



American Journal of
**Biochemistry and
Molecular Biology**

ISSN 2150-4210



Academic
Journals Inc.

www.academicjournals.com

Comparative Efficacy of Some Plant Extracts against Fungal Deterioration of Stucco Ornaments in the Mihrab of Mostafa Pasha Ribate, Cairo, Egypt

H.A.M. Afifi

Department of Conservation and Restoration, Faculty of Archaeology, Cairo University, Giza 12613, Egypt

ABSTRACT

The demand for medicinal plants is expanding rapidly where plants are now recognized as a safe, efficient and in expensive fungicides for treatment of stucco ornaments. In this study, three different plants extracts *Anethum graveolens*, *Cymbopogon citrates* and *Juniperus oxycedrus* occurring naturally in the plant essentials were evaluated to inhibit the fungal growth of the stucco ornaments in the Ribate of Mostafa Pasha (Azdomor Al Salehy) that belonged to the Ayyubid period. Three fungal species namely: *Fusarium oxysporum*, *Aspergillus niger*, *Alternaria alternate* were isolated from different sites on the tested stucco ornaments. The toxicity of the three plant extracts concentrations against fungal growth revealed that *Cymbopogon citrates* at 100% recorded the highest significant inhibition zones (6.92, 8.0 and 5.19 cm) in all tested fungal species (*A. niger*, *F. oxysporium* and *A. alternate*), respectively. It is the most effective one in stopping fungal deterioration followed by *Anethum graveolens* fungicide which only retarded fungal growth while *Juniperus oxycedrus* was least effective.

Key words: Stucco ornaments, fungi, mihrab, ribate, *Anethum graveolens*, *Cymbopogon citrates*, *Juniperus oxycedrus*

INTRODUCTION

Growth of microorganisms on decorative stucco cause aesthetic and structural damage. Stucco may be colonized by communities of organisms that interact with the substrate at different stages (Ciferri, 1999). It must be pointed out that the biodeterioration of a given artifact commonly results from the complex interaction established by the microorganisms co-existing simultaneously (Warscheid and Braams, 2000). Suitable temperatures and moisture content combined with rich nutrients in the medieval substrate create suitable conditions for reproduction of quiescent fungal spores. Humidity and heat permit for hyphae germination that penetrate the substrate, with mechanically destructive effects (McNamara and Mitchell, 2005).

As colonization proceeds, the smooth surface of the painting is modified: Pigments initially compact and resistant to attack by various agents become rough and bioreactive (Giullitte, 1995). These events give rise to exfoliation, cracking and loss of the coloured patina (Ciferri, 1999). Certain deteriogenic fungi change cell surface molecules, causing genetic recombinations (Vestrepn *et al.*, 2004). Their metabolic products cause further chemical damage (Angell and Chamberlain, 1991). The capacity of fungi to dissolve carbonates depends on available carbon sources, such as oxalic and citric acids which may mobilize cations with chelating activity (Hirsch *et al.*, 1995). Fungi are an important constituent of microbial endolithic assemblages in moist ecosystems (Golubic *et al.*, 2005). Growth and viability of fungi on building materials under moistening and drying conditions were investigated by Pasanen *et al.* (2000).

Several preventive and remedial methods have been used in tropical environments for control and eradication of microorganisms on stucco monuments. Remedial methods are aimed at the direct elimination by chemical treatments to eliminate and control the growth of biodeteriogens (Curri, 1978; Geweely and Afifi, 2011). Fungal damage on archaeological building materials and their control measures in some museums was studied, where *Aspergillus flavus*, *A. versicolor*, *Aureobasidium* sp. and *Penicillium frequentans* was recorded by Cepero *et al.* (1992) and *Cladosporium* sp. was detected by Warscheid *et al.* (1992) and Kumar and Sharma (1992) evaluates biocides for preservation of architectural building materials.

Natural plant materials are important sources of some of the widely used plant extracts (Salehi Sourmaghi *et al.*, 2006; Thatoi *et al.*, 2008; Ajibesin *et al.*, 2008; Keymanesh *et al.*, 2009). Flowers, leaves, roots and other parts are finally ground and used or the toxic principals are extracted and utilized (Mondal and Khalequzzaman, 2010; Isman, 2006; Benamar *et al.*, 2010). Several investigations have been directed towards the antimicrobial activity of essential oils: (Adebajo *et al.*, 1989; Gangrade *et al.*, 1991; Kishore *et al.*, 1980; Mishra *et al.*, 1990; Skukla and Tripathi, 1987). Leaf extract of lemongrass was significantly inhibited the radial growth of *Aspergillus niger*, *Fusarium moniliforme* (Zaman *et al.*, 1997; ChemMatters, 2004).

