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Research Article

Comparative Study on the Influence of Organic Fertilizer and Soil Amendments on Evening Primrose (*Oenothera biennis* L.)

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Abstract

Background: This study was done in the light of concept safety production of medicinal and aromatic plants. **Materials and Methods:** This study was conducted in open fields for two seasons continuously to compare the effects of organic fertilizer, soil amendments, a mixture of them and chemical NPK fertilizer (control) on growth parameters, floral characteristics and chemical constituents of *Oenothera biennis* L. (evening primrose) plants. **Results:** The outcome data pointed out that mixture of organic fertilizers (cattle manure and humic substances) and soil amendments (zeolite and magnetite) led to significant increment in morphological growth (plant height, number of branches, number of leaves, leaves area, leaves fresh weight and leaves dry weight), floral characteristics (number of flower, number of capsules, seed yield as well as health index) and chemical composition symbolized in plant pigments, total carbohydrates, net photosynthesis, stomatal conductance, water use efficiency, crude protein, total phenolics, N, P, K%, moreover indigenous hormones characterized in Indole Acetic Acid (IAA), gibberellic acid (GA₃), cytokinins (CK) and abscisic acid (ABA), fixed oil content in seeds and oil yield per hectare in comparison with the recommended dose of chemical fertilizer NPK (control) under the same conditions. From an economic view, the feasibility study of experiment clearly indicated that mix of organic and natural fertilizers proven its value and potential to achieve maximum profitable returns. **Conclusion:** These results disclose that mixture organic and natural resources of fertilizers could reduce or replace the addition of chemical fertilizers, accordingly improve the quality and quantity of medicinal and aromatic plants, besides minimizing economic costs and pollution of the agricultural environment.

Key words: Evening primrose, soil amendments, organic fertilizer, growth parameters, chemical composition, economical evaluation

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

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INTRODUCTION

Since the dawn of history, medicinal and aromatic plants were used for healing different diseases in all civilizations around the world. The commercial production of therapeutic and remedial plants has two major ways; chemical or/and organic. Hence, the industrialized agro-ecosystem term emerged, which represents one of the main human agricultural practices with the highest environmental emission of pollutants to the atmosphere, soil and water. On the side, organic way in contrast to synthetic materials means to enhance agroecosystem by application and use where possible natural resources to fulfill or improve crop production¹. Application of manures, composts, humic substances, biofertilizers, zeolites, magnetite feldspar and rock phosphate ores and other natural materials or beneficial microorganisms represent an organic way^{2,3}. So many researchers were reported the importance of manures, humic substances, natural zeolites and magnetite feldspar in organic production of crops represented in ameliorate chemical and physical properties of soil, high available water-holding capacity and high adsorption capacities, decreased soil pH, high cation exchange capacities, also produce long-term soil improvements as well as slow-release fertilizer, increased soil organic matter which enhances absorption of available nutrients and maintained nutrients from leaching or loss⁴⁻⁸.

The genus *Oenothera* consists of 145 species subdivided into 18 sections⁹. *Oenothera biennis* or evening primrose is a biennial plant of the Onagraceae family found in North America and parts of Asia Europe and Africa. It is grown commercially on a wide scale, later years; it has made the transition from being an undomesticated flower and backyard plant to an established agricultural crop¹⁰. The plant contains a range of different essential fatty acids including, palmitic acid, linoleic acid, oleic acid, γ -stearic acid and linolenic acid. The plant has been used to treat a variety of diseases including, atopic eczema, multiple sclerosis, coronary heart disease, psoriasis, diabetic neuropathy, autoimmune conditions, some types of cancer and gastrointestinal symptoms¹¹.

Since there is not needed of much water during the period of vegetation¹². Thus, it could be valuable alternative pharmaceutical purposes and oil crop in the new area (sandy soil) which have limited water resource in Egypt. So, the cultivation of evening primrose requires a proper fertilizer in a sandy soil, which have not; any impact on the environment and is considered an excellent product and to avoid any chemical residues.

Therefore, the key objectives of present study is to evaluate productivity, components and economically feasibility study of *O. biennis* plants yield when cultivated and grown in new reclaimed sandy soils under safety conditions [organic fertilizers (cattle manure and humic substances), soil amendments (zeolite and magnetite) and mixtures of them] comparing with commercial synthetic fertilizers symbolized in NPK.

MATERIALS AND METHODS

The present study was conducted at the experimental farm of Wadi El-Natron, Beheira Governorate at a private farm (newly reclaimed land of the desert) longitude 28°54' E, latitude 28°20' N and altitude 130 m in Egypt, during two consecutive seasons 2014 and 2015. Some physical and chemical analysis of the experimental farm soil (Table 1) was carried out as described by Page et al.¹³ in both seasons before planting. Oenothera biennis L. (Onagraceae) seeds were obtained from the experimental farm of Faculty of Pharmacy, Cairo University and sown in pots 7.5 (26×20 cm) filled with sand and peat (2:1) as a seed bed, at the end of October and harvested in May in both seasons, after 45 days (15-17 cm height) transplanted to open field with a distance of 60 cm between rows and 50 cm spacing between plants in plots with 6×7 m². Natural zeolite loaded with micronutrients as granules used in this study was obtained from Prima Company, Indonesia as shown in Table 2. The cattle manure was acquired from the Animals Production Department, Faculty of Agriculture, Cairo University (Table 3). While natural humic substances (Table 4) obtained from Soil and Water and Environment Research Institute, Agriculture Research Center (ARC). As for magnetite (Table 5) was provided by El-Ahram Company for Mining-Egypt. The experimental design was Randomized Complete Blocks Design (RCBD) with ten replicates.

Land preparation: Before planting, the soil was first mechanically plowed and propelled twice till the soil surface has been settled.

