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Molecular Modelling Analysis of the Metabolism of Zebularine

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Abstract: Zebularine (ZEB) is a pyrimidinone ribinucleoside that is a potent inhibitor of DNA methyltransferases. The inhibition of DNA methylation by ZEB is believed to result from the formation of a covalent adduct between the enzyme and ZEB-substituted DNA. Based on both *in vitro* and *in vivo* activity in mammalian cells, ZEB has been proposed for clinical evaluation as an oral antitumor agent. The compound is quite stable with half-lives of 44 and 68 h at pH 1.0 and 2.0, respectively and shows no evidence of decomposition after more than a week at pH 5. At pH 7.4, the half-life is 508 h. It has been suggested that the enhanced chemical stability of ZEB is responsible for its oral activity. Molecular modelling analyses based on molecular mechanics, semi-empirical (PM3) and DFT (at B3LYP/6-31G* level) calculations show that ZEB and all its metabolites have large LUMO-HOMO energy differences indicating that they will be kinetically inert, thus providing an explanation for the long half-lives at different physiologically relevant pHs. The high solvation energies of ZEB and its metabolites suggest that they can be easily excreted via the urine.

Key words: Zebularine, anticancer drug, leukaemia, DNA methylation, epigenetic silencing, molecular modelling

Introduction

Zebularine (ZEB) is the generic name for 2(1H)-pyrimidinone riboside, a pyrimidinone ribinucleoside that targets epigenetic modulation of DNA methylation (Cheng *et al.*, 2003; Yoo *et al.*, 2004). More precisely, it is a potent inhibitor of DNA methyltransferases (Zhou *et al.*, 2002) that are a class of enzymes involved in the epigenetic silencing of tumour suppressor genes (Yoo *et al.*, 2004). It was originally synthesized as an inhibitor of cytidine deaminase (EC 3.54.5, CDA) (McCormack *et al.*, 1980). ZEB is quite stable with half-lives of 44 and 68 h at pH 1.0 and 2.0, respectively and shows no evidence of decomposition after more than a week at pH 5. At pH 7.4, the half-life is 508 h (Marquez *et al.*, 2003). It has been suggested that the oral activity of ZEB, as applied to the reactivation of silenced *p16* expression, is due to its enhanced chemical stability (Cheng *et al.*, 2003). As a CDA inhibitor, ZEB is found to have a K_i value of 2 μ M, which represents a 10-fold reduction in potency relative to the less stable prototypic inhibitor tetrahydrouridine (THU) (McCormack *et al.*, 1980). The inhibition of DNA methylation by ZEB is believed to result from the formation of a covalent adduct between the enzyme and ZEB-substituted DNA (Ben-Kasus *et al.*, 2005). It has been hypothesized the metabolic activation of ZEB requires it to be phosphorylated and incorporated into DNA. It forms a deoxynucleotide triphosphate that incorporates into DNA at the GCGC target site of DNA methyl transferase. The resulting complex functions as a mechanism-based inhibitor of DNA methylation (Ben-Kasus *et al.*, 2005).

Based on both *in vitro* and *in vivo* activity in mammalian cells resulting from the reactivation of a dormant *p16* tumour suppressor gene (Cheng *et al.*, 2003), ZEB has been proposed for clinical evaluation as an oral antitumour agent (Klecker *et al.*, 2006). ZEB was also found to possess modest antitumour activity against murine B16 melanoma, P388 leukaemia and L1210 leukaemia and showed

activity against when administered i.p. or orally (Driscoll *et al.*, 1991). Furthermore, ZEB enhances the activity of decitabine which is a clinically used DNA methyl transferase inhibitor in both human and murine leukaemia cell lines (Beumer *et al.*, 2006).