The aim of the present study was to investigate the fungal deterioration phenomena of stucco ornaments in Ribate of Mostafa Pasha, Cairo, Egypt which suffer from severe fungal deterioration. Hence, preservation of this material from fungal biodeterioration by using safe fungicide was a national goal in this work.

MATERIALS AND METHODS

Evaluation of the possibility of using plant extracts especially botanical fungicide in preservation of the tested stucco was carried out. Sensitivity of isolated fungal species against three different plant extracts was tested.

Source of isolation and field observation: Ribate of Mostafa Pasha has many stucco decoration in the mihrabs. These decorations suffered from different forms of deterioration grades as discoloration black to dark green due to fungal colonies. It is clear and scattered at different parts particularly in the damped parts. Many cracks and lost parts at different altitudes in the stucco decorations are observed.

Isolation and identification of fungi from tested sample: Samples from markedly damaged stucco surfaces were collected. Swabbing with sterile cotton swabs and scalpel from markedly damaged surfaces with visible colonies of microscopic fungi was carried out. In the laboratory, swab samples were shaken mechanically for 10 min in 10 mL sterile distilled water and 1 mL aliquots of the resulting suspensions used to prepare spread plates on Czapeck's Dox agar in order to isolate as wide a range of microfungi as possible. Plates were incubated in the dark at laboratory temperature (25°C) for 7 days and the microscopic fungi were identified using the diagnostic keys of Booth (1977), Gilman (1957), Barnett and Hunter (1972) and Moubasher (1993).

Plant extracts inhibition zone: Control of fungal growth on stucco decoration from the mihrab of Mostafa Pasha Ribate by plant extracts inhibition zone was performed. Three plant extracts were used in protecting stucco from biodeterioration namely: *Anethum graveolens*, *Cymbopogon citrates*, *Juniperus oxycedrus*. Aliquots of about 15 mL of Czapeck's Dox agar were dispersed into sterile

Table 1: Surveys of genera and species of fungi isolated from stucco samples

Fungal species	Count of species	Frequency of occurrence
<i>Aspergillus niger</i>	16	High
<i>Fusarium oxysporum</i>	9	Moderate
<i>Alternaria alternata</i>	5	Low
Total count	30	
No. of species	3	

petri dishes. Each dish was fissured by using sterilized cork borer (1 cm diameter hole) at its center and 1 mL of each plant extract was added.

Each plant extract was used in five concentrations 10, 30, 50, 70 and 100% and was put in each hole and each petri dish was inoculated with tested fungi which cut from the colony margin of 2-4 day old cultures. The plates were incubated for 24 h intervals, after which the diameters of inhibition zone (in cm) were measured.

RESULTS AND DISCUSSION

Frequency of occurrence of fungal species isolated from Stucco Ornaments in the Mihrab of Mostafa Pasha Ribate: Three deteriorating fungal species were isolated from different sites on stucco. Genus *Aspergillus* contributed the broadest spectra where *A. niger* showed highest occurrence as it occurred in the all tested sites on stucco. This result agree with Abdel-Hamid and Ouf (1990) who recorded that *Aspergillus* genus was the broadest deteriorating factor causing damage of materials and also Bisht (1995) said that the most common bio-deteriorating fungal species were *Aspergillus*, *Cladosporium*, *Alternaria* and *Penicillium*. On the other hand, *Fusarium oxysporum* was isolated in moderate occurrence and *Alternaria alternata* was low occurrence (Table 1).

It is distinctly possible that the minerals may have been contaminated by airborne spores. Hyvarinen *et al.* (2004) stated that mold growth of *Aspergillus* spp. on moistened damage building materials with numbers was between 10 and 10⁸ cfu g⁻¹ and associated with adverse health effects. Eight microfungi on 21 different types of building material was investigated and only *Penicillium*, *Aspergillus* were a dominant species (Nielsen *et al.*, 2004). The record of fungal occurrence on the building material points to the presence of dung (Van Geel *et al.*, 2003). Fungi were isolated from the damaged building materials could use organic substances of restoration material as nutrient sources for growth on inorganic building materials, it is in fact the interrelationships which play the main role within microbial communities of this type (Karpovich-Tate and Rebrikova, 2002).

Comparative assessment of microbiological deterioration of constructional material which is part of the cultural heritage was achieved by Herrera *et al.* (2004).