Fertilizers added and their combinations

Chemical fertilizers: Chemical fertilizers as recommended dose according to the Ministry of Agriculture and Land Reclamation were added at the rate of 474 kg ha⁻¹ as ammonium nitrate (33%) divided into two doses, the first was added after 2 weeks from planting, while the second was added 4 weeks later, both calcium superphosphate

Table 1: Chemica	al characteristics c	of the organic fer	Table 1: Chemical characteristics of the organic fertilizers cattle manure (CM) applied to <i>O. biennis</i> L.	iure (CM) appl.	ied to <i>O. bienn.</i>	s L.							
Organic	Moisture	Organic	Organic	z	N:O	NO ₃ -N	NH3-N	Ь	×	Fe	Zn	ŋ	Mn
characteristics	content (%)	matter (%)	carbon (%)	(%)	ratio	(mdd)	(mdd)	(%)	(%)	(mdd)	(mdd)	(mdd)	(mdd)
Seasons													Ī
1st	9.40	62.98	31.68	1.88	17.4 :1	173.90	49.40	0.29	1.18	1865.50	192.60	44.30	90.80
2nd	9.10	62.24	36.10	1.83	19.7:1	206.80	63.60	09:0	1.12	2690.90	179.80	43.20	159.20
Table 2: Chemic	Table 2: Chemical composition and physical properties of zeolite	nd physical prop	erties of zeolite										
(%) JJ	SiO ₂	TiO ₂	AI_2O_3	Fe ₂ O ₃	FeO	MnO	MgO	CaO	Na_2O	K ₂ O	SrO	P_2O_3	ΙΟΊ
	45.50	2.81	13.30	5.40	8.31	0.51	6.30	9.52	2.83	0.87	0.22	0.67	3.76
TE (ppm)	Ba	S	ڻ	Se	ŋ	Zn	Zr	qN	ïZ	Rb	>		
	10	1.2	35	0.8	19	64	257	13	55	15	22		Ī
						Physical	Physical properties						
Particle specific	Particle specific gravity (whole rock)	; K)	Ambient	Ambient moisture		Hardness	Hardness (mohs scale)		Cati	on Exchange C	Cation Exchange Capacity (CEC) meq/100 g ANZ38 power	eq/100 g ANZ	38 power
2.3	2.3 T m ⁻³		Nomir	Nominal <5%			5.0				147 average	4)	
CC: Chemical co	mposition (%), TE	: Trace elements,	CC: Chemical composition (%), TE: Trace elements, LOI: Loss on ignition	tion									
Table 3: Some c	Table 3: Some chemical composition of humic substances	ition of humic su	bstances										
					Cher	Chemical composition	tion						
Total nitrogen (N)		Phosphorus (P)	Potassium (K)	\S	Calcium (Ca)		Magnesium (Mg)	3)	Sulfur (S)	S)	Iron (Fe)		Boron (B)
1 400/	ď	70720	,000 C		1 000/		7007		/000		/000		/0200

Physical properties Color Black to silvery gray Streak Luster Metallic to sub-metallic Diaphaneity Opaque Mohs hardness 5-6.5 Specific gravity 5.2 Diagnostic properties Strongly magnetic, color, streak, octahedral crystal habit Tenacity Brittle Fracture Subcon choidal to uneven Transparency Opaque Crystal system Isometric

Table 4: Physical properties and chemical composition of magnetite

(15.5%) at the rate of 474 kg ha^{-1} and potassium sulfate (48%) at the rate of 118.5 kg fed⁻¹ were added 1 day before planting.

The most important ore of iron

Chemical composition

Mn, Zn, Mg (trace)

Fe++, Fe++

Cattle manure and zeolite: Both were added to the soil manually, where cattle manure at (47.4 m³ ha⁻¹) as well as zeolite (497.7 kg ha⁻¹), both were added 15 days before planting.

Magnetite: Added to the soil (355.5 kg ha⁻¹), added 1 week before planting.

Humic substances: Applied as solution 2.37 L ha⁻¹ at transplanting date, then 30 and 60 days, respectively per season. All agricultural practices were followed as recommended during both seasons.

Treatments: The treatments used in this study were as follows:

- NPK fertilizers as the control
- Cattle manure
- Zeolite

Chemical formula

 Fe_3O_4

- Humic substances
- Magnetite
- Mixture (Cattle manure+Zeolite+Humic substances+ Magnetite)

Data recorded

Growth parameters: Plant height (cm), the number of branches per plant, the number of leaves per plant, leaves area centimeter square per plant, leaves fresh weight (g) and leaves dry weight (g) were recorded.

Floral characteristics: A number of flowers per plant at 50% from opening, the number of capsules per plant, seed yield per hectare as well as health index were recorded.

The health index was determined using the following equation¹⁴:

Health index =
$$\frac{\text{Stem diameter} \times \text{dry weight}}{\text{Stem height}}$$

Chemical analysis

Total chlorophylls and carotenoid: Total chlorophylls and carotenoid contents (mg g^{-1} fresh weight) were measured by the spectrophotometer and calculated according to the equation described by Moran¹⁵.

Total carbohydrates (%): In plant leaves and seeds were determined by the phosphomolybdic acid method as reported by AOAC¹⁶.

Net photosynthesis, stomatal conductance and water use efficiency: Measurements of net photosynthesis on an area basis (μ mol CO₂ m⁻² sec⁻¹), leaf stomatal conductance (mol H₂O m⁻² sec⁻¹) and water use efficiency of five different leaves per treatment was monitored using a LICOR 6400 (Lincoln, Nebraska, USA) infrared gas analyzer (IRGA). Light intensity (Photosynthetically active radiation, PAR) within the sampling chamber was set at 1500 μ mol m⁻²sec⁻¹, using a Li-6400-02B LED light source (LI-COR). The CO₂ flow into the chamber was maintained at a concentration of 400 μ mol mol⁻¹ using an LI-6400-01 CO₂ mixer (LI-COR).

Nitrogen and crude protein: The total nitrogen content of the dried material (leaves and seeds) was determined by using the modified-micro-Kjeldahl method as described by AOAC¹⁶. The nitrogen percentage was multiplied by 6.25 to estimate the crude protein percentages in leaves and seeds.

Phosphorus: Phosphorus was determined calorimetrically by using the chlorostannate molybdophosphoric blue color method in sulfuric acid according to Jackson¹⁷.

Potassium: Potassium concentrations were determined by using the flame photometer apparatus (CORNING M 410, Germany)¹⁸.

Total phenolics content in seed: Soluble phenolics were extracted 5 times from defatted ground seed into aqueous 80% (v/v) methanol (at a ratio of 1:1, w/v) at room temperature for 1 h using an orbital shaker at 250 rpm. The mixture was centrifuged at 1750×g for 10 min and the supernatants were collected, combined, evaporated to near dryness under vacuum at <40°C and lyophilized. Phenolic acid present in the crude extract was fractionated into free and bound forms according to the procedure described by Kozlowska *et al.*¹⁹ and Zadernowski²⁰.

