anti-oxidant activities, plants such as rosemary plant have phenolic compounds like rosmarinic acid and rosmarol, thymol and carvacrol derived from thyme and oregano⁴⁸. Pepper (*Piper nigrum*) and red chilli (*Capsicum frutescens*) also have antioxidant properties. Fruits and some of the

plants like ginger, curcuma, anise, coriander and green tea (rich in flavonoid) these plants are having well known effect on their antioxidative action⁴⁹. The mode of antioxidative properties of the plant additives through their effect (protection) of feed lipids from Reactive Oxygen Species (ROS), this is main compound for formation of free radicals, it leads to lipid per oxidation, for this reason, the bioactive compounds reduce the ROS formation and prevent lipid peroxidation and prevent the feed/product damage. Some of the researchers found that phytobiotics have phenolic compounds, have the ability to improve the oxidative stability of the product derived from animals like poultry meat⁵⁰⁻⁵² eggs⁵³ and pork⁵⁴. Addition of rosmarin and garlic oil in broiler diet act as anti-oxidant property and have positive effect on glutathione redox system of liver in chickens⁵⁵. Feeding fenugreek, garlic and pepper 10 g kg⁻¹ to broilers diet, reduced the tyrosine and thiobarbituric acid reactive substances in broiler meat, because garlic act as a antioxidant that inhibit the synthesis of fatty acid in the liver and lipid oxidation⁵⁶.

PHYTOCHEMICAL ACTS AS INSECTICIDE

Chemicals have been intentionally fed to chickens for the purpose of controlling house fly (*Musca domestica* L.) larvae infesting poultry manure. Polybor 3 and trolene (ronnel) in the laying mash of hens and observed the effect on the fly population. Oregano exhibits insecticidal activity⁵⁷. *Musca domestica* L. (Diptera: Muscidae), is a serious health threat to human beings and livestock by transmitting many infectious diseases⁵⁸. High cost of chemical pesticides and the environmental hazards as a result of pesticide usage have encouraged scientists to seek less hazardous and cheaper pesticide groups. Considerable efforts have been made to synthesize an alternative to overcome this problem⁵⁹. Botanical products have become more prominent in assessing current and future pest control alternatives. Over the past two decades, surveys of plant families have discovered sources of new botanical insecticides, which could possibly meet some of this demand. For instance, the potential of neem products is being conducted internationally and these products are widely used. Accordingly, botanical insecticides based on natural compounds from plants, are expected to be a possible alternative. They tend to have broad-spectrum activity, relative specificity in their mode of action and easy to process and use. They also tend to be safe for animals and the environment⁶⁰.

Some of the herb's having the larvicidal activity against the housefly, *Musca domestica* L. such as, *Azadirachta indica*, *Cichorium intybus*, *Citrus aurantiifolia*, *Conyzaa egyptiaca*, *Eucalyptus globulus* (fruits and leaves), *Opuntia vulgaris*,

Piper nigrum, *Punica granatum*, *Saccharum* sp., *Salix mucronata*, *Sonchus oleraceus* and *Zea mays* and also the *Piper nigrum* was the most toxic (LC₅₀ = 50.1 ppm) and the *Punica granatum* extract (LC₅₀ = 213.9 ppm) has the least toxic to *Musca domestica*⁶¹.

PHYTOBIOTICS AS FEED ADDITIVES IN POULTRY

Nowadays plant derived products used alternative or replace to antibiotic growth promoter. The extract from thyme, rosmery, sage and the carvacrol blend capsaicin, cinnamaldehyde using this bio active compounds can improved feed digestibility in broilers⁶². Plant based bio active components have digestion stimulating properties, antimicrobial activity, increase enzyme secretion it leads to improve digestibility of nutrients, increase feed intake.

The mode of action of phytobiotics is described through their potential effect on gut microbial population, through their potential reduction in pathogenic microorganism. Improve the digestive capacity of intestine may be the indirect effect of stabilizing the intestinal microbes. Components from the phytobiotics reduce the animal's immune stress during dangerous situation and increase the availability and absorption of essential nutrients from intestine, thereby improve the intake of feed, feed conversion efficiency and increased the growth rate. Supplementation of essential oils related to birds growth performance is still controversial⁶². Even though feeding of birds with essential oils among different age groups improved the growth performance^{9,22,63}. Effect of cinnammon and thyme in broilers diet have positive effect on birds feed intake, feed conversion efficiency, health status, live weight gain and performance traits⁶⁴.

Plant bioactive components like polysaccharide extracted from mushroom and herbs increased the growth of immune organs such as spleen thymus and bursal weight in both normal and immune-inhibition treated chickens⁶⁵. Some of the plant derivative polysaccharides are extracted from mushrooms and herbs (*Tremella fuciformis* and *Lentinus edodes* and *Astragalus membranaceus* Radix) acts as potential alternatives for health and antimicrobial growth promoters. Feeding garlic, ginger and their combination to broilers, 1% garlic significantly (p>0.05) increased body weight gain⁶⁶. Supplementation of garlic and black pepper combination significantly (p>0.01) increased feed intake, weight gain and feed efficiency in broilers⁶⁷.

