



American Journal of **Food Technology**

ISSN 1557-4571



Academic
Journals Inc.

www.academicjournals.com

Review Article

Calotropis procera Extract for Wagashi Cheese Making in West Africa: A Review of Applications and Perspectives for Other Plant Coagulants Utilization

¹Mahamadou Elhadji Gounga, ²Rayanatou Issa Ado, ³Mamadou Lamarana Souaré and ⁴Salamatou Sow

¹Faculty of Agronomy and Environmental Sciences, Dan Dicko Dankoulodo University of Maradi, Po Box 465 ADS Maradi, Niger

²World Food Programme, Maradi Sub-Office, Maradi, Niger

³Department of Food Technology and Control, Higher Institute of Science and Veterinary Medicine of Dalaba, Dalaba, Guinea

⁴Department of Linguistics and Language Sciences, Faculty of Arts and Human Sciences, Abdou Moumouni University of Niamey, Niamey, Niger

Abstract

Plant extracts are used for centuries in traditional cheese manufacturing worldwide. This paper proposed to review the use of *Calotropis procera* extract in wagashi cheese manufacturing and the opportunities that are studied. After a brief reminder of the socioeconomic importance of the wagashi cheese for West African countries, the review discusses its biochemical composition and microbiological characteristics. The *Calotropis procera* enzymes and their proteolytic activities were described with a special attention paid on their caseinolytic and milk clotting properties. The microstructure and texture of milk-coagulated curds by *Calotropis procera* extracts we presented and discussed from recent studies.

Key words: *Calotropis procera*, plant extracts, wagashi cheese, biochemical composition, proteolytic activities, milk clotting, West Africa

Citation: Gounga, M.E., R. Issa Ado, M.L. Souaré and S. Sow, 2024. *Calotropis procera* extract for wagashi cheese making in West Africa: A review of applications and perspectives for other plant coagulants utilization. Am. J. Food Technol., 19: 1-14.

Corresponding Author: Mahamadou Elhadji Gounga, Faculty of Agronomy and Environmental Sciences, Dan Dicko Dankoulodo University of Maradi, Po Box 465 ADS Maradi, Niger

Copyright: © 2024 Mahamadou Elhadji Gounga *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Milk is a highly demanding food, due to its high contents in water, proteins and lactose. In West Africa, livestock breeder's communities essentially consume fresh milk. Processes such as fermentation to make curd milk, churning to make butter and coagulation to make cheese, are mostly used to process the milk production in order to obtain products of long shelf life. Cheese is an excellent source of protein, fat and minerals such as calcium, magnesium and sodium^{1,2}. Coagulation of milk is an important step in cheese making that determines the success of the process and the quality of the product³.

Calf rennet is the main coagulant use for cheese manufacturing. However, milk clotting enzymes of microbial and plant origin exist. The plant enzymatic extracts are used in the manufacture of many artisanal cheeses around the world (Table 1). In some countries, extracts of *Cynara cardunculus* flowers are used to make traditional cheeses as described in literature⁴. *Gibna bayda*, the mostly consumed cheese in Soudan is traditionally manufactured by coagulating milk with *Solanum dubium* extracts⁵. *Withania coagulans* fruits extract possess the ability to clot milk and are used in India and Iran to make locally soft cheeses⁶⁻⁹.

Several works took an interest in the milk clotting activity of *Carica papaya* leaves extracts. Since then, many investigations have been carried out on milk clotting properties of various plant species used in traditional cheese manufacturing¹⁶. Innovative results have been published, showing the main interest accorded by scientists to the milk clotting properties of plant extracts^{7,8,21}.

In Africa, plant extracts easily replace calf rennet in traditional cheese manufacturing. The use of industrial rennet from animal or microbial origin is limited by supplying, financial constraints and conservation. Consequently, local cheese producers have recourse to traditional methods of calf rennet extraction or use of plant extract for milk coagulation

during traditional cheese manufacturing. In addition, the cheese manufacturing in Africa is largely dictated by traditional practices transmitted from generation to generation and the use of rudimentary equipment that did not allow industrial rennet utilization²². Therefore, local cheese producers found an alternative in the utilization of plant extracts as coagulant. In West African countries like Nigeria and Benin, the plant extract of *Calotropis procera* is used in the manufacture of the wagashi cheese^{23,24}. The plant extracts have the advantage of being available and easy to use on artisanal scale. They do not require special conservation methods or specific abilities instead of industrial rennet and often grow wild near breeding places.

The milk clotting activity of plant extracts is due to the presence of chymosin like proteases that are able to hydrolyze casein micelles in milk, causing by that, their reorganization and the formation of the curd³. The milk clotting activity of plant extracts of an enzyme depends on its source, the extraction method employed, the degree of purity and the part of the plant that is used^{25,26}. The main milk clotting enzymes isolated from plants are cysteine proteases (ficine from *Ficus carica*, papain from *Carica papaya*, calotropin from *Calotropis procera*, zingibain from *Zingiber officinale*). However, enzymes belonging to the group of serine (cucumisin from *Cucumis melo*, streblin from *Streblus asper*, dubiumin from *Solanum dubium*) and aspartic proteases (cardosins from *Cynara cardunculus*, calotropin from *Calotropis procera*) have also been highlighted in their performance of milk coagulation²⁷. The Table 2 presented the characteristics of some milk clotting enzymes from plant origin.

This paper reviews the use of *Calotropis procera* extract in the manufacture of the traditional West African cheese and briefly present last studies on the milk clotting properties of *Carica papaya*⁴⁵ leaves and *Balanites aegyptiaca* fruits²¹ that are new tracks for milk clotting extracts for traditional cheese making in West Africa.

Table 1: Plant extracts used for cheese making

Plants	Family	Samples	Milk	Cheese	Country	References
<i>Solanum dubium</i>	Solanaceae	Leaves fruits	Cow milk	<i>Gibna bayda</i>	Sudan	Kheir <i>et al.</i> ⁵
<i>Cynara cardunculus</i>	Asteraceae	Flowers	Sheep milk	Torta del Casar	Spain	Sousa and Malcata ¹⁰
				Serra da Estrela	Portugal	Silva and Malcata ¹¹
<i>Cynara scolymus</i>	Asteraceae	Flowers	Buffalo milk	Tallaga cheese	Egypt	El-Kholy <i>et al.</i> ¹²
				Jeben	Algeria	Nouani <i>et al.</i> ¹³
<i>Zingiber officinalis</i>	Zingiberaceae	Rhizome	Cow milk	Jiang zhuang nai	Japan	Hashim <i>et al.</i> ¹⁴
			Buffalo milk	Jiangzhinai		Huang <i>et al.</i> ¹⁵
<i>Calotropis procera</i>	Asclepiadaceae	Leaves	Cow milk	Warankashi	Nigeria	Aworh and Nakai ¹⁶
		Latex		Wagashi	Benin	Aworh and Muller ¹⁷
<i>Ficus carica</i>	Moraceae	Latex	Goat milk	Caciocotta	Italy	Faccia <i>et al.</i> ¹⁸
<i>Streblus asper</i>	Moraceae	Latex	Cow milk	Dadih	Malaysia	Yazid <i>et al.</i> ¹⁹
<i>Withania coagulans</i>	Solanaceae	Fruits	Sheep milk		Iran	Pezeshki <i>et al.</i> ²⁰

Table 2: Characteristics of some milk clotting proteases from plants origins

Plant	Enzyme	Type	Mass weight (KDa)	Optimal pH	Optimal temperature (°C)	References
<i>Cucumis melo</i>	Cucumisin	Serine protease	50	7.1	70	Kaneda and Tominaga ²⁸
			68.4	9	60-70	Gagaoua <i>et al.</i> ²⁹
				7.1	37	Uchikoba and Kaneda ³⁰
<i>Cucumis trigonus</i>			67	11	70	Asif-Ullah <i>et al.</i> ³¹
<i>Streblus asper</i>	Streblin	Serine protease	64	9	65	Tripathy <i>et al.</i> ³²
<i>Calotropis procera</i>	Procerain	Cysteine proteases	28.8	7-9	55-60	Dubey and Jagannadham ³³
	Procerain B		25.7	6.5-8.5	40-60	Singh <i>et al.</i> ³⁴
	Proteinase II		25	7	50	Aworh <i>et al.</i> ³⁵
	Proteinase I		30	8	50	
<i>Cynara cardunculus</i>	Cardosin A	Aspartic proteases	31+15	5.1	-	Verissimo <i>et al.</i> ³⁶
	Cardosin B		34+14	5.1	70	Heimgartner <i>et al.</i> ³⁷
<i>Cynara scolymus</i>	Cynarase A	Aspartic proteases	30-15	5	60	Llorente <i>et al.</i> ³⁸
	Cynarase B			5	70	Sidrash <i>et al.</i> ³⁹
<i>Zingiber officinalis</i>	Zingibain	Cysteine protease	36	5.5	60	Hashim <i>et al.</i> ⁴⁰
			31	5.58-5.22	70	Huang <i>et al.</i> ¹⁵
<i>Euphorbia milii</i>	Miliine	Serine protease	-	9	45	Moro <i>et al.</i> ⁴¹
<i>Euphorbia nivulia</i>	Nivuliine II	-	43.42	6-6.5	55	Badgujar and Mahajan ⁴²
<i>Opuntia ficus-indica</i>	-	Cysteine protease	23.2	5.2	70	Teixeira <i>et al.</i> ⁴³
<i>Solanum dubium</i>	Dubiumine	Serine protease	-	10	70	Kheir <i>et al.</i> ⁵
			66	11	70	Ahmed <i>et al.</i> ⁴⁴
<i>Carica papaya</i>	Papain	Cysteine protease	23.4	5-9	50	Monti <i>et al.</i> ⁴⁵
<i>Ficus carica</i>	Ficin	Cysteine protease	23	7	-	Devaraj <i>et al.</i> ⁴⁶
<i>Ficus religiosa</i>	Religiosin	Cysteine protease	43.4	8-8.5	50	Kumari <i>et al.</i> ⁴⁷
	Religiosin B	Serine protease	63	8-8.5	-	Kumari <i>et al.</i> ⁴⁸

