



International Journal of
**Agricultural
Research**

ISSN 1816-4897



Academic
Journals Inc.

www.academicjournals.com

Phytotoxic Effects of Multi-purpose Tree Species on Germination and Growth of *Parthenium hysterophorus* L.

Mulatu Wakjira, Gezahegn Berecha and Befekadu Bulti

Department of Horticulture and Plant Sciences, College of Agriculture and Veterinary Medicine, Jimma University, P.O. Box 307, Jimma, Ethiopia

Corresponding Author: Mulatu Wakjira, Department of Horticulture and Plant Sciences, College of Agriculture and Veterinary Medicine, Jimma University, P.O. Box 307, Jimma, Ethiopia Tel: +251 (0) 471110102 Fax: +251 (0) 471110934

ABSTRACT

The current study was conducted to test the inhibitory potential of aqueous extracts and dry shoot residues of four multipurpose tree species (*Albizia gummifera*, *Azadirachta indica*, *Melia azedarach* and *Sesbania sesban*) on seed germination and seedling growth of an invasive alien weed *Parthenium hysterophorus* under laboratory and greenhouse conditions. Leaf aqueous extracts and dry residues of all the multi-purpose tree species drastically inhibited germination and seedling growth of parthenium. Parthenium radicle was more inhibited than plumule in Petri dishes. Inhibitory effects increased with increasing aqueous extract concentration and residue amount. Soil surface-placed leaf residues exerted strong reduction on germination and growth of parthenium than soil-incorporated residues. In contrast to the higher inhibitory effects in Petri dishes, lower inhibitory effects were observed by the aqueous extracts of the multi-purpose tree species in pot-culture. Therefore, further investigations will be required under greenhouse and field conditions for pragmatic recommendation of species selection in the frame of multi-purpose tree species-mediated ecological management of parthenium weed.

Key words: Aqueous extracts, dry leaf residues, germination, growth, invasive alien weed, *Parthenium hysterophorus*, phytotoxicity

INTRODUCTION

Invasive Alien Species (IAS) are widely recognized among the greatest threats to biodiversity and productivity. They are becoming major challenges for countries like Ethiopia striving for food security. Parthenium (*Parthenium hysterophorus* L., Asteraceae) is one of the IAS registered in Ethiopia. It is an aggressive noxious weed, native to the Americas but now widely spread in Asia, Africa and Australia (Evans, 1997). Parthenium weed was introduced accidentally into Ethiopia in the mid-1970s and it was first reported from Ethiopia in 1988 at Dire-Dawa and Harerge, eastern Ethiopia (Seifu, 1990). Subsequently it spreads rapidly in all regions of the country, along roads and railways, through grazing areas and arable lands, adversely threatening crop production, animal husbandry and biodiversity (Tefera, 2002). The successful spread of parthenium in so many parts of the world including Ethiopia has mainly been attributed to its allelopathic properties, due to which it outcompetes crops and pasture species (Singh *et al.*, 2003, 2005).

Parthenium is a noxious weed because of its allelopathic effect (Wakjira *et al.*, 2005; Kohli *et al.*, 2006; Wakjira, 2009), its strong competitiveness with both crops and pasture for soil

moisture and nutrients (Singh *et al.*, 2003, 2005) and the hazard it poses to humans and animals (Wiesner *et al.*, 2007) and biodiversity (Tefera, 2002). Currently, parthenium is considered as the most serious weed in both arable and grazing lands as it caused severe crop losses (Tamado and Milberg, 2000). In Ethiopia, grain yield losses from 40 to 97% were recorded when parthenium is left uncontrolled throughout the growing season of sorghum (Tamado *et al.*, 2002b).

Currently, parthenium is one of the noxious weeds threatening crop production in Ethiopia. The rapid spread of parthenium in Ethiopia would be a bigger risk to the expansion and sustainable production of many crops (Tefera, 2002; Wakjira *et al.*, 2005; Wakjira, 2009; Wakjira *et al.*, 2009). Control of parthenium is therefore crucial to boost productivity of crops and maintain the integrity of both agro-biodiversity and ecosystem complex of the country. Various control methods attempted world-wide in controlling parthenium have their own pros and cons. Hand-weeding mostly used by small-scale farmers is more difficult due to the allergic effects of parthenium on human body (Tefera, 2002; Wiesner *et al.*, 2007). Mechanical control on the other hand is rather costly in terms of machinery, labour and time requirement. Furthermore, resource poor farmers of Ethiopia cannot afford the purchase of herbicides and the use of herbicides is unsafe in terms of health and environmental considerations and not allowed in organic farming systems. Therefore, other options must be sought for ecological management of parthenium weed in Ethiopia. One such option is the use of allelopathic plants that suppress the germination and growth of parthenium.

The use of phytotoxic plants as an ecological weed management tool will play an important role in future weed control and crop productivity. The phytotoxic/allelopathic compounds can be used as natural herbicides and they are less disruptive of the global ecosystem than are synthetic agrochemicals (Khalid *et al.*, 2002). Recent investigations indicated the potential use of allelopathic plants in parthenium weed management (Anjum *et al.*, 2005; Javaid *et al.*, 2005; Shafique *et al.*, 2005; Javaid and Anjum, 2006).

In Ethiopia, most of the research work regarding parthenium concentrated on its distribution (Tamado and Milberg, 2000), germination ecology (Tamado *et al.*, 2002a), impacts on sorghum (Tamado *et al.*, 2002b), allelopathic effects (Tefera, 2002; Wakjira *et al.*, 2005, 2009), biological control using fungi (Tessema, 2002) and management by utilization (Wakjira *et al.*, 2009). Little attempts have been made to investigate and exploit the ecologically-based management methods like the use of allelopathic plant species including the multi-purpose tree species in Ethiopia. Therefore, the objective of the present study was to test the inhibitory potential of aqueous extracts and dry shoot residues of four multipurpose tree species (*Albizia gummifera*, *Azadirachta indica*, *Melia azedarach* and *Sesbania sesban*) on seed germination and seedling growth of an invasive alien weed parthenium under laboratory and greenhouse conditions.

MATERIALS AND METHODS

General experimental set-up: Laboratory and greenhouse experiments were executed at Jimma University, Jimma, Ethiopia from January 2007 to December 2009. The study site is located at about 7°, 41' N latitude and 36°, 50' E longitude at an altitude of 1710 m above sea level. The mean minimum and maximum temperatures in the laboratory and greenhouse were 20 and 23°C and 12 and 25°C during the experimental period. The relative humidity ranged from 31 to 90%. The soil was clay-loam in texture with an organic matter content of 3-5% and a pH of 5.6-6.5.