GROWTH-PROMOTING EFFICACY

The growth promoting feed additives helps to reduce the stress in critical situation and helps to improve the immune

status of host animal and increase the absorption of nutrients which can help to improve the better growth of birds. Phytobiotic feed additives stabilize the feed hygiene and beneficially affect the intestinal microbial ecosystem through controlling the harmful pathogens⁶⁸. A mixture of essential oils from anise, oregano and citrus peels along with an antibiotic growth promoter and noticed decreased microbial activity in terminal ileum, cecum and colon, reduced chyme content of volatile fatty acids, reduced bacterial colony count as well as biogenic amines. Relief from the microbial activity and its related product from small intestine because production of volatile fatty acids counteracts intestinal pH stabilization and required for digestive enzyme activity. Formation of biological amines is causing toxicity via decarboxylation of limiting essential amino acids such as cadavarine from lysine and scatol from tryptophan⁶⁹. Using phyto-genic feed additives can alter the morphological changes in intestinal tissues and possible benefits to the digestive tract. Phyto-genic feed additives increased the villi length and reduced crypt depth in the jejunum and colon in broilers^{70,71}. In small intestine digestive capacity was improved due to use of these feed additives, this has the indirect side effect that stabilizing the microbial eubiosis in the gastro intestinal system. Organic acids and antibiotics an improved pre-cecal digestive capacity reduces the flux of fermentable matter into the hindgut and thus lessens the post ileal microbial growth and the excretion of bacterial matter in faeces in turkey⁷². As bacterial protein is the dominant fraction of total fecal protein, an improved pre-cecal digestive capacity may result indirectly in an increased apparent digestibility of dietary protein. Improvement in feed efficiency was concluded by adding a medicinal mixture (including 10 g kg⁻¹ oregano, 5 g kg⁻¹ *Ziziphora* and 5 g kg⁻¹ peppermint) to broilers diet that can show the improvement of feed conversion ratio is related with improvement of crude protein digestibility⁷³. Addition of 4 g kg⁻¹ black seed to broilers diet improved FCR without affecting body weight gain⁷⁴. Hydrophylic extract of liquid fresh green tea at the level of 0.1 or 0.2 g kg⁻¹ in broiler diet it increased body weight, feed efficiency, carcass weight and dressed weight⁷⁵. Adding green tea by products in broiler and layer diets improved the productive performance and in serum and yolk it reduces the cholesterol content⁷⁶. Inclusion of thyme essential oil in Japanese quail diet may increase the cumulative body weight gain, cumulative feed intake and cumulative feed conversion efficiency¹⁰. The active principle of thyme oil act as digestibility enhancer, maintain the important microbial flora content in gut system, stimulate the endogenous digestive enzyme secretion and thus improving the growth performance in poultry. Inclusion of ginger powder (0.5, 1 and 1.5%) in broiler diet showed increased

breast and thigh muscle yield and reduced abdominal fat content at 1.5% ginger powder inclusion level⁷⁷. Dietary thyme essential oil in broiler diet showed higher weight gain at 0.15 mL L⁻¹ of water⁷⁸. Inclusion of black pepper, fenugreek and garlic in broiler diet at 10 g kg⁻¹, black pepper fed group showed higher eviscerated percentage due to digestive stimulant effect and fenugreek reduced abdominal fat content in broiler meat, due to the anti-cholesterimic effect⁷⁹.

GUT MORPHOLOGY

Broilers supplemented with peppermint essential oil had higher duodenal crypt depth⁸⁰. Thyme essential oil in Japanese quail diet can help to increased *Lactobacillus* and decreased *E. coli* in the ileum¹⁰. The essential oils (carvacrol, 24.5%), 1,8-cineole (20.1%), camphor (12.1%) and thymol (6.0%) in broiler diet showed longer intestine and heavier caecal weight⁸¹. Oregano (*Origanum vulgare* L.) essential oil in broiler diet at the rate of (300, 600 and 1200 mg kg⁻¹ of diet) showed increase the useful bacteria (lactic acid bacteria) 1200 mg kg⁻¹ dose and reduce the harmful bacteria (*Escherichia coli*) at 300 mg kg⁻¹ oregano essential oil⁸². Thyme and cinnamom at 0.5 and 1% inclusion level favourably altered antimicrobial balance (reduced total bacteria count and coli form group in jejunum and large intestine) in broilers gastro intestinal tract⁶⁶. Addition thyme essence in water had positive increase in desirable bacteria and reduced unfavourable bacteria⁷⁸. Dietary supplementation of ginger, garlic at 1% level, villus height, length, crypt and depth ratio was significantly ($p < 0.05$) increased at 1% ginger. It increased intestinal absorptive area⁶⁶.

