

# American Journal of **Drug Discovery** and **Development**

ISSN 2150-427X



American Journal of Drug Discovery and Development 3 (4): 225-234, 2013 ISSN 2150-427x / DOI: 10.3923/ajdd.2013.225.234 © 2013 Academic Journals Inc.

# Nimesulide-phosphatidylcholine Complex for Improvement of Solubility and Dissolution

<sup>1</sup>Ajay Semalty and <sup>2</sup>Yuveraj Singh Tanwar

<sup>1</sup>Department of Pharmaceutical Sciences, H.N.B. Garhwal University, Srinagar (Garhwal), 246174, Uttarakhand, India

<sup>2</sup>B.N. College of Pharmacy, Udaipur, 313002, Rajasthan, India

Corresponding Author: Ajay Semalty, Department of Pharmaceutical Sciences, H.N.B. Garhwal University, Srinagar (Garhwal), 246 174, Uttarakhand, India Tel: +91 1346 252174 Fax: +91 1346 252174

### ABSTRACT

Nimesulide, a potent non steroidal anti-inflammatory drug, is a highly selective cyclooxygenase-2 (COX-2) inhibitor. Being a class II drug (according to biopharmaceutical classification system or BCS) its poor aqueous solubility results in low bioavailability. Moreover, its absorption is dissolution rate limited. It also shows hepatic and gastrointestinal toxicity in long term use. Therefore, to improve solubility, dissolution (hence the bioavailability) and to reduce toxic effects of nimesulide, its phospholipid complex was prepared. The prepared phospholipid complex was evaluated for drug loading, solubility, Scanning Electron Microscopy (SEM), Infrared absorption (FT-IR), Differential Scanning Calorimetry (DSC), X-ray powder diffractometry (X-RPD) and in vitro dissolution study. The aqueous solubility of nimesulide was improved significantly in the complex. In the SEM phospholipid complex was found to be fluffy and porous with rough surface morphology. FT-IR, DSC and X-RPD data confirmed the formation of the complex. The prepared phospholipid complex showed significantly improved dissolution profile. It was concluded that the phospholipid complexation of nimesulide like BCS class II drugs may be a very effective, reliable and safe approach to improve the solubility and dissolution of drugs.

**Key words:** Nimesulide, phosphatidylcholine, FT-IR, differential scanning calorimetry, X-RPD, scanning electron microscopy, solubility

# INTRODUCTION

Dissolution of drug is directly dependent on the aqueous solubility of the drug. The drugs which have the water solubility less than the 10 mg mL<sup>-1</sup> (over the pH range of 1-7 at 37°C) show the potential bioavailability problems. Dissolution and solubility are the two important properties which play an important role in formulation development of the drugs (Wells, 2006; Pose-Vilarnovo et al., 2001; Ozkan et al., 2000). The bioavailability of the drugs which show the dissolution rate limited absorption may be improved by improving their aqueous solubility. Various techniques and dosage forms or drug delivery systems have been designed and adopted for improving the solubility and dissolution of drugs. These techniques include supercritical fluid process, micronization, solid dispersion, cyclodextrin complexes and phospholipid complexes etc. (Perrut et al., 2005; Semalty et al., 2011; Pralhad and Rajendrakumar, 2004; Babu and Pandit, 2004; Rawat and Jain, 2004; Sajeesh and Sharma, 2006; Semalty et al., 2010b).

Out of these, the complexation technique has been employed more precisely to improve the solubility and the dissolution of poor water soluble drugs (Semalty *et al.*, 2009a; Bhati *et al.*, 2012).

Fig. 1(a-b): (a) Nimesulide and (b) Phosphatidylcholine,  $R_1$  and  $R_2$  = Long chain of fatty acid

Among the complexation techniques the phospholipid complexation and cyclodextrin complexation are the two most widely investigated approached for improving the solubility. In the various previous studies it was reported that developing the drugs as lipid complexes (also called pharmacosomes) may prove to be a potential approaches to improve solubility and to minimize the GI toxicity of drugs (Semalty et al., 2009b, 2010a) In the phospholipid complexation the drug and a phospholipid (Fig. 1b) are treated in certain molar ratio (generally 1:1 or 1:2) to yield an amphiphilic complex with improved solubility, permeability and dissolution profile.

