

American Journal of **Biochemistry and Molecular Biology**

ISSN 2150-4210



American Journal of Biochemistry and Molecular Biology 1 (3): 291-309, 2011 ISSN 2150-4210 / DOI: 10.3923/ajbmb.2011.291.309 © 2011 Academic Journals Inc.

Bioactive Compounds from Endophytes and their Potential in Pharmaceutical Effect: A Review

Baby Joseph and R. Mini Priya

Interdisciplinary Research Centre, Malankara Catholic College, Mariagiri, Kaliakkavilai, Kanyakumari District, Tamil Nadu, 629153, South India

Corresponding Author: Baby Joseph, Interdisciplinary Research Centre, Malankara Catholic College, Mariagiri, Kaliakkavilai, Kanyakumari District, Tamil Nadu, 629153, South India

ABSTRACT

Endophytes are microorganisms that live inside the host plant tissues which have novel metabolites exhibiting a variety of biological activities against different diseases. In fact, a significant number of interesting natural bioactive compounds have been reported in the last years. The microbial biotransformation process is a novel alternative method to obtain bioactive compounds. In this review, some aspects in the phytochemistry of endophytes producing Phytochemicals and its pharmaceutical effects are discussed.

Key words: Bioactive compounds, endophytes, antimicrobial effect, antioxidant effect, anticancer effect, insecticidal effect, antidiabetic effect, immunosuppressive effect

INTRODUCTION

Many complementary and alternative medicines have enjoyed increased popularity in recent decades (Joseph and Priya, 2011). Drugs derived from natural sources play a significant role in the prevention and treatment of human diseases. About 61% of new drugs developed between 1981 and 2002 were based on natural products and they have been very successful especially in the areas of infectious disease and cancer (Cragg and Newman, 2005). More than 90% of the terms recorded in Indian medical literature are derived from plant sources (Joseph and Priya, 2010). For the fast two years, there has been an increasing interest in the investigation of different novel natural bioactive products from plants (Joseph et al., 2010). Recent trends, however, show that the discovery rate of active novel chemical entities is declining (Lam, 2007). Therefore, there is a need to bio-prospect new sources and if possible from less explored regions and habitats to maximize the discovery of novel bioactive metabolites.

Endophytes are microorganisms that include bacteria and fungi living within plant tissues without causing any immediate overt negative effects have been found in every plant species examined to date and recognized as the potential sources of novel natural products for exploitation in medicine, agriculture and industry with more bioactive natural products isolated from the microorganisms (Bacon and White, 2000; Strobel and Daisy, 2003; Kumar and Sagar, 2007). Endophytes are ubiquitous with rich biodiversity, which have been found in every plant species examined to date. It is noteworthy that, of the nearly 3, 00,000 plant species that exist on the earth, each individual plant is the host to one or more endophytes (Strobel and Daisy, 2003). In this view of the special colonization in certain hosts, it is estimated that there may be as many as

1 million different endophyte species. However, only a handful of them have been described (Andrew and Hirano, 1991), which means the opportunity to find new and targeting natural products from interesting endophytic microorganisms among myriads of plants in different niches and ecosystems is great. Some of the endophytes are the chemical synthesizers in inside the plants (Owen and Hundley, 2004).

Many of them are capable of synthesizing bioactive compounds that can be used by plants for defense against human pathogens and some of these compounds have been proven useful for novel drug discovery. Recent studies have reported hundreds of natural products including substance of alkaloids, terpenoids, flavonoids, steroids, etc. from endophytes. Up to now, most of the natural products from endophytes are antibiotics, anticancer agents, biological control agents and other bioactive compounds by their different functional roles. Thus far, they have not been widely explored for therapeutic properties. A single endophyte may be able to produce not one but several bioactive metabolites. As a result, the role of endophytes in the production of novel structures for exploitation in medicine is receiving increased attention (Wang et al., 2000; Ezra et al., 2004a; Gunatilaka, 2006).

A small amount of endophytes have been studied, recently, several research groups have been motivated to evaluate and elucidate the potential of these microorganisms applied on biotechnological processes focusing on the production of bioactive compounds. The production of bioactive substances by endophytes is directly related to the independent evolution of these microorganisms, which may have incorporated genetic information from higher plants, allowing them to better adapt to plant host and carry out some functions such as protection from pathogens, insects and grazing animals (Strobel, 2003). Endophytes are chemical synthesizer inside plants (Owen and Hundley, 2004), in other words, they play a role as a selection system for microbes to produce bioactive substances with low toxicity toward higher organisms (Strobel, 2003). Bioactive natural compounds produced by endophytes have been promising potential usefulness in safety and human health concerns, although there is still a significant demand of drug industry for synthetic products due to economic and time-consuming reasons (Strobel et al., 2004). Problems related to human health such as the development of drug resistance in human pathogenic bacteria, fungal infections and life threatening virus claim for new therapeutic agents for effective treatment of diseases in human, plants and animals that are currently unmet (Strobel and Daisy, 2003; Strobel, 2003; Zhang et al., 2005). Recent review by Cragg and Newman (2007) presented a list of all approved agents from 1981 to 2006, from which a significant number of natural drugs are produced by microbes and/or endophytes. Endophytes provide a broad variety of bioactive secondary metabolites with unique structure, including alkaloids, benzopyranones, chinones, flavonoids, phenolic acids, quinones, steroids, terpenoids, tetralones, xanthones and others (Tan and Zou, 2001). Such bioactive metabolites find wide-ranging application as agrochemicals, antibiotics, immunosuppressants, antiparasitics, antioxidants anticancer agents and (Gunatilaka, 2006).

Methods to obtain bioactive compounds include the extraction from a natural source, the microbial production via fermentation, or microbial transformation. Extraction from natural sources presents some disadvantages such as dependency on seasonal, climatic and political features and possible ecological problems involved with the extraction, thus calling for innovative approaches to obtain such compounds (Bicas et al., 2009). Hence, biotechnological techniques by using different microorganisms appear promising alternatives for establishing an inexhaustible, cost-effective and renewable resource of high-value bioactive products and aroma compounds. The biotransformation

method has a huge number of applications (Borges *et al.*, 2009), for instance, it has been extensively employed for the production of volatile compounds (Bicas *et al.*, 2009; Bicas *et al.*, 2008; Krings *et al.*, 2006). These volatile compounds possess not only sensory properties, but other desirable properties such as antimicrobial (vanillin, essential oil constituents), antifungal and antiviral (some alkanolides), antioxidant (eugenol, vanillin), somatic fat reducing (nootkatone), blood pressure regulating (2-[E]-hexenal), anti-inflammatory properties (1,8-cineole) and others (Berger, 2009).

The intent of this review is to provide insights into the phytochemistry of endophytes producing phytochemicals, biological effects of endophytes producing bioactive compounds, the importance of including endophytic microbes for novel drugs and the microbial biotransformation process as a novel alternative method to obtain bioactive compounds. This review, however, also describes these compounds by different functions and pharmaceutical potential for human use.

PHYTOCHEMISTRY

Tan and Zou (2001) believe the reason why some endophytes produce certain phytochemicals originally characteristic of the host might be related to a genetic recombination of the endophyte with the host that occurs in evolutionary time. This is a concept that was originally proposed as a mechanism to explain why the endophytic fungus *T. andreanae* may be producing paclitaxel (Stierle *et al.*, 1993). Thus, if endophytes can produce the same rare and important bioactive compounds as their host plants, this would not only reduce the need to harvest slowgrowing and possibly rare plants but also preserve the world's ever diminishing biodiversity. Furthermore, it is recognized that a microbial source of a valued product may be easier and more economical to produce, effectively reducing its market price (Strobel and Daisy, 2003).

All aspects of the biology and interrelatedness of endophytes with their respective hosts is a vastly under investigated and exciting field. Thus, more background information on a given plant species and its microorganismal biology would be exceedingly helpful in directing the search for bioactive products. Currently, no one is quite certain of the role of endophytes in nature and what appears to be their relationship to various host plant species. While some endophytic fungi appear to be ubiquitous (e.g., Fusarium species, Pestalotiopsis species and Xylaria species), one cannot definitively state that endophytes are truly host specific or even systemic within plants any more than one can assume that their associations with plants are chance encounters. Frequently, many endophytes (biotypes) of the same species are isolated from the same plant and only one of the endophytes will produce a highly biologically active compound in culture (Li et al., 1996). A great deal of uncertainty also exists between what an endophyte produces in culture and what it may produce in nature. It does seem apparent that the production of certain bioactive compounds by the endophyte in situ may facilitate the domination of its biological niche within the plant or even provide protection to the plant from harmful invading pathogens. This may be especially true if the bioactive product of the endophyte is unique to it and is not produced by the host. Seemingly, this would more easily facilitate the study of the role of the endophyte and its role in the plant. Furthermore, little information exists relative to the biochemistry and physiology of the interactions of the endophyte with its host plant. It would seem that many factors changing in the host as related to the season and age, environment and location may influence the biology of the endophyte. Indeed, further research at the molecular level must be conducted in the field to study endophyte interactions and ecology. The ecological awareness of the role these organisms play in nature will provide the best clues for targeting particular types of endophytic bioactivity with the greatest potential for bio-prospecting (Strobel and Daisy, 2003).