ANTI COCCIDIAL ACTIVITY

Photobiotic are having anticoccidial activity. *Eimeria* (an intracellular protozoan) belonging to the phylum Apicomplexa cause avian coccidiosis, which is an economically important disease in the poultry industry⁸³. In five week old chicken with supplementation of green tea at 0.5 and 2% level it reduces the oocytes in faecal material at 38.5 and 51.5% level⁶³.

IMMUNE STIMULATORY EFFECT

Certain plant derived polysaccharides have (adjuvant effect) immunostimulatory substances⁸⁴. Phyto-genic compounds have prebiotic effect it helpful to improve or beneficial effect on gut health by improving the gut associated lymphoid tissue response (immunological effect)

directly or short chain fatty acid mediation such as lactic acid producing bacteria⁸⁵. Polysaccharide compounds derived from mushroom (*Tremella fuciformis*, *Lentinus endodes*) and herbs (*Astragalus membranaceus* Radix) used in broiler chicken, these compounds act as immune modulator or enhancer to improve the innate or adaptive immunity or specific immunity (both cellular and humoral immunity⁶⁵. Dietary fructo-oligosaccharide changes the intestinal microflora environment and leads to regulate the secretion of immunoglobulin A (IgA) in murine Peyer's patches cell in intestinal mucosa⁸⁶.

CONTROL OF OCHRATOXIN BY PHYTOCHEMICAL

Azadirachta indica (neem) extracts had effect on mycelial growth, sporulation, morphology and OTA production by *P. verrucosum* and *P. brevicompactum*⁸⁷. Sesquiterpenes are high in volatile essential oils of lemon balm plant and have a bitter principal to them. These compounds have been found to have very high anti-fungal properties.

BLOOD CHARACTERISTICS

Supplementation of 20 g kg⁻¹ black cumin in broiler diet it reduces the triglyceride and cholesterol level in blood. Raising the level of green tea from 0.25-0.75% significantly decreased blood lipid fractions and increased high-density lipoprotein (HDL) in Japanese quail⁸⁸ and reduced LDL-cholesterol content in broilers⁸⁹. Dietary inclusion of thyme oil to Japanese quail reduces the serum glucose, triglycerides and total cholesterol. Blood glucose level is a biochemical indicator of stress from in this thyme oil reduced the blood glucose, so it could be justified that stress lowering effect¹⁰. Thyme essential oil reduce synthesis of hepatic 3-hydroxy-3-methylglutaryl coenzyme A (HMGCoA) reductase this is the enzyme responsible for cholesterol synthesis¹¹. Dietary inclusion of ginger root powder in broilers diet at the level of 2% reduces the serum cholesterol level⁹⁰. Dietary inclusion of ginger in broiler diet (0.2%) lowering the effect on blood serum cholesterol, triglycerides and glucose also potent effect antioxidant action and antistress effect³⁵. Inclusion of 1.5% ginger powder in broiler diet reduced the serum glucose, triglyceride and cholesterol level⁷⁷.

SAFETY ISSUES OF PHYTOBIOTICS

Phytobiotics contain many pharmacologically active components which play a major role in the defence system of

the plant. From this point of view safety concerns cannot be excluded, although, phytobiotics are generally assumed as Generally Recognized As Safe (GRAS)⁹¹. Phytobiotics contain as well irritating ingredients which may be harmful to both animals and humans. Furthermore, undesired residues may not be excluded in a natural product. In several experiments it was observed that bioactive ingredients or their metabolites may be transferred to tissues⁹². Some concerns are reported for capsaicin (cancer causing), cyanide containing ingredients, carvacrol (in oregano) and glycosides. The content of capsaicin in chilli (paprika) is limited by EU regulation (only regulated for paprika as food additive E160c, <250 mg kg⁻¹; EU, 1995) due to probable toxic effects. Few information is available on metabolites deposited as residues to tissues resulting in impaired sensory attributes. In general, for probable negative effects the dose is important. High inclusion levels of oregano may affect the Minimum Inhibitory Concentration (MIC) for antibiotics¹². For rats the LD₅₀ value was reached for oregano oil for a dose 100 times the normal supplementation level in feed. The observed results indicate the necessity to check phytobiotics thoroughly for potential safety issues.

CONCLUSION

The anti-microbial more specifically the anti-bacterial spectrum activity of phyto-chemicals is well documented worldwide; although, their mechanism of action is not clearly described yet. Their antioxidant activity and hypo-lipidemic property of these plant derived compounds are being explored now in both animals as well as in human nutrition. These phytobiotics also impart readily acceptable flavour to the products especially meat and eggs. They also aid in digestive process via stimulating the digestive secretions throughout the gastro-intestinal tract thereby increasing the overall digestibility of the nutrients and reduce the environmental pollution. Their antifungal activity can be used in preservation or improving the keeping quality of the raw as well as the finished feeds free of mycotoxin contamination and insects attack. These phytobiotics favourably alters the gut microbial population and improves the overall performance of the individual. They also improve the shelf life of products mainly of preserved meat and eggs. With their wide range of activities these phytobiotics can be considered as a new group of feed additive for better growth rather than simply looking them as an alternate to antibiotics in food production industry.