The lipid complexes are prepared with phosphatidylcholine (PC, Fig. 1b). PC is an integral part of the cell membrane exists in zwitterionic form. PCs are studied extensively due to their interaction with several physiologically active compounds. PCs are amphiphilic molecules which yield the product of improved solubility and permeability when complexed with drugs with poor solubility and/or permeability. PC is not only a passive carrier in drug delivery but is itself a natural component with well investigated and reported clinical efficacy for various liver diseases (Semalty et al., 2010b; Kidd, 1996, 2002).

Nimesulide (4'-nitro-2'-phenoxy methane sulfonanilide) is one of the potent non steroidal anti-inflammatory drugs (NSAIDs). It is a selective cyclooxygenase-2 (COX-2) inhibitor and is 5-16 folds selective for COX-2 than COX-1 (Fig. 1a) (Bishnoi et al., 2005; Ferrari et al., 1993; Singla et al., 2000). It is very sparingly soluble in water (about 0.01 mg mL<sup>-1</sup>). The low solubility leads to very low dissolution and hence the poor bioavailability. Nimesulide is also associated with gastrointestinal disturbances and hepatic toxicity as the most frequent side effects on its long term administration (Tan et al., 2007; Davis and Brogden, 1994; Bjarnason and Thjodleifsson, 1999). Due to the poor aqueous solubility and wettability of nimesulide it is always a challenge to formulate its oral or parenteral dosage forms. As the absorption of nimesulide is solubility (and hence the dissolution) rate limited, increasing the aqueous solubility of nimesulide may be a potential approach to improve its dissolution and hence the bioavailability.

Various studies have reported that phospholipid complexes of NSAIDs improve the permeation across the bio membranes, reduced toxicities and thereby improve their bioavailability and GI safety (Khazaeinia and Jamali, 2003). When used in drug dosage form or delivery system, the phospholipid covers the surface of the mucus as a hydrophobic protective coat and hence protects the GI tissues (Goddard *et al.*, 1990; Lichtenberger *et al.*, 1983).

Therefore, the preparing the lipid complex may not only improve the aqueous solubility and dissolution rate (and hence the bioavailability) but also reduce its local gastrointestinal toxicities. For fulfilling these objectives nimesulide-phospholipid complex was prepared in the present study. The complex thus prepared was evaluated physico-chemically for drug content, chemical interaction (FT-IR), thermal analysis (DSC), crystallinity (X-RPD), Surface Morphology (SEM), solubility and dissolution study.

#### MATERIALS AND METHODS

**Materials:** Nimesulide (98%) was obtained from Panacea Biotech, Delhi (India). Soya phosphatidylcholine (LIPOID S-80) was obtained as a gift sample from LIPOID, Germany. All other chemical reagents were of analytical grade.

**Method of preparation:** To prepare the complex nimesulide and Soya phosphatidylcholine (PC) were taken in 1:1 molar ratio and dissolved in 30 mL of dichloromethane in a 100 mL round bottom flask. The solvent was evaporated off in a rotary vacuum evaporator (Perfit 5600, India) under vacuum at 40°C. The dried residue obtained is the resultant complex which was stored in vacuum desiccators.

**Drug content:** Nimesulide-PC complex equivalent to 50 mg of nimesulide was weighed. To the weighed complex 100 mL of pH 6.8 phosphate buffer was added in a volumetric flask. After the continuous stirring on a magnetic stirrer (Remi, 5MLH, India) for 24 h at room temperature samples were taken, filtered, diluted suitably and then analyzed spectrophotometrically (Lambda 25, Perkin Elmer, USA) at 390 nm to determine the drug content.

**Infrared spectroscopy (FTIR):** The IR spectra were recorded on a Perkin Elmer FT-IR, RX-1 spectrophotometer in KBr pellets.

**Differential scanning calorimetry (DSC):** DSC study was performed for the samples of nimesulide, phosphatidylcholine and the prepared complex using a 2910 Modulated Differential Scanning Calorimeter V4.4E (TA Instrument, USA). The investigations were carried out over the temperature range 0-300°C (@ 10°C min<sup>-1</sup>).