Plant extracts sensitivity: Three plant extracts namely: *Anethum graveolens*, *Cymbopogon citratus* and *Juniperus oxycedrus* have been applied on the isolated mould colonization from deteriorated stucco decorations. The data revealed that a significant decrease in the growth of all isolated fungal species with progressive increasing in the concentration of the all three tested plant extracts. The lowest concentrations (10-30%) of the tested *A. graveolens* and *J. oxycedrus* extracts have no effect on the growth of three tested fungal species.

The most resistant tested fungal species was *Alternaria alternata*, where the lowest inhibition zone (2.1 cm) was occurred with 100% concentration of *A. graveolens* extract, on the other hand

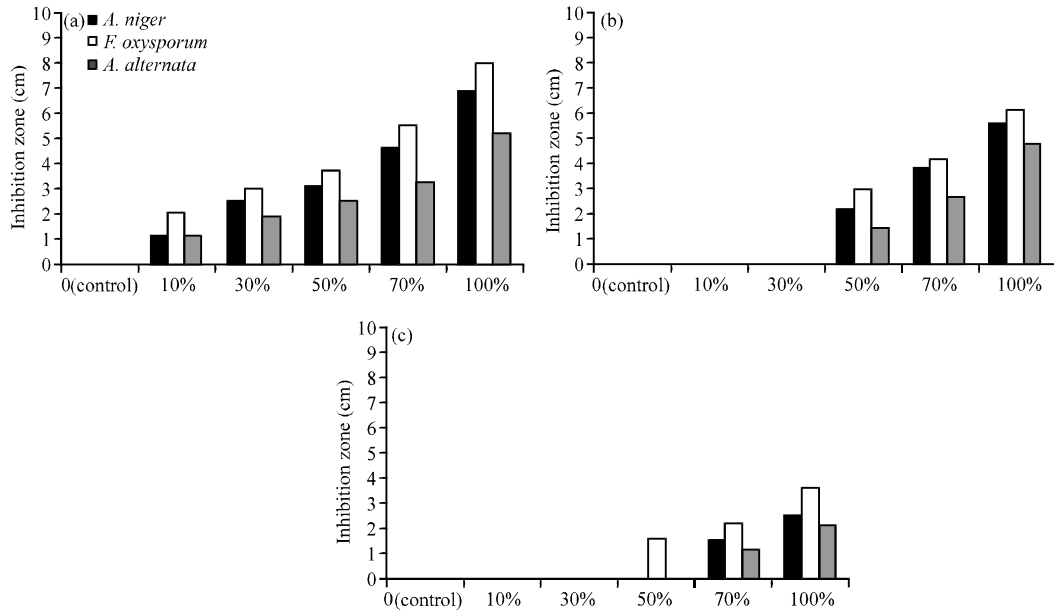


Fig. 1 (a-c): Effect of different concentration of (a) *Cymbopogon citrates*, (b) *Juniperus oxycedrus* and (c) *Anethum graveolens*, extracts on the growth of the three isolated deteriorated fungal species, LSD at 0.05 = 0.50, 1.54 and 2.11, respectively

Fusarium moniliforme was considered the most sensitive tested fungal species, where the maximum significant inhibition zone (8.0 cm) was showed with *C. citratus* at 100% concentration (Fig. 1a, b). This may refer to the spores of *Alternaria* are pigmented and more resistant to the inhibitory effect of the tested plant extracts where certain correlations between morphology of the spores and their susceptibility to inhibitor were indicated. This finding agree with that obtained by Geweely (2009) and Hibben and Stotzky (1969) who stated that the spores were more sensitive to inhibition are relatively hyaline, while the most resistant spores were large and pigmented.

Fungicides sensitivity on the isolated fungal species (fungicides inhibition zone): The obtained data revealed that *Cymbopogon citrates* at 100% recorded the highest significant inhibition zones (6.92, 8.0 and 5.19 cm) in all tested fungal species (*A. niger*, *F. oxysporium* and *A. alternata*, respectively) Fig. 1a. The obtained data was agree with that obtained by Fiori *et al.* (2000) who stated that the plant species responsible for the greatest inhibition of the mycelial growth of microorganisms was *C. citratus*. The highest inhibitory effect of *Cymbopogon citratus* may be referred to the presence of antifungal protein in the extract. Linthorst and Van Loon (1991) stated that low molecular weight proteins or peptides in the plant extract have antifungal activities. They were believed to be involved in a defense mechanism against pathogenic fungi by inhibiting microbial growth through binding to chitin or increasing the permeability of the microbial membranes or cell wall. According to the explanation of Farag *et al.* (1989), it appears that there is a relationship between the chemical structure of the most abundant compounds in the plant extract and the antimicrobial activity. Fiori *et al.* (2000) indicated that the plant species responsible for the greatest inhibition of the mycelial growth of microorganisms was *Cymbopogon citratus*. Tzortzakis and Economakis (2007) tested the antifungal activity of *Cymbopogon citratus* oil against *Aspergillus niger*. According to the explanation of Farag *et al.* (1989), it appears that there is a