Use of *Calotropis procera* extract for traditional wagashi manufacturing

History and socioeconomic importance of wagashi cheese

in West Africa: Wagashi is the name given to the traditional West African soft cheese produced in both savannah and forest. It is the West African cheese per excellence considering the importance of its production and the geographical area of its marketing i.e., Benin, Nigeria, Ghana, Togo and all border countries.

The name “wagashi” dates from 17th century, a period during which the Sahelian communities, i.e., Hausa, Songhay and Fulani settled in Northern Benin and adapted themselves and their activities to its weather conditions. The abundance of water and the availability of pasture all year round created the favorable conditions for a great milk production. The traditional commerce of butter and rotten milk practiced by Fulani women did not allow to flow out the overfull of milk. However, the forest populations were not major consumers of milk products because of their eating habits and frequent allergies to lactose. Pastoralists therefore had to find a way to sell their overfull of milk. It is in this context that wagashi was borne. Its manufacture was inspired by a Songhay knowledge as the Songhay of Gao and Tombouctou areas were already manufacturing a traditional soft cheese named “gassi” or “gashi”. The “gassi” cheese was manufactured with goat milk using animal rennet for the milk clotting.

Wagashi was certainly produced to meet the demand of numerous Songhay living beside the Fulani since the fall of the Songhay Empire. With time and practice, the cheese manufacturing was improved and the animal rennet was replaced by *Calotropis procera* extract. Wagashi became that a known and appreciated local cheese²⁴.

The name wagashi is composed of two parts: “*waa*” which means milk in Songhay language and “*gash*” which means shuddering²⁴. The name’s composition refers to the cheese manufacturing process in which an extract of *Calotropis procera* was added to a heated milk. At the coagulation time and under the effect of heating, the milk gives an effect of shuddering during the formation of the curd. Fulani adapted the name to their vernacular language and call the cheese “*gasire*” in singular and “*gasijje*” in plural.

The trade of wagashi, even as a traditional and local product is very lucrative and its marketing chain is important and well-organized. The cheese is produced in camps by Fulani women and sold in rural markets to sedentary women who collect and resale back the cheese in large urban markets. The cheese’s trade chain is thus in such a way that sale is from rural camps to rural markets, then from rural markets to large urban centers and finally from large urban centers to neighboring countries. Therefore, through wagashi trade, the Fulani in the wagashi production areas became very rich through earning more benefits than Sahelian²⁴.



Fig. 1(a-c): (a) *Calotropis procera* latex tree, (b) Flowers, fruit and (c) Seeds

Currently, the demand for the cheese is increasing due to urban growth and modernization of its cooking methods. The promoters of Beninese gastronomy are continually making propositions to value the cheese. Thus, wagashi is now valorized in modern restaurants menus that propose new dishes like “Fulani cheese salad” and “Fulani cheese skewers”. The official tables also, integrate wagashi as a local specialty.

***Calotropis procera* plant:** *Calotropis procera*, commonly known as sodom apple, dead sea apple, swallow wort or giant milkweed, is a plant from Asclepiadaceae family widespread in West and East Africa, but also in Asia, especially in India, Saudi Arabia and Pakistan^{49,50}. The plant is a soft wood shrub, growing in arid and semi-arid areas and does not exceed 6 m high⁵¹. The leaves are opposite, large, green and covered by a white film-coat when young (Fig. 1a). The inflorescence is dense, multiflowered and appear axillary or terminal. The flowers are pentamerous, white and purple colored (Fig. 1b). The seeds are ovate, brown colored, flattened silky and overcome by a white light fluff (Fig. 1c).

Calotropis procera is a latex plant used in many traditional medicines for its therapeutically properties⁵¹. The plant extracts possess antimicrobial⁵²⁻⁵⁵, antihelminthic⁵⁶⁻⁵⁸, antipyretic⁵⁰ and anti-inflammatory activities^{59,60}. In India, the decoction of the plant is used in traditional medicine to treat dysentery, fever, rheumatism, asthma and as expectorant and purgative⁶¹. Investigations in the Kingdom of Saudi Arabia reported that the different parts of the plant, including the

roots, are commonly used to combat fever, constipation, muscular pain and spasm⁵¹. In Nigeria, local communities especially in the north, use *Calotropis procera* to treat infectious diseases, indigestion and rheumatism^{52,62}. At the opposite of these virtues, the latex of *Calotropis procera* have been reported to induce ocular toxicity⁶³, burning and throat edema when ingested and inflammation of skin and mucous membranes⁵⁶. Among all the parts of the plant, stems and roots have higher toxicity probably because of their high content in latex⁶².

Wagashi cheese

Areas of production and availability: In West Africa, there is traditionally two types of cheeses i.e., the dried one, named “Tchuku”, a specialty of nomadic Tuareg and Fulani communities and the fresh one, named “wagashi”, manufactured by Fulani and some Yoruba northern communities¹⁷. In the Tchuku process, coagulation of the milk is obtained by adding an abomasum aqueous extract to whole cow, sheep or goat milk. The Tuareg cheese basin is with Soudan, the only area in Sub Saharan Africa where the use of abomasum extract for cheese making is well mastered by populations²². In the manufacture of wagashi, a plant extract of *Calotropis procera* leaves is used as milk coagulant with whole cow milk. The curd obtained is white with a sweet milk taste (Fig. 2a). A stain with sorghum extract gives it a red color reminiscent of that of the meat that it easily replaces in many traditional dishes^{64,65} (Fig. 2b).

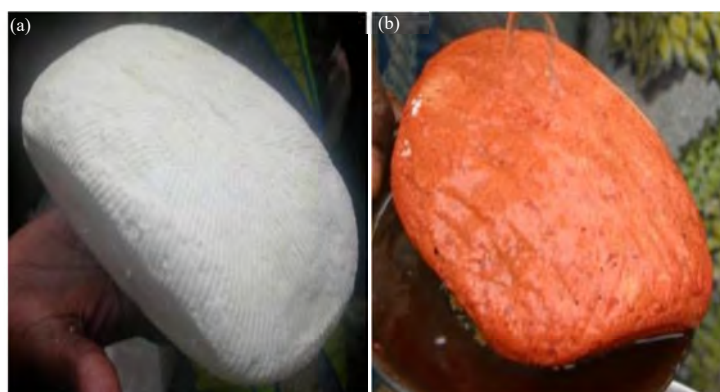


Fig. 2(a-b): (a) Fresh and (b) Colored (B) traditional wagashi

Red color is due to the coloration by the *Sorghum panicum* extract during manufacturing

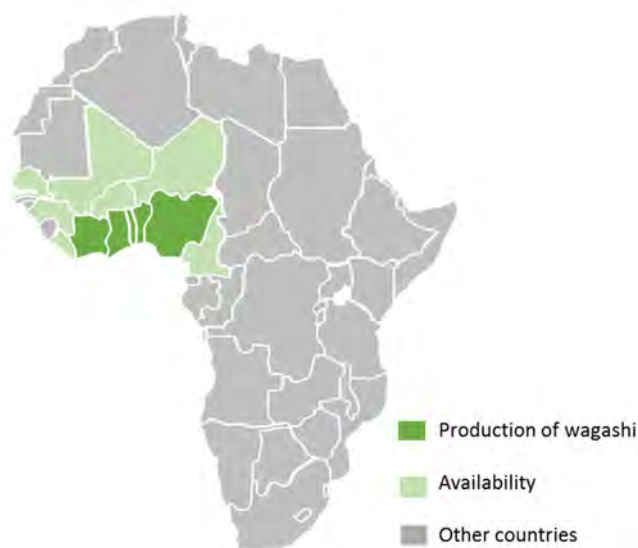


Fig. 3: Areas of production and availability of Wagashi in West Africa

The traditional West African soft cheese manufactured with *Calotropis procera* extract as coagulant is largely produced in Nigeria, Ghana, Ivory Coast, Togo and the Republic of Benin but is also available in many areas of West Africa, especially in countries that share borders with the cheese's areas of production (Fig. 3). The cheese takes practically the same name in the sub region: « Warankashi » or « Wara » in Nigeria, « Waragashi » or « Woagashi » in the Republic of Benin and « Wagashi » in Ghana⁶⁶.