Fresh leaf samples of tree species commonly grown in Ethiopia (*Albizia gummifera*, *Azadirachta indica*, *Melia azedarach* and *Sesbania sesban*) were collected at their vegetative stage. Except *A. indica*, leaves of all other tree species were collected from Jimma area. The *A. indica* leaves were collected from Ziway, Ethiopia. The leaves were air-dried and ground in a mill of 2 mm sieve size.

Mature parthenium seeds were collected from naturally growing parthenium plants around the Jimma University campus. The seeds were air-dried and threshed by hand and their viability was selected. Seeds were surface-sterilized by shaking them for five minutes in 1% sodium hypochlorite (NaOCl) solution and washed with distilled water for three minutes immediately before use. For the laboratory experiment, 9 cm diameter Petri dishes were used. In the case of lathhouse experiments, 1 L pots were used. Sieved agricultural soil obtained from Eladale Research Farm of Jimma University was used for the pot experiments.

Experiments 1 and 2 were carried out to evaluate the inhibitory effects of aqueous extracts of the four multi-purpose tree species on parthenium germination and growth under laboratory and greenhouse conditions. Three, 6 and 9 g of each of the ground material of the test species were soaked in 100 mL distilled water (w/v) for 24 h at room temperature (21 to 22°C). The extracts were filtered through three layers of cheesecloth and designated as 3, 6 and 9% leaf aqueous extracts, respectively and stored in dark conical flasks until use.

Experiments 3 and 4 were carried out with dry leaf residues incorporated into the soil (Experiment 3) and placed on the soil surface (Experiment 4) of the four multi-purpose tree species under greenhouse conditions. The aims of these experiments were to investigate the variations in inhibitory potential of the ground leaf materials from the four tree species when incorporated into the soil and placed on the soil surface on the emergence and growth of parthenium.

Experiment 1: Effects of leaf aqueous extracts in the laboratory: Fifty sterilized parthenium seeds were placed in Petri dishes lined with double layers of filter paper, a thick filter paper (T415) was placed on the bottom and a thin filter paper (Whatman No. 1) on the top. The filter paper was moistened with 10 mL of the 3, 6 or 9% leaf aqueous extracts and distilled water (control). The treatments were replicated four times in a Completely Randomized Design (CRD). The Petri dishes and pots were rotated every third day to avoid unforeseen variations within the laboratory and greenhouse, respectively. The experiment was conducted twice.

Experiment 2: Effects of leaf aqueous extracts in pot-culture: Fifty sterilized parthenium seeds were sown per pot and 10 mL of the 3, 6 or 9% leaf aqueous extracts were added per pot. The control treatments received 10 mL distilled water per pot. The treatments were laid down in a Randomized Complete Block Design (RCBD) with four replications. The experiment was executed twice.

Experiment 3: Effects of soil-incorporated dry leaf residues: Ground leaves were thoroughly mixed in one litre pots filled with 1 kg sieved agricultural soil. A pot filled with same soil without dry leaf residues was used as control. Therefore, the treatments include 3, 6 and 9 g dry leaf residues of the four multi-purpose tree species and control (without dry leaf residue). The treatments were laid down in RCBD with three replications. The experiments were executed twice.

Experiment 4: Effects of soil surface-placed dry leaf residues: Three, 6 and 9 g of the dry leaf residues of the four tree species were placed on the surface of pots filled with sieved agricultural soil. A pot filled with the same volume of sieved agricultural soil without dry leaf residues was used as control. The treatments were laid down in RCBD with four replications. The experiments were executed twice.

Measurements: In the laboratory experiment with aqueous extracts, germinated seeds were counted daily from 10th to 14th days after sowing (DAS). Emerged seedlings were counted daily from 8th day to 17th DAS in the case of greenhouse experiments with aqueous extracts. In the greenhouse experiment with dry leaf residues, emerged seedlings were counted from 7th to 13th DAS. Germination/emergence percentages were determined from the number of germinated seeds or emerged seedlings. Germination/emergence rate (G/ER) is the number of seeds germinated or seedlings emerged from the seeds sown per Petri dish or per pot and was calculated by Maguire (1962) as:

$$\text{G/ER (seeds day}^{-1}\text{)} = \frac{\text{No. of normal seedlings}}{\text{Days of first count}} + \dots + \frac{\text{No. of normal seedlings}}{\text{Days of final count}}$$

Radicle and plumule lengths (mm) were measured on five randomly selected seedlings per Petri dish in laboratory experiments. In greenhouse experiments seedlings were thinned to ten per pot at 20 DAS and five seedlings were taken at random from five different positions from the middle of the pot four weeks after sowing. The five sample seedlings were separated into root and shoot parts and root and shoot lengths (mm) were measured for both experiments with aqueous extracts and dry leaf residues. In the case of dry leaf residues, the numbers of true leaves per plant were counted and leaf area per plant (cm²) was determined by scanning the leaves using a portable leaf area meter (ADC AM300). Root and shoot fresh weights were determined per plant using a sensitive balance. Root and shoot dry weight per plants was determined after drying in an oven at 70°C for 24 h. The inhibition (-) or stimulation (+) percentage compared to control was calculated using Chung *et al.* (2001):

$$\text{Inhibition (-) or stimulation (+) percentage (\%)} = \left(\frac{\text{Treatment} - \text{Control}}{\text{Control}} \right) * 100$$

Data analysis: For each of the response variables, the validity of model assumptions was verified by examining the residuals. The average data were analyzed using Analysis of Variance (ANOVA). Two phases of statistical analysis were employed. First, whether the interaction and/or the main effects of the fixed factors of interest were significant was analyzed. Second, treatment combinations having significant interactions and main effects were further compared. Multiple comparisons of means were performed using Fisher's protected Least Significant Difference (LSD) test. All statistical procedures were performed using GenStat statistical package, GenStat for Windows 12th Edition (VSN International Ltd., 2009).