relationship between the chemical structure of the most abundant compounds in the essential oils and the anti-microbial activity. The data in Fig. 1b showed that applying of 100% *J. oxycedrus* on the tested fungi gave the moderate inhibitory effect. On the other hand *A. graveolens* had low inhibitory effect against all tested fungal species (Fig. 1c). Building materials are shown to be deteriorated by a variety of fungi and the control was achieved by Nalli *et al.* (2006), Schieweck *et al.* (2005), Do *et al.* (2005) and Clausen and Vina (2005). Tzortzakis and Economakis (2007) tested the antifungal activity of *Cymbopogon citratus* oil against *Aspergillus niger*. Differences in sensitivity to fungicide also existed between strains of the same species (Dannenhauer *et al.*, 1983).

CONCLUSION

Mostafa Pasha Ribate belonged to the Ayyubid period has many stucco ornaments in the two mihrabs. These decorations have many colored spots due to some fungal deterioration. Three of plant extracts were examined against the deteriorated tested fungal species. *Cymbopogon citratus* was the best one for controlling all tested fungal species. Treatment of the stucco ornaments of the mihrab must be carried out by applying *Cymbopogon citratus* extract, where the highest inhibition zones were showed in *A. niger*, *F. oxysporium* and *A. alternata*, respectively. It was the most preferable inhibitor to prevent fungal growth instead of using chemicals fungicides to avoid the unfavorable side effect of chemicals on the properties of archeological stone and public health and environment.

ACKNOWLEDGMENT

I would like to express my deepest thanks to Dr. Neveen S.I. Geweely, Associate professor of microbiology, Department of Botany, Faculty of Science, Cairo University her assistance and helpful discussions.

REFERENCES

- Abdel-Hamid, H.E. and S.A. Ouf, 1990. Study on fungi isolated from old manuscript pages. Agric. Ain Shames Univ. Cairo, Egypt, Ann. Agric. Sci., 2: 635-652.
- Adebajo, A.C., K.J. Oloke and A.J. Aladesanmi, 1989. Anti-microbial activities and microbial transformation of volatile oils of *Eugenia uniflora*. Fitoterapia, 60: 415-455.
- Ajibesin, K.K., N. Rene, D.N. Bala and U.A. Essiett, 2008. Antimicrobial activities of the extracts and fractions of *Allanblackia floribunda*. Biotechnology, 7: 129-133.
- Angell, P. and A.H.L. Chamberlain, 1991. The role of extracellular products in copper colonization. Int. Biodeterior., 27: 135-143.
- Barnett, H.L. and B.B. Hunter, 1972. Illustrated of Imperfect Fungi. 3rd Ed., Burgess Publishing Co., Minneapolis, Minnesota.
- Benamar, H., W. Rached, A. Derdour and A. Marouf, 2010. Screening of algerian medicinal plants for acetylcholinesterase inhibitory activity. J. Biol. Sciences, 10: 1-9.
- Bisht, A.S., 1995. Effect of climate on museum objects and its control. Biodeterioration of cultural heritage some case studies. Proceedings of 3rd International Symposium on Restoration and Conservation of Monuments. (ISRCM'95), Hyderabad, India, pp: 1-7.
- Booth, C., 1977. The Genus *Fusarium*. Common wealth Mycological Institute, New Surrey, England.