Cheese processing: The West African soft cheese is usually manufactured on artisanal scale by women from Fulani and Yoruba breeders communities^{17,22}. The cheese processing is

not well standardized and use traditional equipment and empirical methods⁶⁷. The amount of leaves required for clotting a known quantity of milk is not well determined⁶⁸. However, the expertise of traditional manufacturers limits the overdoses of *Calotropis procera* extract during the cheese processing⁶⁸. All the parts of the plant excepted the roots possess the ability to clot milk and are employed for the coagulant extraction.

For traditional Wagashi processing, after milking, fresh whole cow milk is directly heated on wood fire without any standardization. The extraction of the coagulant is made by grinding young leaves of *Calotropis procera* in a little amount of milk or water and filtering. The filtrate obtained is

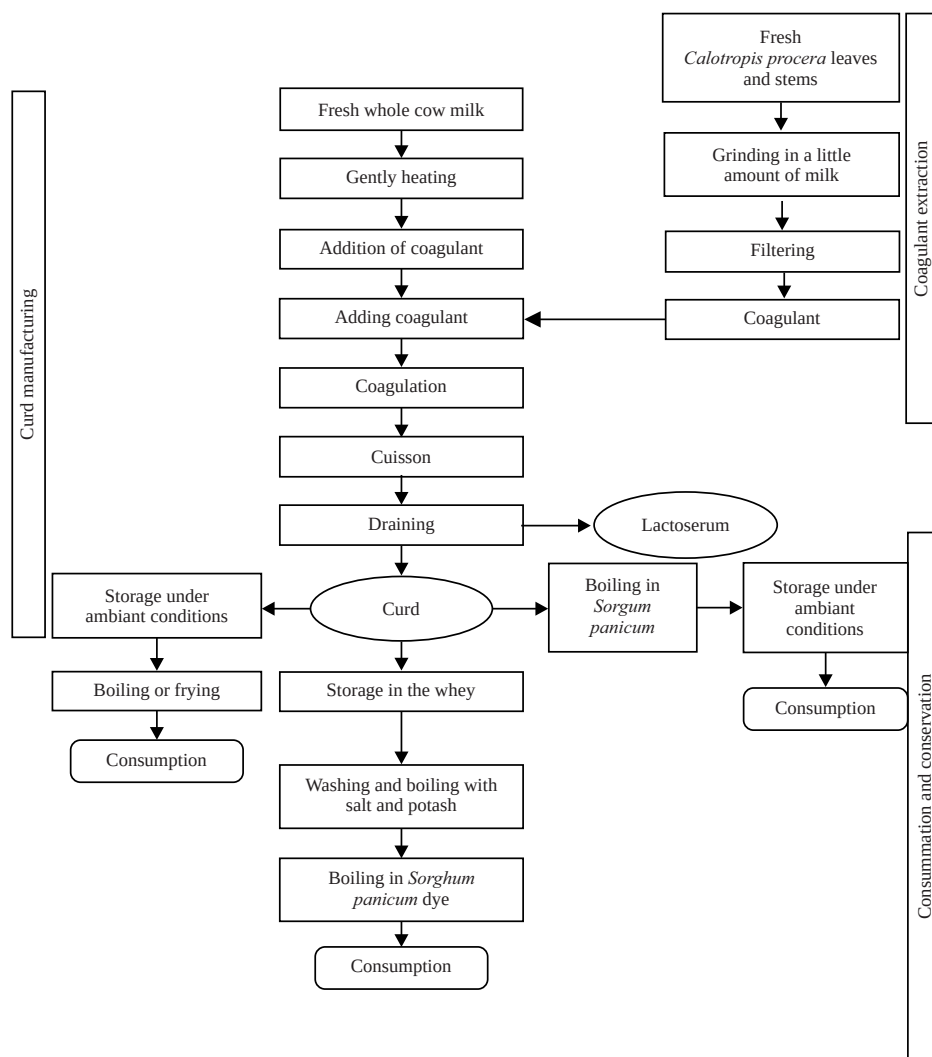


Fig. 4: Wagashi cheese traditional processing

incorporated to a whole cow milk heated on wood fire as described in our previous work. Then the heating continues by increasing fire intensity, until the separation of the curd and the whey. This results in a clear yellowish or greenish whey and crumbly curd with a sweet and creamy taste. The curd obtained, without being pressed, is drained for hours in different types of containers^{22,69}. The operation is completed after obtaining a cheese with good physico-chemical and rheological properties. The traditional processing of Wagashi cheese was described in Fig. 4. A yield of about 1 kg of cheese for 5 liters of whole cow milk is classically obtained⁷⁰. The cheese pieces are stored in the whey collected from the cheese making operation. The cheese is sometimes boiled, salted and colored with red sorghum (*Sorghum panicum*) extract that gives to the product a red color comparable to that of meat⁶⁴. The coloring operation provides a protective

cheese crust to wagashi and enhances its shelf life for few days.

Consumption and nutritional composition: The wagashi cheese is locally appreciated and consumed as is or incorporated to traditional into traditional food recipes. In rural areas, it is becoming the main supplier of proteins of animal origin, thus replacing meat or fish⁷¹. Wagashi cheese is consumed fresh, within 2 or 3 days after manufacture without ripening^{70,72}. Wagashi easily finds its place in numerous traditional sauces to replace meat or fish or in combination with them. It can also be fried and eaten as fried cake or sandwich filling⁷³. Furthermore, the West African soft cheese also finds way in the preparation of celebration and official receptions tables. This increases the local demand of the cheese.

The nutritional evaluation has revealed variable contents in protein, lipid and minerals. These variations can be attributed to the cheese processing method, the quality of the milk used, the storage period and the analytical methods used⁶⁸. As the cheese processing is not standardized, its biochemical and nutritional composition varies a lot. For wagashi samples collected from local markets in Nigeria, the protein and fat contents varies from 20.13 and 4.10% to 37.8 and 39%, respectively^{68,74}. Ojedapo *et al.*⁷⁵ have proved that salting increases the total solid content of the cheese and extend its shelf life. All the minerals contained in milk are present in the cheese but their concentrations vary also from one sample to another. In spite of these variations, wagashi can be considered as a dietary supplement especially for populations whose supply is poor in protein, minerals and probiotics^{70,76}. On the other hand, Mahami *et al.*⁷⁷ and Badmos *et al.*⁷⁸ reported that *Moringa oleifera* extracts supplementation during wagashi manufacturing enhanced the protein, fat and mineral contents of the cheese from 14.50, 28.15 and 0.87% to 18.50, 34.04 and 0.94%, respectively.

However, as for many other plant proteases, the great inconvenience of *Calotropis procera* extract is its proteolytic activity which leads to a bitter taste in the cheese especially in its fresh state^{76,79}. Fortunately, the fermentation that occurs within 24 to 48 hrs after the cheese manufacturing confers a sweet acidic taste that characterizes the cheese and hides the bitter taste when present.

Microbial quality and conservation: Fresh wagashi has a very short shelf life and can't be stored more than 3 days under ambient conditions as traditionally done^{67,70-81}. In the traditional conditions of storage, fermentation occurs quickly, probably due to the action of lactic bacteria⁸⁰ and grants an acidic taste to the cheese.

Microbial analysis of the cheese as reported by Belewu *et al.*⁷⁰ revealed the presence of many fungi in both fresh and boiled-colored wagashi cheeses. Alalade and Adeneye reported a coliform bacterial count of about 475.75 CFU/g in cheese samples collected from local sellers in Nigeria⁶⁸. Other studies showed the presence of pathogenic microorganisms (*E. coli* O157:H7 and *Listeria monocytogenes*) with counts exceeding the international standards in marketed wagashi cheese⁸². Poor hygienic conditions, the warm climate in which the cheese is stored and the vulnerability of dairy products to microbial growth are factors that explain the higher levels of the milk and cheese contamination⁸³. At the artisanal scale, frying in vegetable oil

or boiling with salt and potash followed by coloration with sorghum (*Sorghum vulgare*) extract are used to extend the shelf life of the cheese⁷². The products once treated can be stored at ambient temperature for 2 to 3 months without any modification of their organoleptic properties.