X-ray powder diffractometry (XRPD): To assess the crystallinity, XRPD of all the samples were performed using Bruker Axs-D8 Discover Powder X-ray diffractometer (Germany). The scanning was performed in the range of 5-50° of  $2\theta$  in step scan mode (step width  $1^{\circ}$ min<sup>-1</sup>).

Scanning electron microscopy (SEM): To assess the surface morphology of the prepared complex as compared to its components SEM of the complex was performed using JEOL JSM 5600.

**Solubility study:** To determine the effect of complexation on solubility, solubility of nimesulide, PC and its complex was determined in distilled water at 25±0.1°C (Semalty *et al.*, 2013b).

Dissolution study (in vitro drug release): The dissolution studies were carried out in a USP XXIII, eight station dissolution test apparatus, type II (8DR, VEEGO, India) at 100 rpm and at 37°C using pH 6.8 phosphate buffer (900 mL) as media. The complex equivalent to 50 mg of nimesulide was taken for the study and its comparison was done with the dissolution of plain nimesulide (50 mg). Samples of dissolution fluid were withdrawn at different intervals and replaced with the equal volume of fresh media. Withdrawn samples were filtered, diluted suitably and then analysed spectrophotometrically.

#### RESULTS AND DISCUSSION

In the present study the complex showed 92% w/w drug content of nimesulide. High loading of drug in the complex make the use of complex practically possible to deliver the therapeutic dosage effectively.

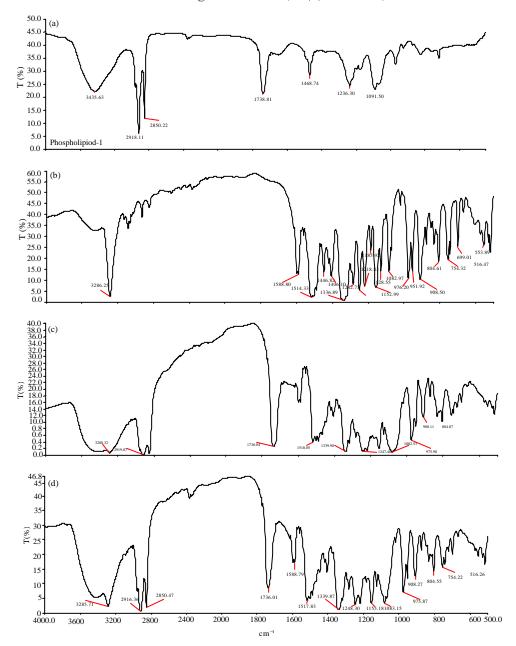


Fig. 2(a-d): IR Spectra: (a) Phospholipid, (b) Nimesulide, (c) Nimesulide-phospholipid complex and (d) Physical mixture

**FTIR:** The possible interaction between nimesulide and PC (phosphatidylcholine) in the phospholipid complex was studied by FTIR (Fig. 2). PC showed characteristic peaks at 3435 cm<sup>-1</sup> (Hydroxyl stretching); 2918 and 2850 cm<sup>-1</sup> (C-H stretching of long fatty acid chain); 1738 cm<sup>-1</sup> (carbonyl stretching of the fatty acid ester); 1236 cm<sup>-1</sup> (P = O stretching band); 1091 cm<sup>-1</sup> (P-O-C stretching) and 970 cm<sup>-1</sup> (N<sup>+</sup>(CH<sub>3</sub>)<sub>3</sub> stretching). Nimesulide showed the characteristic peaks at 3286 cm<sup>-1</sup> for the amino (N-H stretching and at 1514 cm<sup>-1</sup> for N = O stretching.

The FTIR of the complex showed significant changes in the characteristic absorption peaks of nimesulide. The peaks of amino (N-H) and nitro (N = O) group at  $3286~{\rm cm}^{-1}$  and  $1514~{\rm were}$  shifted to higher wave number in the complex. On the other hand the characteristic absorption peaks of

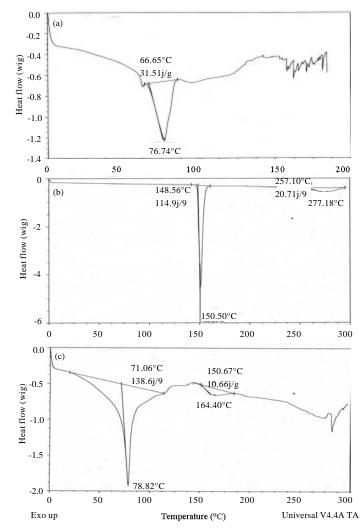


Fig. 3(a-c): DSC Thermograms: (a) Phospholipid, (b) Nimesulide and (c) Nimesulide-phospholipid complex

P = O and P-O-C of the phosphatidylcholine were broadened and also changed their position in the FTIR spectra of the complex. The FTIR spectra of the physical mixture was showing the same characteristic peaks of nimesulide and PC without any significant change and this indicated no interaction in between them.