BIOPHARMACEUTICAL EFFECTS OF ENDOPHYTES

The following section shows effects of bioactive compounds obtained from endophytic microbes and their potential in the pharmaceutical and agrochemical areas.

Antimicrobial effect: Metabolites bearing antibiotic activity can be defined as low-molecular-weight organic natural substances made by microorganisms that are active at low concentrations against other microorganisms (Guo et al., 2008). Endophytes are believed to carry out a resistance mechanism to overcome pathogenic invasion by producing secondary metabolites (Tan and Zou, 2001). So far, studies reported a large number of antimicrobial compounds isolated from endophytes, belonging to several structural classes like alkaloids, peptides, steroids, terpenoids, phenols, quinines and flavonoids (Yu et al., 2010). The discovery of novel antimicrobial metabolites from endophytes is an important alternative to overcome the increasing levels of drug resistance by plant and human pathogens, the insufficient number of effective antibiotics against diverse bacterial species and few new antimicrobial agents in development, probably due to relatively unfavorable returns on investment (Yu et al., 2010; Song, 2008). The antimicrobial compounds can be used not only as drugs by humankind but also as food preservatives in the control of food spoilage and food-borne diseases, a serious concern in the world food chain (Liu et al., 2008).

Many bioactive compounds, including antifungal agents, have been isolated from the genus Xylaria residing in different plant hosts, such as sordaricin with antifungal activity against Candida albicans (Pongcharoen et al., 2008), mellisol and 1,8- dihydroxynaphthol 1-O-aglucopyranoside with activity against herpes simplex virus type 1 (Pittayakhajonwut et al., 2005), multiplolides A and B with activity against Candida albicans (Boonphong et al., 2001). The bioactive compound isolated from the culture extracts of the endophytic fungus Xylaria sp. YX-28 isolated from Ginkgo biloba L. was identified as 7-amino-4-methylcoumarin (Liu et al., 2008). The compound presented broad-spectrum inhibitory activity against several food-borne and food spoilage microorganisms including S. aureus, E. coli, S. typhia, S. typhimurium, S. enteritidis, A. hydrophila, Yersinia sp., V. anguillarum, Shigella sp., V. parahaemolyticus, C. albicans, P. expansum and A. niger, especially to A. hydrophila and was suggested to be used as natural preservative in food (Liu et al., 2008). Another strain F0010 of the endophytic fungus Xylaria sp., from Abies holophylla was characterized as a producer of griseofulvin, a spirobenzofuran antifungal antibiotic agent used for the treatment of human and veterinary animals mycotic diseases (Park et al., 2005). They evaluated and reported high antifungal activity in vivo and in vitro of the endophyte-produced griseofulvin against plant pathogenic fungi, controlling effectively the development of various food crops. Aliphatic compounds, frequently detected in cultures of endophytes, often show biological activities. Four antifungal aliphatic compounds were characterized from stromata of E. typhina on P. pratense (Koshino et al., 1989). Two novel ester metabolites isolated from an endophyte of the eastern larch presented antimicrobial activity. One compound was toxic to spruce budworm (Choristoneura fumiferana Clem.) larvae and the other may serve as potent antibacterial agent against Vibrio salmonicida, Pseudomonas aeruginosa and Staphylococcus aureus (Findlay et al., 1997a). Chaetomugilin A and D with antifungal activities were isolated from an endophytic fungus C. globosum collected from Ginkgo biloba (Qin et al., 2009). Cytosporone B and C were isolated from a mangrove endophytic fungus, *Phomopsis* sp. They inhibited two fungi C. albicans and F. oxysporum with the MIC value ranging from 32 to 64 mg Ml $^{-1}$ (Huang et al., 2008).

Chlorinatedmetabolites such as (·) mycorrhizin A, (+)- cryptosporiopsin isolated from endophytic Pezicula strains were reported as strongly fungicidal and herbicidal agents and to a lesser extent, as algicidal and antibacterial agents (Schulz et al., 1995). Similarly, two other new chlorinated benzophenone derivatives, Pestalachlorides A and B, from the plantendophytic fungus Pestalotiopsis adusta, proven to display significant antifungal activity against three plant pathogenic fungi, Fusarium culmorum, Gibberella zeae and Verticillium albo-atrum (Li et al., 2008a). The production of Hypericin, a naphthodianthrone derivative and Emodin believed to be the main precursor of hypericin by the endophytic fungus isolated from an Indian medicinal plant was reported. Both compounds demonstrated antimicrobial activity against several bacteria and fungi including Staphylococcus aureus sp., aureus, Klebsiella pneumoniae sp., ozaenae, Pseudomonas aeruginosa, Salmonella enterica sp., Enteric and Escherichia coli and fungal organisms Aspergillus niger and Candida albicans (Kusari et al., 2008).

An endophytic Streptomyces sp., from a fern-leaved grevillea (Grevillea pteridifolia) in Australia was described as a promising producer of novel antibiotics, kakadumycin A and echinomycin. Kakadumycin A is structurally related to echinomycin, a quinoxaline antibiotic and presents better bioactivity than echinomycin especially against Grampositive bacteria and impressive activity against the malarial parasite Plasmodium falciparum (Castillo et al., 2003). Another novel endophytic Streptomyces SUK 06 from Thottea grandiflora in Malaysia was reported bioactive secondary metabolites with ethyl acetate have killing activity against Bacillus subtilis, Pseudomonas aeruginosa, Bacillus cereus, Pleisiomonas shigelloides and MRSA. Nevertheless, there were some antifungal activity measured against Fusarium solani, Aspergillus fumigatus, Pythium ultimum, Phytophthora erythroseptica and Geothrichum candidum (Ghadin et al., 2008). More than 50% of endophytic fungi strains residing in Quercus variabilis possessed growth inhibition against at least one pathogenic fungus or bacteria. Cladosporium sp., displaying the most active antifungal activity, was investigated and found to produce a secondary metabolite known as brefeldin A, a lactone with antibiotic activity. Results showed brefeldin A to be more potent than the positive control in antifungal activity (Wang et al., 2007).

Coronamycin, a peptide antibiotic produced by an endophytic fungi Streptomyces sp., isolated Monstera sp., is active against pythiaceous fungi, the human fungal pathogen Cryptococcus neoformans and the malarial parasite, Plasmodium falciparum (Ezra et al., 2004b). Production of lipopeptide pumilacidin, an antifungal compound, by B. pumilus isolated from cassava cultivated by Brazilian Amazon Indian tribes was described for the first time (De Melo et al., 2009). The compounds 2-hexyl-3-methyl-butanodioic acid and cytochalasin D were isolated from the endophytic fungus Xylaria sp., Isolated from Brazilian Cerrado and presented antifungal activity (Cafeu et al., 2005). Two new bioactive metabolites, ethyl 2, 4-dihydroxy-5, 6dimethylbenzoate and phomopsilactone were isolated from an endophytic fungus Phomopsis cassiae from Cassia spectabilis and displayed strong antifungal activity against two phytopathogenic fungi, Cladosporium cladosporioides and C. sphaerospermum (Silva et al., 2005). The polyketide citrinin produced by endophytic fungus Penicillium janthinellum from fruits of Melia azedarach, presented 100% antibacterial activity against Leishmania sp. (Marinho et al., 2005). Among the 12 secondary metabolites produced by the endophytic fungi Aspergillus fumigatus CY018 isolated from the leaf of Cynodon dactylon, asperfumoid, fumigaclavine C, fumitremorgin C, physcion and helvolic acid were shown to inhibit Candida albicans (Liu et al., 2004). Endophyte Verticillium sp., isolated from roots of wild Rehmannia glutinosa produced two compounds 2, 6-Dihydroxy-2methyl-7- (prop-1E-enyl)-1-benzofuran-3(2H)-one, reported for the first time and ergosterol peroxide with clear inhibition of the growth of three pathogens including *Verticillium* sp., (You *et al.*, 2009).

Another fascinating use of antibiotic products from endophytic fungi is the inhibition of viruses. Two novel human cytomegalovirus protease inhibitors, cytonic acids A and B have been isolated from the solid-state fermentation of the endophytic fungus Cytonaema sp., Their structures as p-tridepside isomers were elucidated by mass spectrometry and NMR methods (Guo et al., 2000). An endophytic fungus Pestalotiopsis theae of an unidentified tree on Jianfeng Mountain, China, was capable of producing Pestalotheol C with anti-HIV properties (Li et al., 2008b). It is apparent that the potential for the discovery of compounds, from endophytes, having antiviral activity is in its infancy. The fact, however, that some compounds have been found is promising. The main limitation in compound discovery is probably related to the absence of appropriate antiviral screening systems in most compound discovery programs (Strobel and Daisy, 2003).