RESULTS

Experiment 1 Effects of leaf aqueous extracts in the laboratory: In Petri dishes, leaf aqueous extracts of the four multi-purpose tree species reduced germination percentage, germination rate, radicle length and plumule length of parthenium (average reductions of 59, 60, 63 and 55%, respectively) (Fig. 1). Leaf aqueous extracts of *A. indica*, *M. azedarach*, *S. sesban* and *A. gummifera* reduced germination percentage (by 67, 59, 56 and 54%, respectively), germination rate (by 65, 63, 62 and 49%, respectively), radicle length (by 67, 64, 71 and 50%, respectively) and plumule length (by 60, 65, 55 and 41%, respectively). Though the multi-purpose tree species

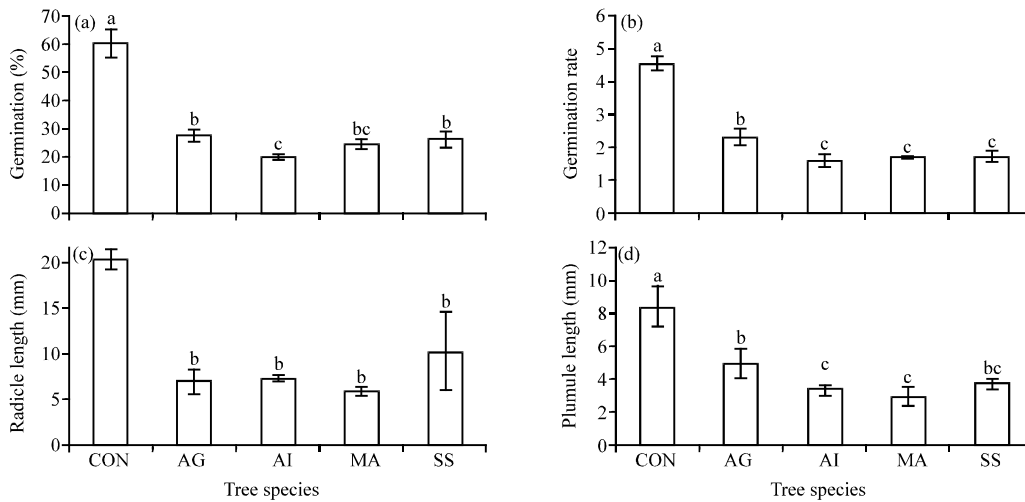


Fig. 1: Main effects of leaf aqueous extracts of four tree species (CON: Control, AG: *Albizia gummifera*, AI: *Azadirachta indica*, MA: *Melia azedarach* and SS: *Sesbania sesban*) on germination percentage (a), germination rate (b), radicle length (c) and plumule length (d) in Petri dishes. Means followed by different letters differ significantly ($p < 5\%$) as established by Fisher's protected LSD-test (LSD = 5.37, 0.49, 4.401 and 1.56, respectively). Vertical bars represent mean values \pm SE

Table 1: Main effects of leaf aqueous extract concentrations of four tree species on Germination Percentage (GP), Germination Rate (GR), Radicle Length (RL) and Plumule Length (PL) in Petri dishes

Aqueous extract concentration (%)	GP (%)	GR (seeds day ⁻¹)	RL (mm)	PL (mm)
0	60.0a ¹	4.6a	20.4a	8.4a
3	23.8 (-60)b	1.7 (-62)b	6.4 (-69)b	4.7 (-44)b
6	10.5 (-83)c	0.8 (-82)c	3.6 (-83)bc	1.9 (-77)c
9	4.3 (-93)d	0.2 (-95)d	0.03 (-100)c	0.1 (-99)d
LSD	5.370	0.4911	4.401	1.556

¹ Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly ($p < 5\%$) as established by Fisher's protected LSD-test

provided strong reduction on germination and growth of parthenium compared to control, the differences between the tree species were not large.

Compared to control, leaf aqueous extract concentrations from the four multi-purpose tree species drastically reduced germination percentage (-79%) and rate (-80%) and radicle (-84%) and plumule lengths (-74%) on average basis (Table 1). For all the response variables, there was an increase in inhibitory effect with an increase in aqueous extract concentration. Consequently, at the highest concentration (9%), nearly complete failure of germination and growth of parthenium was observed (97%).

The amount of reduction on germination and growth of parthenium provided by the main effects of leaf aqueous extract concentration (-80%) was higher than that provided by the main effects of multi-purpose tree species (-60%) (Fig. 1 and Table 2). In both cases, comparable reductions were observed for emergence and growth whereas radicles were reduced more than plumules.

Table 2: Main effects of leaf aqueous extracts of four tree species on emergence percentage (EP), emergence rate (ER), root length (RL) and shoot length (SL) in pot-culture

Tree species	EP (%)	ER (seeds day ⁻¹)	RL (mm)	SL (mm)
Control	46.1a	30.8a	34.8a	6.11a
<i>Albizia gummifera</i>	41.1 (-11)bc	25.5 (-17)b	27.7 (-20)b	4.97 (-19)b
<i>Azadirachta indica</i>	41.9 (-9)b	24.5 (-21)b	28.3 (-19)b	4.78 (-22)bc
<i>Melia azedarach</i>	40.4 (-13)bc	23.7 (-23)b	25.7 (-26)b	4.34 (-29)bc
<i>Sesbania sesban</i>	38.4 (-17)c	24.4 (-21)b	26.0 (-25)b	4.16 (-32)c
LSD	3.133	2.619	4.174	0.778

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly (p < 5%) as established by Fisher's protected LSD-test

Table 3: Main effects of leaf aqueous extract concentrations of four tree species on Emergence Percentage (EP), Emergence Rate (ER), Root Length (RL) and Shoot Length (SL) in pot-culture

Aqueous extract concentration (%)	EP (%)	ER (seeds day ⁻¹)	RL (mm)	SL (mm)
0	46.1a ¹	30.8 a	34.8a	6.11a
3	41.4 (-10)b	25.0 (-19)b	29.0 (-17)b	5.19 (-15)b
6	39.2 (-15)b	22.9 (-26)b	23.9 (-32)c	3.90 (-36)c
9	34.9 (-24)c	19.4 (-37)c	20.1 (-42)c	3.04 (-50)d
LSD	3.133	2.619	4.174	0.778

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly (p < 5%) as established by Fisher's protected LSD-test

Experiment 2 Effects of leaf aqueous extracts in pot-culture: In contrast to the results obtained under laboratory conditions in Petri dishes, lower inhibitions were observed on emergence and growth of parthenium under greenhouse conditions in pot-culture. Evidently, lower effects were observed by the main effects of the multi-purpose tree species and aqueous extract concentrations under greenhouse conditions (Table 2, 3). Aqueous extracts from the four multi-purpose tree species reduced emergence percentage (-12%), emergence rate (-20%), root length (-23%) and shoot length (-25%) of parthenium compared to control on average (Table 2). Though the multi-purpose tree species reduced germination and growth of parthenium compared to control, the differences between the inhibition effects of the tree species were very small. The amount of reduction provided by the multi-purpose tree species (20% on average) on emergence and growth of parthenium was relatively lower compared to the reduction provided by the aqueous extract concentrations (27% on average) (Table 2 and 3). Compared to control, aqueous extract concentrations from the four multi-purpose tree species reduced parthenium emergence percentage (-17%), emergence rate (-27%), radicle length (-30%) and plumule length (-34%) (Table 3). Similar to the results under laboratory conditions, for all the variables, there was an increase in inhibitory effect with an increase in aqueous extract concentration.