Therefore, the formation of nimesulide-PC complex was indicated due to interaction of amino and nitro group of nimesulide with polar end of PC. The results of FTIR are well supported by the previous studies done with various PC complexes of drugs (Singh *et al.*, 2011; Semalty *et al.*, 2012).

**DSC:** DSC of all the samples (nimesulide, PC and its complex) were performed to investigate the thermal behavior of the drug. In the DSC study nimesulide (Fig. 3) exhibited sharp endothermic peak at 150.50°C ( $\Delta H_f = 114.9 \text{ J g}^{-1}$ ) corresponding to the melting of nimesulide. PC showed an endothermic peak at 76.74 °C ( $\Delta H_f = 31.51 \text{ J g}^{-1}$ ). The nimesulide-PC complex exhibited a sharp new peak at 78.82°C ( $\Delta H_f = 138.6 \text{ J g}^{-1}$ ) which showed interactions between nimesulide and PC and confirming the formation of a complex. Various studies also support these results in which complex do not show the peak corresponding to the components of the complex and rather show a

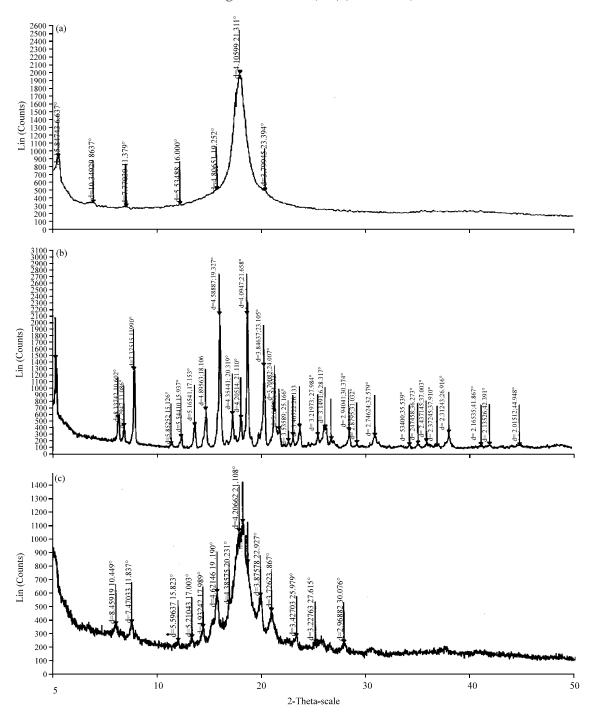


Fig. 4(a-c): X-RPD Pattern: (a) Phospholipid, (b) Nimesulide and (c) Nimesulide-phospholipid complex

entirely new peak (Singh et~al., 2011; Semalty et~al., 2012; Li et~al., 2008; Xiao et~al., 2006; Maiti et~al., 2007; Kumar et~al., 2008).

XRPD: To assess the crystallinity XRPD of nimesulide, phosphatidylcholine and the complex was performed (Fig. 4). Nimesulide showed intense diffraction peaks of crystallinity and suggested that

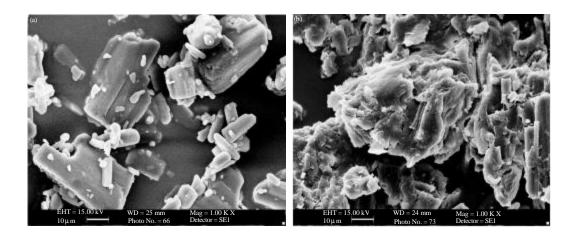


Fig. 5: Scanning electron micrographs, (a) Nimesulide and (b) Nimesulide-Pc complex