Another interesting aspect of this research, newly described endophytic fungus *Muscodor albus* from small limbs of *Cinnamomum zeylanicum* effectively inhibits and kills certain other fungi and bacteria by producing a mixture of volatile compounds (Worapong *et al.*, 2001; Strobel *et al.*, 2001). This mixture mimicked the antibiotic effects of the volatile compounds produced by the fungus. It was also used to gain positive identification of the ingredients of the fungal volatile compounds (Strobel *et al.*, 2001). Each of the five classes of volatile compounds produced by the fungus had some inhibitory effect against the test fungi and bacteria, but none was lethal. The most effective class of inhibitory compounds was the esters, of which isoamyl acetate was the most biologically active. The ecological implications and potential practical benefits of the mycofumigation effects of *M. albus* are very promising given the fact that soil fumigation utilizing methyl bromide. This fungus is just as effective in causing inhibition and death of test microbes in the laboratory as *M. albus* (Worapong *et al.*, 2002). In addition, for the first time, a non-muscodor species, a *Gliocladium* sp., was discovered to be a volatile antibiotic producer.

Another important aspect of this research is inhibition of plant pathogens which is relevant in agriculture field. Many endophytic species produce antibiotic substances (Schulz and Boyle, 2005; Strobel et al., 2002; Wang et al., 2007). Liquid extracts from endophyte cultures have been found to inhibit the growth of several species of plant pathogenic fungi (Liu et al., 2001; Park et al., 2005; Inacio et al., 2006; Kim et al., 2007). If such compounds where produced by endophytes in plants, this could constitute a defense mechanism against fungal pathogens. Experiments where plant protection against pathogenic fungi is observed after the inoculation of plants with endophytes, as well as after the application of endophytic culture filtrates, suggest that the endophyte may produce an antifungal compound or a substance that induces plant defense mechanisms in the plant. This is the case with Chaetomium and Phoma endophytes of wheat, when these fungi were previously inoculated in plants, reduced severity of foliar disease caused by Puccinia and Pyrenophora sp., was observed and, the same protective effect was observed when only endophytic culture filtrates were applied to the plants (Dingle and McGee, 2003; Istifadah and McGee, 2006). Rajendran et al. (2008) reported that the management of Basal Stem Rot disease sixty endophytic, rhizosphere strains was isolated from coconut, other crops and virgin soils. The strains showed high growth promotion were subjected to Ganoderma mycelium inhibition study in vitro. The strains EPC5 and EPC8 were showed high growth promotion and strong inhibition to Ganoderma pathogen.

When a mixture of six species of endophytes frequently isolated from cacao (*Theobroma cacao* L.) trees was used to inoculate leaves of endophyte-free seedlings of this plant species, the severity

of a leaf disease caused by a *Phytophthora* sp., was significantly reduced in endophyte-inoculated leaves. A mechanism of induced plant resistance did not seem to be involved, because differences in disease severity were observed between endophyte-inoculated and non-inoculated leaves of the same plant. In this case, the protection against a pathogen could be the result of direct competition among endophytes already present in leaves and the pathogen (Arnold *et al.*, 2003). For instance, most tissue available for infection may be already occupied, or endophytes may produce zones of inhibition restricting the entry of other fungi.

Endophyte infection may alter plant biochemistry in a way that defense mechanisms against pathogens are induced. *Piriformospora indica* Sav. Verma, Aj. Varma, Rexer, G. Kost and P. Franken are a root endophyte with a wide host range, including several species of cereals and *Arabidopsis*. Barley plants inoculated with this endophyte have shown resistance to a vascular (*Fusarium culmorum* (W.G. Sm.) Sacc.) and a leaf pathogen [*Blumeria graminis* (DC) Speer], in addition to an increase in yield and salt stress tolerance (Waller *et al.*, 2005). The protection against the leaf pathogen appears to be mediated by a mechanism of induced resistance, because in the pathogen-inoculated plants there is a defense response involving the death of host cells.

Some endophytes may be mycoparasites. Acremonium strictum W. Gams is an endophyte which has been frequently isolated from Dactylis glomerata L. and other grasses (Marquez et al., 2007), recently it has been shown that this fungus is a mycoparasite of Helminthosporium solani Durieu and Mont., a potato pathogen (Rivera-Varas et al., 2007). A significant increase in resistance to dollar spot disease, caused by Sclerotinia homoeocarpa F.T. Benn., has been observed in Festuca rubra L. cultivars infected by Epichloë festucae Leuchtm., Schardl and M.R. Siegel. (Clarke et al., 2006). Cultivars of several turfgrass species infected by Epichloe and Neotyphodium endophytes are commercially available at the present time. The efficient vertical transmission of these endophytes has allowed the production of infected seed at a commercial scale. Since Neotyphodium and Epichloe infected cultivars have shown increased resistance to herbivores, plant pathogens and some conditions of abiotic stress, the use of such symbiotic cultivars can result in a reduction in the use of insecticides and fungicides in lawns (Brilman, 2005).

Antioxidant effect: Many antioxidant compounds possess anti-inflammatory, antiatherosclerotic, antitumor, antimutagenic, anticarcinogenic, antibacterial, or antiviral activities in higher or lower level (Owen et al., 2000; Cozma, 2004; Halliwell, 1994; Mitscher et al., 1996; Sala et al., 2002). Natural antioxidants are commonly found in medicinal plants, vegetables and fruits. However, it has been reported that metabolites from endophytes can be a potential source of novel natural antioxidants (Liu et al., 2007) evaluated the antioxidant activity of an endophytic Xylaria sp., isolated from the medicinal plant Ginkgo biloba. The results collected indicated that the methanol extract exhibited strong antioxidant capacity due to the presence of phenolics and flavonoids compounds among 41 identified compounds. Huang and coworkers investigated the antioxidant capacities of endophytic fungal cultures of medicinal Chinese plants and its correlation to their total phenolic contents. They suggested that the phenolic content were the major antioxidant constituents of the endophytes (Huang et al., 2007). Pestacin and isopestacin, 1, 3-dihydro isobenzofurans, were obtained from the endophytic fungus Pestalotiopsis microspora isolated from a plant growing in the Papua New Guinea, Terminalia morobensis (Harper et al., 2003; Strobel et al., 2002). Besides antioxidant activity, pestacin and isopestacin also presented antimycotic and antifungal activities, respectively. Pestacin is believed to have antioxidant activity 11 times greater than Trolox, a vitamin E derivative, primarily via cleavage of an unusually reactive C-H bond and to a lesser extent, O-H abstraction (Harper *et al.*, 2003). Isopestacin possess antioxidant activity by scavenging both superoxide and hydroxy free radicals in solution, added to the fact that isopestacin is structurally similar to the flavonoids (Strobel *et al.*, 2002).

Liu et al. (2009) reported for the first time, the capacity of endophytic microorganisms to produce polysaccharides with antioxidant. The bacterium endophyte Paenibacillus polymyxa isolated from the root tissue of Stemona japonica Miquel, a traditional Chinese medicine, produced exopolysaccharides (EPS) that demonstrated strong scavenging activities on superoxide and hydroxyl radicals. Graphislactone A, a phenolic metabolite isolated from the endophytic fungus Cephalosporium sp., IFB-E001 residing in Trachelospermum jasminoides, demonstrated to have free radical-scavenging and antioxidant activities in vitro stronger than the standards, Butylated Hydroxytoluene (BHT) and ascorbic acid, coassayed in the study (Song et al., 2005).

Anticancer effect: Some evidences that bioactive compounds produced by endophytes could be alternative approaches for discovery of novel drugs, since many natural products from plants, microorganisms and marine sources were identified as anticancer agents (Firakova et al., 2007). The anticancer properties of several secondary metabolites from endophytes have been investigated recently. Following, some examples of the potential of endophytes on the production of anticancer agents are cited.

The diterpenoid Taxol (also known in the literature as paclitaxel) have generated more attention and interest than any other new drug since its discovery, possibly due to its unique mode of action compared to other anticancer agents (Gangadevi and Muthumary, 2008; Firakova et al., 2007). This compound interferes with the multiplication of cancer cells, reducing or interrupting their growth and spreading. FDA (Food and Drug Administration) has approved Taxol for the treatment of advanced breast cancer, lung cancer and refractory ovarian cancer (Cremasco et al., 2009). Taxol was firstly isolated from the bark of trees belonging to Taxus family (Taxus brevifolia), its most common source (Wani et al., 1971). Nevertheless, these trees are rare, slow growing and produce small amount of Taxol, which explain its high price in the market when obtained by this natural source (Gangadevi and Muthumary, 2008). Besides, in the context of environmental degradation, the use of plant source as unique option have limited the supply of this drug due to the destructive collection of yew trees (Guo et al., 2008). Several reports about Taxol anticancer properties were published since its discovery (Lu et al., 2007; Kakolyris et al., 2006; Peltier et al., 2006), as well as other sources for production of Taxol have been investigated in the last decade. The isolation of Taxol-producing endophyte Taxomyces andreanae has provided an alternative approach to obtain a cheaper and more available product via microorganism fermentation (Stierle et al., 1993). After that, Taxol has also been found in a number of different genera of fungal endophytes either associated or not to yews, such as Taxodium distichum (Li et al., 1996), Wollemia nobilis (Strobel et al., 1997), Phyllosticta spinarum (Kumaran et al., 2008), Bartalinia robillardoides (Gangadevi and Muthumary, 2008), Pestalotiopsis terminaliae (Gangadevi and Muthumary, 2009), Botryodiplodia theobromae (Pandi et al., 2010).