Endogenous phytohormones: Freeze-dried plant herbs (equivalent 6 g fresh weight) were ground to a fine powder in a mortar and pestle. The powdered material was extracted 3 times $(1\times3 \text{ and } 2\times1 \text{ h})$ with methanol (80% v/v,15 mL g⁻¹ fresh weight), supplemented with butylated hydroxytoluene (2,6-Di-tert-butyl-p-cresol) as an antioxidant at 4°C in darkness. The extract was centrifuged at 4000 rpm. The supernatant was transferred into flasks wrapped with aluminium foil and the residue was twice extracted again. The supernatants were combined and the volume was reduced to 10 mL at 35°C under vacuum. The aqueous extract was adjusted to pH 8.6 and extracted three times with an equal volume of pure ethyl acetate. The combined alkaline ethyl acetate extract was dehydrated over anhydrous sodium sulfate then filtered. The filtrate was evaporated to dryness under vacuum at 35°C and redissolved in 1 mL absolute methanol. The methanol extract was used after methylation for the determination of cytokinins (CK)²¹. The remaining aqueous extract was acidified to pH 2.6 and extracted as previously described by ethyl acetate. The methanol extract was used after methylation according to Fales et al.²² for determination of gibberellic acid (GA), abscisic acid (ABA) and indole-acetic acid (IAA). The quantification of the endogenous phytohormones was carried out with anti-unicum gas-liquid chromatography, 610 series, equipped with flame ionization detector according to the method described by Vogel²³. The fractionation of phytohormones was conducted using a coiled glass column (1.5 m \times 4 mm) packed with 1% OV-17. Gasses flow rates were 30, 30 and 330 mL min⁻¹, for nitrogen, hydrogen and air, respectively. The peaks identification and quantification of phytohormones were performed by using external authentic hormones and a Microsoft program to calculate the concentrations of the identified peaks.

Fixed oil content (Percentage of the seeds): Fixed oil extracted from seeds by using a Soxhlet apparatus. The oil

percentage was determined according to the methylation (change fixed oil into fatty acid) and GLC analysis was also recorded by G.C. mass in Medicinal and Aromatic Plant Laboratory Dokki (ARC) according to Kinsella²⁴ then oil yield per hectare was estimated in a hectare.

Methylation of fatty acid: Gas-liquid chromatographic analysis of fatty acid was done on methyl ester which was prepared and purified by the method of Kinsella²⁴ with some modifications. The methyl ester was prepared by refluxing the liberated fatty acids of *Oenothera* seeds with sulfuric acid (5 mL 1% v/v) in dried methanol for 30 min at 55°C. The fatty acid methyl esters were extracted several times with ether. The combined ether extracts were dried over anhydrous sodium sulfate, filtered and concentrated at 55°C.

GLC of fatty acid methyl esters: Separation of fatty acid methyl esters was carried out using capillary column, which contained 15% diethyl glycol succinate DEGS. The injector port and flame ionization detector were set at 240°C. The flow rate of carrier gas, nitrogen was 10 mL min⁻¹. The gas chromatograph (Perkin-Elmar model 8310) had a temperature program from 100-190°C with interment rate of 7°C min⁻¹. The initial and final time were identified according to their retention time compared to those of authentic samples.

Statistical analysis: The experimental design was Randomized Complete Blocks Design (RCBD) with 10 replicates. The data were analyzed using ANOVA at 5% significance level, the difference between treatments, then analyzed using Duncan Multiple Range Test (DMRT)²⁵ at 5%.

Economic evaluation: The yield components were calculated and economic analysis was performed using the following equations proposed by FAO²⁶ and Mubashir *et al.*²⁷:

- Gross income = Yield × price
- Profitable Return (PR) = Gross income-Total production
- PR% over control = PR-control treatments
- Benefit Cost Ratio (BCR) = PR over control/Total production cost
- Investment Factor (IF) = Gross income/Total production cost
- (IF) must equal or more than 3

RESULTS AND DISCUSSION

Growth parameters: Growth parameters of *O. biennis* [plant height, number of branches, number of leaves, leave area, leaves fresh and dry weight per plant (g)] and floral characteristics (number of flowers, number of capsules, seed yield as well as health index) represented in Table 6 showed significant variation with mixture application treatment (cattle manure+zeolite+humic substances+magnetite). In particular, mixture application treatment was significantly increased the number of branches (83%), the number of leaves (91%) and leaves dry weight per plant (83%) than the control, in both seasons. These results are in accordance with the findings of Ahmed *et al.*²⁸ on roselle plants, Ramadan² on cabbage and Salama⁶ on berseem clover and annual rye-grass.

This could be attributed to the positive effects of mixture application treatment (zeolite, magnetite, cattle manure and humic acid) on the improve physical and chemical properties of soil, high available water-holding and high adsorption capacities, decreased soil pH, higher CEC, increased soil organic matter which enhance absorption of available nutrients, in turn, enhanced plant growth and production, may affect phytohormone production leading to improve cell activity and plant growth^{2,29-34}.

Chemical composition

Total carbohydrates and photosynthetic pigments in leaves of *Oenothera biennis***:** Values of total carbohydrates and

photosynthetic pigments (carotenoids content and total chlorophyll) in leaves of *O. biennis* showed significant differences in comparison with the control treatment, whereas only in some treatments the contents of few minerals were similar or lower to control (Table 7). The most effective treatment in this respect was the mixture application treatment as plants for this treatment contained 22.55% total carbohydrates, 0.80 and 2.76 mg g⁻¹ carotenoids content and total chlorophyll, while, control plant gave 16.78%, 0.57 and 1 mg g⁻¹, in both successive seasons, respectively.

Increased total carbohydrates and photosynthetic pigments in mixture application treatment as compared to other treatments may be due to many biotic and abiotic factors are influencing the production of isoprenoid substrates for carotenogenesis³⁵. In addition, mixture treatment caused a considerable change in the soil pH that would have a significant increase in macronutrients availability (N, K, Mg, P and Ca) to plant roots, increased their uptake which increased