REFERENCES

1. Kamel, C., 2001. Tracing Modes of Action and the Roles of Plant Extracts in Non-Ruminants. In: Recent Advances in Animal Nutrition, Garnsworthy, P.C. and J. Wiseman (Eds.). Nottingham University Press, Nottingham, UK., pp: 135-150.
2. Athanasiadou, S., J. Githiori and I. Kyriazakis, 2007. Medicinal plants for helminth parasite control: Facts and fiction. *Animal*, 1: 1392-1400.
3. Windisch, W. and A. Kroismayr, 2007. Natural phytobiotics for health of young piglet and poultry: Mechanisms and application. *Poult. Sci.*, 86: 643-643.
4. Kamel, C., 2000. A novel look at a classic approach of plant extracts. *Feed Mix*, 3: 19-23.
5. Bote, C.J.L., 2004. Bioflavonoid's effects reach beyond productivity. *Feed Mix.*, 12: 12-15.
6. Chattopadhyay, D., G. Arunachalam, L. Ghosh, K. Rajendran, A.B. Mandal and S.K. Bhattacharya, 2005. Antipyretic activity of *Alstonia macrophylla* Wall ex A. DC: An ethnomedicine of Andaman Islands. *Pharm. Pharm. Sci.*, 8: 558-564.
7. Hashemi, S.R., I. Zulkifli, M. Hair-Bejo, M. Karami and A.F. Soleimani, 2009. The effects of *Euphorbia hirta* and acidifier supplementation on growth performance and antioxidant activity in broiler chickens. Proceedings of the 21st Veterinary Association Malaysia (VAM) Congress, August 7-9, 2009, Port Dickson, Malaysia, pp: 215-217.
8. Franco-Jimenez, D.J., S.E. Scheideler, R.J. Kittok, T.M. Brown-Brandl, L.R. Robeson, H. Taira and M.M. Beck, 2007. Differential effects of heat stress in three strains of laying hens. *J. Applied Poult. Res.*, 16: 628-634.
9. Cross, D.E., R.M. McDevitt, K. Hillman and T. Acamovic, 2007. The effect of herbs and their associated essential oils on performance, dietary digestibility and gut microflora in chickens from 7 to 28 days of age. *Br. Poult. Sci.*, 48: 496-506.
10. Khaksar, V., M.M. van Krimpen, H. Hashemipour and M. Pilevar, 2012. Effects of thyme essential oil on performance, some blood parameters and ileal microflora of Japanese quail. *J. Poult. Sci.*, 49: 106-110.
11. Lee, K.W., H. Everts, H.J. Kappert, K.H. Yeom and A.C. Beynen, 2003. Dietary carvacrol lowers body weight gain but improves feed conversion in female broiler chickens. *J. Applied Poult. Res.*, 12: 394-399.
12. Mountzouris, K.C., V. Paraskevas and K. Fegeros, 2009. Phytogenic Compounds in Broiler Nutrition. In: Phytogenics in Animal Nutrition, Steiner, T. (Ed.). Nottingham University Press, Nottingham, ISBN-13: 9781904761716.
13. Windisch, W., K. Schedle, C. Piltzner and A. Kroismayr, 2008. Use of phytogenic products as feed additives for swine and poultry. *J. Anim. Sci.*, 86: E140-E148.
14. Cowan, M.M., 1999. Plant products as antimicrobial agents. *Clin. Microbiol. Rev.*, 12: 564-582.
15. Gopi, M., K. Karthik, H.V. Manjunathachar, P. Tamilmahan and M. Kesavan *et al.*, 2014. Essential oils as a feed additive in poultry nutrition. *Adv. Anim. Vet. Sci.*, 2: 1-7.
16. Chung, K.T., S.E. Stevens Jr., W.F. Lin and C.I. Wei, 1993. Growth inhibition of selected food-borne bacteria by tannic acid, propyl gallate and related compounds. *Lett. Applied Microbiol.*, 17: 29-32.
17. Karou, D., A. Savadogo, A. Canini, S. Yameogo and C. Montesano *et al.*, 2005. Antibacterial activity of alkaloids from *Sida acuta*. *Afr. J. Biotechnol.*, 4: 1452-1457.
18. Morrissey, J.P. and A.E. Osbourn, 1999. Fungal resistance to plant antibiotics as a mechanism of pathogenesis. *Microbiol. Mol. Biol. Rev.*, 63: 708-724.
19. Lee, K.W., H. Everts and A.C. Beynen, 2004. Essential oils in broiler nutrition. *Int. J. Poult. Sci.*, 3: 738-752.
20. Jamroz, D., J. Orda, C. Kamel, A. Wiliczekiewicz, T. Wertelecki and J. Skorupinska, 2003. The influence of phytogenic extracts on performance, nutrient digestibility, carcass characteristics and gut microbial status in broiler chickens. *J. Anim. Feed Sci.*, 12: 583-596.
21. Mitsch, P., K. Zitterl-Eglseer, B. Kohler, C. Gabler, R. Losa and I. Zimpernik, 2004. The effect of two different blends of essential oil components on the proliferation of *Clostridium perfringens* in the intestines of broiler chickens. *Poult. Sci.*, 83: 669-675.
22. Conner, D.E., 1993. Naturally Occurring Compounds. In: Antimicrobials in Foods, Davidson, P. and A.L. Branen (Eds.). Marcel Dekker, Inc., New York, ISBN: 0-8247-8906-7, pp: 441-468.
23. Farag, R.S., A.Z.M.A. Badei, F.M. Hewedi and G.S.A. El-Baroty, 1989. Antioxidant activity of some spice essential oils on linoleic acid oxidation in aqueous media. *J. Am. Oil Chem. Soc.*, 66: 792-799.
24. Helander, I.M., H.L. Alakomi, K. Latva-Kala, T. Mattila-Sandholm and I. Pol *et al.*, 1998. Characterization of the action of selected essential oil components on gram-negative bacteria. *J. Agric. Food Chem.*, 46: 3590-3595.
25. Bowles, B.L. and A.J. Miller, 1993. Antibotulinal properties of selected aromatic and aliphatic aldehydes. *J. Food Prod.*, 56: 788-794.
26. Fritz, Z., A. Schleider and S. Kinal, 1993. Effect of substituting milfoil, St. Johnswort and lovage for antibiotics on chicken performance and meat quality. *J. Anim. Feed Sci.*, 2: 189-195.
27. Aksit, M., E. Goksoy, F. Kok, D. Ozdemir and M. Ozdogan, 2006. The impacts of organic acid and essential oil supplementations to diets on the microbiological quality of chicken carcasses. *Arch. Geflugelkd.*, 70: 168-173.
28. Newton, S.M., C. Lau, S.S. Gurcha, G.S. Besra and C.W. Wright, 2002. The evaluation of forty-three plant species for *in vitro* antimycobacterial activities; isolation of active constituents from *Psoralea corylifolia* and *Sanguinaria canadensis*. *J. Ethnopharmacol.*, 79: 57-67.