- Cepero, A.P., J. Martinez, A.S. Castro and J. Machado, 1992. The biodeterioration of cultural property in the republic of Cuba: A review of some experiences. Proceedings of the 2nd International Conference on Biodeterioration of Cultural Property. (ICBCP'92), International Communications Specialists, pp: 479-487.
- ChemMatters, 2004. Teacher's Guide. American Chemical Society, USA.
- Ciferri, O., 1999. Microbial degradation of paintings. *Applied Environ. Microbiol.*, 65: 879-885.
- Clausen, C.A. and W.Y. Vina, 2005. Azole-based antimycotic agents inhibit mold on unseasoned pine. *Int. Biodeterior. Biodegrad.*, 55: 99-102.
- Curri, S.B., 1978. Biocide testing and enzymological studies on damaged stone and fresco surfaces: Preparation of antibiograms. *Biochem. Exp. Biol.*, 15: 97-104.
- Dannenhauer, H., A. Resz and F. Grossmann, 1983. Experiments with alzodef (cyanamide) as a medium for soil disinfection in vegetable and ornamental plant cultivation. *Zeitsch Pflanzenkrankheiten und pflanzenschutz*, 90: 468-478.
- Do, J., S. Hun, S. Hyoungseok and S. Yangseob, 2005. Antifungal effects of cement mortars with two types of organic antifungal agents. *Cem. Concr. Res.*, 35: 371-376.
- Farag, R.S., Z.Y. Dawz, F.M. Hewedi and G.S. El-Barotyl, 1989. Antimicrobial activity of some Egyptian Spice essential oils. *J. Food Prot.*, 52: 665-667.
- Fiori, A.C.G., K.R.F. Schwan-Estrada, J.R. Stangarlin, J.B. Vida, C.A. Scapim, M.E.S. Cruz and S.F. Pascholati, 2000. Antifungal activity of leaf extracts and essential oils of some medicinal plants against *Didymella bryoniae*. *J. Phytopathol.*, 148: 483-487.
- Gangrade, S.K., R.D. Shrivastava, O.P. Sharma, N.K. Jain and K.C. Trivedi, 1991. *In vitro* anti-fungal effect of the essential oils. *Indian Perfum*, 35: 46-49.
- Geweely, N.S., 2009. Novel inhibition of some pathogenic fungal and bacterial species by new synthetic phytochemical coumarin derivatives. *Ann. Microbiol.*, 59: 359-368.
- Geweely, N.S.I. and H.A.M. Afifi, 2011. Bioremediation of some deterioration products from sandstone of archeological karnak temple using stimulated irradiated alkalo-thermophilic purified microbial enzymes. *Geomicrobiol. J.*, 28: 56-67.
- Gilman, J.C., 1957. *A Manual of Soil Fungi*. 2nd Edn., The Iowa State College Press, Ames, Iowa, USA., pp: 450.
- Giullitte, O., 1995. Bioreceptivity: A new concept for building ecology studies. *Sci. Total Environ.*, 167: 215-220.
- Golubic, S., R. Gudrun and L. Therese, 2005. Endolithic fungi in marine ecosystems. *Trends Microbiol.*, 13: 229-235.
- Herrera, L.K., A. Carlos, G. Patricia, G.D.S. Sandra and V. Hector, 2004. International biodeterioration and biodegradation. Proceedings of the 12th International Biodeterioration and Biodegradation Symposium (Biosorption and Bioremediation III). (IBBS'04), Elsevier, pp: 135-141.
- Hibben, C.R. and G. Stotzky, 1969. Effects of ozone on the germination of fungus spores. *Can. J. Microbiol.*, 15: 1187-1196.
- Hirsch, P., F.E.W. Eckhardt and R.J. Palmer, 1995. Fungi active in weathering of rock and stone monuments. *Can. J. Bot.*, 73: 1384-1390.
- Hyvarinen, A., T. Meklin, A. Vepsalainen and A. Nevalainen, 2004. Fungi and actinobacteria in moisture-damaged building materials concentrations and diversity. *Int. Biodeterioration Biodegradation*, 49: 27-37.