It should be noted that ZEB has been found to have poor oral bioavailability in rhesus monkey, Fisher rat and CD2F1 mouse (Holleran *et al.*, 2005) which may be due to its rapid hepatic metabolism to uridine that is further metabolized to uracil (Kleckler *et al.*, 2006). Biochemically ZEB behaves like an analogue of cytosine but with its own unique properties (Ben-Kasus *et al.*, 2005). The 2-(1H)-pyrimidinone ring of ZEB lacks the amino group normally found in the 4-position of the nucleobase cytosine that makes it susceptible to nucleophilic addition of water across the 3,4-double bond to form a covalent hydrated C-4 adduct.

Figure 1 summarizes metabolism of ZEB. It includes: oxidation of ZEB to uridine by aldehyde oxidase, removal of its ribose moiety and that from uridine by uridine phosphorylase (EC 2.4.2.3)

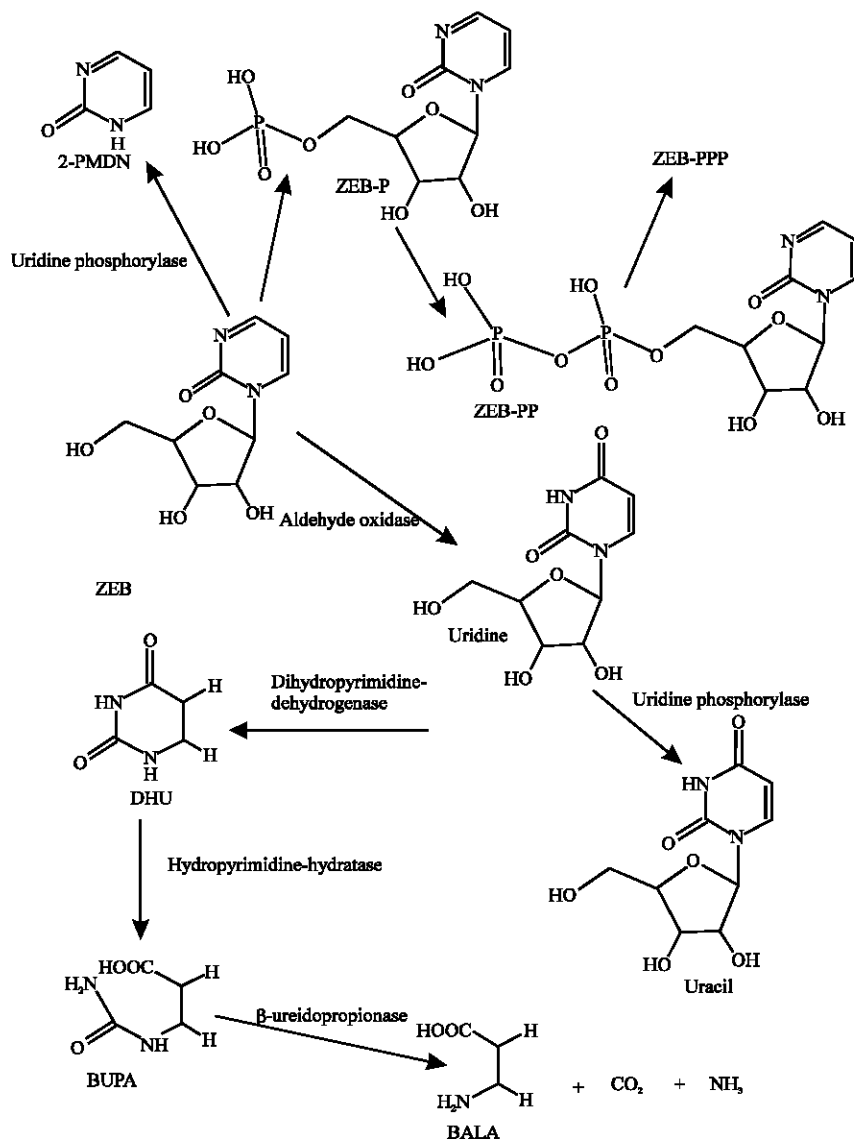


Fig. 1: Metabolic pathways for ZEB and its metabolites (Based on Kleckler *et al.*, 2006)