Experiment 3 Effects of soil-incorporated dry leaf residues: Soil-incorporated dry leaf residues of the four multi-purpose tree species provided a significant reduction on emergence percentage (-23%), emergence rate (-28%), root length (-10%), leaf area (-15%) and root fresh weight (-25%) of parthenium (Table 4). Shoot fresh weight was inhibited by *A. indica* and *S. sesban* (by 44 and 4%, respectively) while *M. azedarach* and *A. gummifera* stimulated shoot fresh weight by 46 and 16%, respectively (Table 4). In relation to inhibition of emergence, the four tree species ranked in the order of *S. sesban* > *A. indica* > *M. azedarach* > *A. gummifera*. However, the effects on growth of parthenium were inconsistent.

Table 4: Main effects of soil-incorporated dry leaf residues of four tree species on Emergence Percentage (EP) and rate (ER), Leaf Area (LA) and Root Fresh Weight (RFW) and Shoot Fresh Weight (SFW) in pot-culture

Tree species	EP (%)	ER (seeds day ⁻¹)	RL (mm)	LA (cm ² plant ⁻¹)	RFW (g plant ⁻¹)	SFW (g plant ⁻¹)
Control	74.7a ¹	12.4a	30.5a	235.1a	0.7ab	0.31ab
<i>Albizia gummifera</i>	59.0 (-21)b	10.3 (-17)b	28.7 (-6)ab	192.9 (-18)b	0.5 (-27)bc	0.35 (+16)a
<i>Azadirachta indica</i>	54.7 (-27)bc	8.2 (-34)cd	26.6 (-13)c	144.7 (-39)c	0.4 (-41)c	0.17 (-45)b
<i>Melia azedarach</i>	58.7 (-21)bc	9.2 (-25)bc	26.7(-13)bc	230.6 (-2)ab	0.8 (-10)a	0.45 (+46)a
<i>Sesbania sesban</i>	54.0 (-28)c	7.6 (-38)d	27.9 (-9)bc	227.8 (-3)ab	0.4 (-42)c	0.29 (-4)ab
LSD	4.382	1.395	2.039	41.95	0.2328	0.1676

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly (p<5%) as established by Fisher's protected LSD-test

Table 5: Main effects of soil-incorporated dry leaf residues of four tree species on Emergence Percentage (EP) and rate (ER), Root Length (RL), Leaf Area (LA) and Root Fresh Weight (RFW) in pot-culture

Residue amount (g)	EP (%)	ER (seeds day ⁻¹)	RL (mm)	LA (cm ² plant ⁻¹)	RFW (g plant ⁻¹)
0	74.7a ¹	12.4a	30.5a	235.1a	0.71a
3	57.3 (-23)b	9.1 (-26)b	28.9 (-5)a	222.2 (-6)ab	0.61 (-13)ab
6	52.0 (-30)c	7.9 (-37)b	26.0 (-15)b	187.8 (-20)bc	0.46 (-35)bc
9	42.3 (-43)d	6.1 (-51)c	24.5 (-20)b	150.9 (-36)c	0.35 (-51)c
LSD	4.382	1.395	2.039	41.95	0.2328

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly (p<5%) as established by Fisher's protected LSD-test

Soil-incorporated residue amounts provided greater reduction compared to the main effects of the multi-purpose tree species (average reductions of 27 and 20%, respectively) (Table 4, 5). Compared to control, the amounts of soil-incorporated dry leaf residues from the four multi-purpose tree species reduced emergence percentage (-32%), emergence rate (-38%), root length (-13%), leaf area (-21%) and root fresh weight (-33%) on an average (Table 5). For all the response variables, there was an increase in inhibitory effect on parthenium emergence and growth with an increase in the amount of soil-incorporated dry leaf residue. Evidently, the highest residue amount (9 g) provided reduction on emergence percentage, emergence rate, root length, leaf area and root fresh weights of 43, 51, 20, 36 and 51%, respectively.

Experiment 4 Effects of soil surface-placed dry leaf residues: The analysis of variance for germination percentage and rate confirmed that there were significant two-way interactions between multi-purpose tree species and amounts of dry leaf residues placed on the soil surface (p<0.001) (Fig. 2). This clearly indicated that multiplicative effects occur between tree species and amounts of soil-surface placed dry leaf residues in inhibiting emergence of parthenium, both percentage and rate.

Soil surface-placed dry leaf residues of all the multi-purpose tree species provided strong reductions on emergence percentage (-43%) and emergence rate (-46%) compared to control (Fig. 2). Application of *A. gummifera*, *A. indica*, *M. azedarach* and *S. sesban* on the soil surface reduced emergence percentage by 33, 40, 80 and 74%, respectively (Fig. 1a). Emergence rate was reduced by surface application of dry leaf residues of *A. gummifera*, *A. indica*, *M. azedarach* and *S. sesban* by 34, 46, 86 and 80%, respectively (Fig. 2b). Except for *A. gummifera* on emergence rate, there was an increase in inhibitory effect on the emergence of parthenium with an increase in the amount of leaf residue placed on the soil surface. For *A. gummifera*, there were no significant

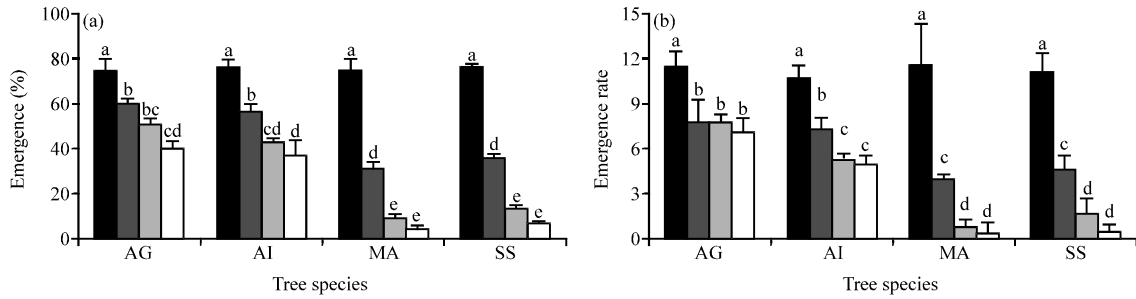


Fig. 2: Effects of soil surface-placed dry leaf residues of four tree species (CON: Control, AG: *Albizia gummifera*, AI: *Azadirachta indica*, MA: *Melia azedarach* and SS: *Sesbania sesban*) at different amounts: 0 g (black), 3 g (grey), 6 g (stripes) and 9 g (white) on parthenium emergence (%) and rate (seeds per day) in pot-culture. Means followed by different letters differ significantly ($p < 5\%$) as established by Fisher's protected LSD-test (LSD = 13.09 and 1.81, respectively). Vertical bars represent mean values \pm SE