Natural and chemical preservatives have been tested in order to reduce the microbial load and extend the shelf life of wagashi. Belewu *et al.*⁸⁴ revealed the effect of eucalyptus and lemon grass oil. The lemon grass has proved its positive effect on reducing the microbial growth and thus enhancing the quality and the shelf life of the cheese. Ashayé *et al.*⁸¹ have used *Aframomum danielli* and succeeded in extending the shelf life of the cheese from 3 to 6 days under cold temperature without important modifications in the organoleptic characteristics. Salting, which is traditionally used to extend the shelf life and improve the organoleptic characteristics of wagashi cheese has also proved it's effective on the yield and nutritional composition of the cheese⁸⁰. Alalade and Adeneye⁶⁸ studied the short-term frozen conditions and found that this method of preservation reduced the coliform microflora amount in the cheese without any modification to the nutritional value. Freezing can thus extend the shelf life of the cheese and make it safer for consumption.

Texture and microstructure: The microscopic observation by scanning electron microscopy of curds coagulated by the plant extract of *Calotropis procera* have revealed spherical particles of about 200 nm size that is in accordance with the micellar size^{85,86} (Fig. 5). In spite of the proteolysis causing the destabilization of the micellar balance and the formation of the curd, casein micelles that had not been destroyed were aggregated together to form a network during the coagulation of milk by the plant extract⁸⁷. The comparison of such curds with those coagulated with chymosin in similar conditions highlights slight differences. The network appeared more compact in the curd obtained with *Calotropis procera* latex compared to that coagulated with chymosin, in which black spaces corresponding to whey pockets were frequently observed, with a more regular gel organization than in the case of *Calotropis procera* extract⁸⁵.

The texture profile analysis and crush test performed on curds coagulated by a plant extract of *Calotropis procera* have revealed the values of hardness, cohesiveness and elasticity of 5.6 and 1.2 N, respectively⁸⁸. Ado *et al.*⁸⁷ have demonstrated that homogenization of processed milks increases the curds hardness and decreases their adhesiveness.

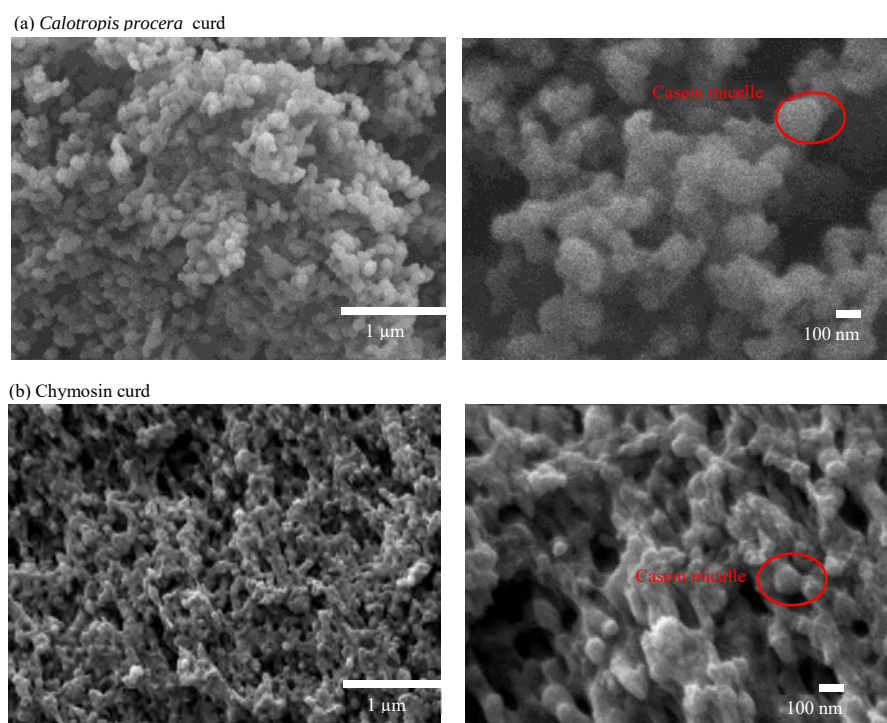


Fig. 5(a-b): Scanning electron micrographs of the milk coagulum obtained with (a) *Calotropis procera* extract and (b) Chymosin curd

Proteases from *Calotropis procera* latex: It has been extracted, purified and characterized two proteases from the latex of *Calotropis procera*. The first one (Procerain), is a cysteine protease (28.8 KDa) with an optimum pH and temperature of 8 and 60, respectively³³. The second one is cysteine protease (Procerain B) with different characteristics from those of the first one⁸⁹. At the same time, the presence of three novel cysteine proteases of 26.213, 26.133 and 25.086 KDa distinct from procerain and procerain B has been reported (Ramos *et al.*⁹⁰). The milk clotting activity of *Calotropis procera* extract is probably due to the joint action of all these proteases and this explain the great proteolytic activity of the plant extract⁸⁵.

Cysteine proteases are characterized by a dyad of cysteine and histidine in their catalytic site^{27,91}. In plants, cysteine proteases are involved in protein maturation, degradation and rebuilding in response to various external stimuli such as stress⁹¹⁻⁹³. Oseni and Ekperegini²⁵ demonstrated that cysteine and calcium ions addition enhance the proteolytic activity of the enzyme which is conserved over a wide range of pH and temperature^{25,33}. The proteolytic activity of the two characterized proteases is effective on a variety of certain intact and denaturated proteins^{35,94}. Rayanatou *et al.*⁸⁵ have

reported that the mechanism causing milk coagulation by the leaves extract of *Calotropis procera* was a nonspecific hydrolysis of κ casein which probably leads to the destabilization of casein micelles and their rearrangement in gel. Authors have also highlighted the hydrolysis of α_{s1} , α_{s2} and β caseins but assumed that these mechanisms were probably not at the origin of the milk coagulation property of the plant extract and would be due to the presence of a large number of proteases in the extract.

New tracks for traditional cheese making in West Africa:

***Carica papaya* leaves:** *Carica papaya*, commonly named pawpaw or papaya is an evergreen tree of 3 to 10 m in height, which develops in tropical and equatorial regions. It is essentially cultivated for its fruits but additionally for its latex and leaves which are reported to have many nutritional and therapeutic properties⁹⁵. Papaya grows normally as a single stemmed tree. The leaves are green, large and palmately lobed⁹⁶. The flowers appear in the axils of leaves. Papaya fruits are big and oval with a central seed cavity that contains a large quantity of seeds of black color⁹⁵. The flesh of the fruit, initially green change to yellow or orange at the maturity of the fruit⁹⁶.

All the parts of the plant are used for treatment of many diseases⁹⁷⁻⁹⁹. The leaves, fruits and seeds have shown anti-bacterial activity^{100,101}. The latex and leaves of the plant have been reported to possess anti-helminthic activity^{102,103} and the dried seeds have shown effectiveness against human intestinal parasitosis¹⁰⁴. In many traditional medicines, the plant is used in the treatment of infectious diseases such as malaria, dengue and jaundice¹⁰⁵. The papaya plant, more especially young leaves, exhibit anti-oxidant¹⁰⁶ and anti-tumor¹⁰⁷ activities. Papaya fruits are rich in dietary fiber, provitamin A, proteins and minerals⁹⁵. This rich composition underlies a large nutraceutical and pharmacological properties such as digestive, diuretic, anti-hypertensive and hepatoprotective^{97,99}.

The latex of *Carica papaya* contains many enzymes but the mostly studied is papain, a proteolytic enzyme widely exploited in the industrial sector¹⁰⁸. Papain (EC 3.4.22.2) is a cysteine protease of 23.4 KDa, active in a wide range of pH (5-9) and temperature (40-90 °C)¹⁰⁹. Papain exhibit a high reaction speed, about thrice that of *Calotropis procera* extract¹⁷. In the food industry, papain is essentially used for tenderizing meat and clarifying beer¹⁰⁹.

In Nigeria, *Carica papaya* leaves are sometimes used as *Calotropis procera* surrogate to make traditional wagashi cheese⁷⁹. However, *Calotropis procera* extracts were preferred as they give a white curd with a sweet milk taste instead of *Carica papaya* leaves that slightly colored the curd in green and give them a pronounced vegetable taste⁸³. Recently, essays have been conducted to optimize the proteolytic and milk-clotting activity of *Carica papaya* leaves extract¹¹⁰. More so, clotting and proteolytic activities are investigated and discussed in relation to their application on the different cheesemaking products¹¹¹. Diouf *et al.*¹¹² have used different concentrations of fresh *Carica papaya* leaves extracts for clotting raw cow milk. The results obtained have shown that milk clotting times decreased with increasing concentration of *Carica papaya* leaves extracts i.e., the shortest time was observed with the highest concentration but the curd obtained exhibited color (greenish) and taste (bitterness) defaults¹¹². Adetunji and Salawu⁷⁹ have shown that *Carica papaya* leaves extract can easily replace *Calotropis procera* which is commonly used for traditional cheese making in West Africa. The product, even if slightly less rich in protein and fat than the traditional wagashi, presents an interesting content in minerals such as Zinc and Copper⁷⁹. Akinyolé and Adewumi¹¹³ have compared *Carica papaya* and *Calotropis procera* milk clotting activities using sheep milk and shown that *Carica papaya* leaves extract is more effective than *Calotropis procera*. This suggested that this plant enzyme has

more affinity for sheep milk than cow milk, which open an opportunity for production of new traditional products manufacturing.