Organic-C (%) EC (dS m⁻¹ at 25 0.38 0.15 Total-N (%) 0.02 pH (soil paste) 7.8 0.45 SO 0.33 <u>_</u> **Texture** grade Sand HCO3-1.22 Physical properties Chemical properties HCO3-Anion (meq L⁻¹) Ő Clay (%) 1.99 0.22 ╧ Silt (%) 0.52 Na^{+} 7.21 ¥± 0.33 Cations (med L⁻¹) Sand (%)

able 5: Some physical and chemical properties of experimental farm

			Σ	No. of			Leave	eave area			Leaves dry	s dry			
	Plant hei	Plant height (cm)	branci pl	branches per plant	No. of leaves per plant (cm²)	aves (cm²)	per plant (g)	olant ۱)	Leaves fresh weight per	eaves fresh weight per	weight per plant plant (g)	er plant : (g)	No. of flowers per plant	wers nt	No. of capsule per plant
Treatments	1st	1st 2nd	1st		1st	2nd	1st		1st		1st		1st		1 1st
NPK control	116.8 ^b	116.8 ^b 123.3 ^b	9.0 _a	9.5ª	132.5 ^b	136.0 ^b	532.5 ^b	538.8 ^b	143.7 ^b	145.5 ^b	30.7 ^b	31.8 ^b	217 ^b	235 ^b	245 ^b
Cattle manure	113.5°	118.4 ^b	8.6ª	8.8 _a	126.6 ^b	131.9⁵	510.4€	540.7 ^b	134.4€	138.6₺	29.1 ^b	30.3 ^b	190€	228 ^b	236^{b}
Zeolite	110.1⁵	116.5€	7.4⁵	7.5 ^b	122.5€	125.6€	498.6€	529.2€	132.3€	135.5€	27.5€	27.9°	101⁴	155€	191€
Humic substances	115.6 ^b	120.9⁰	9.2ª	9.6	129.3 ^b	135.6 ^b	530.2 ^b	541.1 ^b	140.4⁵	143.0♭	30.1 ^b	30.8 ^b	209 ^b	230₀	233 ^b
Magnetite	112.2€	115.7	7.0 ^b	7.4 ^b	119.2⁵	124.8€	500.8€	519.3°	129.5⁴	131.8	25.5€	25.8	_p 68	142€	188€
Mix	126.3ª	126.3 ^a 129.5 ^a	10.5ª	11.7ª	145.5ª	151.0ª	763.6ª	781.0ª	156.5ª	160.2ª	37.2ª	38.5ª	260ª	288°	428ª

the number of chloroplast per cell as well as photosynthetic efficiency and increased sugar content in plants^{36,37}. Also, the positive influences of HA on chlorophylls and sugar contents could be mainly due to hormone-like activities of the humic acids through their involvement in cell respiration, photosynthesis, oxidative phosphorylation, protein synthesis and various enzymatic reactions^{38,39}. There are many reports which are in agreement with the present findings indicating that increased nitrogen supply elevated chlorophyll contents in *Clematis vitalba* leaves⁴⁰. Also, Al-Sahaf³ reported that chicken manure increased pigment in four *Brassica* vegetables. Moreover, Farouk *et al.*⁴¹ found that cucumber accumulated the highest carbohydrate content due to treatment with chitosan.

Net photosynthesis, stomatal conductance and water use efficiency: The highest significant values of net photosynthesis, stomatal conductance and water use efficiency (18.77 μ mol CO₂ m⁻² sec⁻¹, 1.65 mol H₂O m⁻² sec⁻¹ and 16.65 μ mol mmol⁻¹) were obtained from treatment of mixture application comparison with control plants (14.91 μ mol CO₂ m⁻² sec⁻¹, 1.45 mol H₂O m⁻² sec⁻¹, 13.15 μ mol mmol⁻¹) (Fig. 1) in both seasons.

Bittelli *et al.*⁴² concluded that the application of chelators increased significantly root growth as root length, which increased the plant's absorption ability. Moreover, application of chitosan assisted in conserving water the plants by closing the stomata and decreasing transpiration hence increasing relative water content in the leaves. Similar results were reported by Soliman and Mahmoud⁴³ on *Adansonia digitata*.

Indigenous hormones at stem elongation and flowering stage of *Oenothera biennis*: The obtained data in this concern clearly revealed that the highest values of IAA, GA₃ and CK hormones were recorded with mixture treatment in comparison with both control and all other treatments. Vice versa in the case of ABA hormone which records the lowest value with mixture treatment comparing with all other treatments (Table 8).

The increase in hormones by application of mixture treatment could be due to increasing the microbial populations resulting from adding cattle manure and humic acid in soils that led to the production of plant growth regulators by microorganisms^{44,45} or due to the effects of humates⁴⁶. In contrast, Mato *et al.*⁴⁷ concluded that the application of HA inhibits indole acetic acid (IAA) oxidase,

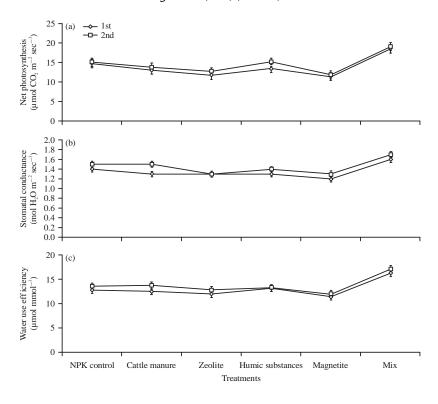


Fig. 1(a-c): Effect of different fertilizers and soil amendments on (a) Net photosynthesis, (b) Stomatal conductance and (c) Water use efficiency of *O. biennis* during both seasons 2014 (1st) and 2015 (2nd)

Table 7: Effect of different fertilizers and soil amendments on total carbohydrates and photosynthetic pigments of *O. biennis* during two seasons 2014 (1st) and 2015 (2nd)

	in lea	oohydrates ves (%)	(mg g ⁻¹ fre	3	(mg g ⁻¹ fre	lorophyll esh weight)
Treatments	 1st	2nd	 1st	 2nd	 1st	2nd
NPK control	8.23 ^b	8.55b	0.25°	0.32 ^b	0.47 ^b	0.53 ^b
Cattle manure	7.63 ^b	8.48 ^b	0.23 ^c	0.31 ^b	0.38 ^c	0.53 ^b
Zeolite	6.76 ^c	7.69 ^b	0.19 ^d	0.28 ^c	0.33 ^c	0.37 ^c
Humic substances	8.13 ^b	8.45 ^b	0.30 ^b	0.35 ^b	0.44 ^b	0.55 ^b
Magnetite	6.78 ^c	7.72 ^b	0.20 ^c	0.25°	0.33 ^c	0.35°
Mix	10.24 ^a	12.31ª	0.39a	0.41a	1.35 ^a	1.41ª

Means with the same letter in a column are not significantly different by p = 5%

Table 8: Effect different fertilizers and soil amendments on hormones at stem elongation and flower of O. biennis during two seasons 2014 (1st) and 2015 (2nd)