Table 6: Main effects of soil surface-placed dry leaf residues of four tree species on Root Length (RL), Leaf Number (LN), Root Fresh Weight (RFW) and Root Dry Weight (RDW) in pot-culture

Tree species	RL (mm)	LN	RFW (g plant ⁻¹)	RDW (g plant ⁻¹)
Control	30.7a ¹	8.6a	0.70a	0.122a
<i>Albizia gummifera</i>	28.9 (-6)a	8.3 (-3)a	0.45 (-35)b	0.098 (-19)ab
<i>Azadirachta indica</i>	24.7 (-20)b	8.2 (-4)a	0.67 (-4)a	0.136 (+12)a
<i>Melia azedarach</i>	29.0 (-5)a	7.8 (-9)a	0.36 (-48)b	0.071 (-41)b
<i>Sesbania sesban</i>	24.8 (-19)b	6.4 (-26)b	0.35 (-50)b	0.073 (-40)b
LSD	3.914	0.883	0.2112	0.04772

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly ($p < 5\%$) as established by Fisher's protected LSD-test

differences between all amounts of soil surface-placed dry leaf residues in reducing parthenium emergence rate (Fig. 2b). The amounts of soil surface-placed dry leaf residues reduced emergence percentage by 57% whereas emergence rate by 62%.

Soil surface-placed dry leaf residues from the four multi-purpose tree species reduced root length (-13%), leaf number (-10%) and root fresh weight (-34%) of parthenium on average compared to control (Table 6). Root dry weight was reduced by *M. azedarach*, *S. sesban* and *A. gummifera* (by 41, 40 and 19%, respectively) whereas stimulated by *A. indica* (by 12%). The amounts of surface-placed dry leaf residues exerted more reduction (-39%) on emergence and growth of parthenium compared to the main effects of the multi-purpose tree species (-29%). Compared to control, the amounts of soil surface-placed dry leaf residues from the four multi-purpose tree species provided reductions on root length (-17%), leaf number (-14%), leaf area (-31%), root fresh weight (-46%) and shoot dry weight (-42%) (Table 7). For all the response variables, there was an increase in inhibitory effect on emergence and growth of parthenium with an increase in the amount of dry leaf residue placed on the soil surface.

DISCUSSION

Plant invasions are threatening biodiversity and productivity worldwide and are becoming the major challenges for developing countries like Ethiopia striving to achieve food security.

Table 7: Main effects of soil surface-placed dry leaf residue amounts of four tree species on Root Length (RL), Leaf Area (LA), Leaf Number (LN), Root Fresh Weight (RFW) and Shoot Dry Weight (SDW) in pot-culture

Residue amount (g)	RL (mm)	LA (cm ² plant ⁻¹)	LN	RFW (g plant ⁻¹)	SDW (g plant ⁻¹)
0	30.7a ¹	226.7a	8.6a	0.69a	0.076a
3	30.3 (-1)a	198.7 (-12)ab	8.1 (-6)ab	0.46 (-34)b	0.051 (-34)b
6	24.8 (-19)b	151.9 (-33)bc	7.3 (-15)bc	0.31 (-47)b	0.046 (-40)b
9	21.5 (-30)b	121.5 (-46)c	6.7 (-22)c	0.36 (-56)b	0.036 (-52)b
LSD	3.914	55.45	0.883	0.2112	0.02251

¹Values in parenthesis indicate percentage reduction (-) or stimulation (+) compared to control. Means followed by different letters within one column differ significantly (p<5%) as established by Fisher's protected LSD-test

Parthenium is one of the IAS spreading at an alarming rate in Ethiopia. Due to increased societal concerns about the environmental and human health impacts together with inability of the small-scale farmers to buy herbicides, efforts are being made worldwide to reduce the heavy reliance on synthetic herbicides through the use of alternative methods. The use of weed suppressive multi-purpose tree species in agro-ecosystem is among these alternatives as it has the potential to retard germination and growth of weeds. Inhibition of germination could minimize the number of weed plants that compete with the crop species and that grow and add seed to the seed bank. Furthermore, weed plants that germinated at slower rates are often smaller and less competitive. This could give an opportunity for the crop species to outcompete the weeds (Jefferson and Pennacchio, 2003). The aim of the present study was to evaluate different multi-purpose tree species for their inhibitory potential in suppressing germination and growth of an invasive alien weed parthenium. To achieve this objective, four sets of experiments were executed using leaf aqueous extracts and dry residues of four multi-purpose tree species under the laboratory conditions in Petri dishes and under the greenhouse conditions in pot-culture.

Effects of multi-purpose tree species: There were variations among the multi-purpose tree species in their inhibitory potential in the present study. This finding is in line with previous findings that indicated different plants have different inhibitory potential (Stoll *et al.*, 2006; Adler and Chase, 2007; Price *et al.*, 2008). The plausible mechanism for inhibition of germination and growth could be due to reduced rate of cell division and elongation attributed to allelochemicals (Javaid and Anjum, 2006) or could be physical interference by surface-placed residues with the upward movement of the emerging seedlings and the downward penetration of light to germinating seeds (Teasdale and Mohler, 2000).

Compared to control, leaf aqueous extracts of all of the multi-purpose tree species greatly reduced germination percentage and rate and radicle and plumule lengths of parthenium under laboratory conditions in Petri dishes (Fig. 1). However, variations were evident among the tree species in the extent of reduction they provided. Regarding reduction of germination, the four tree species ranked in the order of *A. indica* > *M. azedarach* > *S. sesban* > *A. gummifera* whereas in reduction of growth they were ranked in the order of *M. azedarach* > *A. indica* > *S. sesban* > *A. gummifera*. In the case of aqueous extracts in pot-culture, *S. sesban* and *M. azedarach* provided greater reduction of both emergence and growth compared to *A. indica* and *A. gummifera* (Table 2). In relation to inhibition of emergence provided by soil-incorporated leaf residues, the four tree species ranked in the order of *S. sesban* > *A. indica* > *M. azedarach* > *A. gummifera*. The effects on growth of parthenium were inconsistent though *A. indica* and *S. sesban* still played a significant role in reduction of growth. However, soil-incorporated residues from *M. azedarach* and

A. gummifera strongly stimulated shoot fresh weight (Table 4). In the case of soil surface-placed residues, *M. azedarach* and *S. sesban* provided more reduction on both germination and growth of parthenium than *A. indica* and *A. gummifera* (Fig. 2 and Table 6). Furthermore, *A. indica* slightly stimulated root dry weight (Table 6).