Another important anticancer compound is the alkaloid Camptothecin, a potent antineoplastic agent which was firstly isolated from the wood of *Camptotheca acuminata* Decaisne (Nyssaceae) in China (Wall et al., 1966). Camptothecin and 10-hydroxycamptothecin are two important precursors for the synthesis of the clinically useful anticancer drugs, topotecan and irinotecan (Uma et al., 2008). Although it's potential use in medical treatments, the unmodified Camptothecin suffers from drawbacks that compromise its applications due to very low solubility in aqueous media

and high toxicity (Li et al., 2006; Kehrer et al., 2001). On the other hand, some Camptothecin derivatives retain the medicinal properties and can show other benefits without causing over drawbacks in some cases (Kusari et al., 2009a; Jew et al., 1999). Therefore, it is desirable to develop strategies for isolation, mixture separation and production of Camptothecin and its analogues from novel endophytic fungal sources. The anticancer properties of the microbial products Camptothecin and two analogues (9-methoxycamptothecin and 10-hydroxycamptothecin) were already reported. The products were obtained from the endophytic fungi Fusarium solani isolated from Camptotheca acuminate (Kusari et al., 2009b). Several reports have described other Camptothecin producing endophytes (Shweta et al., 2010; Liu et al., 2010; Rehman et al., 2008; Amna et al., 2006; Puri et al., 2005). Since then, endophytes have been included inmany studies purposing new approaches for drug discovery. Ergoflavin, a dimeric xanthene linked in position 2, belongs to the compound class called ergochromes and was described as a novel anticancer agent isolated from an endophytic fungi growing on the leaves of an Indian medicinal plant Mimusops elengi (Sapotaceae) (Deshmukh et al., 2009). Secalonic acid D, a mycotoxin also belonging to ergochrome class, is known to have potent anticancer activities. It was isolated from the mangrove endophytic fungus and observed high cytotoxicity on HL60 and K562 cells by inducing leukemia cell apoptosis (Zhang et al., 2009).

Phenylpropanoids have attracted much interest for medicinal use as anticancer, antioxidant, antimicrobial, anti-inflammatory and immunosuppressive properties (Korkina, 2007). Despite the phenylpropanoids belong to the largest group of secondary metabolites produced by plants, reports showed the production of such compounds by endophytes. The endophytic Penicillium brasilianum, found in root bark of Melia azedarach, promoted the biosynthesis of phenylpropanoid amides (Fill et al., 2010). Likewise, two monolignol glucosides, coniferin and syringin, are produced not only by the host plant, but were also recognized by the endophytic Xylariaceae species as chemical signals during the establishment of fungus-plant interactions (Chapela et al., 1991). Koshino and coworkers characterized two phenylpropanoids and lignan from stromata of Epichloe typhina on Phleum pretense (Koshino et al., 1988). Lignans are other kinds of anticancer agents originated as secondary metabolites through the shikimic acid pathway and display different biological activities that make them interesting in several lines of research (Gordaliza et al., 2004). Although their molecular backbone consists only of two phenylpropane units (C6-C3), lignans show enormous structural and biological diversity, especially in cancer chemotherapy (Korkina, 2007). Podophyllotoxin and analogs are clinically relevant mainly due to their cytotoxicity and antiviral activities and are valued as the precursor to useful anticancer drugs like etoposide, teniposide and etopophos phosphate (Kusari et al., 2009a; Kour et al., 2008). The aryl tetralin lignans, such as podophyllotoxin, are naturally synthesized by Podophyllum sp., however, alternative sources have been searched to avoid endangered plant. Another study showed a novel fungal endophyte, Trametes hirsute, that produces podophyllotoxin and other related anyl tetralin lignans with potent anticancer and properties (Puri et al., 2006). Novel microbial sources of Podophyllotoxin were reported from the endophytic fungi Aspergillus fumigatus Fresenius isolated from Juniperus communis L. Horstmann (Kusari et al., 2009b), Phialocephala fortinii isolated from Podophyllum peltatum (Eyberger et al., 2006) and Fusarium oxysporum from Juniperus recurva (Kour et al., 2008).

Wagenaar et al. (2000) reported identification of three novel cytochalasins, bearing antitumor activity from the endophyte *Rhinocladiella* species. Extensive experiments identified these new compounds as 22-oxa-12-cytochalasins. Torreyanic acid is an unusual dimeric quinone isolated from

the endophytic fungus Pestalotiopsis microspora from T. taxifolia (Florida torreya) and was proven to have selective cytotoxicity 5 to 10 times more potent in cell lines that are sensitive to protein kinase C agonists and causes cell death by apoptosis (Lee et al., 1996). Gliocladicillins A and B were reported as effective antitumor agents in vitro and in vivo, since they induced tumor cell apoptosis and showed significant inhibition on proliferation of melanoma B16 cells implanted into immunodeficient mice (Chen et al., 2009). Crude Extracts of Alternaria alternata, an endophytic fungus isolated from Coffea Arabica L., displayed moderate cytotoxic activity towards HeLa cells in vitro, when compared to the dimethyl sulfoxide (DMSO) treated cells (Fernandes et al., 2009). The investigation of endophytic actinomycetes associated with pharmaceutical plants in rainforest reported 41 microorganisms from the genus Streptomyces displayed significant antitumor activity against HL-60 cells, A549 cells, BEL-7404 cells and P388D1 cells (Li et al., 2008c). The screening of endophytic fungi isolated from pharmaceutical plants in China showed that 13.4% endophytes were cytotoxic on HL-60 cells and 6.4% on KB cells (Huang et al., 2001). Finally, other compounds with anticancer properties isolated from endophytic microbes were reported such as cytoskyrins, phomoxanthones A and B, photinides A-F, rubrofus arin B and (+)- epiepoxydon (Brady et al., 2000; Isaka et al., 2004; Ding et al., 2009; Song et al., 2004; Klemke et al., 2004).

Insecticidal effect: Several endophytes are known to have anti-insect properties. Nodulisporic acids, novel indole diterpenes that exhibit potent insecticidal properties against the larvae of the blowfly, work by activating insect glutamate-gated chloride channels. The first nodulisporic compounds were isolated from an endophyte, a Nodulisporium sp., from the plant Bontia daphnoides. This discovery has since resulted in an intensive search for more Nodulisporium sp., or other producers of more-potent nodulisporic acid analogues (Demain, 2000). Insect toxins have also been isolated from an unidentified endophytic fungus from wintergreen (Gaultheria procumbers). The two new compounds, 5-hydroxy-2-(1'-hydroxy-5'-methyl- 4'-hexenyl) benzofuran and 5-hydroxy-2-(1-oxo-5-methyl-4- hexenyl) benzofuran, both show toxicity to spruce budworm and the latter is also toxic to the larvae of spruce budworm (Findlay et al., 1997b). Another endophytic fungus, Muscodor vitigenus, from a liana (Paullina paullinioides), yields naphthalene as its major product. Naphthalene, the active ingredient in common mothballs, is a widely exploited insect repellant. M. vitigenus shows promising preliminary results as an insect deterrent and has exhibited potent insect repellency against the wheat stem sawfly (Cephus cinctus) (Daisy et al., 2002a, b). As the world becomes wary of ecological damage done by synthetic insecticides, endophytic research continues for the discovery of powerful, selective and safe alternatives.

Antidiabetic effect: A nonpeptidal fungal metabolite (L-783,281) was isolated from an endophytic fungus (*Pseudomassaria* sp.,) collected from an African rainforest near Kinshasa in the Democratic Republic of the Congo (Zhang *et al.*, 1999). This compound acts as insulin mimetic and unlike insulin, is not destroyed in the digestive tract and may be given orally. Oral administration of L-783,281 to two mouse models of diabetes resulted in significant lowering of blood glucose levels. These results may lead to new therapies for diabetes (Zhang *et al.*, 1999).

Immunosuppressive effect: The endophytic fungus Fusarium subglutinans, isolated from T. wilfordii, produces the immunosuppressive but noncytotoxic diterpene pyrones subglutinol A and B (Lee et al., 1995). Subglutinol A and B are equipotent in the mixed lymphocyte reaction

assay and thymocyte proliferation assay, with a 50% inhibitory concentration of 0.1 µM In the same assay systems, the famed immunosuppressant drug cyclosporine is roughly as potent in the mixed lymphocyte reaction assay and 104 more potent in the thymocyte proliferation assay. Still, the lack of toxicity associated with subglutinols A and B suggests that they should be explored in greater detail (Lee et al., 1995). The Microbiology Department at Sandoz Ltd. developed a computer-aided evaluation program to screen and evaluate fungi for bioactivity. The program can recognize and eliminate from study common fungi producing known compounds and thereby direct attention to the evaluation of rare samples, which are more likely to produce metabolites with novel bioactivity. This approach resulted in the discovery of the fungus Tolypocladium inflatum, from which cyclosporine, a hugely beneficial immunosuppressant, was isolated (Borel and Kis, 1991). This example perfectly depicts the current aim of many investigators to seek out rare endophytes from interesting and uncommon hosts and environments (Strobel and Daisy, 2003).