Table 1: Solubility study ( $H_2O/n$ -Octanol) at 25°C

Sample	Aqueous solubility ( $\mu g \ mL^{-1}$ )	n-Octanol solubility ( $\mu g \ mL^{-1}$ )
Nimesulide	9.67±0.902	160.60 <b>8</b> ±0.076
Nimesulide complex	0.104±1.025	124.902±0.234

Data expressed as mean values and standard deviations ( $\pm$ SD), n = 3

the drug is in crystalline state. PC showed a major single diffraction peak. In the nimesulide-phospholipid complex, the sharp and intense peaks characteristic to the crystalline nimesulide were not observed at all. There was only a large diffraction peak s in which it was not possible to distinguish the characteristic peaks of nimesulide. This confirmed the presence of nimesulide in amorphous state in the complex rather than its original crystalline state. It was also evident that XRPD data also well supported the results of DSC studies confirming the interaction resulting in the formation of the complex. Previous studies done with insulin, diclofenac, aceclofenac etc., well supported these results (Cui et al., 2006; Semalty et al., 2009a, 2010c; Singh et al., 2012b).

**SEM:** To determine the surface morphology SEM micrographs of nimesulide and its complex were obtained (Fig. 5). The pure nimesulide was showing its characteristic small crystals of regular shape with a smooth surface. But the complex showed crystals with blunt faces. The surface of the complex were non porous and rough. The characteristic rough surface morphology might have contributed to the improved solubility and the dissolution of nimesulide from the complex.

**Solubility study:** Aqueous solubility of nimesulide was found to be improved significantly in the complex (Table 1). This increase in the solubility of the complex may be explained by its amorphous characteristics and reduction in molecular crystallinity of nimesulide.

The amorphous nature of the complex (as confirmed by XRPD); typically rough surface morphology (as confirmed by SEM) and changes brought about due to complexation (as confirmed by FTIR and DSC) might have been responsible for the improvement in solubility (Semalty *et al.* 2013a; Singh *et al.*, 2013).

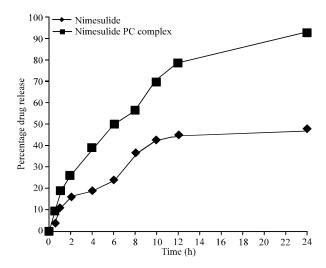


Fig. 6: Dissolution pattern of Nimesulide and its complex

**Dissolution study:** The complex of nimesulide showed a better dissolution profile than its free state (Fig. 6). Nimesulide showed only 47.65% drug release at the end of 24 h. But the nimesulide PC complex showed 92.76% drug release in dissolution study. The release might have been improved due to complexation which resulted in improved solubility of the drug. The improved surface morphology and amorphization induced by the complexation might have resulted in improved dissolution of nimesulide from its complex (Singh *et al.*, 2012a).

#### CONCLUSION

To improve the solubility and dissolution of nimesulide its lipid complex was prepared. It was concluded that the lipid complexation of nimesulide can lead to change in its state of crystallinity (amorphizing), its thermal behaviour, solubility and the dissolution profile. The lipid complexes might also be helpful in improving oral absorption of drug (class II drugs) with reduced toxicities and improved gastrointestinal safety (important with respect to NSAIDs).

## ACKNOWLEDGMENTS

The authors are thankful to Government of India, UGC Research Grant 37-643/2009. The authors acknowledge Panacea Biotech, Delhi (India) for giving nimesulide and LIPOID GmbH Germany for providing the gift sample of phosphatidylcholine for the research work. Facilities provided by UGC-DAE Consortium for Scientific Research, Indore (India), are gratefully acknowledged.

#### REFERENCES

Babu, R.J. and J.K. Pandit, 2004. Effect of cyclodextrins on the complexation and transdermal delivery of bupranolol through rat skin. Int. J. Pharm., 271: 155-165.

Bhati, L.K., G. Tiwari, R. Tiwari and V. Kumar, 2012. Enhancement of complexation efficiency of meloxicam using binary and ternary solid systems: Formulation considerations. Am. J. Drug Discov. Develop., 2: 17-31.

Bishnoi, M., C.S. Patil, A. Kumar and S.K. Kulkarni, 2005. Protective effect of nimesulide (COX inhibitor), AKBA (5-LOX inhibitor) and their combination in aging associated abnormalities in mice. Meth. Find. Exp. Clin. Pharmacol., 27: 465-470.