This finding is supported by previous reports that demonstrated about the allelopathic effects of *A. indica* (Xuan *et al.*, 2004; Shafique *et al.*, 2005; Al-Charchafchi *et al.*, 2007; Rahnavard *et al.*, 2008), *M. azedarach* (Hong *et al.*, 2003, 2004) and *S. sesban* (Mubarak *et al.*, 2009) on different plants. For *A. gummifera*, findings that indicate about its allelopathic effects are nonexistent. Hence, it was for the first time that we have systematically evaluated and demonstrated the allelopathic inhibitory effects of *A. gummifera* on germination and growth of parthenium.

Effects of quantity: aqueous extract concentrations and dry leaf residue amounts: Both under laboratory conditions in Petri dishes and under greenhouse conditions in pot-culture, there was an increase in inhibitory effect on both germination and growth of parthenium with an increase in extract concentration from the four multi-purpose tree species. This was in agreement with several previous authors for many different plants including *A. indica* (Tefera, 2002; Xuan *et al.*, 2004; Wakjira *et al.*, 2005; Al-Charchafchi *et al.*, 2007; Jafari *et al.*, 2007; Rahnavard *et al.*, 2008; Mutlu and Atici, 2009; Wakjira, 2009). Similar to the aqueous extract concentrations, as the amount of dry shoot residue incorporated and placed on the soil surface increased, inhibitory effect also increased though the variation between the amounts was small. This was in agreement with previous reports for many plants including *A. indica* and *M. azedarach* (Hong *et al.*, 2004; Xuan *et al.*, 2004; Singh *et al.*, 2005; Batlang and Shushu, 2007; Wakjira, 2009).

Effects of residue placement method: The amount and quality of the residue material, soil biological, chemical and physical factors, environmental conditions, extent of tissue disruption and methods of residue placement influence the release of allelochemicals and the consequent residue-mediated inhibitory or stimulatory effects (Mohler, 1996; Liebman and Mohler, 2001; Morra and Kirkegaard, 2002; Kruidhof *et al.*, 2009). In the present study, dry and ground leaves were used both in the incorporation and surface-placed experiments that provided uniform tissue disruption. Compared to soil-incorporated dry leaf residues, the soil surface-placed dry leaf residues drastically inhibited germination and growth of parthenium (Table 4-7, Fig. 2). Our current finding was in agreement with previous findings for other plant species (Ismail and Chong, 2002; Wakjira, 2009). The inhibitory effect as a result of soil incorporation might be explained by the fact that the more inhibitory chemicals were available that caused inhibitory effect (Chon *et al.*, 2005). The possible explanation for the reduction provided by the surface placed residues could be physical interference with the upward movement of the emerging seedlings and the downward penetration of light to germinating seeds (Teasdale and Mohler, 2000) or due to the release of inhibitory allelochemicals.

Effects on radicle and plumule growth: In the laboratory experiment we have demonstrated that radicle growth is more reduced than plumule growth (Fig. 3). This is in line with several other reports on many plants including (Tefera, 2002; Wakjira *et al.*, 2005; Al-Charchafchi *et al.*, 2007; Rahnavard *et al.*, 2008; Wakjira, 2009). The plausible reason could be the radicles are in direct

contact with the leaf aqueous extracts and hence with the inhibitory chemicals (Tefera, 2002; Rietjens and Alink, 2003; Wakjira *et al.*, 2005; Al-Charchafchi *et al.*, 2007; Wakjira, 2009).

Comparison of laboratory and greenhouse experiment results: In the present study, clear, greater and fast inhibition effect by the multi-purpose tree species was observed in Petri dishes compared to pot-culture. Experiments with aqueous extracts in Petri dishes are easy and fast but do not prove the existence of allelopathy under field conditions. Pot soil with residue material is less convenient and more laborious, but presumed to be closer to reality. From the present study the multi-purpose tree species provided a strong reduction on germination and growth of parthenium under laboratory conditions in Petri dishes. This however does not mean that they are also maintained under greenhouse and field conditions. Laboratory screening however helps to select promising species for greenhouse and field evaluation and field studies are pertinent to provide strong evidence for existence of allelopathy-mediated inhibitory potentials (Inderjit and Weston, 2000; Wu *et al.*, 2001; Morgan and Overholt, 2005). In contrast to the results obtained under laboratory conditions Petri dishes, lower inhibitory effects of aqueous extracts of the four multi-purpose tree species were observed under greenhouse conditions in pot-culture. This is congruent with findings of previous workers (Morgan and Overholt, 2005). This could be explained by the complex reactions taking place in the soil. Allelochemicals are subject to various biotic and abiotic processes that reduce their persistence, concentrations, availability and biological activities after they are added into the soil (Huang *et al.*, 1977; Wang *et al.*, 1986; Makino *et al.*, 1996; Blum *et al.*, 1999; Okumura *et al.*, 1999).

Practical considerations and prospects: The rapid spread of parthenium in Ethiopia would be a bigger risk to the expansion and sustainable production of many crops (Tamado *et al.*, 2002b); Tefera, 2002; Tessema, 2002; Wakjira *et al.*, 2005, 2009; Wakjira, 2009). Control of parthenium is therefore crucial to boost productivity of many crops and maintain the integrity of both agro-biodiversity and ecosystem complex of in the country. Various control methods attempted world-wide in controlling parthenium have their own pros and cons. Hand-weeding mostly used by small-scale farmers is more difficult due to the allergic effects of parthenium on human body (Tefera, 2002; Wiesner *et al.*, 2007). Mechanical control on the other hand is rather costly in terms of machinery, labour and time requirement. Furthermore, resource poor farmers of Ethiopia cannot afford the purchase of herbicides and the use of herbicides is unsafe in terms of health and environmental considerations and not allowed in when the farmers want to certify for organic production. Therefore, other system-oriented options must be sought for ecological management of parthenium weed in Ethiopia. One such option is the use of phytotoxic plants that suppress the germination and growth of parthenium. In the current study, the four multi-purpose tree species drastically reduced the germination and growth of parthenium. This could suggest that these multi-purpose tree species produce allelochemicals responsible for inhibitory effects or provide physical suppression on germination and growth of parthenium. Though the trend in pot-culture followed that of the Petri dish result, in contrast to the higher inhibitory effects obtained in Petri dishes, lower inhibitory effects by the multi-purpose tree species were observed in pot-culture. Therefore, further investigations will be required under greenhouse and field conditions for pragmatic recommendation of species selection in the frame of multi-purpose tree species-mediated ecological management of parthenium weed.