***Balanites aegyptiaca* fruits:** *Balanites aegyptiaca*, also known as desert palm, wild date or soap berry, is an indigenous tree growing widely in arid and semi-arid regions of African countries¹¹⁴. It is a multipurpose tree offering wood, food, medicine and cosmetics to many African populations^{21,98,115,116}. *Balanites aegyptiaca* is a multibranched, spiny shrub or tree growing up to 10 m tall^{115,116}. The leaves are small and green, alternative and two-foliate¹¹⁶. The trunk is covered by green or dark brown barks according to the age of the tree. Fruits are drupes, rather long, green when young but turned to yellow when mature. Flowers are small, greenish white and appear axillary or fascicle¹¹⁷.

Balanites aegyptiaca tree is well known by West African populations who use all the parts of the plant more for medicine than food. The fruits are sold in local markets and consumed as sweets¹¹⁵. In countries like Nigeria and Niger, young leaves and flowers of the plant are cooked and mixed with peanuts past to make a delicious local salad¹¹⁷. The kernel produces an edible oil, which is also used by local women as cosmetic to remove dark spots on the facial skin or fight against hair loss.

Next to these nutritional purposes, *Balanites aegyptiaca* is used in many traditional medicines. The plant exhibits anti-microbial activity¹¹⁸⁻¹²⁰ even on antibiotic resistant germs^{118,121}. *Balanites aegyptiaca* extracts also show anti-helminthic, larvicidal¹²², anti-inflammatory¹²³, anti-oxidant¹²⁰, anti-anaphylactic⁹⁸ and anti-carcinogenic¹²⁴ activities. These biological evidences account for the traditional use of the plant as laxative, antifungal and antimicrobial agent¹²⁴ for skin diseases, as hepatoprotective, purgative and in the treatment of characteristics pathologies such as jaundice, yellow fever, diarrhea, hemorrhoid, syphilis and cough^{98,116}.

In Cameroon, the fruits extracts of *Balanites aegyptiaca* are traditionally used to thicken a gruel made with flour and fresh cow milk²¹. This encouraged local researchers to conduct further investigations on the milk clotting activity of the fruits. To solve some processing problems, Beka *et al.*²¹ provided additional steps for disinfecting fruit samples and pasteurizing milk to reduce the bio burn of the final product, clarifying the extract to improve the coloration of the product, extract concentration to optimize the clotting time and flavoring with *Zingiber officinalis* and *Ocimum gratissimum* to palliate the bitter taste of the cheese. Two proteases with milk clotting activity have been isolated and characterized i.e. a serine and an aspartic protease, that are responsible of the milk clotting activity of the fruits²¹.

Aspartic proteases, also known as acidic proteases are characterized by two amino acid aspartic in their catalytic site⁹¹. *In vitro* experiences show that aspartic proteases in plants are involved in protein synthesis or degradation, thus playing certainly a role in plant senescence, stress response, apoptosis and reproduction^{91,93}.

Serine proteases contains a reactive serine residue located in their active site that is essential for the function of the enzyme. This active site contains also one Asp and one His amino acid which join the Serine residue to form a "catalytic triad"⁹². Serine proteases achieve quite the same roles in plants as aspartic proteases⁹³.

CONCLUSION

Calotropis procera leaves extract is the main plant coagulant used to make cheese in West Africa. Cheese making in this part of the world is largely dictated by the availability of milk and traditional empirical practices inherited from generation to generation. In general, African traditional foods and their processing methods have received limited researches. However, investigations conducted on traditional products are essential to provide information to how their production and preservation can be improved and thereby contribute to improve food and nutritional security and reduce poverty in African countries.

SIGNIFICANCE STATEMENT

In West Africa, cheese is becoming more and more part of the dietary habits of a large part of the population. The wagashi cheese replace fish or meat in numerous traditional food recipes. The use of industrial rennet from animal or microbial origin in cheese processing is limited by supplying, financial constraints and conservation. The plant extracts have the advantage of being available and easy to use on artisanal scale. This study reviews the use of *Calotropis procera* extract in the manufacture of the traditional West African cheese and present last studies on the milk clotting properties of other plant leaves and fruits that are new tracks for milk clotting extracts for traditional cheese making in West Africa.

REFERENCES

1. Fox, P.F., P.L. McSweeney, T.M. Cogan and T.P. Guinee, 2004. Cheese: Chemistry, Physics and Microbiology: General Aspects. 3rd Edn., Academic Press, New York, ISBN: 9780080500935, Pages: 640.
2. Stocco, G., A. Summer, C. Cipolat-Gotet, M. Malacarne, A. Cecchinato, N. Amalfitano and G. Bittante, 2021. The mineral profile affects the coagulation pattern and cheese-making efficiency of bovine milk. J. Dairy Sci., 104: 8439-8453.
3. Legg, A.K., A.J. Carr, R.J. Bennett and K.A. Johnston, 2017. General Aspects of Cheese Technology. In: Cheese: Chemistry, Physics and Microbiology, McSweeney, P.L.H., P.F. Fox, P.D. Cotter and D.W. Everett (Eds.), Academic Press, United States, ISBN: 978-0-12-417012-4, pp: 643-675.
4. Roserio, L.B., M. Barbosa, J.M. Ames and R.A. Wilbey 2003. Cheesemaking with vegetable coagulants-The use of *Cynara* L. for the production of ovine milk cheeses. Int. J. Dairy Technol., 56: 76-85.
5. Kheir, S.E.O., O.A.O.E. Owni and M.O.M. Abdalla, 2011. Comparison of quality of Sudanese white cheese (*Gibna bayda*) manufactured with *Solanum dubium* fruit extract and rennet. Pak. J. Nutr., 10: 106-111.
6. Reyhaneh, S., M. Javad and A.J. Mohammad, 2014. The effect of *Withania coagulans* as a coagulant on the quality and sensorial properties of tofu. Afr. J. Food Sci., 8: 112-115.
7. Salehi, M., M.R. Aghamaali, R.H. Sajedi, S.M. Asghari and E. Jorjani, 2017. Purification and characterization of a milk-clotting aspartic protease from *Withania coagulans* fruit. Int. J. Biol. Macromol., 98: 847-854.
8. Qazalbash, M.A., T. Masud, S. Sammi, R.S. Khan and A. Latif, 2018. Effect of different storage conditions on coagulating properties and cheese quality of *Withania coagulans* extract. Int. J. Dairy Technol., 71: 654-662.
9. Jooyandeh, H., A. Ghasemi, M. Hojjati and B. Nasehi, 2020. Utilization of *Withania coagulans* extract as rennet replacer on color and physicochemical characteristics of ultrafiltered Iranian white cheese. FSCT, 17: 1-13.
10. Sousa, M.J. and F.X. Malcata, 2002. Advances in the role of a plant coagulant (*Cynara cardunculus*) *in vitro* and during ripening of cheeses from several milk species. Lait, 82: 151-170.
11. Silva, S.V. and F.X. Malcata, 1998. Proteolysis of ovine caseins by cardosin A, an aspartic acid proteinase from *Cynara cardunculus* L. Lait, 78: 513-519.
12. El-Kholy, W., A.B. Abd El-Khalek, S.H.S. Mohamed, M.T. Fouad and J.M. Kassem, 2016. Tallaga cheese as a new functional dairy product. Am. J. Food Technol., 11: 182-192.
13. Nouani, A., E. Dako, A. Morsli, N. Belhamiche, S. Belbraouet, M.M. Bellal and A. Dadie, 2009. Characterization of the purified coagulant extracts derived from artichoke flowers (*Cynara scolymus*) and from the fig tree latex (*Ficus carica*) in light of their use in the manufacture of traditional cheeses in Algeria. J. Food Technol., 7: 20-29.
14. Hashim, M.M., M. Dong, M.F. Iqbal, W. Li and X. Chen, 2011. Ginger protease used as coagulant enhances the proteolysis and sensory quality of Peshawari cheese compared to calf rennet. Dairy Sci. Technol., 91: 431-440.