- Isman, M.B., 2006. Botanical insecticides, deterrents and repellents in modern agriculture and an increasingly regulated world. *Annu. Rev. Entomol.*, 51: 45-66.
- Karpovich-Tate, N. and N.L. Rebrikova, 2002. Microbial communities on damaged frescoes and building materials in the cathedral of the nativity of the virgin in the pafnutii-borovskii monastery. *Russia Int. Biodeterior.*, 27: 281-296.
- Keymanesh, K., J. Hamed, S. Moradi, F. Mohammadipana and S. Sardari, 2009. Antibacterial, antifungal and toxicity of rare Iranian plants. *Int. J. Pharmacol.*, 5: 81-85.
- Kishore, N., S.K. Singh and N.K. Dubey, 1980. Fungitoxic activity of essential oil of *Juniperus communis*. *Indian Perfum.*, 33: 25-29.
- Kumar, R. and R.K. Sharma, 1992. Conservation of deul of the Lord Jagannath temple, Puri (Orissa), India: A case study. *Proceedings of the 7th International Congress on Deterioration and Conservation of Stone, (ICDCS'92), Portugal*, pp: 1471-1480.
- Linthorst, H.J.M. and L.C. Van Loon, 1991. Pathogenesis related proteins of plants. *Crit. Reine Plant Sci.*, 10: 123-150.
- McNamara, C.J. and R. Mitchell, 2005. Microbial deterioration of historic stone. *Front. Ecol. Environ.*, 3: 445-451.
- Mishra, D.N., A.K. Mishra and V. Dixit, 1990. Mycotoxic properties of the essential oil of eucalyptus against some dermatophytes. *Vegetos*, 3: 182-185.
- Mondal, M. and M. Khalequzzaman, 2010. Toxicity of naturally occurring compounds of plant essential oil against *Tribolium castaneum* (Herbst). *J. Biol. Sci.*, 10: 10-17.
- Moubasher, A.H., 1993. *Soil Fungi in Qatar and other Arab Countries*. Center of Scientific and Applied Research, University of Qatar, Doha, Qatar, ISBN-13: 9992121025, pp: 566.
- Nalli, S., O.J. Horn, A.R. Grochowalski, D.G. Cooper and J.A. Nicell, 2006. Origin of 2-ethylhexanol as a VOC. *Environ. Pollut.*, 140: 181-185.
- Nielsen, K.F., G. Holm, L.P. Uttrup and P.A. Nielsen, 2004. Mould growth on building materials under low water activities. Influence of humidity and temperature on fungal growth and secondary metabolism. *Int. Biodeterior. Biodegrad.*, 54: 325-336.
- Pasanen, A.L., J.P. Kasanen, S. Rautiala, M. Ikaheimo, J. Rantamaki, H. Kaariainen and P. Kalliokoski, 2000. Fungal growth and survival in building materials under fluctuating moisture and temperature conditions. *Int. Biodeterior. Biodegrad.*, 46: 117-127.
- Salehi Sourmaghi, M.H., G. Amin, N. Samadi, F. Hemmati and P. Sarkhail, 2006. Chemical composition and antimicrobial activity of essential oil of *Salvia spinosa* L. *Asian J. Plant Sci.*, 5: 654-656.
- Schieweck, A., B. Lohrengel, N. Siwinski, C. Genning and T. Salthammer, 2005. Organic and inorganic pollutants in storage rooms of the Lower Saxony State Museum Hanover. Germany. *Atmos. Environ.*, 39: 6098-6108.
- Skukla, H.S. and S.C. Tripathi, 1987. Anti-fungal substance in the essential oil of anise (*Pimpinella anisum* L.). *Agric. Biol. Chem.*, 51: 1991-1993.
- Thatoi, H.N., S.K. Panda, S.K. Rath and S.K. Dutta, 2008. Antimicrobial activity and ethnomedicinal uses of some medicinal plants from simlipal biosphere reserve, Orissa. *Asian J. Plant Sci.*, 7: 260-267.
- Tzortzakis, N.G. and C.D. Economakis, 2007. Antifungal activity of lemongrass (*Cymbopogon citratus* L.) essential oil against key post harvest pathogens. *Innovative Food Sci. Emerg. Technol.*, 8: 253-258.

- Van Geel, B., J. Buurman, O. Brinkkemper, J. Schelvis, A. Aptroot, G. Van Reenen and T. Hakbijl, 2003. Environmental reconstruction of a Roman Period settlement site in Uitgeest (The Netherlands), with special reference to coprophilous fungi. *J. Archaeolog. Sci.*, 30: 873-883.
- Vestrepen, K.J., T.B. Reynolds, G.R. Fink, 2004. Origins of variation in the fungal cell surface. *Nat. Rev. Microbiol.*, 2: 533-540.
- Warscheid, T., D. Barros, T.W. Becker, S. Eliasaro and G. Grote *et al.*, 1992. Biodeterioration studies on soapstone, quartzite and sandstone of historical monuments in Brazil and Germany: Preliminary results and evaluation for restoration practices. Proceedings of the 7th International Congress on Deterioration and Conservation of Stone, June 15-18, Lisbon, Portugal, pp: 491-500.
- Warscheid, T. and J. Braams, 2000. Biodeterioration of stone: A review. *Int. Biodeterior Biodegrad.*, 46: 343-368.
- Zaman, M.A., A.K.M. Saleh, G.M.M. Rahman and M.T. Islam, 1997. Seed-borne fungi of mustard and their control with indigenous plant extracts. *Bangladesh J. Plant Pathol.*, 13: 25-28.