ACKNOWLEDGMENTS

We thank Ethiopian Institute of Agricultural Research (EIAR) for funding and Jimma University College of Agriculture and Veterinary Medicine (JUCAVM) for laboratory and greenhouse facilities. We are greatly indebted to Daniel Damtew for his assistance during execution of the laboratory and greenhouse experiments and Arafat Kassahun for data entry.

REFERENCES

- Adler, M.J. and C.A. Chase, 2007. Comparison of the allelopathic potential of leguminous summer cover crops: Cowpea, sunn hemp and velvetbean. *Hortscience*, 42: 289-293.
- Al-Charchafchi, F., I. Al-Nabhani, H. Al-Kharousi, F. Al-Quraini and A. Al-Hanai, 2007. Effect of aqueous extract of *Azadirachta indica* (Neem) leaves on germination and seedling growth of *Vigna radiata* (L.). *Pak. J. Biol. Sci.*, 10: 3885-3889.
- Anjum, T., R. Bajwa and A. Javaid, 2005. Biological Control of *Parthenium* I: Effect of *Imperata cylindrical* on distribution, germination and seedling growth of *Parthenium hysterophorus* L. *Int. J. Agric. Biol.*, 7: 448-450.
- Batlang, U. and D.D. Shushu, 2007. Allelopathic activity of sunflower (*Helianthus annuus* L.) on growth and nodulation of bambara groundnut (*Vigna subterranean* (L.) Verdc.). *J. Agron.*, 6: 541-547.
- Blum, U., S.R. Shafer and M.E. Lehman, 1999. Evidence for inhibitory allelopathic interactions involving phenolic acids in field soils: Concepts vs. an experimental model. *Crit. Rev. Plant Sci.*, 18: 673-693.
- Chon, S.U., H.G. Jang, D.K. Kim, Y.M. Kim, H.O. Boo and Y.J. Kim, 2005. Allelopathic potential in lettuce (*Lactuca sativa* L.) plants. *Sci. Hort.*, 106: 309-317.
- Chung, I.M., J.K. Ahn and S.J. Yun, 2001. Assessment of allelopathic potential of barnyard grass (*Echinochola crus-galli*) on rice (*Oryza sativum* L.) cultivars. *Crop Protect.*, 20: 921-928.
- Evans, H.C., 1997. *Parthenium hysterophorus*: A review of its weed status and the possibilities for biological control. *Biocont. News Inform.*, 18: 89-98.
- Hong, N.H., T.D. Xuan, T. Eiji, T. Hiroyuki, M. Mitsuhiro and T.D. Khanh, 2003. Screening for allelopathic potential of higher plants from Southeast Asia. *Crop Prot.*, 22: 829-836.
- Hong, N.H., T.D. Xuan, E. Tsuzuki, H. Terao, M. Matsuo and T.D. Khanh, 2004. Weed control of four higher plant species in paddy rice fields in Southeast Asia. *J. Agron. Crop Sci.*, 190: 59-64.
- Huang, P.M., T.S.C. Wang, M.K. Wang, M.H. Wu and N.W. Hsu, 1977. Retention of phenolic acids by non-crystalline hydroxy-aluminium and iron compounds and clay minerals of soil. *Soil Sci.*, 123: 213-219.
- Inderjit and L.A. Weston, 2000. Are laboratory bioassays for allelopathy suitable for prediction of field response. *J. Chem. Ecol.*, 26: 2111-2118.
- Ismail, B. and T.V. Chong, 2002. Effects of aqueous extracts and decomposition of *Mikania micrantha* H.B.K. debris on selected agronomic crops. *Weed Biol. Manage.*, 2: 31-38.
- Jafari, L., B. Kholdebarin and E. Jafari, 2007. Phytotoxic effects of *Chenopodium album* L. Water extract on higher plants. *Am. J. Plant Physiol.*, 2: 221-226.
- Javaid, A., T. Anjum and R. Bajwa, 2005. Biological control of *Parthenium* II: Allelopathic effect of *Desmostachya bipinnata* on distribution and early seedling growth of *Parthenium hysterophorus* L. *Int. J. Biol. Biotech.*, 2: 459-463.
- Javaid, A. and T. Anjum, 2006. Control of *Parthenium hysterophorus* L. by aqueous extracts of allelopathic grasses. *Pak. J. Bot.*, 38: 139-145.