(Klecker *et al.*, 2006) to produce 2-pyrimidinone (2-PMDN) and uracil, respectively; reduction of uracil to dihydrouracil (DHU) catalysed by dihydropyrimidine dehydrogenase (EC 1.3.1.2) and subsequent hydrolysis of DHU by dihydropyrimidine hydratase to β -ureidopropionic acid (BUPA) and further hydrolysis of BUPA to carbon dioxide, ammonia and β -alanine (BALA) by β -ureidopropionase (Voet and Voet, 1995). Since incorporation into DNA appears to be a requirement for activity of ZEB, it has been suggested that ZEB must undergo phosphorylation followed by subsequent conversion to the corresponding 2'-deoxynucleotide before it can be incorporated (Ben-Kasus *et al.*, 2005). No information is currently available on any side-effects of ZEB.

In this study, molecular modelling analyses have been carried out using the program (Spartan, 2002) to investigate the stability of ZEB and its metabolite with the aim of providing significant information on their toxicity (or lack of it). In particular, the study would show whether ZEB and its metabolites could compromise antioxidant status of the cell and/or could induce DNA damage. Modelling analyses have also been carried out of mono-, di- and tri-phosphorylated adducts of ZEB namely ZEB-P, ZEB-PP and ZEB-PPP.

Computational Methods

The geometries of ZEB and its metabolites uridine, uracil, DHU, BUPA, BALA, 2-PMDN, ZEB-P, ZEB-PP and ZEB-PPP have been optimized based on molecular mechanics, semi-empirical and DFT calculations, using the molecular modelling program Spartan '02. Molecular mechanics calculations were carried out using MMFF force field.

Semi-empirical calculations were carried out using the routine PM3. The order of calculations: molecular mechanics followed by semi-empirical followed by DFT assisted in reaching global minimum. It was found that when semi-empirical calculations were not preceded by molecular mechanics calculations, structure could be embedded in a local minimum. DFT calculations were carried using the program Spartan '02 at B3LYP/6-31G* level. In optimization calculations, a RMS gradient of 0.001 was set as the terminating condition. Although RMS value gradient of 0.001 will not be sufficiently low for vibrational analysis, it is believed to be sufficient for calculations associated with electron energy levels. For the optimized structures, single point calculations were carried to give heat of formation, enthalpy, entropy, free energy, dipole moment, solvation energy, energies for HOMO and LUMO. The gradient in single point calculation as a fraction was found to be less than 0.001. The study was carried out in the School of Biomedical Sciences, The University of Sydney during January to April 2006.

Results and Discussion

Table 1 gives the total energy, heat of formation as per PM3 calculation, enthalpy, entropy, free energy, dipole moment, energies of HOMO and LUMO as per both PM3 and DFT calculations for ZEB and its metabolites uridine, uracil, 2-PMDN, DHU, BUPA, BALA, ZEB-P, ZEB-PP and ZEB-PPP. Figure 2-10 give the regions of negative electrostatic potential (greyish-white envelopes) in (a), HOMOs (where red indicates HOMOs with high electron density) in (b), LUMOs in (c) and density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral) in (d) as applied to the optimized structures of the optimized structures of ZEB and its metabolites uridine, uracil, 2-PMDN, DHU, BUPA, BALA, ZEB-P and ZEB-PP. The figure of ZEB-PPP has not been included.

The calculated solvation energies of ZEB and its metabolites uridine, uracil, DHU, BUPA and BALA from PM3 calculations in kcal mol⁻¹ are, respectively -21.45, -20.02, -14.15, -14.35, -13.64, -20.35 and -14.59, respectively. The values for ZEB-P, ZEB-PP and ZEB-PPP could not be calculated.