- Bjarnason, I. and B. Thjodleifsson, 1999. Gastrointestinal toxicity of non-steroidal antiinflammatory drugs: The effect of nimesulide compared with naproxen on the human gastrointestinal tract. Rheumatology, 38: 24-32.
- Cui, F., K. Shi, L. Zhang, A. Tao and Y. Kawashima, 2006. Biodegradable nanoparticles loaded with insulin-phospholipid complex for oral delivery: Preparation, *in vitro* characterization and *in vivo* evaluation. J. Control. Release, 114: 242-250.
- Davis, R. and R.N. Brogden, 1994. Nimesulide. An update of its pharmacodynamic and pharmacokinetic properties and therapeutic efficacy. Drugs, 48: 431-454.
- Ferrari, P.G., U. Zanetti, F. Scalvini, D. Rossi and I. Scaricabarozzi, 1993. A controlled clinical study of the efficacy and tolerability of nimesulide vs naproxen in maxillo-facial surgery. Drugs, 46: 171-173.
- Goddard, P.J., Y.C. Kao and L.M. Lichtenberger, 1990. Luminal surface hydrophobicity of canine gastric mucosa is dependent on a surface mucous gel. Gastrointerology, 98: 361-370.
- Khazaeinia, T. and F. Jamali, 2003. A comparison of gastrointestinal permeability induced by diclofenac-phospholipid complex with diclofenac acid and its sodium salt. J. Pharm. Pharm. Sci., 6: 352-359.
- Kidd, P.M., 1996. Phosphatidylcholine: A superior protectant against liver damage. Altern. Med. Rev., 1: 258-274.
- Kidd, P.M., 2002. Phosphatidylcholine. (Monograph). In: Alternative Medicine Review Monographs, Czap, K. (Ed). Thorne Research Inc, Dover, ID, ISBN-13: 978-0972581509, pp: 310-315.
- Kumar, M., M. Ahuja and S.K.Sharma, 2008. Hepatoprotective study of curcumin-soya lecithin complex. Sci. Pharm., 76: 761-774.
- Li, Y., D.J. Yang, S.L. Chen, S.B. Chen and A.S.C. Chan, 2008. Comparative physicochemical characterization of phospholipids complex of puerarin formulated by conventional and supercritical methods. Pharm. Res., 25: 563-577.
- Lichtenberger, L.M., L.A. Graziani, E.J. Dial, B.D. Butler and B.A. Hills, 1983. Role of surface active phospholipids in gastric cytoprotection. Science, 219: 1327-1329.
- Maiti, K., K. Mukherjee, A. Gantait, B.P. Saha and P.K. Mukherjee, 2007. Curcumin-phospholipid complex: Preparation, therapeutic evaluation and pharmacokinetic study in rats. Int. J. Pharm., 330: 155-163.
- Ozkan, Y., T. Atay, N. Dikmen, A. Isimer and H.Y. Aboul-Enein, 2000. Improvement of water solubility and in vitro dissolution rate of gliclazide by complexation with β-cyclodextrin. Pharm. Acta Helv., 74: 365-370.
- Perrut, M., J. Jung and F. Leboeuf, 2005. Enhancement of dissolution rate of poorly-soluble active ingredients by supercritical fluid processes: Part I: Micronization of neat particles. Int. J. Pharm., 288: 3-10.
- Pose-Vilarnovo, B., I. Perdomo-Lopez, M. Echezarreta-Lopez, P. Schroth-Pardo, E. Estrada and J.J. Torres-Labandeira, 2001. Improvement of water solubility of sulfamethizole through its complexation with β- and hydroxypropyl-β-cyclodextrin: Characterization of the interaction in solution and in solid state. Eur. J. Pharm. Sci., 13: 325-331.
- Pralhad, T. and K. Rajendrakumar, 2004. Study of freeze-dried quercetin-cyclodextrin binary systems by DSC, FT-IR, X-ray diffraction and SEM analysis. J. Pharm. Biomed. Anal., 34: 333-339.