BIOACTIVE COMPOUNDS PRODUCTION BY BIOTRANSFORMATION PROCESS

Biotransformation can be defined as the use of biological systems to produce chemical changes on compounds that are not their natural substrates (Borges et al., 2008). The microbial growth, sustenance and reproduction depends on the availability of a suitable form of reduced carbon source, used as chemical energy, which under normal conditions of culture broth are the common sugars. Nevertheless, microorganisms are believed to have no limit to adapt to new environments and to metabolize various foreign substrates to carbon and nitrogen sources (Doble et al., 2004). A molecule can be modified by transforming functional groups, with or without degradation of carbon skeleton. Such modifications result in the formation of novel and useful products not easily prepared by chemical methods (Borges et al., 2009). The biotransformation process provides a number of advantages over chemical synthesis. The process can be carried out under mild conditions like ambient temperature and without the need of high pressure and extreme conditions, thus reducing undesired byproduct, energy needs and cost (Suresh et al., 2006). The region-and stereo-selectivity of the process allows the production of enantiomerically pure compounds, eliminating the need for complicated separation and purification steps (Borges et al., 2008). Besides, the reactions occur under ecologically acceptable conditions, with lower emission of industrial resides and production of biodegradable resides and products, thus reducing the environmental problems (Bicas et al., 2009; Aleu and Collado, 2001).

Finally, the products obtained by biotransformation process can be labeled as natural. On the other hand, chemical synthesis often result in environmentally unfriendly production processes and lacks substrate selectivity, possibly causing the formation of undesirable reaction mixtures and reducing process efficiency and increasing downstream cost (Longo and Sanroman, 2006). Therefore, biotransformation is a useful method for production of novel compounds; enhancement in the productivity of a desired compound; overcoming the problems associated with chemical analysis; leading to basic information to elucidate the biosynthetic pathway (Suresh et al., 2006). For this reason, biotransformation using microbial cultures and/or their enzymatic systems alone has received increasing attention as a method for the conversion of lipids, monoterpenes, diterpenes, steroids, triterpenes, alkaloids, lignans and some synthetic chemicals, carrying out stereospecific and stereoselective reactions for the production of novel bioactive molecules with some potential for pharmaceutical and food industries (Borges et al., 2009; Figueiredo et al., 1996; Pimentel et al., 2010).

CONCLUSION

Endophytes are a poorly investigated group of microorganisms that represent an abundant and dependable source of bioactive and chemically novel compounds with potential for exploitation in a wide variety of medical, agricultural and industrial areas. The mechanisms through which endophytes exist and respond to their surroundings must be better understood in order to be more predictive about which higher plants to seek, study and spend time isolating micro floral components. This may facilitate the product discovery processes. Although work on the utilization of this vast resource of poorly understood microorganisms has just begun, it has al ready become obvious that an enormous potential for organism, product and utilitarian discovery in this field holds exciting promise. This is witnessed by the discovery of a wide range of products and microorganisms that already hold inkling for future prospects as mentioned in this report. However, the application of microorganisms by the food and pharmaceutical industries to obtain compounds of interest is still modest, considering the great availability of useful microorganisms.

ACKNOWLEDGMENT

The authors acknowledges to Management of Malankara Catholic College (MCC) for their all efficient support for our research. We thank Research Team of Interdisciplinary Research Centre (MCC), for critically reviewing the manuscript and helpful discussions. We express appreciation to Mr. S. Murugan providing financial support for some of the work reviewed in this report.

REFERENCES

- Aleu, J. and I.G. Collado, 2001. Biotransformations by Botrytis species. J. Mol. Catalysis, 13: 77-93.
 Amna, T., S.C. Puri and V. Verma, 2006. Bioreactor studies on the endophytic fungus entrophospora infrequens for the production of an anticancer alkaloid camptothecin. Can. J. Microbiol., 52: 189-196.
- Andrew, J.H. and S.S. Hirano, 1991. Microbial Ecology of Leaves. Brock/Spinger Series, Springer Verlag, London.
- Arnold, A.E., L.C. Mejia, D. Kyllo, E.I. Rojas, Z. Maynard, N. Robbins and E.A. Herre, 2003. Fungal endophytes limit pathogen damage in a tropical tree. PNAS USA., 100: 15649-15654.
- Bacon, C.W. and J.F. White, 2000. Microbial Endophytes. Marcel Dekker, New York, pp: 341-388. Berger, R.G., 2009. Biotechnology of flavours-the next generation. Biotechnol. Lett., 31: 1651-1659.
- Bicas, J.L., A.P. Dionisio and G.M. Pastore, 2009. Bio-oxidation of terpenes: An approach for the flavor industry. Chem. Rev., 109: 4518-4531.
- Bicas, J.L., F.F.C. Barros, R. Wagner, H.T. Godoy and G.M. Pastore, 2008. Optimization of R-(+)-a-terpineol production by the *Biotransformation* of R-(+)-limonene. J. Industrial Microbiol. Biotechnol., 35: 1061-1070.
- Boonphong, S., P. Kittakoop, M. Isaka, D. Pittayakhajonwut, M. Tanticharoen and Y. Thebtaranonth, 2001. Multiplolides A and B, new antifungal 10-membered lactones from *Xylaria* multiplex. J. Nat. Prod., 64: 965-967.
- Borel, J.F. and Z.L. Kis, 1991. The discovery and development of *Cyclosporine*. Transplant. Proc., 23: 1867-1874.
- Borges, K.B., W.D.S. Borges, M.T. Pupo and P.S. Bonato, 2007. Endophytic fungi as models for the *Stereoselective biotransformation* of thioridazine. Applied Microbiol. Biotechnol., 77: 669-674.
- Borges, K.B., W.D.S. Borges, M.T. Pupo and P.S. Bonato, 2008. Stereoselective analysis of thioridazine-2-sulfoxide and thioridazine-5-sulfoxide: An investigation of rac-thioridazine *Biotransformation* by some endophytic fungi. J. Pharmac. Biomed. Anal., 46: 945-952.

- Borges, K.B., W.S. Borges, R. Duran-Paton, M.T. Pupo, P.S. Bonato and I.G. Collado, 2009. Stereoselective *Biotransformations* using fungi as biocatalysts. Tetrahedron Asymmetry, 20: 385-397.
- Brady, S.F., M.P. Singh, J.E. Janso and J. Clardy, 2000. Cytoskyrins A and B, new BIA active *Bisanthraquinones* isolated from an endophytic fungus. Org. Lett., 2: 4047-4049.
- Brilman, L.A., 2005. Endophytes in Turfgrass Cultivars. In: Neotyphodium in Cool Season Grasses, Roberts, C.A., C.P. West and D.E. Spiers (Eds.). Blackwell Publishing, Iowa, USA., pp: 341-349.
- Cafeu, M.C., G.H. Silva, H.L. Teles, V.S. da Bolzani, A.R. Araujo, M.C.M. Young and L.H. Pfenning, 2005. Antifungal compounds of *Xylaria* sp., an endophytic fungus isolated from Palicourea marcgravii (Rubiaceae). Quimica Nova, 28: 991-995.
- Castillo, U., J.K. Harper, G.A. Strobel, J. Sears and K. Alesi *et al.*, 2003. Kakadumycins, novel antibiotics from Streptomyces sp. NRRL 30566, an endophyte of *Grevillea pteridifolia*. FEMS Microbiol. Lett., 224: 183-190.
- Chapela, I.H., O. Petrini and L. Hagmann, 1991. Monolignol glucosides as specific recognitionmessengers in fungus-plant symbioses. Physiological Mol. Plant Pathol., 39: 289-298.
- Chen, Y., H. Guo, Z. Du, X.Z. Liu, Y. Che and X. Ye, 2009. Ecology-based screen identifies new metabolites from a Cordyceps-colonizing fungus as cancer cell proliferation inhibitors and apoptosis inducers. Cell Proliferation, 42: 838-847.
- Clarke, B.B., Jr. J.F. White, H. Hurley, M.S. Torres, S. Sun and D.R. Huff, 2006. Endophyte-mediated suppression of dollar spot disease in fine fescues. Plant Dis., 90: 994-998.
- Cozma, L.S., 2004. The role of antioxidant therapy in cardiovascular disease. Curr. Opin. Lipidol., 15: 369-371.
- Cragg, G.M. and D.J. Newman, 2005. Biodiversity: A continuing source of novel drug leads. Pure Appl. Chem., 77: 7-24.
- Cragg, M.G. and D.J. Newman, 2007. Natural products as sources of new drugs over the last 25 years. J. Nat. Prod., 70: 461-477.
- Cremasco, M.A., B.J. Hritzko and N.H. Linda Wang, 2009. Experimental purification of paclitaxel from a complex mixture of taxanes using a simulated moving bed. Brazil. J. Chem. Eng., 26: 207-218.
- Daisy, B., G. Strobel, D. Ezra, U. Castillo, G. Baird and W.M. Hess, 2002a. Muscodor vitigenus anam. sp. nov., an endophyte from *Paullinia* paullinioides. Mycotaxon, 84: 39-50.
- Daisy, B.H., G.A. Strobel, U. Castillo, D. Ezra, J. Sears, D. Weaver and J.B. Runyon, 2002b. Naphthalene, an insect repellent, is produced by Muscodor vitigenus, a novel endophytic fungus. Microbiology, 148: 3737-3741.
- De Melo, F.M.P., M.F. Fiore, L.A.B. de Moraes, M.F. Silva-Stenico and S. Scramin *et al.*, 2009. Antifungal compound produced by the cassava endophyte *Bacillus pumilus* MAIIIM4A. Sci. Agricola, 66: 583-592.
- Demain, A.L., 2000. Microbial Natural Products: A Past with a Future. In: Biodiversity: New Leads for Pharmaceutical and Agrochemical Industries, Wrigley, S.K., M.A. Hayes, R. Thomas, E.J.T. Chrystal and N. Nicholson (Eds.). The Royal Society of Chemistry, Cambridge, United Kingdom, pp: 3-16.
- Deshmukh, S.K., P.D. Mishra and A. Kulkarni-Almeida, 2009. Anti-inflammatory and anticancer activity of ergoflavin isolated from an endophytic fungus. Chem. Biodiversity, 6: 784-789.
- Ding, G., Z. Zheng, S. Liu, H. Zhang, L. Guo and Y. Che, 2009. Photinides A-F, cytotoxic benzofuranone-derived γ-lactones from the plant endophytic fungus Pestalotiopsis photiniae. J. Nat. Prod., 73: 942-945.

