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## Screening of Antibiotic Resistant Inhibitors from Indian Traditional Medicinal Plants Against *Streptococcus mutans*

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**Abstract:** Aqueous and ethanolic extracts of ten plants used in traditional Indian medicine for treatment of an infectious nature were screened for antibacterial activity by disc diffusion method against *Streptococcus mutans*. Among them the ethanol extracts of *Allium sativum* bulb exhibited high degree of inhibition followed by *Azadirachta indica* leaf extracts. The aqueous extracts of plants did not show any antibacterial activity against *Streptococcus mutans*.

**Key words:** Antibacterial activity, *Streptococcus mutans*, traditional medicine, high degree of inhibition, dental caries

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### INTRODUCTION

Human infections caused by Gram-positive bacterial pathogens are increasingly difficult to treat, predominantly due to emergence of drug resistant bacterial strains. One such Gram-positive bacterial organism is *Streptococcus mutans* causing dental caries in humans (Hamada and Slade, 1980; Akhtar and Bhakuni, 2004). This bacterium adheres firmly to the smooth tooth surfaces and produces sticky water insoluble dextran from dietary sucrose, forming plaque which facilitates the accumulation of microorganisms. *Streptococcus mutans* and other organisms in the plaque produce organic acids such as lactic acid that gradually destroy the enamel and form a cavity (Bhattacharya *et al.*, 2003). Elimination of *Streptococcus mutans* would reduce the incidence of dental caries. Extensive efforts have been made to find an active agent against dental caries. However *Streptococcus mutans* was found to be resistant to many of the antibacterial agents viz., Penicillin, amoxicillin, cefuroxin, tetracycline and erythromycin (Jarvinen *et al.*, 1993; Bhattacharya *et al.*, 2003). In addition they may lead to side effects including gastrointestinal problems (Craig, 1998). These drawbacks justifies further research and development of natural antimicrobial agents that are effective and safe for the host.

It has been well documented that traditional medicinal plants confer considerable antibacterial activity against various microorganisms (Jonathan *et al.*, 2000). Many plants were reported to inhibit the growth of many oral bacteria (Pack *et al.*, 1998), particularly *Streptococcus mutans* and control plaque and thus prevent caries have been investigated (Jagtap and Karkera, 2000; Margan *et al.*, 2001). Use of plant based alternatives for oral health has been successfully promoted for example, the use of antibacterial chewsticks has been widely advocated by health agencies where their use is culturally acceptable (Almas, 1999; Jagtap and Karkera, 2000). In the present study an attempt has been made to enrich the knowledge of antibacterial activity of ten Indian traditional medicinal plants that have been screened for antistreptococci activity.

### MATERIALS AND METHODS

#### Ethnobotany and Plant Collection

The ethnobotanical survey was conducted in Shevaroy and Kolli hills of Eastern Ghats of Tamil Nadu (Fig. 1), South India from January to July 2000. The comparison between medicinal and

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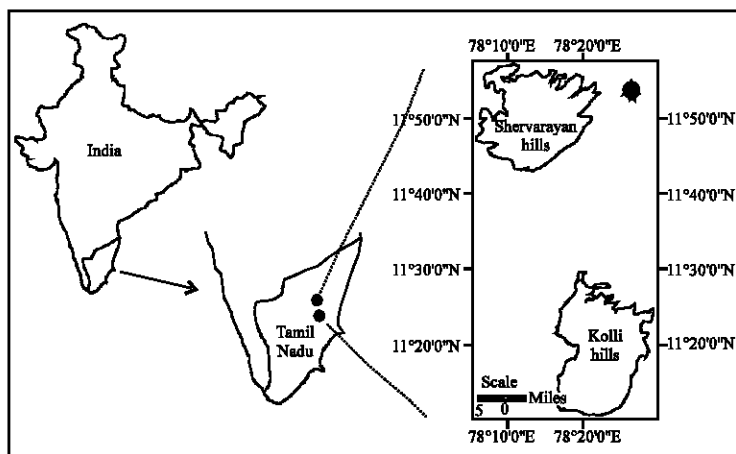


Fig. 1: Location map of Kolli and Shevaroy hills

non-medicinal plants is based on research in the community namely Malayalis of the same areas. Based on their information and literature we have collected 12 medicinal plants for antibacterial activity against *Streptococcus mutans*.