	GA3 (μg g ⁻¹ fresh weight)		IAA (μg g ⁻¹ f	fresh weight)	CK (µg g⁻¹	fresh weight)	ABA (μg g ⁻¹	fresh weight)
Treatments	 1st	2nd	1st	2nd	 1st	2nd	 1st	2nd
NPK control	13.27 ^b	14.82 ^b	10.09 ^b	11.14 ^b	9.12 ^b	9.62 ^b	7.24 ^b	6.22 ^b
Cattle manure	13.08 ^b	14.11 ^b	9.58 ^c	11.25 ^b	9.05⁵	9.68 ^b	7.98 ^b	6.41 ^b
Zeolite	10.24 ^c	11.18 ^c	9.26°	9.71°	8.69 ^c	8.94°	9.06ª	7.63ª
Humic substances	13.25 ^b	14.55 ^b	10.55 ^b	11.78 ^b	9.11 ^b	9.60 ^b	7.09 ^b	6.45 ^b
Magnetite	9.85⁵	11.71°	9.13€	9.42€	8.57 ^c	8.85°	9.17ª	7.07 ^a
Mix	16.71ª	16.89ª	13.26ª	13.67ª	11.81ª	12.21 ^a	5.47 ^c	4.38°

Means with the same letter in a column are not significantly different by p = 5%

thereby hindering the destruction of this plant growth hormone. In addition, Maheshwari⁴⁸ reported that magnetic treatments also affected by phytohormone production.

Total carbohydrates, protein, total phenolics content and some macro-nutrients in seed: Application of mixture treatment increased significantly total carbohydrates, protein

and total phenolics content in seed as compared to control plants in the both seasons (Table 9). Similar results were obtained by Khalil and El-Aref⁴⁹ and El-Shayeb⁵⁰.

Data in the Table 9 clearly indicate that mixture application treatment provided higher N, P and K% in seeds relative to control. The increased nutrient uptake due to the mixture treatment would be attributed to increasing root volume and surface area together would have led to the more nutrient uptake by providing better means for greater absorption as well as enhancement of microbial activity and reduced nutrient losses in the soil. In addition, the prevention of nutrient fixation in the soil, which led to increased availability of nutrient to the plants⁵¹. Similarly, several studies reported that humic acid, manure or zeolite applications increased, most macro-nutrient concentrations in many plants⁵²⁻⁵⁵.

Seed yield, health index, oil percentage and oil yield: The mixture application treatment produced the highest

significant seed yield, health index, oil percentage and oil yield values were 1291.06 kg ha⁻¹, 1.66, 23.80% and 265.32 L ha⁻¹, respectively, in both seasons (Table 10, Fig. 2). While, on the contrary, the control treatment produced the lowest significant seed yield, health index, oil percentage and oil yield values were 1111.17 kg ha⁻¹, 1.39, 20.24% and 145.95 L ha⁻¹, respectively, in both seasons.

The increase in seed yield, health index, oil percentage and oil yield of *O. biennis* plants in this study could be explained by increase the microbial populations resulting from adding cattle manure and humic acid in soils. These microorganisms can produce materials that may affect plant growth, such as substances acting as plant hormone analogs or growth regulators^{44,45}. Also, Mansour⁵⁶ and Mosa *et al.*⁵⁷ reported that this increase might be attributed to stimulating effect of magnetite on plant growth and the absorption of N, P, K and Ca. The previous result was supported by Habashy and Laila⁵⁸ on wheat crop with humic acid; Shehata *et al.*⁵⁹ on strawberries with compost, amino and

Table 9: Effect of different fertilizers and soil amendments on total carbohydrates, protein, total phenolic content, macro-nutrients in seeds of *O. biennis* during two seasons 2014 (1st) and 2015 (2nd)

	Total cark	oohydrates			•	nenolics in seeds						
		(g/100 g)		seeds (%)	. 5	dry matter)	N% in s		P% in		,	seeds
Treatments	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd	1st	2nd
NPK control	14.27 ^b	15.34 ^b	13.41 ^b	14.22 ^b	203.4 ^b	215.8 ^b	2.18 ^b	2.14 ^b	2.12 ^a	2.27ª	1.89 ^b	2.55ª
Cattle manure	12.11 ^c	13.79 ^c	12.15 ^c	13.26 ^c	191.6⁵	200.5 ^b	1.95℃	2.10 ^b	1.91°	2.21a	1.77b	2.34ª
Zeolite	11.73 ^d	12.96 ^d	10.68 ^d	11.31 ^d	177.3°	180.5€	1.07 ^c	1.20€	2.10 ^a	2.25ª	2.12ª	2.73a
Humic substances	14.70 ^b	15.11 ^b	13.35 ^b	14.05 ^b	197.8⁵	217.2ь	2.09b	2.17 ^b	1.97 ^c	2.18a	1.95⁵	2.40a
Magnetite	11.39 ^d	12.62 ^d	10.51 ^d	11.08 ^d	154.7 ^d	161.3 ^d	1.02€	1.17 ^c	2.06 ^b	2.11a	1.68 ^b	2.41a
Mix	17.18ª	18.47ª	17.32a	17.69ª	288.7ª	309.5ª	3.07a	3.15ª	2.79a	2.91ª	2.94ª	2.97ª

Means with the same letter in a column are not significantly different by p = 5%

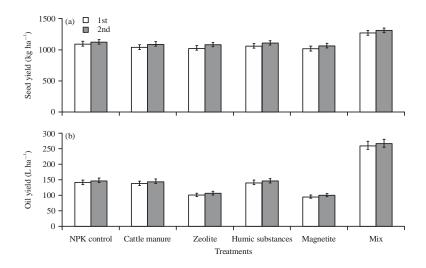


Fig. 2(a-b): Effect of different fertilizers and soil amendments on the (a) Seed yield and (b) Oil yield of *O. biennis* during both seasons 2014 (1st) and 2015 (2nd)

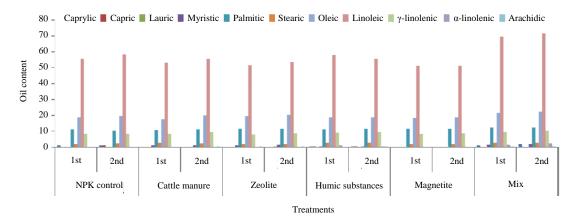


Fig. 3: Effect of different fertilizers and soil amendments on the major compounds of oil of *O. biennis* during both seasons 2014 (1st) and 2015 (2nd)

Table 10: Effect of different fertilizers and soil amendments on the chemical composition of O. biennis during two seasons 2014 (1st) and 2015 (2nd)