15. Huang, X.W., L.J. Chen, Y.B. Luo, H.Y. Guo and F.Z. Ren, 2011. Purification, characterization and milk coagulating properties of ginger proteases. *J. Dairy Sci.*, 94: 2259-2269.
16. Aworh, O.C. and S. Nakai, 1986. Extraction of milk clotting enzyme from sodom apple (*Calotropis procera*). *J. Food Sci.*, 51: 1569-1570.
17. Aworh, O.C. and H.G. Muller, 1987. Cheese-making properties of vegetable rennet from sodom apple (*Calotropis procera*). *Food Chem.*, 26: 71-79.
18. Faccia, M., G. Picariello, A. Trani, P. Loizzo, G. Gambacorta, C. Lamacchia and A. di Luccia, 2012. Proteolysis of Caciocotta cheese made from goat milk coagulated with caprigin (*Ficus carica sylvestris*) or calf rennet. *Eur. Food Res. Technol.*, 234: 527-533.
19. Yazid, A.M., H.M. Fong, A.B. Sipat, M. Shuhaimi and Y.M.A. Idris, 1999. Proteolysis of milk and casein fractions by *Streblus asper* (Kesinai) extract. *Pak. J. Biol. Sci.*, 2: 654-659.
20. Pezeshki, A., J. Hesari, A.A. Zonoz and B. Ghambarzadeh, 2011. Influence of *Withania coagulans* protease as a vegetable rennet on proteolysis of Iranian UF white cheese. *J. Agr. Sci. Tech.*, 13: 567-576.
21. Beka, R.G., F. Krier, M. Botquin, V.D. Guiama and P. Donn *et al.*, 2014. Characterisation of a milk-clotting extract from *Balanites aegyptiaca* fruit pulp. *Int. Dairy J.*, 34: 25-31.
22. O'Connor, C.B., 1993. Traditional Cheesemaking Manual. International Livestock Centre for Africa, Addis Ababa, Ethiopia, ISBN-13: 9789290532736, Pages: 43.
23. Akinyosoye, V.O., 2007. Demand for dairy products in Nigeria: Evidence from the Nigerian. *J. Rural Econ. Dev.*, 16: 13-26.
24. Mazou, M., F.P. Tchobo, R.G. Degnon, G.A. Mensah and M.M. Soumanou, 2012. Effect of temperature and salt on the quality of waragashi cheese during storage in Benin Republic. *Afr. J. Food Sci.*, 16: 494-499.
25. Oseni, O.A. and M.M. Ekperigin, 2013. Partial characterization of proteolytic and milk clotting enzymes in sodom apple *Calotropis procera* (Ait.) R.Br. (Asclepiadaceae) plant. *Am. J. Biochem. Mol. Biol.*, 3: 256-263.
26. Pagthinathan, M., H.M. Ghazali, A.M. Yazid and H.L. Foo, 2019. Extraction, purification and characterisation of a milk-clotting protease from 'kesinai' (*Streblus asper* Lour.) leaves. *Int. Food Res. J.*, 26: 913-922.
27. Shah, M.A., S.A. Mir and M.A. Paray, 2014. Plant proteases as milk-clotting enzymes in cheesemaking: A review. *Dairy Sci. Technol.*, 94: 5-16.
28. Kaneda, M. and N. Tominaga, 1975. Isolation and characterization of a proteinase from the sarcocarp of melon fruit. *J. Biochem.*, 78: 1287-1296.
29. Gagaoua, M., F. Ziane, S.N. Rabah, N. Boucherba, A.A.K.E.H. El-Okki, A. Bouanane-Darenfed and K. Hafid, 2017. Three phase partitioning, a scalable method for the purification and recovery of cucumisin, a milk-clotting enzyme, from the juice of *Cucumis melo* var. *reticulatus*. *Int. J. Biol. Macromol.*, 102: 515-525.
30. Uchikoba, T. and M. Kaneda, 1996. Milk-clotting activity of cucumisin, a plant serine protease from melon fruit. *Appl. Biochem. Biotechnol.*, 56: 325-330.
31. Asif-Ullah, M., K.S. Kim and Y.G. Yu, 2006. Purification and characterization of a serine protease from *Cucumis trigonus* Roxburghi. *Phytochemistry*, 67: 870-875.
32. Tripathi, P., R. Tomar and M.V. Jagannadham, 2011. Purification and biochemical characterisation of a novel protease streblin. *Food Chem.*, 125: 1005-1012.
33. Dubey, V.K. and M.V. Jagannadham, 2003. Procerain, a stable cysteine protease from the latex of *Calotropis procera*. *Phytochemistry*, 62: 1057-1071.
34. Singh, A.N., A.K. Shukla, M.V. Jagannadham and V.K. Dubey, 2010. Purification of a novel cysteine protease, procerain B, from *Calotropis procera* with distinct characteristics compared to procerain. *Process Biochem.*, 45: 399-406.
35. Aworh, O.C., V. Kasche and O.O. Apampa, 1994. Purification and some properties of sodom-apple latex proteinases. *Food Chem.*, 50: 359-362.
36. Verissimo, P., C. Faro, A.J.G. Moir, Y. Lin, J. Tang and E. Pires, 1996. Purification, characterization and partial amino acid sequencing of two new aspartic proteinases from fresh flowers of *Cynara cardunculus* L. *Eur. J. Biochem.*, 235: 762-768.
37. Heimgartner, U., M. Pietrzak, R. Geertsens, P. Brodelius, A.C. da Silva Figueiredo and M.S.S. Pais, 1990. Purification and partial characterization of milk clotting proteases from flowers of *Cynara cardunculus*. *Phytochemistry*, 29: 1405-1410.
38. Llorente, B.E., C.B. Bruti, C.L. Natalucci and N.O. Caffini, 1997. Partial characterization of a milk clotting proteinase isolated from artichoke (*Cynara scolymus* L., asteraceae). *Acta Farm. Bonaerense*, 16: 37-42.
39. Sidrach, L., F. Garcia-Canovas, J. Tudela and N.J. Rodriguez-Lopez, 2005. Purification of cynarases from artichoke (*Cynara scolymus* L.): Enzymatic properties of cynarase A. *Phytochemistry*, 66: 41-49.
40. Hashim, M.M., D. Mingsheng, M.F. Iqbal and C. Xiaohong, 2011. Ginger rhizome as a potential source of milk coagulating cysteine protease. *Phytochemistry*, 72: 458-464.
41. Moro, L.P., H. Cabral, D.N. Okamoto, I. Hirata, M.A. Juliano, L. Juliano and G.O. Bonilla-Rodriguez, 2013. Characterization, subsite mapping and N-terminal sequence of miliin, a serine-protease isolated from the latex of *Euphorbia milii*. *Process Biochem.*, 48: 633-637.
42. Badgujar, S.B. and R.T. Mahajan, 2012. Comparison of cysteine proteases of four laticiferous plants and characterization of *Euphorbia nivulia* Buch.-Ham. latex glycosylated cysteine peptidase. *Indian J. Nat. Prod. Resour.*, 3: 152-160.
43. Teixeira, G., A.R. Santana, M.S. Pais and A. Clemente, 2000. Enzymes of *Opuntia ficus-indica* (L.) Miller with potential industrial applications. *Appl. Biochem. Biotechnol.*, 88: 299-312.