29. Burt, S., 2004. Essential oils: Their antibacterial properties and potential applications in foods: A review. *Int. J. Food Microbiol.*, 94: 223-253.
30. Okitoi, L.O., H.O. Ondwasy, D.N. Siamba and D. Nkurumah, 2007. Traditional herbal preparations for indigenous poultry health management in Western Kenya. *Livestock Res. Rural Dev.*, Vol. 19.
31. Trombetta, D., F. Castelli, M.G. Sarpietro, V. Venuti and M. Cristani *et al.*, 2005. Mechanisms of antibacterial action of three monoterpenes. *Antimicrob. Agents. Chemother.*, 49: 2474-2478.
32. Kirkpinar, F., H.B. Unlu and G. Ozdemir, 2011. Effects of oregano and garlic essential oils on performance, carcass, organ and blood characteristics and intestinal microflora of broilers. *Livest. Sci.*, 137: 219-225.
33. Murali, N., G.S. Kumar-Phillips, N.C. Rath, J. Marcy and M.F. Slavik, 2012. Effect of marinating chicken meat with lemon, green tea and turmeric against foodborne bacterial pathogens. *Int. J. Poult. Sci.*, 11: 326-332.
34. Yassin, M.A., A.M.A. El-Samawaty, M. Moslem, A. Bahkali and K.A. Abd-El Salam, 2011. Fungal biota and occurrence of aflatoxigenic *Aspergillus* postharvest corn grains. *Fresenius Environ. Bull.*, 20: 903-909.
35. Mohamed, A.B., M.A.M. Al-Rubaei and A.Q. Jalil, 2012. Effect of ginger (*Zingiber officinale*) on performance and blood serum parameters of broiler. *Int. J. Poult. Sci.*, 11: 143-146.
36. Akgul, A. and M. Kivanc, 1988. Inhibitory effects of selected Turkish spices and oregano components on some foodborne fungi. *Int. J. Food Microbiol.*, 6: 263-268.
37. Omaf, 2004. Pesticide storage, handling and application. Ontario Ministry of Agriculture, Food and Rural Affairs. California, USA.
38. Owino, P.O. and S.W. Waudu, 1992. Medicinal plants of Kenya: Effects on *Meloidogyne incognita* and the growth of okra. *Afro-Asian J. Nematol.*, 2: 64-66.
39. Anjorin, S.T. and E.A. Salako, 2009. The status of pesticidal plants and materials identification in Nigeria. *Niger. J. Plant Prot.*, 23: 25-32.
40. Afzal, R., S.M. Mughal, M. Munir, K. Sultana, R. Qureshi, M. Arshad and M.K. Laghari, 2010. Mycoflora associated with seeds of different sunflower cultivars and its management. *Pak. J. Bot.*, 42: 435-445.
41. Abou-Bakr, S., 2011. Effect of some plant extracts on fungal and aflatoxin production. *Int. J. Acad. Res.*, 3: 116-120.
42. Tagoe, D.N.A., H.D. Nyarko and R. Akpaka, 2011. A comparison of the antifungal properties of onion (*Allium cepa*), ginger (*Zingiber officinale*) and garlic (*Allium sativum*) against *Aspergillus flavus*, *Aspergillus niger* and *Cladosporium herbarum*. *Res. J. Med. Plant*, 5: 281-287.
43. Bhatnagar, D., H.J. Zeringue and S.P. McCormick, 1990. Neem leaf extracts inhibit aflatoxin biosynthesis in *Aspergillus flavus* and *Aspergillus parasiticus*. Proceedings of the USDA Neem Workshop, April 16-17, Beltsville, Maryland, pp: 118-127.