	Health index	·	Oil seeds (%)	
Treatments	1st	2nd	1st	2nd
NPK control	1.18	1.60	19.85 ^b	20.62 ^b
Cattle manure	1.10	1.10	18.36°	19.95 ^b
Zeolite	0.99	0.95	17.75 ^d	18.59 ^c
Humic substances	1.17	1.14	19.42⁵	20.88 ^b
Magnetite	0.88	0.86	17.22 ^d	18.34 ^c
Mix	1.56	1.57	22.68ª	24.91ª

Means with the same letter in a column are not significantly different by p = 5%

Table 11: Effect of economical evaluation of different treatments

						Profitable		<u> </u>
					Profitable return	return increase		
	Total yield	Total production	Gross income	Profitable return	over control	percentage		
Treatments	(L ha ⁻¹)	$cost (US $ ha^{-1})$	$(US\$ ha^{-1})$	(PR) (US $$ ha^{-1}$)	(US\$ ha ⁻¹)	$(US\$ ha^{-1})$	BCR	IF
NPK control	148.6	463.3	19543	19079.0				42.1
Cattle manure	146.5	443.1	19227	18784.0	295.0	3.64	0.66	43.3
Zeolite	108.0	403.8	14184	13780.6	5299.0	65.60	13.10	35.1
Humic substances	147.2	369.2	19540	19171.4	91.7	1.13	0.24	52.7
Magnetite	101.8	294.3	13238	12944.4	6135.2	76.00	20.80	44.9
Mix	268.4	497.7	35618	43653.0	24573.3	305.00	49.30	71.5

humic acids and Mohamed *et al.*⁶⁰ on orange trees with magnetite and some biofertilizers application.

Oil components: Concerning the effect of different treatments on fatty acids resulted from chromatographic analysis of oil components (Fig. 3). It could be concluded that the main fatty acids represented in caprylic, myristic, palmitic, stearic, oleic, linoleic, γ -linolenic and α -Linolenic gave the utmost amount comparing with either control plants or all other treatments in both first and second season. Contrary to that, the highest amount of capric acid (0.24-1.08) in both seasons was obtained from control plants. With reference to lauric acid,

which recorded the highest amount (0.38-0.47) with zeolite treatment in comparison with both control and all other treatments.

The increment of major oil content with mixture treatment could be explained on the basis of available elements, vitamins, gibberellins, cytokines, hormone-like substances, amino acids and sugars that lead to an increase in biochemical processes within the plant (the luxury of metabolism) consequently an increase in fatty acid content. These results concurred with those of Azim⁶¹ on *Oenothera biennis*, who stated that, application of NPK caused variation in fatty acid content, Abd El-Latif⁶² on caraway plants,

reported that organic manure affected oil content and Mahmoud⁶³ on yarrow plants who declared that zeolite and organic fertilizers greatly influenced essential oil content.

Economic evaluation: The data in Table 11 clearly revealed that mixture treatment realized a maximum production cost of 210 US\$ ha⁻¹, while the minimum production cost resulted from magnetite treatment. Generally, from data analysis, the cost of commercial production of *O. biennis* differed with variable types of treatments, meanwhile, all treatments fulfilled reasonable profitability because of their (IF)²⁶ more than 3. The maximum profitable return of 18419 was attained when mix treatment applied. Concerning Benefit Cost Ratio (BCR), the value of BCR was economical with the application of mix since it recorded a value of 49.3. Regarding, Investment Factor (IF), data showed that the highest IF of 71.5 was recorded when mix treatment applied and gross income 15029 US as well.

CONCLUSION

The addition of mixture application (cattle manure+zeolite+humic substances+magnetite) to the soil and plant improved growth parameters and chemical compositions of *O. biennis* plants that led to decrease the addition of chemical fertilizer rate to the level that could replace it with mentioned mix treatment and reduce the production cost. Further developments in using such safety and natural products in this sector could have large-scale economic implications and multiple benefits for consumers, producers, farmers and the ecosystem. Therefore, a mentioned mixture application can be considered as an economical fertilization for *O. biennis* plants.

SIGNIFICANCE STATEMENTS

Best results with a mixture of organic fertilizers and soil amendments which increased:

- Growth parameters (Plant height, the number of branches, number of leaves per plant, leaves an area, leaves fresh and dry weight)
- Floral characteristics (Number of flowers per plant, the number of capsules per plant, seed yield per hectare as well as health index)
- Chemical compositions (total chlorophylls and carotenoid, total carbohydrates, net photosynthesis,

- stomatal conductance and water use efficiency, N, P, K%, crude protein, total phenolic content in seed and endogenous phytohormones)
- The increment of seed yield, health index, oil percentage and oil yield
- The oil content of seeds in a mixture of organic fertilizers and soil amendments which gave the utmost amount comparing with either control plants or all other treatments especially y-linolenic acid content

REFERENCES

- FAO., 2004. The scope of organic agriculture, sustainable forest management and ecoforestry in protected area management. Working Paper No. 18, Environment and Natural Resources Service Sustainable Development Department, Rome, Italy, pp: 53.
- Ramadan, M.E., 2012. Effect of sulphur, magnetite and potassium humate on productivity and quality of cabbage under Ras Sudr conditions. Ph.D. Thesis, Ain Shams University, Egypt.
- Al-Sahaf, F.H., 2013. Photosynthetic pigments affected by fertilizer source in four brassica vegetables. J. Agric. Sci. Technol. B, 3: 246-253.
- Fahramand, M., H. Moradi, M. Noori, A. Sobhkhizi, M. Adibian, S. Abdollahi and K. Rigi, 2014. Influence of humic acid on increase yield of plants and soil properties. Int. J. Farm. Allied Sci., 3: 339-341.
- 5. Makungwe, M., 2014. Evaluation of the potential of zeolite as a soil conditioner for two Zambian soils. M.Sc. Thesis, University of Zambia, Zambia.
- 6. Salama, H.S.A., 2015. Interactive effect of forage mixing rates and organic fertilizers on the yield and nutritive value of berseem clover (*Trifolium alexandrinum* L.) and annual ryegrass (*Lolium multiflorum* Lam.). Agric. Sci., 6: 415-425.
- Fuchs, J.G. and W.J.M. Cuijpers, 2016. Compost Types, Feedstocks and Composting Methods. In: Handbook for Composting and Compost Use in Organic Horticulture, Van der Wurff, A.W.G., J.G. Fuchs, M. Raviv and A.J. Termorshuizen (Eds.). BioGreenhouse COST Action FA, USA., ISBN: 9789462577497, pp: 29-43.
- Khomami, M.A., G.M. Mammadov, F.A. Chokami and S. Sedaghathoor, 2016. Growth and reproductive performance of *Eisenia fetida* in cow manure, cow manure+sugarcane bagasse and cow manure+sawdust waste. Applied Ecol. Environ. Res., 14: 237-247.
- Agrawal, A.A., A.P. Hastings, M.T.J. Johnson, J.L. Maron and J.P. Salminen, 2012. Insect herbivores drive real-time ecological and evolutionary change in plant populations. Science, 338: 113-116.