44. Ahmed, I.A.M., I. Morishima, E.E. Babiker and N. Mori, 2009. Dubiumin, a chymotrypsin-like serine protease from the seeds of *Solanum dubium* Fresen. *Phytochemistry*, 70: 483-491.
45. Monti, R., C.A. Basilio, H.C. Trevisan and J. Contiero, 2000. Purification of papain from fresh latex of *Carica papaya*. *Braz. Arch. Biol. Technol.*, 43: 501-507.
46. Devaraj, K.B., P.R. Kumar and V. Prakash, 2008. Purification, characterization, and solvent-induced thermal stabilization of ficin from *Ficus carica*. *J. Agric. Food Chem.*, 56: 11417-11423.
47. Kumari, M., A. Sharma and M.V. Jagannadham, 2012. Religiosin B, a milk-clotting serine protease from *Ficus religiosa*. *Food Chem.*, 131: 1295-1303.
48. Kumari, M., A. Sharma and M.V. Jagannadham, 2010. Decolorization of crude latex by activated charcoal, purification and physico-chemical characterization of religiosin, a milk-clotting serine protease from the latex of *Ficus religiosa*. *J. Agric. Food Chem.*, 58: 8027-8034.
49. Gupta, S., B. Gupta, K. Kapoor and P. Sharma, 2012. Ethnopharmacological potential of *Calotropis procera*: An overview. *Int. Res. J. Pharm.*, 3: 19-22.
50. Mossa, J.S., M. Tariq, A. Mohsin, A.M. Ageel, M.A. Al-Yahya, M.S. Al-Said and S. Rafatullah, 1991. Pharmacological studies on aerial parts of *Calotropis procera*. *Am. J. Chin. Med.*, 19: 223-231.
51. Roy, S., R. Sehgal, B.M. Padhy and V.L. Kumar, 2005. Antioxidant and protective effect of latex of *Calotropis procera* against alloxan-induced diabetes in rats. *J. Ethnopharmacol.*, 102: 470-473.
52. Kareem, S.O., I. Akpan and O.P. Ojo, 2008. Antimicrobial activities of *Calotropis procera* on selected pathogenic microorganisms. *Afr. J. Biomed. Res.*, 11: 105-110.
53. Kawo, A.H., A. Mustapha, B.A. Abdullahi, L.D. Rogo, Z.A. Gaiya and A.S. Kumurya, 2009. Phytochemical properties and antibacterial activities of the leaf and latex extracts of *Calotropis procera* (Ait.f.) Ait.f. *Bayero J. Pure Appl. Sci.*, 2: 34-40.
54. Ahmad, N., F. Anwar, S. Hameed and M.C. Boyce, 2011. Antioxidant and antimicrobial attributes of different solvent extracts from leaves and flowers of Akk [*Calotropis procera* (Ait.) Ait. F.]. *J. Med. Plants Res.*, 5: 4879-4887.
55. Saadabi, A.M.A., N.M. Ali, H.I. Mohammed, F.N. Alsafi and H.B. Mustafa, 2012. An *in vitro* antimicrobial activity of *Calotropis procera* (Ait.) R.Br. extracts on certain groups of pathogenic microorganisms. *Res. J. Med. Sci.*, 6: 13-17.
56. Kumar, V.L. and Y.M. Shivkar, 2004. Involvement of prostaglandins in inflammation induced by latex of *Calotropis procera*. *Mediators Inflammation*, 13: 151-155.
57. Shivkar, Y.M. and V.L. Kumar, 2003. Anthelmintic activity of latex of *Calotropis procera*. *Pharm. Biol.*, 41: 263-265.
58. Murti, Y., A.P. Singh and D. Pathak, 2013. *In-vitro* anthelmintic & cytotoxic potential of different extracts of *Calotropis procera* leaves. *Asian J. Pharm. Clin. Res.*, 6: 14-16.
59. Arya, S. and V.L. Kumar, 2005. Antiinflammatory efficacy of extracts of latex of *Calotropis procera* against different mediators of inflammation. *Mediators Inflammation*, 4: 228-232.
60. Babu, A.R.S. and S.S. Karki, 2011. Anti-inflammatory activity of various extracts of roots of *Calotropis procera* against different inflammation models. *Int. J. Pharm. Pharm. Sci.*, 3: 191-194.
61. Sharma, A.K., R. Kharb and R. Kaur, 2011. Pharmacognostical aspects of *Calotropis procera* (Ait.) R. Br. *Int. J. Pharm. Biol. Sci.*, 2: 480-488.
62. Mainasara, M.M., B.L. Aliero, A.A. Aliero and M. Yakubu, 2012. Phytochemical and antibacterial properties of root and leaf extracts of *Calotropis procera*. *Niger. J. Basic Appl. Sci.*, 20: 1-6.
63. Waikar, S. and V.K. Srivastava, 2015. *Calotropis* induced ocular toxicity. *Med. J. Armed Forces India*, 71: 92-94.
64. Akogou, F.U.G., H.M.W. den Besten, A.P.P. Kayode, V. Fogliano and A.R. Linnemann, 2018. Antimicrobial evaluation of red, phytoalexin-rich sorghum food biocolorant. *PLoS ONE*, Vol. 13. 10.1371/journal.pone.0194657.
65. Anihouvi, D.G.H., Y.E. Kpoclou, M.A. Massih, O.H.I. Afé and M.F. Assogba *et al*, 2019. Microbiological characteristics of smoked and smoked-dried fish processed in Benin. *Food Sci. Nutr.*, 7: 1821-1827.
66. Nyamakwere, F., G. Esposito, K. Dzama and E. Raffrenato, 2021. A review of artisanal cheese making: An African perspective. *South Afr. J. Anim. Sci.*, 51: 296-309.
67. Adetunji, V.O. and O.O. Babalobi, 2011. A comparative assessment of the nutritional contents of 'Wara' a West African soft cheese using *Calotropis procera* and *Cymbopogon citratus* as coagulants. *Afr. J. Food Agric. Nutr. Dev.*, 11: 5573-5585.
68. Alalade, O.A. and J.A. Adeneye, 2006. The effect of storage period on the chemical composition and coliform microflora of wara cheese. *Int. J. Dairy. Sci.*, 1: 126-130.
69. Fakayode, S.B., E.O. Olorunsanya, L.O.E. Nwauwa, T.M. Yusuf and O.O. Oyeleye, 2012. Economics of local cow milk products marketing in Kwara State, Nigeria. *J. Agric. Food Sci.*, 10: 35-50.
70. Belewu, M.A., K. Belewu and C.C. Nkwunonwo, 2005. Effect of biological and chemical preservatives on the shelf life of West African soft cheese. *Afr. J. Biotechnol.*, 4: 1076-1079.
71. Aboudoulaye, M.C.D. and S. Kaya, 2020. Characterisation and standardisation of Wangashi cheese production steps. *Sci. Polonorum Technologia Aliment.*, 19: 375-386.
72. Adegoke, G.O., E.N. Nse and A.O. Akanni, 1992. Effects of heat, processing time and pH on the microflora, aflatoxin content and storability of "wara", a soft white cheese. *Food Nahrung*, 36: 259-264.
73. Omotosho, O.E., G. Oboh and E.E.J. Iweala, 2011. Comparative effects of local coagulants on the nutritive value, *in vitro* multienzyme protein digestibility and sensory properties of Wara cheese. *Int. J. Dairy Sci.*, 6: 58-65.

74. Lawal, A.K. and O.M. Adediji, 2013. Nutritional and elemental analysis of warankasi (fermented milk product) sold in Lagos metropolis. *Int. Res. J. Biotechnol.*, 4: 112-116.
75. Ojedapo, L.O., G.O. Tona, S.R. Amao and J.A. Adeneye, 2014. Yield, composition and coagulation time of unsalted and salted soft cheese prepared from the milk of White Fulani cows. *Int. J. Curr. Microbiol. Appl. Sci.*, 3: 378-383.
76. Anihouvi, E.S. and H. Kesenkas, 2022. Wagashi cheese: Probiotic bacteria incorporation and significance on microbiological, physicochemical, functional and sensory properties during storage. *LWT*, Vol. 155. 10.1016/j.lwt. 2021.112933.
77. Mahami, T., F.C.K. Ocloo, S.T. Odonkor, C. Owulah and S.A. Coffie, 2012. Preliminary study on the influence of *Moringa* seed extracts supplementation on the yield and quality of cottage cheese. *Int. J. Recent Trends Sci. Technol.*, 2: 4-8.
78. Badoms, A.H.A., A.M.A. El-Imam and D.J. Ajiboye, 2014. The effect of crude leaf extracts of *Moringa oleifera* on the bacterial, nutritional and sensory properties of West African soft cheese. *Wayamba J. Anim. Sci.*, 6: 939-946.
79. Adetunji, V.O. and O.T. Salawu, 2008. West African soft cheese 'wara' processed with *Calotropis procera* and *Carica papaya*: A comparative assessment of nutritional values. *Afr. J. Biotechnol.*, 7: 3360-3362.
80. Joseph, J.K. and F.A. Akinyosoye, 1997. Comparative studies on red sorghum extract and other chemicals as preservatives for West African soft cheese. *Int. Dairy J.*, 7: 193-198.
81. Ashaye, O.A., O.O. Taiwo and G.O. Adegoke, 2006. Effect of local preservative (*Aframomum daniellii*) on the chemical and sensory properties of Warankashi. *Afr. J. Agric. Res.*, 1: 10-16.
82. Adetunji, V.O. and M.I. Arigbede, 2011. Occurrence of *E. coli* O157:H7 and *Listeria monocytogenes* and identification of hazard analysis critical control points (HACCPs) in production operations of a typical tropic cheese wara and yoghurt. *Pak. J. Nutr.*, 10: 796-804.
83. Adetunji, V.O., 2011. Effects of processing on antibiotic residues (streptomycin, penicillin-G and tetracycline) in soft cheese and yoghurt processing lines. *Pak. J. Nutr.*, 10: 792-795.
84. Belewu, M.A., A.A. El-Imam, K.D. Adeyemi and S.A. Oladunjoye, 2012. Eucalyptus oil and lemon grass oil: Effect on chemical composition and shelf-life of soft cheese. *Environ. Nat. Resour. Res.*, 2: 114-118.
85. Rayanatou, I.A., E.H.G. Mahamadou, G. Garric, M. Harel-Oger and A. Leduc *et al.*, 2017. Physico-chemical characterization of dairy gel obtained by a proteolytic extract from *Calotropis procera*-A comparison with chymosin. *Food Chem.*, 232: 405-412.
86. Horne, D.S., 2008. Casein Micelle Structure and Stability. In: *Milk Proteins*, Singh, H., M. Boland and A. Thompson, (Eds.), Academic Press, Cambridge, Massachusetts, ISBN 978-0-12-374039-7, pp: 133-162.
87. Ado, R.I., C. Lopez, V. Lechevalier, M.E. Gouna and B. Robert *et al.*, 2018. Dairy curd coagulated by a plant extract of *Calotropis procera*: Role of fat structure on the chemical and textural characteristics. *Food Res. Int.*, 105: 694-702.
88. Belewu, M.A., 2001. Nutritional and rheological evaluation of West African soft cheese made from plant coagulant (*Calotropis procera*) during storage. *J. Food Technol. Africa*, 6: 93-95.
89. Singh, A.N., P. Yadav and V.K. Dubey, 2013. cDNA cloning and molecular modeling of procerain B, a novel cysteine endopeptidase isolated from *Calotropis procera*. *PLoS ONE*, Vol. 8. 10.1371/journal.pone.0059806.
90. Ramos, M.V., E.S. Araujo, T.L. Juca, A.C.O. Monteiro-Moreira and I.M. Vasconcelos *et al.*, 2013. New insights into the complex mixture of latex cysteine peptidases in *Calotropis procera*. *Int. J. Biol. Macromol.*, 58: 211-219.
91. Simoes, I. and C. Faro, 2004. Structure and function of plant aspartic proteinases. *Eur. J. Biochem.*, 271: 2067-2075.
92. Antao, C.M. and F.X. MacLcata, 2005. Plant serine proteases: Biochemical, physiological and molecular features. *Plant Physiol. Biochem.*, 43: 637-650.
93. van der Hoorn, R.A.L., 2008. Plant proteases: From phenotypes to molecular mechanisms. *Annu. Rev. Plant Biol.*, 59: 191-223.
94. Grudkowska, M. and B. Zagdanska, 2004. Multifunctional role of plant cysteine proteinases. *Acta Biochim. Pol.*, 51: 609-624.
95. Milind, P. and Gurditta, 2011. Basketful benefits of papaya. *Int. Res. J. Pharm.*, 2: 6-12.
96. Yogiraj, V., P.K. Goyal, C.S. Chauhan, A. Goyal and B. Vyas, 2014. *Carica papaya* Linn: An overview. *Int. J. Herbal Med.*, 2: 1-8.
97. Krishna, K.L., M. Paridhavi and J.A. Patel, 2008. Review on nutritional, medicinal and pharmacological properties of papaya (*Carica papaya* Linn). *Nat. Prod. Radiance*, 7: 364-373.
98. Patil, S.D., S.V. Ahale and S.J. Surana, 2011. Evaluation of antiasthmatic and antianaphylactic activity of *Balanites aegyptiaca* (Delile), (Balanitaceae). *Asian J. Pharm. Clin. Res.*, 4: 52-55.
99. Aravind, G., D. Bhowmik, S. Duraivel and G. Harish, 2013. Traditional and medicinal uses of *Carica papaya*. *J. Med. Plants Stud.*, 1: 7-15.
100. Orhue, P.O. and A.R.M. Momoh, 2013. Antibacterial activities of different solvent extracts of carica papaya fruit parts on some gram positive and gram negative organisms. *Int. J. Herbs Pharmacol. Res.*, 2: 42-47.
101. Aruljothi, S., C. Uma, P. Sivagurunathan and M. Bhuvaneshwari, 2014. Investigation on antibacterial activity of *Carica papaya* leaf extracts against wound infection-causing bacteria. *Int. J. Res. Stud. Biosci.*, 2: 8-12.
102. Adetunji, V.O. and O.T. Salawu, 2010. Efficacy of ethanolic leaf extracts of *Carica papaya* and *Terminalia catappa* as molluscicides against the snail intermediate hosts of schistosomiasis. *J. Med. Plants Res.*, 4: 2348-2352.