#### Extraction of Plant Materials

The collected plant materials were separated into leaf, root, bulb and bark and shade dried at 37°C for 7 days. The extracts were prepared using the solvents viz., water and ethanol. Dried plant materials (20 g) were extracted with 100 mL of water and ethanol for 72 h. Each mixture was stirred every 24 h using a sterile glass rod. At the end of fraction each extract was passed through Whatman No.1 filter paper. The filtrate obtained was concentrated in vacuo using rotary evaporator.

#### Test Bacteria

The test bacterium *Streptococcus mutans* was collected from Clinical Microbiology Laboratory, Sea Horse Hospital, Tiruchirappalli. Brain Heart Infusion (BHI, Difco) was used for the culture of *Streptococcus mutans*.

#### Antibacterial Assay

The disc diffusion assay of Iennette (1985) as described by Rosanaivo and Ratsimananga-Urverg (1993) and Rabe and Van Staden (1997) were used with modification to determine the growth rates of bacterium with plants extracts. A diluted bacterial cultures (200 µL) was spread over Muller Hinton Agar Plates using sterile glass L-rod. Two hundred microliter of each extract was applied per filter paper disc (Whatman No.1, 6 mm diameter) and was allowed to dry before placing them on the agar plates in petriplates. Each extract was tested in triplicate (3 discs/plate) and the plates were incubated at 37°C for 24 h and size of the growth zones were recorded. Calculations were carried out in triplicate with their mean values and standard deviation by using formula given by Gupta (1977).

## RESULTS AND DISCUSSION

The ethanolic bulb extract of *Allium sativum* was the most active against *Streptococcus mutans*. It showed high degree of inhibition (25.6 mm) followed by *Azadirachta indica* leaf extracts exhibited significant inhibition (24.4 mm) against the test bacterium. The ethanolic extracts of *Embelia ribes* fruit

Table 1: Antibacterial activity of ten Indian Traditional medicinal plants

Family and Botanical name	Local name	Parts used	Anti-infective uses	Location/ Tribals	Solvent used	Inhibition zone Mean±SD
Liliaceae <i>Allium sativum</i>	Venkaya Poonda	Bulb	Antiseptic	Kolli Hills, Malayalis	Aqueous Ethanol	- 25.6±0
Meliaceae <i>Azadirachta indica</i>	Veppam	Leaves	Skin infections, oral infections	Kolli Hills, Malayalis	Aqueous Ethanol	- 24.4±0
Myrsinaceae <i>Embelia ribes</i>	Vizhalari	Fruits	Antiseptic	Shevaroy hills, Malayalis	Aqueous Ethanol	- 23.5±0
Papavaraceae <i>Argemone maxicana</i>	Bramma Thundu	Leaves	Skin infections	Shevaroy hills, Malayalis	Aqueous Ethanol	- 22.7±0
Rubiaceae <i>Citrus medica</i>	Ezhumitchai	Roots	Asthma, Tumours, Skin infections	Shevaroy hills, Malayalis	Aqueous Ethanol	- 22.0±0
Mimosaceae <i>Acacia nilotica</i>	Shivakaay	Bark	Skin diseases and Scabies	Kolli Hills, Malayalis	Aqueous Ethanol	- 22.3±0
Asclepiadaceae <i>Calotropis gigantea</i>	Erikku	Boiled leaves	Palalyses, Tumours	Kolli Hills, Malayalis	Aqueous Ethanol	- 22.0±0
Alangiaceae <i>Alangium salvifolium</i>	Alangi	Leaf	Dog bite, fever, skin diseases	Kolli Hills, Malayalis	Aqueous Ethanol	- 22.3±0
Lamiaceae <i>Ocimum sanctum</i>	Tuzhasi	Leaf	Skin diseases, Dental caries	Kolli Hills, Malayalis	Aqueous Ethanol	- -
Menispermaceae <i>Tinospora cordifolia</i>	Chenthil Kodi	Leaf	Dyspepsia, Skin diseases, fever	Shevaroy hills, Malayalis	Aqueous Ethanol	- -

showed moderate inhibition (23.5 mm). It was noted that the *Streptococcus mutans* was less susceptible to the extracts of *Citrus medica* roots (22 mm), *Acacia nilotica* barks (22.3 mm) and *Calotropis gigantea* leaves whereas the leaf residues of *Ocimum sanctum* and *Tinospora cordifolia* did not show any antibacterial effect on *Streptococcus mutans*. Complete absence of measurable inhibitory action was observed in the aqueous extracts of ten Indian Traditional medicinal plants (Table 1).