- Dingle, J. and P.A. Mcgee, 2003. Some endophytic fungi reduce the density of pustules of Puccinia recondita f.sp. tritici in wheat. Mycol. Res., 107: 310-316.
- Doble, M., A.K. Kruthiventi and V.G. Gaikar, 2004. Biotransformations and Bioprocesses. Marcel Dekker, New York, NY, USA.
- Eyberger, A.L., R. Dondapati and J.R. Porter, 2006. Endophyte fungal isolates from Podophyllum peltatum produce podophyllotoxin. J. Nat. Prod., 69: 1121-1124.
- Ezra, D., U.F. Castillo, G.A. Strobel, W.M. Hess and H. Porter *et al.*, 2004a. Coronamycins, peptide antibiotics produced by a verticillate Streptomyces sp. (MSU-2110) endophytic on Monstera sp., Microbiol., 150: 785-793.
- Ezra, D., W.M. Hess and G.A. Strobel, 2004b. New endophytic isolates of Muscodor albus, a volatile-antibiotic-producing fungus. Microbiol., 150: 2023-2031.
- Fernandes, M.D.R.V., T.A.C. Silva, L.H. Pfenning, D.L.M. Aparecida and D.C.C. Miguel *et al.*, 2009. Biological activities of the fermentation extract of the endophytic fungus Alternaria alternata isolated from *Coffea arabica* L. Brazilian J. Pharmac. Sci., 45: 677-685.
- Figueiredo, A.C., M.J. Almendra, J.G. Barroso and J.J.C. Scheffer, 1996. Biotransformation of monoterpenes and sesquiterpenes by cell suspension cultures of *Achillea millefolium* L. ssp. Millefolium. Biotechnol. Lett., 18: 663-668.
- Fill, T.P., B.F. da Silva and E. Rodrigues-Fo, 2010. Biosynthesis of phenylpropanoid amides by an endophytic Penicillium brasilianum found in root bark of Melia azedarach. J. Microbiol. Biotechnol., 20: 622-629.
- Findlay, J.A., G. Li and J.A. Johnson, 1997a. Bioactive compounds from an endophytic fungus from eastern larch (Larix laricina) needles. Can. J. Chem., 75: 716-719.
- Findlay, J.A., S. Bethelezi, G. Li and M. Sevek, 1997b. Insect toxins from an endophyte fungus from wintergreen. J. Nat. Prod., 60: 1214-1215.
- Firakova, S., M. Sturdikova and M. Muckova, 2007. Bio-active secondary metabolites produced by microorganisms associated with plants. Biologia, 62: 251-257.
- Gangadevi, V. and J. Muthumary, 2008. Taxol, an anticancer drug produced by an endophytic fungus Bartalinia robillardoides Tassi, isolated from a medicinal plant, Aegle marmelos Correa ex Roxb. World J. Microbiol. Biotechnol., 24: 717-724.
- Gangadevi, V. and J. Muthumary, 2009. Taxol production by Pestalotiopsis terminaliae, an endophytic fungus of Terminalia arjuna (arjun tree). Biotechnol. Applied Biochem., 52: 9-15.
- Ghadin, N., N.M. Zin, V. Sabaratnam, N. Badya, D.F. Basri, H.H. Lian and N.M. Sidik, 2008. Isolation and characterization of a novel endophytic streptomyces SUK 06 with antimicrobial activity from malaysian plant. Asian J. Plant Sci., 7: 189-194.
- Gordaliza, M., P.A. Garcia, J.M. Miguel Del Corral, M.A. Castro and M.A. Gomez Zurita, 2004. Podophyllotoxin: Distribution, sources, applications and new cytotoxic derivatives. Toxicon, 44: 441-459.
- Gunatilaka, A.A.L., 2006. Natural products from plant-associated microorganisms: Distribution, structural diversity, bioactivity and implication of their occurrence. J. Nat. Prod., 69: 509-526.
- Guo, B., J. Dai, S. Ng, Y. Huang, C. Leong, W. Ong and B.K. Carte, 2000. Cytonic acids A and B: novel tridepside inhibitors of hCMV protease from the endophytic fungus Cytonaema species. J. Nat. Prod., 63: 602-604.
- Guo, B., Y. Wang, X. Sun and K. Tang, 2008. Bio-active natural products from endophytes: A review. Applied Biochem. Microbiol., 44: 136-142.

- Halliwell, B., 1994. Free radicals, antioxidants and human disease: Curiosity, cause, or consequence. Lancet, 344: 721-724.
- Harper, J.K., A.M. Arif, E.J. Ford, G.A. Strobel and D.P. Tomer *et al.*, 2003. Pestacin: A 1,3-dihydro isobenzofuran from Pestalotiopsis microspora possessing antioxidant and antimycotic activities. Tetrahedron, 59: 2471-2476.
- Huang, Y., J. Wang, G. Li, Z. Zheng and W. Su, 2001. Antitumor and antifungal activities in endophytic fungi isolated from pharmaceutical plants Taxus mairei, Cephalataxus fortunei and Torreya grandis. FEMS Immunol. Med. Microbiol., 31: 163-167.
- Huang, W.Y., Y.Z. Cai, J. Xing, H. Corke and M. Sun, 2007. A potential antioxidant resource: Endophytic fungi from medicinal plants. Econ. Bot., 61: 14-30.
- Huang, Z., X. Cai, C. Shao, Z. She and X. Xia *et al.*, 2008. Chemistry and weak antimicrobial activities of phomopsins produced by mangrove endophytic fungus Phomopsis sp., ZSU-H76. Phytochemistry, 69: 1604-1608.
- Inacio, M.L., G.H. Silva, H.L. Teles, H.C. Trevisan and A.J. Cavalheiro *et al.*, 2006. Antifungal metabolites from Colletotrichum gloeosporioides, an endophytic fungus in Cryptocarya mandioccana Nees (Lauraceae). Bioch. Syst. Ecol., 34: 822-824.
- Isaka, M., A. Jaturapat, K. Rukseree, K. Danwisetkanjana, M. Tanticharoen and Y. Thebtaranonth, 2004. Phomoxanthones A and B, novel xanthone dimers from the endophytic fungus Phomopsis species. J. Nat. Prod., 64: 1015-1018.
- Istifadah, N. and P.A. McGee, 2006. Endophytic Chaetomium globosum reduces development of tan spot in wheat caused by Pyrenophora tritici-repentis. Aust. Plant Path., 35: 411-418.
- Jew, S.S., H.J. Kim, M.G. Kim, E.Y. Roh and C.I. Hong et al., 1999. Synthesis and In vitro cytotoxicity of hexacyclic Camptothecin analogues. Bioorganic Med. Chem. Lett., 9: 3203-3206.
- Joseph, B. and R.M. Priya, 2010. Preliminary phytochemicals of *Psidium guajava* L. leaf of methanol extract and its cytotoxic study on HeLa cell lines. Inventi Rapid Ethnopharmacol., Vol. 1.
- Joseph, B., R.M. Priya P.A.M. Helen and S. Sujatha, 2010. Bio-active compounds in essential oil and its effects of antimicrobial, cytotoxic activity from the *Psidium guajava* (L.) Leaf. J. Adv. Biotechnol., 9: 10-14.
- Joseph, B. and R.M. Priya, 2011. Phytochemical and biopharmaceutical aspects of *Psidium guajava* (L.) essential oil: A review. Res. J. Med. Plant, 5: 432-442.
- Kakolyris, S., A. Agelidou, N. Androulakis, A. Geroyianni and V. Georgoulias *et al.*, 2006. Cisplatin plus etoposide chemotherapy followed by thoracic irradiation and paclitaxel plus cisplatin consolidation therapy for patients with limited stage small cell lung carcinoma. Lung Cancer, 53: 59-65.
- Kehrer, D.F.S., O. Soepenberg, W.J. Loos, J. Verweij and A. Sparreboom, 2001. Modulation of camptothecin analogs in the treatment of cancer: A review. Anti-Cancer Drugs, 12: 89-105.
- Kim, H.Y., G.J. Choi, H.B. Lee, S.W. Lee and H.K. Kim *et al.*, 2007. Some fungal endophytes from vegetable crops and their anti-oomycete activities against tomato late blight. Lett. Appl. Microbiol., 44: 332-337.
- Klemke, C., S. Kehraus, A.D. Wright and G.M. Konig, 2004. New secondary metabolites from the marine endophytic fungus Apiosporamontagnei. J. Nat. Prod., 67: 1058-1063.
- Korkina, L.G., 2007. Phenylpropanoids as naturally occurring antioxidants: From plant defense to human health. Cell Mol. Biol., 53: 15-25.
- Koshino, H., S.I. Terada and T. Yoshihara, 1988. Three phenolic acid derivatives from stromata of Epichloe typhina on Phleum pretense. Phytochemistry, 29: 1333-1338.