103. Kanthal, L.K., P. Mondal, S. de, S. Jana, S. Aneela and K. Satyavathi, 2012. Evaluation of anthelmintic activity of *Carica papaya* latex using pheritima posthuma. Int. J. Life Sci. Pharma Res., 2: 10-12.
104. Okeniyi, J.A.O., A. Tinuade, Ogunlesi, O.A. Oyelami and L.A. Adeyemi, 2007. Effectiveness of dried *Carica papaya* seeds against human intestinal parasitosis: A pilot study. J. Med. Food, 10: 194-196.
105. Kala, C.P., 2012. Leaf juice of *Carica papaya* L.: A remedy of dengue fever. Med. Aromat. Plants, Vol. 1. 10.4172/2167-0412.1000109.
106. Maisarah, A.M., A.B. Nurul, R. Asmah and O. Fauziah, 2013. Antioxidant analysis of different parts of *Carica papaya*. Int. Food Res. J., 20: 1043-1048.
107. Otsuki, N., N.H. Dang, E. Kumagai, A. Kondo, S. Iwata and C. Morimoto, 2010. Aqueous extract of *Carica papaya* leaves exhibits anti-tumor activity and immunomodulatory effects. J. Ethnopharmacol., 127: 760-767.
108. Azarkan, M., A. El Moussaoui, D. van Wuytswinkel, G. Dehon and Y. Looze, 2003. Fractionation and purification of the enzymes stored in the latex of *Carica papaya*. J. Chromatogr. B, 790: 229-238.
109. Amri, E. and F. Mamboya, 2012. Papain, a plant enzyme of biological importance: A review. Am. J. Biochem. Biotechnol., 8: 99-104.
110. Hachana, Y., I. Frija, R. Maoual, I. El Akram Znaidi and R. Fortina, 2021. Use of papaya leaf extract for the production of fresh goat's cheese as an alternative to animal rennet [German]. J. Food Saf. Food Qual., 72: 185-191.
111. Nicosia, F.D., I. Puglisi, A. Pino, C. Caggia and C.L. Randazzo, 2022. Plant milk-clotting enzymes for cheesemaking. Foods, Vol. 11. 10.3390/foods11060871.
112. Diouf, L., A.N. Mallaye, M. Mbengue, A. Kane and A. Diop, 2012. *Carina papaya* leaves: A substitute for animal rennet in cheese-making tradition. J. Nat. Prod. Plant Res., 2: 517-523.
113. Akinloye, A.M. and O.O. Adewumi, 2014. Effects of local coagulants on the yield of cheese using cow and sheep milk. Int. J. Dev. Sustainability, 3: 150-161.
114. Okia, C.A., J. Kwetegyeka, P. Okiror, J.M. Kimondo, Z. Teklehaimanot and J. Obua, 2013. Physico-chemical characteristics and fatty acid profile of desert date kernel oil in Uganda. Afr. Crop Sci. J., 21: 723-734.
115. Sagna, M.B., A. Diallo, P.S. Sarr, O. Ndiaye and D. Goffner *et al*, 2014. Biochemical composition and nutritional value of *Balanites aegyptiaca* (L.) del fruit pulps from Northern Ferlo in Senegal. Afr. J. Biotechnol., 13: 336-342.
116. Kumawat, B.K., M. Gupta, T. Chand and Y. Singh, 2012. Preliminary phytochemical investigation on leaves of *Balanites aegyptiaca* (L.) Delile. Res. J. Pharm. Biol. Chem. Sci., 3: 762-768.
117. Umar, K.J., L. Abubakar, B. Alhassan, S.D. Yahaya, L.G. Hassan, N.A. Sani and M.U. Muhammad, 2014. Nutritional profile of *Balanites aegyptiaca* flower. Studia Univ., 24: 169-173.
118. Shahid, M., N. Jahan, A. Shahzad, A. Sahai, S. Sharma and S. Parveen, 2012. Antimicrobial potential of *Balanites aegyptiaca* (L.) del, *Stevia rebaudiana* (Bert.) bertonii, *Tylophora indica* (Burm.f.) merrill, and *Cassia sophera* (Linn.). Open Conf. Proc. J., 3: 63-69.
119. Anywar, G., H. Oryem-Origa and M. Kamatenesi-Mugisha, 2014. Antibacterial and antifungal properties of some wild nutraceutical plant species from Nebbi District, Uganda. J. Pharm. Res. Int., 4: 1753-1761.
120. Kahsay, T., A. Mulugeta and C.R. Unnithan, 2014. Antioxidant and antibacterial activities of *Balanites aegyptiaca* delil from Northern Ethiopia. Am. J. Pharm. Tech. Res., 4: 416-422.
121. Tula, M.Y., T.B. Danchal, F.O. Iruolaje and G.A. Onyeje, 2014. Studies on phytochemical constituents and antibacterial potentials of extracts of *Balanites aegyptiaca* (Del.) parts on antibiotic resistant bacterial isolates. Eur. J. Med Plants, 4: 854-864.
122. Chapagain, B.P. and Z. Wiesman, 2005. Larvicidal activity of the fruit mesocarp extract of *Balanites aegyptiaca* and its saponin fractions against *Aedes aegypti*. Dengue Bull., 29: 203-207.
123. Suky, T.M.G., B. Parthipan, C. Kingston and V.R. Mohan, 2011. Anti-inflammatory activity of aerial part of *Balanites aegyptiaca* (L.) Del against carrageenan induced paw oedema. Int. J. PharmTechnol. Res., 3: 639-643.
124. Ibrahim, O.H.M., A.D. Al-Qurashi, K.A. Asiry, M.A.A. Mousa, N.A. Alhakamy and K.A.M. Abo-Elyousr, 2022. Investigation of potential *in vitro* anticancer and antimicrobial activities of *Balanites aegyptiaca* (L.) delile fruit extract and its phytochemical components. Plants, Vol. 11. 10.3390/plants11192621.