44. Awuah, R.T., 1996. Possible utilization of plant products in grain storage. Proceedings of the Workshop on Mycotoxins in Food in Africa, Nov. 6-10, International Institute of Tropical Agriculture, Benin, pp: 32-32.
45. Hasan, M.M., S.P. Chowdhury, Shahidul Alam, B. Hossain and M.S. Alam, 2005. Antifungal effects of plant extracts on seed-borne fungi of wheat seed regarding seed germination, Seedling health and vigour index. *Pak. J. Biol. Sci.*, 8: 1284-1289.
46. Deans, S.G. and P.G. Waterman, 1993. Biological Activities of Volatile Oils. In: *Volatile Oil Crops: Their Biology, Biochemistry and Production*, Hay, R.K.M. and P.G. Waterman (Eds.). Longman, London, UK., pp: 97-111.
47. Wei, A. and T. Shibamoto, 2007. Antioxidant activities and volatile constituents of various essential oils. *J. Agric. Food Chem.*, 55: 1737-1742.
48. Cuppett, S.L. and C.A. Hall, 1998. Antioxidant activity of the Labiatae. *Adv. Food. Nutr. Res.*, 42: 245-271.
49. Nakatani, N., 1994. Antioxidants from Spices and Herbs. In: *Food Phytochemicals for Cancer Prevention II. Teas, Spices and Herbs*, Huang, M.J., T. Osawa, C.T. Ho and R. Rosen (Eds.). American Chemical Society, Washington DC., USA.
50. Basmacioglu, H., O. Tokusoglu and M. Ergul, 2004. The effect of oregano and rosemary essential oils or alpha-tocopherol acetate on performance and lipid oxidation of meat enriched with n-3 PUFA's in broilers. *South Afr. J. Anim. Sci.*, 34: 197-210.
51. Giannenas, I., P. Florou-Paneri, N.A. Botsoglou, E. Christaki and A.B. Spais, 2005. Effect of supplementing feed with oregano and/or α -tocopheryl acetate on growth of broiler chickens and oxidative stability of meat. *J. Anim. Feed Sci.*, 14: 521-535.
52. Florou-Paneri, P., I. Giannenas, E. Christaki, A. Govaris and N. Botsoglou, 2006. Performance of chickens and oxidative stability of the produced meat as affected by feed supplementation with oregano, vitamin C, vitamin E and their combinations. *Arch. Ge ugelkd.*, 70: 232-240.
53. Botsoglou, N.A., A. Govaris, E.N. Botsoglou, S.H. Grigoropoulou and G. Papageorgiou, 2003. Antioxidant activity of dietary oregano essential oil and alpha-tocopheryl acetate supplementation in long-term frozen stored Turkey meat. *J. Agric. Food Chem.*, 51: 2930-2936.
54. Janz, J.A.M., P.C.H. Morel, B.H.P. Wilkinson and R.W. Purchas, 2007. Preliminary investigation of the effects of low-level dietary inclusion of fragrant essential oils and oleoresins on pig performance and pork quality. *Meat Sci.*, 75: 350-355.
55. Ancsin, Z., M. Erdelyi and M. Mezes, 2009. Effect of rosemary and garlic oil supplementation on glutathione redox system of broiler chickens. *Acta Biol. Szegediensis*, 53: 19-21.
56. Kirubakaran, A., M. Moorthy, R. Chitra and G. Prabakar, 2016. Effect of Fenugreek garlic and black pepper supplementation on chemical properties of fresh and frozen broiler meat. *Indian Vet. J.*