- Koshino, H., T. Yoshihara, S. Sakamura, T. Shimanuki, T. Sato and A. Tajimi, 1989. Novel C-11 epoxy fatty acid from stromata of Epichloe typhina on Phleum pretense. Agric. Biol. Chem., 53: 2527-2528.
- Kour, A., A.S. Shawl and S. Rehman, R.K. Khajuria and V. Verma *et al.*, 2008. Isolation and identification of an endophytic strain of Fusarium oxysporum producing podophyllotoxin from Juniperus recurva. World J. Microbiol. Biotechnol., 24: 1115-1121.
- Krings, U., B. Hardebusch, D. Albert, R.G. Berger, M. Marostica Jr and G.M. Pastore, 2006. Odor-active alcohols from the fungal transformation of a-farnesene. J. Agric. Food Chem., 54: 9079-9084.
- Kumar, S. and A. Sagar, 2007. Microbial associates of Hippophae rhamnoides (Seabuckthorn) Plant Pathol. J., 6: 299-305.
- Kumaran, R.S., J. Muthumary and B.K. Hur, 2008. Production of taxol from, an endophytic fungus of Cupressus sp. Eng. Life Sci., 8: 438-446.
- Kusari, S., M. Lamshoft, S Zuhlke and M. Spiteller 2008. An endophytic fungus from Hypericum perforatum that produces hypericin. J. Nat. Prod., 71: 1059-1063.
- Kusari, S., M. Lamshoft and M. Spiteller, 2009a. Aspergillus fumigatus Fresenius, an endophytic fungus from *Juniperus communis* L. Horstmann as a novel source of the anticancer pro-drug deoxypodophyllotoxin. J. Applied Microbiol., 107: 1019-1030.
- Kusari, S., S. Zuhlke and M. Spiteller, 2009b. An endophytic fungus from Camptotheca acuminata that produces camptothecin and analogues. J. Nat. Prod., 72: 2-7.
- Lam, K.S., 2007. New aspects of natural products in drug discovery. Trends Microbiol., 15: 279-289.
 Lee, J., E. Lobkovsky, N.B. Pliam, G.A. Strobel and J. Clardy, 1995. Subglutinols A and B: immunosuppressive compounds from the endophytic fungus Fusarium subglutinans. J. Org. Chem., 60: 7076-7077.
- Lee, J.C., G.A. Strobel, E. Lobkovsky and J.C. Clardy, 1996. Torreyanic acid: A selectively cytotoxic quinone dimer from the endophytic fungus Pestalotiopsis microspora. J. Org. Chem., 61: 3232-3233.
- Li, J.Y., G. Strobel, R. Sidhu, W.M. Hess and E.J. Ford, 1996. Endophytic taxol-producing fungi from bald cypress, Taxodium distichum. Microbiology, 142: 2232-2236.
- Li, E., L. Jiang, L. Guo, H. Zhang and Y. Che, 2008a. Pestalachlorides A-C, antifungal metabolites from the plant endophytic fungus Pestalotiopsis adusta. Bioorganic Med. Chem., 16: 7894-7899.
- Li, E., R. Tian, S. Liu, X. Chen, L. Guo and Y. Che, 2008b. Pestalotheols A-D, bioactive metabolites from the plant endophytic fungus Pestalotiopsis theae. J. Nat. Prod., 71: 664-668.
- Li, J., G.Z. Zhao, H.H. Chen, H.B. Wang and S. Qin *et al.*, 2008c. Antitumour and antimicrobial activities of endophytic streptomycetes from pharmaceutical plants in rainforest. Lett. Applied Microbiol., 47: 574-580.
- Li, Q.Y., Y.G. Zu, R.Z. Shi and L.P. Yao, 2006. Review camptothecin: Current perspective. Curr. Med. Chem., 23: 2021-2039.
- Liu, C.H., W.X. Zou, H. Lu and R.X. Tan, 2001. Antifungal activity of Artemisia annua endophyte cultures against phytopathogenic fungi. Biotechnology, 88: 277-282.
- Liu, J., J. Luo, H. Ye, Y. Sun, Z. Lu and X. Zeng, 2009. Production, characterization and antioxidant activities in vitro of exopolysaccharides from endophytic bacterium Paenibacillus polymyxa EJS-3. Carbohydrate Polymers, 78: 275-281.
- Liu, J.Y., Y.C. Song, Z. Zhang, L. Wang, Z.J. Guo, W.X. Zou and R.X. Tan, 2004. Aspergillus fumigatus CY018, an endophytic fungus in Cynodon dactylon as a versatile producer of new and bioactive metabolites. J. Biotechnol., 114: 279-287.

- Liu, K., X. Ding, B. Deng and W. Chen, 2010. 10- Hydroxycamptothecin produced by a new endophytic Xylaria sp., M20, from Camptotheca acuminate. Biotechnol. Lett., 32: 689-693.
- Liu, X., M. Dong, X. Chen, M. Jiang, X. Lv and G. Yan, 2007. Antioxidant activity and phenolics of an endophytic Xylaria sp. from Ginkgo biloba. Food Chem., 105: 548-554.
- Liu, X., M. Dong, X. Chen, M. Jiang, X. Lv and J. Zhou, 2008. Antimicrobial activity of an endophytic Xylaria sp.YX-28 and identification of its antimicrobial compound 7-amino-4-methylcoumarin. Applied Microbiol. Biotechnol., 78: 241-247.
- Longo, M.A. and M.A. Sanroman, 2006. Production of food aroma compounds: Microbial and enzymatic methodologies. Food Technol. Biotechnol., 44: 335-353.
- Lu, H., B. Li, Y. Kang, L. Li and C. Xu, et al., 2007. Paclitaxel nanoparticle inhibits growth of ovarian cancer xenografts and enhances lymphatic targeting. Cancer Chemother. Pharmacol., 59: 175-181.
- Marinho, A.M.R., E. Rodrigues-Filho, M.D.L.R. Moitinho and L.S. Santos, 2005. Biologically active polyketides produced by Penicillium janthinellum isolated as an endophytic fungus from fruits of Melia azedarach. J. Brazilian Chem. Soc., 16: 280-283.
- Marquez, S.S., G.F. Bills and I. Zabalgogeazcoa, 2007. The endophytic mycobiota of the grass Dactylis glomerata. Fung Divers, 27: 171-195
- Mitscher, L.A., H. Telikepalli, E. McGhee and D.M. Shankel, 1996. Natural antimutagenic agents. Mutation Res., 350: 143-152.
- Owen, N.L. and N. Hundley, 2004. Endophytes-the chemical synthesizers inside plants. Sci. Progress, 87: 79-99.
- Owen, R.W., W.E. Giacosa, R. Hull, B.S. Haubner and H. Bartsch, 2000. The antioxidant/anticancer potential of phenolic c ompounds Isolated from olive oil. Eur. J. Cancer, 36: 1235-1247.
- Pittayakhajonwut, P., R. Suvannakad, S. Thienhirun, S. Prabpai, P. Kongsaeree and M. Tanticharoen, 2005. An anti-herpes simplex virus-type 1 agent from Xylariamellisii (BCC 1005). Tetrahedron Lett., 46: 1341-1344.
- Pandi, M., R. Manikandan and J. Muthumary, 2010. Anticancer activity of fungal taxol derived from Botryodiplodia theobromae Pat., an endophytic fungus, against 7, 12 dimethyl benz(a)anthracene (DMBA)-induced mammary gland carcinogenesis in Sprague dawley rats. Biomed. Pharmacother., 64: 48-53.
- Park, J.H., G.J. Choi, H.B. Lee, K.Y. Cho and K. Jin-Cheol *et al.*, 2005. Griseofulvin from Xylaria sp. strain F0010, an endophytic fungus of Abies holophylla and its antifungal activity a gainst plant pathogenic fungi. J. Microbiol. Biotechnol., 15: 112-117.
- Peltier, S., J.M. Oger, F. Lagarce, W. Couet and J.P. Benoit, 2006. Enhanced oral paclitaxel bioavailability after administration of paclitaxel-loaded lipid nanocapsules. Pharmac. Res., 23: 1243-1250.
- Pimentel, M.R., G. Molina, A.P. Dionisio, M.R.M. Junior and G.M. Pastore, 2010. The use of endophytes to obtain bioactive compounds and their application in biotransformation process. Biotechnol. Res. Int., 2011: 11-11.
- Pongcharoen, W., V. Rukachaisirikul, S. Phongpaichit, T. Kuhn, M. Pelzing, J. Sakayaroj and W.C. Taylor, 2008. Metabolites from the endophytic fungus Xylaria sp. PSU-D14. Phytochemistry, 69: 1900-1902.
- Puri, S.C., A. Nazir, R. Chawla, R.K. Sharma and G.N. Qazi *et al.*, 2006. The endophytic fungus Trametes hirsute as a novel alternative source of podophyllotoxin and related aryl tetralin lignans. J. Biotechnol., 122: 494-510.