Table 1: Calculated thermodynamic and other parameters of ZEB and its metabolites

Molecule	Calculation type	Total energy	Heat of	Enthalpy	Entropy	Solvation energy	Free energy
		(kcal mol ⁻¹ / atomic unit*)	formation (kcal mol ⁻¹)				
ZEB	PM3	-199.89	-178.45	126.02	117.89	-21.45	90.87
	DFT	-796.49		129.09	112.50	-23.22	95.45
Uridine	PM3	-256.24	-236.23	129.44	124.52	-20.02	92.32
	DFT	-871.73		133.12	119.23	-19.21	97.57
Uracil	PM3	-81.09	-66.95	56.66	81.97	-14.15	32.22
	DFT	-414.82		58.65	79.07	-11.21	35.08
2-PMDN	PM3	-22.47	-8.12	53.04	75.55	-14.35	30.52
	DFT	-339.56		54.66	73.99	-11.04	32.60
DHU	PM3	-98.28	-84.64	71.13	83.57	-13.64	46.22
	DFT	-416.05		73.90	81.56	-10.98	49.58
BUPA	PM3	-149.77	-129.42	89.60	99.17	-20.35	60.03
	DFT	-492.44		91.74	95.16	-18.83	63.37
BALA	PM3	-110.89	-97.10	71.38	83.01	-14.59	46.62
	DFT	-323.75		72.53	80.56	-13.69	48.51
ZEB-P	PM3		-380.83	143.82	146.89		100.02
	DFT	-1361.17		145.04	145.12		101.77
ZEB-PP	PM3		-596.83	161.74	176.73		109.05
	DFT	-1931.88		163.82	175.58		111.50
ZEB-PPP	PM3		-811.06	180.04	195.46		121.77
	DFT	-2499.61		182.23	194.23		124.35

Molecule	Calculation type	Area (Å ²)	Volume (Å ³)	Dipole	HOMO (eV)	LUMO (eV)	LUMO-HOMO (eV)
				moment (debye)			
ZEB	PM3	213.21	190.00	6.0	-9.42	-0.70	8.72
	DFT	207.28	187.73	5.5	-6.29	-1.35	4.94
Uridine	PM3	223.88	197.56	3.6	-9.69	-0.43	9.26
	DFT	220.34	195.49	3.8	-6.72	-1.08	5.64
Uracil	PM3	123.72	101.88	3.9	-9.79	-0.56	9.23
	DFT	122.92	101.17	4.3	-6.87	-1.17	5.70
2-PMDN	PM3	114.60	94.66	5.3	-9.21	-0.71	8.50
	DFT	113.74	93.93	6.0	-6.54	-1.64	4.90
DHU	PM3	128.68	106.19	3.7	-10.33	0.20	10.53
	DFT	127.95	105.51	3.9	-7.12	0.40	7.52
BUPA	PM3	156.89	124.48	4.8	-10.49	0.53	11.02
	DFT	151.37	123.08	5.7	-7.19	0.21	7.40
BALA	PM3	116.31	90.25	3.3	-9.89	1.01	10.90
	DFT	116.11	90.47	3.4	-6.40	0.19	6.59
ZEB-P	PM3	272.45	236.94	7.89	-9.26	-0.56	8.70
	DFT	254.42	231.71	6.5	-6.67	-1.82	4.85
ZEB-PP	PM3	332.26	283.53	7.3	-9.21	-0.64	8.57
	DFT	315.80	278.40	7.7	-6.38	-1.61	4.77
ZEB-PPP	PM3	377.90	327.19	6.9	-9.20	-0.88	8.32
	DFT	359.00	321.04	6.8	-6.91	-2.11	4.80

* In atomic units from DFT calculations

The dipole moments of ZEB, uracil, 2-PMDN, DHU, BUPA, BALA, ZEB-P, ZEB-PP and ZEB-PPP from DFT calculations are 5.5, 3.8, 4.3, 6.0, 3.9, 5.7, 3.4, 6.5, 7.7 and 6.8 indicating that ZEB and all its metabolites are highly polar. The results indicate ZEB and its metabolites would be soluble in water and therefore would be easily excreted in the urine.