- 10. Ahmad, Z. and P. Lapinskas, 1998. Evening primrose-A plant of nutritional and pharmacological importance. Proceedings of the Novel Crops Symposium, November 1998, Islamabad.
- 11. ACS., 2009. Evening primrose. American Cancer Society (ACS). http://www.cancer.org/docroot/ETO/content/ETO_5_3X_E vening_Primrose.asp?sitearea=ETO
- 12. Hoppe, B., 2009. Handbuch des arznei-und gewurzpflanzenbaus-band 5: Arznei-und gewurzpflanzen L-Z. Verein fur Arznei-Und Gewurzpflanzen, Bernburg, pp: 646-654.
- Page, A.L., R.H. Miller and D.R. Keeney, 1982. Methods of Soil Analysis, Part 2: Chemical and Mineralogical Properties.
 2nd Edn., American Society of Agronomy, Madison, Wisconsin, USA.
- Fan, X.X., Z.G. Xu, X.Y. Liu, C.M. Tang, L.W. Wang and X.L. Han, 2013. Effects of light intensity on the growth and leaf development of young tomato plants grown under a combination of red and blue light. Scient. Hortic., 153: 50-55.
- Moran, R., 1982. Formulae for determination of chlorophyllous pigments extracted with N,N-dimethylformamide. Plant Physiol., 69: 1376-1381.
- AOAC., 1990. Official methods of Analysis. 14th Edn., Association of Official Analytical Chemists, Washington DC., Pages: 1094.
- 17. Jackson, M.L., 1973. Soil Chemical Analysis. Printice-Hall of India, New Delhi, pp: 144-197, 381.
- 18. Al-Khayri, J.M., 2002. Growth, proline accumulation and ion content in sodium chloride-stressed callus of date palm. *In Vitro* Cell. Dev. Biol.-Plant, 38: 79-82.
- Kozlowska, H., D.A. Rotkiewicz, R. Zadernowski and F.W. Sosulski, 1983. Phenolic acids in rapeseed and mustard. J. Am. Oil Chem. Soc., 60: 1119-1123.
- 20. Zadernowski, R., 1987. Studies on phenolic compounds of rapeseed flours. Acta Academiae Agriculturae Technicae Olsteniensis Technologia Alimentorum, 21: 3-55.
- 21. Plamer, M.V., R. Hogan and P.F. Wareing, 1981. Cytokinins metabolism *Phaseolus vuglaris* L. identification of endogenous cytokinins and metabolism of (8-C¹⁴) dihrozeatin in stems of decapitated plants. Palnta, 153: 297-302.
- 22. Fales, H.M., T.M. Jaouni and J.F. Babashak, 1973. Simple device for preparing ethereal diazomethane without resorting to codistillation. Anal. Chem., 45: 2302-2303.
- 23. Vogel, A.I., 1975. A Text Book of Practical Organic Chemistry. 3rd Edn., English Language Book Socity and Longman Group Ltd., USA., pp: 197-596.
- 24. Kinsella, J.E., 1966. Metabolic patterns of the fatty acids of *Periplaneta americana* (L.) during its embryonic development. Can J. Biochem., 44: 247-258.
- 25. Duncan, D.B., 1955. Multiple range and multiple F tests. Biometrics, 11: 1-42.
- 26. FAO., 2000. Fertilizers and their Use. 4th Edn., Food and Agriculture Organization, Rome, Pages: 34.

- 27. Mubashir, M., S.A. Malik, A.A. Khan, T.M. Ansari, S. Wright, M.V. Brown and K.R. Islam, 2010. Growth, yield and nitrate accumulation of irrigated carrot and okra in response to nitrogen fertilization. Pak. J. Bot., 42: 2513-2521.
- Ahmed, Y.M. E.A. Shalaby and N.T. Shanan, 2011. The use of organic and inorganic cultures in improving vegetative growth, yield characters and antioxidant activity of Roselle plants (*Hibiscus sabdariffa* L.). Afr. J. Biotechnol., 10: 1988-1996.
- 29. Mumpton, F.A., 1999. *La roca magica*: Uses of natural zeolites in agriculture and industry. Proc. Natl. Acad. Sci. USA., 7: 3463-3470.
- 30. Aleksiev, B., E. Djourova and Z. Milakovsk-Vergilova, 2000. Zeolitic rocks from the Mezek Mountain, NE Rhodopes. Geologiya, 92: 159-165.
- 31. Fiorentino, G., R. Spaccini and A. Piccolo, 2006. Separation of molecular constituents from a humic acid by solid-phase extraction following a transesterification reaction. Talanta, 68: 1135-1142.
- 32. Kavoosi, M., 2007. Effects of zeolite application on rice yield, nitrogen recovery and nitrogen use efficiency. Commun. Soil Sci. Plant Anal., 38: 69-76.
- 33. Castrillon, L., Y. Fernandez-Nava, E. Maranon, L. Garcia and J. Berrueta, 2009. Anoxic-aerobic treatment of the liquid fraction of cattle manure. Waste Manage., 29: 761-766.
- 34. Indraratne, S.P., X. Hao, C. Chang and F. Godlinski, 2009. Rate of soil recovery following termination of long-term cattle manure applications. Geoderma, 150: 415-423.
- 35. Cordoba, E., M. Salmi and P. Leon, 2009. Unravelling the regulatory mechanisms that modulate the MEP pathway in higher plants. J. Exp. Bot., 10: 2933-2943.
- 36. Taiz, L. and E. Zeiger, 2006. Plant Physiology. 4th Edn., Sinauer Associates Inc. Publishers, Sunderland, Massachusetts, ISBN-10: 0-87893-856-7, pages: 764.
- Farouk, S., A.A. Mosa, A.A. Taha, H.M. Ibrahim and A.M. El-Gahmery, 2011. Protective effect of humic acid and chitosan on radish (*Raphanus sativus*, L. var. sativus) plants subjected to cadmium stress. J. Stress Physiol. Biochem., 7: 99-116.
- 38. Chen, Y. and T. Aviad, 1990. Effects of Humic Substances on Plant Growth. In: Humic Substances in Soil and Crop Sciences: Selected Readings, MacCarthy, P., C.E. Clapp, R.L. Malcolm and P.R. Bloom (Eds.). Soil. Sci. Soc. Am., Madison, Wisconsin, USA., pp: 161-186.
- 39. Muscolo, A., F. Bovalo, F. Gionfriddo and S. Nardi, 1999. Earthworm humic matter produces auxin-like effects on *Daucus carota* cell growth and nitrate metabolism. Soil Biol. Biochem., 31: 1303-1311.
- 40. Bungard, R.A., D. McNeil and J.D. Morton, 1997. Effects of nitrogen on the photosynthetic apparatus of *Clematis vitalba* grown at several irradiances. Aust. J. Plant Physiol., 24: 205-214.