- Puri, S.G., V. Verma, T. Amna, G.N. Qazi and M. Spiteller, 2005. An endophytic fungus from Nothapodytes foetida that produces camptothecin. J. Nat. Prod., 68: 1717-1719.
- Qin, J.C., Y.M. Zhang, J.M. Gao, M.S. Bai, S.X. Yang, H. Laatsch and A.L. Zhang, 2009. Bioactive metabolites produced by Chaetomium globosum, an endophytic fungus isolated from Ginkgo biloba. Bioorganic Med. Chem. Lett., 19: 1572-1574.
- Rajendran, L., G. Karthikeyan, T. Raguchander and R. Samiyappan, 2008. Cloning and sequencing of novel endophytic Bacillus subtilis from coconut for the management of basal stemrot disease. Asian J. Plant Pathol., 2: 1-14.
- Rehman, S., A.S. Shawl, A. Kour, V. Verma and G.N. Qazi, 2008. An endophytic Neurospora sp., from Nothapodytes foetida producing camptothecin. Applied Biochem. Microbiol., 44: 203-209.
- Rivera-Varas, V.V., T.A. Freeman, N.C. Gudmestad and G.A. Secor, 2007. Mycoparasitism of Helminthosporium solani by Acremonium strictum. Phytopathology, 97: 1331-1337.
- Sala, A., M. Del Carmen Recio, R.M. Giner, S. Manez and H. Tournier *et al.*, 2002. Anti-inflammatory and antioxidant properties of Helichrysum italicum. J. Pharm. Pharmacol., 54: 365-371.
- Schulz, B., J. Sucker, H.J. Aust, K. Krohn, K. Ludewig, P.G. Jones and D. Doring, 1995. Biologically active secondary metabolites of endophytic Pezicula species. Mycological Res., 99: 1007-1015.
- Schulz, B. and C. Boyle, 2005. The endophytic continuum. Mycol. Res., 109: 661-686.
- Shweta, S., S. Zuehlke and B.T. Ramesha, 2010. Endophytic fungal strains of Fusarium solani, from Apodytes dimidiate E. Mey. ex Arn (Icacinaceae) produce camptothecin, 10- hydroxycamptothecin and 9-methoxycamptothecin. Phytochemistry, 71: 117-122.
- Silva, G.H., H.L. Teles, H.C. Trevisan, V.S. Bolzani and M.C.M. Young *et al.*, 2005. New bioactive metabolites produced by Phomopsis cassiae, an endophytic fungus In Cassia spectabilis. J. Braz. Chem. Soc., 16: 1463-1466.
- Song, J.H., 2008. What is new on the antimicrobial horizon. Int. J. Antimicrobial Agents, 32: S207-S213.
- Song, Y.C., H. Li, Y.H. Ye, C.Y. Shan, Y.M. Yang and R.X. Tan, 2004. Endophytic naphthopyrone metabolites are co-inhibitors of xanthine oxidase, SW1116 cell and some microbial growths. FEMS Microbiol. Lett., 241: 67-72.
- Song, Y.C., W.Y. Huang, C. Sun, F.W. Wang and R.X. Tan, 2005. Characterization of graphislactone A as the antioxidant and free radical-scavenging substance from the culture of Cephalosporium sp., IFB-E001, an endophytic fungus in Trachelospermum jasminoides. Biol. Pharmac. Bull., 28: 506-509.
- Stierle, A., G. Strobel and D. Stierle, 1993. Taxol and taxane production by Taxomyces andreanae, an endophytic fungus of Pacific yew. Science, 260: 214-216.
- Strobel, G.A., W.M. Hess, J.Y. Li, E. Ford, J. Sears, R.S. Sidhu and B. Summerell, 1997. Pestalotiopsis guepinii, a taxol-producing endophyte of the wollemi pine, Wollemia nobilis. Aust. J. Bot., 45: 1073-1082.
- Strobel, G.A., E. Dirksie, J. Sears and C. Markworth, 2001. Volatile antimicrobials from a novel endophytic fungus. Microbiology, 147: 2943-2950.
- Strobel, G., E. Ford, J. Worapong, D.M. Grant and P.C. Fung *et al.*, 2002. Isopestacin, an isobenzofuranone from Pestalotiopsis microspora, possessing antifungal and antioxidant activities. Phytochemistry, 60: 179-183.
- Strobel, G.A., 2003. Endophytes as sources of bioactive products. Microbes Infection, 5: 535-544.

- Strobel, G. and B. Daisy, 2003. Bioprospecting for microbial endophytes and their natural products. Microbiol. Mol. Biol. Rev., 67: 491-502.
- Strobel, G., B. Daisy, U. Castillo and J. Harper, 2004. Natural products from endophytic microorganisms. J. Nat. Prod., 67: 257-268.
- Suresh, B., T. Ritu and G.A. Ravishankar, 2006. Biotransformations as Applicable to Food Industries. 2nd Edn., Taylor and Francis, New York, USA., pp: 1655-1690.
- Tan, R.X. and W.X. Zou, 2001. Endophytes: A rich source of functional metabolites. Nat. Prod. Rep., 18: 448-459.
- Uma, S.R., B.T. Ramesha, G. Ravikanth, P.G. Rajesh, R. Vasudeva and K.N. Ganeshaiah, 2008. Chemical Profiling of N. nimmoniana for Camptothecin, an Important Anticancer Alkaloid: Towards the Development of a Sustainable Production System. In: Bioactive Molecules and Medicinal Plants, Ramawat, K.G. and J. Merillion (Eds.). Springer, Berlin, Germany, pp: 198-210.
- Wagenaar, M.M., J. Corwin, G. Strobel and J. Clardy, 2000. Three new cytochalasins produced by an endophytic fungus in the genus Rhinocladiella. J. Nat. Prod., 63: 1692-1695.
- Wall, M.E., M.C. Wani, C.E. Cook, K.H. Palmer, A.T. McPhail and G.A. Sim, 1966. Plant antitumor agents. I. The isolation and structure of camptothecin, a novel alkaloidal leukemia and tumor inhibitor from Camptotheca acuminate. J. Am. Chem, Soc., 88: 3888-3890.
- Waller, F., B. Achatz, T.H. Baltruscha, J. Fodor and K. Becker *et al.*, 2005. The endophytic fungus Piriformospora indica reprograms barley to salt-stress tolerance, disease resistance and higher yield. PNAS USA., 102: 13386-13391.
- Wang, F.W., R.H. Jiao, A.B. Cheng, S.H. Tan and Y.C. Song, 2007. Antimicrobial potentials of endophytic fungi residing in Quercus variabilis and brefeldin A obtained from Cladosporium sp. World J. Microbiol. Biotechnol., 23: 79-83.
- Wang, J., G. Li, H. Lu, Z. Zheng, Y. Huang and W. Su, 2000. Taxol from Tubercularia sp. strain TF5, an endophytic fungus of Taxus mairei. FEMS Microbiol. Lett., 193: 249-253.
- Wani, M.C., H.L. Taylor, M.E. Wall, P. Coggon and A.T. McPhail, 1971. Plant antitumor agents. VI. The isolation and structure of taxol, a novel antileukemic and antitumor agent from Taxus brevifolia. J. Am. Chem. Soc., 93: 2325-2327.
- Worapong, J., G.A. Strobel, B. Daisy, U. Castillo, G. Baird and W.M. Hess, 2002. Muscodor roseus anna. nov. an endophyte from Grevillea pteridifolia. Mycotaxon, 81: 463-475.
- Worapong, J., G.A. Strobel, E.J. Ford, J.Y. Li, G. Baird and W.M. Hess, 2001. Muscodor albus gen. et sp. nov., an endophyte from Cinnamomum zeylanicum. Mycotaxon, 79: 67-79.
- You, F., T. Han, J.Z. Wu, B.K. Huang and L.P. Qin, 2009. Antifungal secondary metabolites from endophytic Verticillium sp. Biochem. Syst. Ecol., 37: 162-165.
- Yu, H., L. Zhang, L. Li, P. Sun and L. Qin *et al.*, 2010. Recent developments and future prospects of antimicrobial metabolites produced by endophytes. Microbiological Res., 165: 437-449.
- Zhang, B., G. Salituro, D. Szalkowski, Z. Li and Y. Zhang *et al.*, 1999. Discovery of small molecule insulin mimetic with antidiabetic activity in mice. Science, 284: 974-977.
- Zhang, J.Y., L.Y. Tao, Y.J. Liang, S. Zhi-Gang and L. Yong-Cheng *et al.*, 2009. Secalonic acid D induced leukemia cell apoptosis and cell cycle arrest of G1 with involvement of GSK-3β/β-catenin/c-Myc pathway. Cell Cycle, 8: 2444-2450.
- Zhang, L., R. An, J. Wang, N. Sun, S. Zhang, J. Hu and J. Kuai, 2005. Exploring novel bioactive compounds from marine microbes. Curr. Opin. Microbiol., 8: 276-281.