Among the plants tested *Allium sativum* was very promising and showed significant inhibition against the test bacterium. Numerous workers (Willis, 1956; Mantis *et al.*, 1979; Jain, 1994) have obtained that garlic juice exhibits strong antibacterial properties and due to these properties garlic juice is very effective against various diseases. It is postulated that the antibacterial properties of garlic juice are due to the inhibition of succinic dehydrogenase via the inactivation of this groups (Srinivasan *et al.*, 2001).

## REFERENCES

- Akhtar, M.S. and V. Bhakuni, 2004. *Streptococcus pneumoniae* hyaluronat lyase: An overview. *Curr. Sci.*, 86: 285-295.
- Almas, K., 1999. The antimicrobial effect of *Azadirachta indica* (Neem) and *Salvadora persica* (Arak) chewing sticks. *Ind. J. Dental Res.*, 10: 23-26.
- Bhattacharya, S., S. Virani, M. Zavro and G.J. Hass, 2003. Inhibition of *Streptococcus mutans* and other oral Streptococci by Hop (*Humulus lupulus* L.) constituents. *Econ. Bot.*, 57: 118-125.
- Craig, A., 1998. Antimicrobial resistance, danger signs all around. *Tennessee Medicine*, 91: 433-455.
- Gupta, S.P., 1977. *Statistical Methods*. S. Chand and Co., New Delhi.
- Hamada, S. and H.D. Slade, 1980. Biology, Immunology and carcinogenicity of *Streptococcus mutans*. *Microbiol. Rev.*, 44: 331-384.
- Iennette, E.H., 1985. *Manual of Clinical Microbiology*. American Association for Microbiology, Washington, DC.
- Jagtap, A.G. and S.G. Karkera, 2000. Extracts of *Juglandaceae regia* inhibits growth *in vitro* adherence, acid production and aggregation of *Streptococcus mutans*. *J. Pharm. Pharmacol.*, 52: 235-242.
- Jain, R.C., 1994. Antibacterial activity of garlic extract. *Ind. J. Med. Microbiol.*, 11: 26-31.

- Jarvinen, H., J. Jenevou and P. Huovinen, 1993. Susceptibility of *Streptococcus mutans* to Chlorhexidine and six other antimicrobial agents. *Antimicrob. Agents Chemother.*, 37: 1158-1159.
- Jonathan, E.K., K.J. Anna and V. Johannes, 2000. Zulu medicinal plants with antibacterial activity. *J. Ethnopharmacol.*, 69: 241-246.
- Mantis, A.J., P.A. Koidis, P.G. Karajonnoglou and A.G. Panetos, 1979. Effect of garlic extract on food poisoning bacteria, *lebensm Wiss U. Technol.*, 12: 330.
- Margan, T.D., A.E. Beezar, J.C. Mitchell and A.W. Bunch, 2001. A microcalorimetric comparison of the *anti-streptococcus mutans* efficacy of plant extracts and antimicrobial agents in oral hygiene formulations. *J. Applied Microbiol.*, 90: 53-58.
- Pack, Y.K., M.H. Koo, J.A. Abreu, M. Ikegaki, J.A. Curry and P.L. Rosalen, 1998. Antimicrobial activity of propolis on oral microorganisms. *Curr. Microbiol.*, 36: 24-28.
- Rabe, T. and Van Staden, 1997. Isolation of an antibacterial sesqui-terpenoid from *Warbugia salutaris*. *J. Ethnopharmacol.*, 73: 171-174.
- Rosanaivo and Ratsimamanga-Urverg, 1993. Biological evaluation of plants with reference of the Malagazy flora, 72-79. Monograph for the IFS-NAPRECA Workshop on Bioassays, Antananavivo, Madagascar.
- Srinivasan, D., T. Sangeetha Nathan, T. Suresh and Lakshmena Perumalsamy, 2001. Antimicrobial plants used in folkloric medicine. *J. Ethnopharmacol.*, 74: 217-220.
- Wills, E.D., 1956. Enzyme inhibition by Allicin, the active principle of garlic. *Biochem. J.*, 63: 514.