- 41. Farouk, S., K.M. Ghoneem and A.A. Ali, 2008. Induction and expression of systematic resistance to downy mildew disease in cucumber plant by elicitors. Egypt. J. Phytopathol., 36: 95-111.
- 42. Bittelli, M., M. Flury, G.S. Campbell and E.J. Nichols, 2001. Reduction of transpiration through foliar application of chitosan. Agric. Forest Meteorol., 107: 167-175.
- 43. Soliman, A.S. and A.M. Mahmoud, 2013. Response of *Adansonia digitata* to compost and zeolite in replacement of chemical fertilization. Am.-Eurasian J. Agric. Environ. Sci., 13: 198-206.
- 44. Brown, G.G., 1995. How do earthworms affect microfloral and faunal community diversity? Plant Soil, 170: 209-231.
- 45. Atiyeh, R.M., S. Lee, C.A. Edwards, N.Q. Arancon and J.D. Metzger, 2002. The influence of humic acids derived from earthworm-processed organic wastes on plant growth. Bioresour. Technol., 84: 7-14.
- 46. Canellas, L.P., F.L. Olivares, A.L. Okorokova-Facanha and A.R. Facanha, 2002. Humic acids isolated from earthworm compost enhance root elongation, lateral root emergence and plasma membrane H⁺-ATPase activity in maize roots. Plant Physiol., 130: 1951-1957.
- 47. Mato, M.C., M.G. Olmedo and J. Mendez, 1972. Inhibition of indoleacetic acid-oxidase by soil humic acids fractionated on sephadex. Soil Biol. Biochem., 4: 469-473.
- 48. Maheshwari, L.B., 2009. Magnetic treatment of irrigation water: evaluation of its effects on vegetable crop yield and water productivity. Ph.D. Thesis, Western Sydney University, New South Wales.
- 49. Khalil, F.A. and K.A.O. EL-Aref, 2001. Bio-fertilizers as partner with mineral N-fertilizer for fertilizing wheat crop cultivar Sids 6. J. Agric. Sci. Mansoura Univ., 26: 8287-8295.
- 50. El-Shayeb, N.S.A., 2009. Physiological studies on *Oenothera biennis* plants. Ph.D. Thesis, Faculty of Science, Benha University, Egypt.
- 51. Raina, J.N. and K.P. Goswami, 1988. Effect of fulvic acid and fulvates on the growth and nutrient uptake by maize plant. J. Indian Soc. Soil Sci., 36: 264-268.
- 52. Butler, T.J. and J.P. Muir, 2006. Dairy manure compost improves soil and increases tall wheatgrass yield. Agron. J., 98: 1090-1096.
- 53. Gevrek, M.N., O. Tatar, B. Yagmur and S. Ozaydin, 2009. The effects of clinoptilolite application on growth and nutrient ions content in rice grain. Turk. J. Field Crops, 14: 79-88.

- 54. Mahmoodabadi, M.R., A. Ronaghi, M. Khayyat and G. Hadarbadi, 2009. Effects of zeolite and cadmium on growth and chemical composition of soybean (*Glycine max* L.). Trop. Subtrop. Agroecosyst., 10: 515-521.
- Yolcu, H., H. Seker, M.K. Gullap, A. Lithourgidis and A. Gunes, 2011. Application of cattle manure, zeolite and leonardite improves hay yield and quality of annual ryegrass (*Lolium multiflorum* Lam.) under semiarid conditions. Aust. J. Crop Sci., 5: 926-931.
- 56. Mansour, E.R., 2007. Effect of some culture practices on cauliflower tolerance to salinity under Ras Suder conditions. M.Sc.Thesis, Ain Shams University, Egypt.
- 57. Mosa, W.F.A.E.G., L.S. Paszt, M. Frac and P. Trzcinski, 2015. The role of biofertilization in improving apple productivity-a review. Adv. Microbiol., 5: 21-27.
- 58. Habashy, N.R. and K.M. Laila, 2005. Effects of organic growth-promoting substances as humic acids and indole-acetic acid on wheat crop with special reference to some chemical composition. Minufiy J. Agric. Res., 30: 1607-1624.
- 59. Shehata, S.A., A.A. Gharib, M.M. El-Mogy, K.F. Abdel Gawad and E.A. Shalaby, 2011. Influence of compost, amino and humic acids on the growth, yield and chemical parameters of strawberries. J. Med. Plants Res., 5: 2304-2308.
- 60. Mohamed, H.M., F.A. Al-Kamar and A.A.M. Abd-Elall, 2013. Effect of magnetite and some biofertilizer application on growth and yield of Valencia orange trees under El-Bustan condition. Nat. Sci., 11: 46-61.
- 61. Azim, G., 2007. Investigations on the effects of harvest methods and storage conditions on yield, quality and germination of evening primrose (*Oenothera biennis* L.) seeds. Ph.D. Thesis, Institute of Crop Science and Plant Breeding, Justus Liebig University Giessen.
- 62. Abd El-Latif, T.A., 2002. Effect of organic manure and biofertilizer on caraway plants (*Carum carvi* L.). J. Agric. Sci. Mansoura Univ., 27: 3463-3472.
- 63. Mahmoud, A.W.M., 2009. Enhancing of growth, essential oil yield and component of yarrow plants (*Achillea millefolium*) grown under safe agriculture conditions. Proceedings of the 1st International Conference on Biological Science, October 16-17, 2009, Yogyakarta, Indonesia, pp: 130-141.