57. Karpouhtsis, I., E. Pardali, E. Feggou, S. Kokkini, Z.G. Scouras and P. Mavragani-Tsipidou, 1998. Insecticidal and genotoxic activities of oregano essential oils. J. Agric. Food Chem., 46: 1111-1115.
58. Khan, M.F. and S.M. Ahmed, 2000. Toxicity of crude neem leaf extract against housefly *Musca domestica* L. Adults as compared with DDVP, dichlorvos. Turk. J. Zool., 24: 219-223.
59. NRC., 2000. The future role of pesticides in U.S. agriculture. Committee on the Future Role of Pesticides in U.S. Agriculture, Board on Agriculture and Natural Resources and Board on Environmental Studies and Toxicology, Commission on Life Sciences, National Academy of Sciences, Washington DC., USA.
60. Belmain, S.R., G.E. Neal, D.E. Ray and P. Golob, 2001. Insecticidal and vertebrate toxicity associated with ethnobotanicals used as post-harvest protectants in Ghana. Food Chem. Toxicol., 39: 287-291.
61. Mansour, S.A., R.F.A. Bakr, R.I. Mohamed and N.M. Hasaneen, 2011. Larvicidal activity of some botanical extracts, commercial insecticides and their binary mixtures against the housefly, *Musca domestica* L. Open Toxinol. J., 4: 1-13.
62. Hernandez, F., J. Madrid, V. Garcia, J. Orengo and M.D. Megias, 2004. Influence of two plant extracts on broilers performance, digestibility and digestive organ size. Poult. Sci., 83: 169-174.
63. Jang, S.I., M.H. Jun, H.S. Lillehoj, R.A. Dalloul, I.K. Kong, S. Kim and W. Min, 2007. Anticoccidial effect of green tea-based diets against *Eimeria maxima*. Vet. Parasitol., 144: 172-175.
64. Al-Kassie, G.A.M., 2010. The effect of thyme and cinnamon on the microbial balance in gastro intestinal tract on broiler chicks. Int. J. Poult. Sci., 9: 495-498.
65. Guo, F.C., H.F.J. Sacelkoul, R.P. Kwakkel, B.A. Williams and M.W.A. Verstegen, 2003. Immunoactive, medicinal properties of mushroom and herb polysaccharides and their potential use in chicken diets. World's Poult. Sci. J., 59: 427-440.
66. Karangiya, V.K., H.H. Savsani, S.S. Patil, D.D. Garg, K.S. Murthy, N.K. Ribadiya and S.J. Vekariya, 2016. Effect of dietary supplementation of garlic, ginger and their combination on feed intake, growth performance and economics in commercial broilers. Vet. World, 9: 245-250.
67. Kirubakaran, A., M. Moorthy, R. Chitra and G. Prabakar, 2016. Influence of combinations of fenugreek, garlic and black pepper powder on production traits of the broilers. Vet. World, 9: 470-474.
68. Roth, F.X. and M. Kirchgessner, 1998. Organic acids as feed additive for young pigs: Nutritional and gastrointestinal effects. J. Anim. Feed Sci., 7: 25-33.
69. Kroismayr, A., J. Sehm, M.W. Pfaffl, K. Schedle, C. Plitzner and W. Windisch, 2008. Effects of avilamycin and essential oils on mRNA expression of apoptotic and inflammatory markers and gut morphology of piglets. Czech J. Anim. Sci., 53: 377-387.
70. Jamroz, D., T. Wiertelcki, M. Houszka and C. Kamel, 2006. Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. J. Anim. Physiol. Anim. Nutr., 90: 255-268.
71. Demir, E., S. Sarica, M.A. Ozcan and M. Suicmez, 2003. The use of natural feed additives as alternatives for an antibiotic growth promoter in broiler diets. Br. Poult. Sci., 44: 44-45.
72. Seskeviciene, J., J. Jankowski and K. Kozlowski, 2005. [Effect of probiotic preparation and phyto-genous feed additive on digestibility of nutrients, metabolizability of gross energy and content of metabolizable energy of a practical feed ration for fattening turkeys]. Archiv Geflugelkunde, 69: 107-109, (In German).
73. Narimani-Rad, M., A. Nobakht, H.A. Shahryar, J. Kamani and A. Lotfi, 2011. Influence of dietary supplemented medicinal plants mixture (ziziphora, oregano and peppermint) on performance and carcass characterization of broiler chickens. J. Med. Plants Res., 5: 5626-5629.
74. Toghyani, M., M. Toghyani, A. Gheisari, G. Ghalamkari and M. Mohammadrezaei, 2010. Growth performance, serum biochemistry and blood hematology of broiler chicks fed different levels of black seed (*Nigella sativa*) and peppermint (*Mentha piperita*). Livestock Sci., 129: 173-178.
75. Eren, G., N. Ocak, A. Altop, S. Cankaya, H. Aksoy and E. Ozturk, 2011. Growth performance, meat quality and caecal coliform bacteria count of broiler chicks fed diet with green tea extract. Asian-Aust. J. Anim. Sci., 24: 1128-1135.
76. Khan, S.H., 2014. The use of green tea (*Camellia sinensis*) as a phyto-genic substance in poultry diets. Onderstepoort J. Vet. Res., 81: 1-8.
77. Barazesh, H., M.B. Pour, S. Salari and T.M. Abadi, 2013. The effect of ginger powder on performance, carcass characteristics and blood parameters of broilers. Int. J. Adv. Bio. Biomed. Res., 1: 1645-1651.
78. Saki, A.A., M. Kalantar and V. Khoramabadi, 2014. Effects of drinking thyme essence (*Thymus vulgaris* L.) on growth performance, immune response and intestinal selected bacterial population in broiler chickens. Poult. Sci. J., 2: 113-123.
79. Kirubakaran, A., M. Moorthy, R. Chitra and G. Prabakar, 2016. Effect of phyto additive supplementation on broiler carcass characteristics. Indian Vet. J.
80. Emami, N.K., A. Samie, H.R. Rahmani and C.A. Ruiz-Feria, 2012. The effect of peppermint essential oil and fructooligosaccharides, as alternatives to virginiamycin, on growth performance, digestibility, gut morphology and immune response of male broilers. Anim. Feed Sci. Technol., 175: 57-64.
81. Bozkurt, M., N. Selek, K. Kucukyilmaz, H. Eren, E. Guven, A.U. Catli and M. Cinar, 2012. Effects of dietary supplementation with a herbal extract on the performance of broilers infected with a mixture of *Eimeria* species. Br. Poult. Sci., 51: 325-332.