- Rawat, S. and S.K. Jain, 2004. Solubility enhancement of celecoxib using β-cyclodextrin inclusion complexes. Eur. J. Pharm. Biopharm., 57: 263-267.
- Sajeesh, S. and C.P. Sharma, 2006. Cyclodextrin-insulin complex encapsulated polymethacrylic acid based nanoparticles for oral insulin delivery. Int. J. Pharm., 325: 147-154.
- Semalty, A., M. Semalty, B.S. Rawat, D. Singh and M.S.M. Rawat, 2009a. Pharmacosomes: The lipid-based new drug delivery system. Expert Opin. Drug Delivery, 6: 599-612.
- Semalty, A., M. Semalty, D. Singh and M.S. Rawat, 2009b. Development and physicochemical evaluation of pharmacosomes of diclofenac. Acta Pharm., 59: 335-344.
- Semalty, A., M. Semalty, B.S. Rawat, D. Singh and M.S.M. Rawat, 2010a. Development and evaluation of pharmacosomes of aceclofenac. Indian J. Pharm. Sci., 72: 576-581.
- Semalty, A., M. Semalty, D. Singh and M.S.M. Rawat, 2010b. Preparation and characterization of phospholipid complexes of naringenin for effective drug delivery. J. Inclusion Phenomena Macrocyclic Chem., 67: 253-260.
- Semalty, A., M. Semalty, M.S.M. Rawat and F. Franceschi, 2010c. Supramolecular phospholipids-polyphenolics interactions: The PHYTOSOME' strategy to improve the bioavailability of phytochemicals. Fitoterapia, 81: 306-314.
- Semalty, A., M. Semalty and M.S.M. Rawat, 2011. Essentials of Pharmaceutical Technology. PharmaMed Press, Hyderabad, India, ISBN 13: 9789381075340, pp: 4-5.
- Semalty, A., M. Semalty, D. Singh and M.S.M. Rawat, 2012. Phyto-phospholipid complex of catechin in value added herbal drug delivery. J. Inclusion Phenom. Macrocyclic Chem., 73: 377-386.
- Semalty, M., M. Panchpuri, D. Singh and A. Semalty, 2013a. Cyclodextrin inclusion complex of racecadotril: Effect of drug-ß-cyclodextrin ratio and the method of complexation. Curr. Drug Discovery Technol.
- Semalty, A., Y.S. Tanwar and M. Semalty, 2013b. Preparation and characterization of cyclodextrin inclusion complex of naringenin and critical comparison with phospholipid complexation for improving solubility and dissolution. J. Therm. Anal. Calorim. 10.1007/s10973-013-3463-y
- Singh, D., M.S.M. Rawat, A. Semalty and M. Semalty, 2011. Gallic acid-phospholipid complex: Drug incorporation and physicochemical characterization. Lett. Drug Design Discovery, 8: 284-291.
- Singh, D., M.S.M. Rawat, A. Semalty and M. Semalty, 2012a. Quercetin-phospholipid complex: An amorphous pharmaceutical system in herbal drug delivery. Curr. Drug Discovery Technol., 9: 17-24.
- Singh, D., M.S.M. Rawat, A. Semalty and M. Semalty, 2012b. Emodin-phospholipid complex. J. Therm. Anal. Calorim., 108: 289-298.
- Singh, D., M.S.M. Rawat, A. Semalty and M. Semalty, 2013. Chrysophanol-phospholipid complex. J. Therm. Anal. Calorim., 111: 2069-2077.
- Singla, A.K., M. Chawla and A. Singh, 2000. Nimesulide: Some pharmaceutical and pharmacological aspects-an update. J. Pharm. Pharmacol., 52: 467-486.
- Tan, H.H., W.M. Ong, S.H. Lai and W.C. Chow, 2007. Nimesulide-induced hepatotoxicity and fatal hepatic failure. Singapore Med. J., 48: 582-585.
- Wells, J., 2006. Pharmaceutical Preformulation: The Physicochemical Properties of Drug Substances. In: Pharmaceutics: The Science and Dosage Form Design, Aulton, M.E. (Ed.). 2nd Edn., Churchil Livingstone, Edinburg, pp: 113-116.
- Xiao, Y., Y. Song, Z. Chen and Q. Ping, 2006. The preparation of silybin-phospholipid complex and the study on its pharmacokinetics in rats. Int. J. Pharma., 307: 77-82.