- Jefferson, L.V. and M. Pennacchio, 2003. Allelopathic effects of foliage extracts from four *Chenopodiaceae* species on seed germination. *J. Arid Environ.*, 55: 275-285.
- Khalid, S., T. Ahmad and R.A. Shad, 2002. Use of allelopathy in agriculture. *Asian J. Plant Sci.*, 1: 292-297.
- Kohli, R.K., D.R. Batish, H.P. Singh and K.S. Dogra, 2006. Status, invasiveness and environmental threats of three tropical American invasive Weeds (*Parthenium hysterophorus* L., *Ageratum conyzoides* L., *Lantana camara* L.) in India. *Biol. Invasions*, 8: 1501-1510.
- Kruidhof, H.M., L. Bastiaans and M.J. Kropff, 2009. Cover crop residue management for optimizing weed control. *Plant Soil*, 318: 169-184.
- Liebman, M. and C.L. Mohler, 2001. Weeds and the Soil Environment. In: *Ecological Management of Agricultural Weeds*, Liebman, M., C.L. Mohler and C.P. Staver (Eds.). Cambridge University Press, Cambridge, UK., pp: 322-374.
- Maguire, J.D., 1962. Speed of germination-aid in selection and evaluation for seedling emergence and vigor. *Crop Sci.*, 2: 176-177.
- Makino, T., Y. Takahashi, Y. Sakurai and M. Nanzyo, 1996. Influence of soil chemical properties on adsorption and oxidation of phenolic acids in soil suspension. *Soil Sci. Plant Nutr.*, 42: 867-879.
- Mohler, C.L., 1996. Ecological bases for the cultural control of annual weeds. *J. Prod. Agric.*, 9: 468-474.
- Morgan, E.C. and W.A. Overholt, 2005. Potential allelopathic effects of Brazilian pepper (*Schinus terebinthifolius* Raddi, Anacardiaceae) aqueous extract on germination and growth of selected Florida native plants. *J. Torrey Bot. Soc.*, 132: 11-15.
- Morra, M.J. and J.A. Kirkegaard, 2002. Isothiocyanate release from soil-incorporated *Brassica* tissues. *Soil Biol. Biochem.*, 34: 1683-1690.
- Mubarak, A.R., D.M.A. Daldoum and A.M. Sayed, 2009. Note on the influence of leaf extracts of nine trees on seed germination, radicle and hypocotyl elongation of maize and sorghum. *Int. J. Agric. Biol.*, 11: 340-342.
- Mutlu, S. and O. Atici, 2009. Allelopathic effect of *Nepeta meyeri* Benth. extracts on seed germination and seedling growth of some crop plants. *Acta Physiol. Plant.*, 31: 89-93.
- Okumura, M., A.B. Filonow and G.R. Waller, 1999. Use of ¹⁴C-labeled *Alfalfa saponins* for monitoring their fate in soil. *J. Chem. Ecol.*, 25: 2575-2583.
- Price, A.J., M.E. Stoll, F.J. Arriaga, J.S. Bergtold, T.S. Kornecki and R.L. Raper, 2008. Effect of cover crop extracts on cotton and radish radicle elongation. *Commun. Biom. Crop Sci.*, 3: 60-66.
- Rahnavard, A., H.R. Mashhadi, H.M. Alizade, S. Sadeghi and Z.Y. Ashrafi, 2008. Study of the allelopathic potential of extracts of *Azadirachta indica* (Neem). *Online J. Biol. Sci.*, 8: 57-61.
- Rietjens, I.M. and G.M. Alink, 2003. Nutrition and health-toxic substances in food. *Ned Tijdschr Geneeskde*, 147: 2365-2370.
- Seifu, W., 1990. *Parthenium hysterophorus* L., a recently introduced noxious weed to Ethiopia. A Preliminary Reconnaissance Survey Report on Eastern Ethiopia. Ministry of Agriculture, East Harerge, Ethiopia.
- Shafique, S., R. Bajwa, A. Javaid and S. Shafique, 2005. Biological control of *Parthenium* IV: suppressive ability of aqueous leaf extracts of some allelopathic trees against germination and early seedling growth of *Parthenium hysterophorus* L. *Pak. J. Weed Sci. Res.*, 11: 75-79.
- Singh, H.P., D.R. Batish, J.K. Pandher and R.K. Kohli, 2003. Assessment of allelopathic properties of *Parthenium hysterophorus* residues. *Agric. Ecosyst. Environ.*, 95: 537-541.

- Singh, H.P., D.R. Batish, J.K. Pandher and R.K. Kohli, 2005. Phytotoxic effects of *Parthenium hysterophorus* on three *Brassica* species. *Weed Biol. Manage.*, 5: 105-109.
- Stoll, M.E., A.J. Price and J.R. Jones, 2006. Cover crop extract effects on radish radicle elongation. *Proceedings of the Southern Conservation Systems Conference*, June 26-28, Amarillo, TX, pp: 184-186.
- Tamado, T. and P. Milberg, 2000. Weed flora in arable fields of eastern ethiopia with emphasis on the occurrence of *Parthenium hysterophorus* L. *Weed Res.*, 40: 507-521.
- Tamado, T., L. Ohlander and P. Milberg, 2002a. Interference by the weed *Parthenium hysterophorus* L. with grain sorghum: Influence of weed density and duration of competition. *Int. J. Pest Manage.*, 48: 183-188.
- Tamado, T., W. Schutz and P. Milberg, 2002b. Germination ecology of the weed *Parthenium hysterophorus* in Eastern Ethiopia. *Ann. Applied Biol.*, 140: 263-270.
- Teasdale, J.R. and C.I. Mohler, 2005. The quantitative relationship between weed emergence and the physical properties of mulches. *Weed Sci.*, 48: 385-392.
- Tefera, T., 2002. Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of *Eragrostis tef*. *J. Agron. Crop Sci.*, 188: 306-310.
- Tessema, T., 2002. Investigation of pathogens for biological control of parthenium (*Parthenium hysterophorus* L.) in Ethiopia. Doctoral Thesis, Humboldt-Universitat zu Berlin, Germany.
- VSN International Ltd., 2009. GenStat for Windows. Statistical Package. 12th Edn., VSN International, Hertfordshire, UK.
- Wakjira, M., G. Berecha and B. Bulti, 2005. Allelopathic effects of *Parthenium hysterophorus* extracts on seed germination and seedling growth of lettuce. *Trop. Sci.*, 45: 159-162.
- Wakjira, M., 2009. Allelopathic effects of *Parthenium hysterophorus* L. on onion germination and growth. *Allelo. J.*, 24: 351-362.
- Wakjira, M., G. Berecha and S. Tulu, 2009. Allelopathic effects of an invasive alien weed *Parthenium hysterophorus* L. compost on lettuce germination and growth. *Afr. J. Agric. Res.*, 4: 1325-1330.
- Wang, T.S.C., P.M. Huang, C.H. Chou and J.H. Chen, 1986. The Role of Soil Minerals in the Abiotic Polymerization of Phenolic Compounds and Formation of Humic Substances. In: *Interactions of Soil Minerals with Natural Organics and Microbes*, Huang, P.M. and M. Schnitzer (Eds.). Soil Science Society of America (SSSA), Madison WI, USA., pp: 251-281.
- Wiesner, M., T. Tessema, A. Hoffmann, P. Wilfried, C. Buettner, I. Mewis and C. Ulrichs, 2007. Impact of the Pan-Tropical weed *Parthenium hysterophorus* L. on human health in Ethiopia. *Utilisation of diversity in land use systems: Sustainable and organic approaches to meet human needs*. Tropentag, October 9-11, 2007, Witzenhausen. http://www.schoenmuth.de/Posterdok/Posterdokumentation-PM2007/2007-Tropentag/wiesner-2007_31.pdf.
- Wu, H., J. Pratley, D. Lemerle, T. Haig and M. An, 2001. Screening methods for the evaluation of crop allelopathic potential. *Bot. Rev.*, 67: 403-415.
- Xuan, T.D., T. Eiji, T. Hiroyuki, M. Mitsuhiro, T.D. Khanh and I.M. Chung, 2004. Evaluation on phytotoxicity of neem (*Azadirachta indica*. A. Juss) to crops and weeds. *Crop Prot.*, 23: 335-345.