82. Roofchae, A., M. Irani, M.A. Ebrahimzadeh and M.R. Akbari, 2011. Effect of dietary oregano (*Origanum vulgare* L.) essential oil on growth performance, cecal microflora and serum antioxidant activity of broiler chickens. Afr. J. Biotechnol., 10: 6177-6183.
83. Giannenas, I.A. and I. Kyriazakis, 2009. Phytobased Products for the Control of Intestinal Diseases in Chickens in the Post Antibiotic Era. In: Phytogenics in Animal Nutrition: Natural Concepts to Optimize Gut Health and Performance, Steiner, T. (Ed.). Nottingham University Press, England, ISBN: 9781904761716, pp: 61-85.
84. Chen, H.L., D.F. Li, B.Y. Chang, L.M. Gong, J.G. Dai and G.F. Yi, 2003. Effects of Chinese herbal polysaccharides on the immunity and growth performance of young broilers. Poult. Sci., 82: 364-370.
85. Hosono, A., A. Ozawa, R. Kato, Y. Ohnishi, Y. Nakanishi, T. Kimura and R. Nakamura, 2003. Dietary fructooligosaccharides induce immunoregulation of intestinal IgA secretion by murine Peyer's patch cells. Biosci. Biotechnol. Biochem., 67: 758-764.
86. Roller, M., G. Rechkemmer and B. Watzl, 2004. Prebiotic inulin enriched with oligofructose in combination with the probiotics *Lactobacillus rhamnosus* and *Bifidobacterium lactis* modulates intestinal immune functions in rats. J. Nutr., 134: 153-156.
87. Mossini, S.A.G., C.C. Arroteia and C. Kimmelmeier, 2009. Effect of neem leaf extract and Neem oil on *Penicillium* growth, sporulation, morphology and ochratoxin A production. Toxins, 1: 3-13.
88. Abdel-Azeem, F.A., 2005. Green tea flowers (*Camellia sinensis*) as natural anti-oxidants feed additives in growing Japanese quail diets. Egypt. Poult. Sci. J., 25: 569-588.
89. Yang, C.J., I.Y. Yang, D.H. Oh, I.H. Bae and S.G. Cho *et al.*, 2003. Effect of green tea by-product on performance and body composition in broiler chicks. Asian-Australasian J. Anim. Sci., 16: 867-872.
90. Zomrawi, W.B., K.A.A. Atti, B.M. Dousa and A.G. Mahala, 2013. The effect of dietary ginger root powder (*Zingiber officinale*) on broiler chicks performance, carcass characteristic and serum constituents. J. Anim. Sci. Adv., 3: 42-47.
91. Mathe, A., 2009. Essential Oils: Biochemistry, Production and Utilisation. In: Phytogenics in Animal Nutrition: Natural Concepts to Optimize Gut Health and Performance, Steiner, T. (Ed.). Nottingham University Press, England, ISBN: 9781904761716.
92. Windisch, W., E. Rohrer and K. Schedle, 2009. Phytogenic Feed Additives to Young Piglets and Poultry. In: Phytogenics in Animal Nutrition-Natural Concepts to Optimize Gut Health and Performance, Steiner, T. (Ed.). Nottingham University Press, Nottingham, UK., pp: 71-76.