ZEB and all its metabolites are found to have relatively large LUMO-HOMO energy differences indicating that they would be kinetically inert. ZEB-P, ZEB-PP, ZEB-PPP and 2-PMDN are expected to be more labile than ZEB and the other metabolites as they have somewhat lower LUMO-HOMO energy differences.

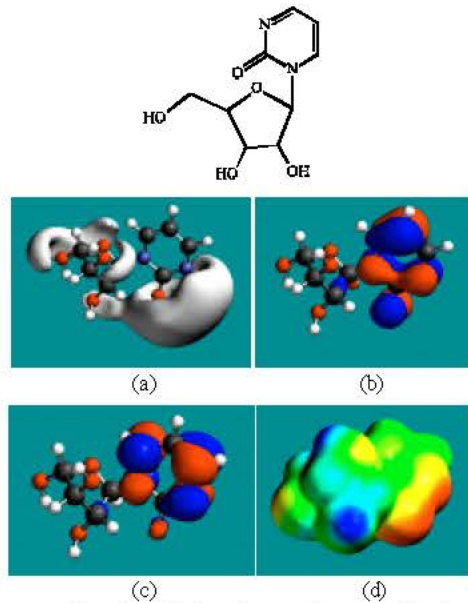


Fig. 2: Structure of ZEB giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

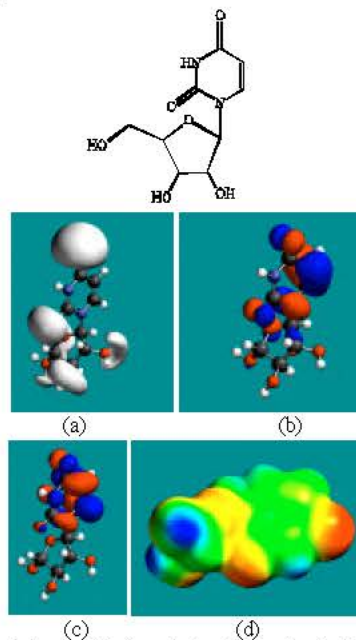


Fig. 3: Structure of uridine giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

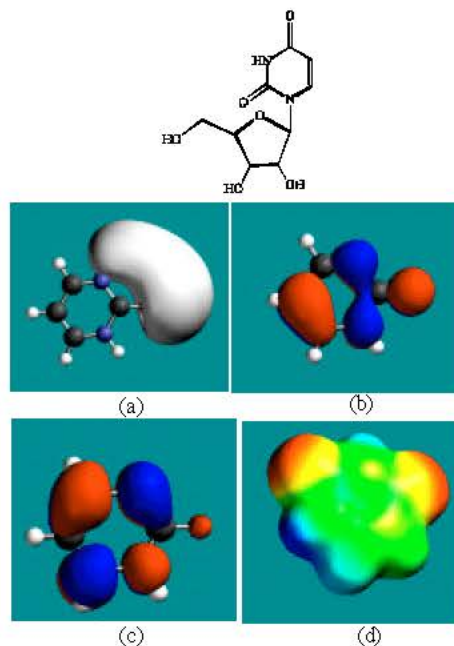


Fig. 4: Structure of uracil giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

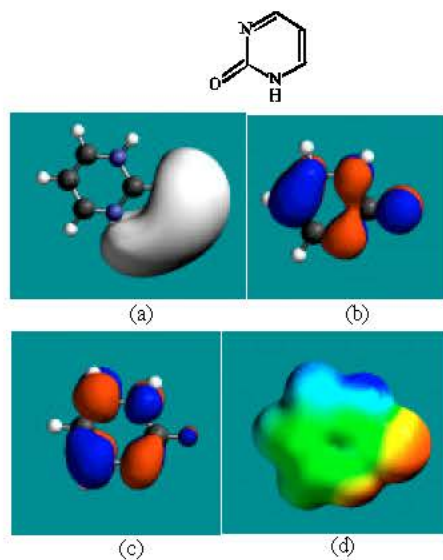


Fig. 5: Structure of 2-PMDN giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

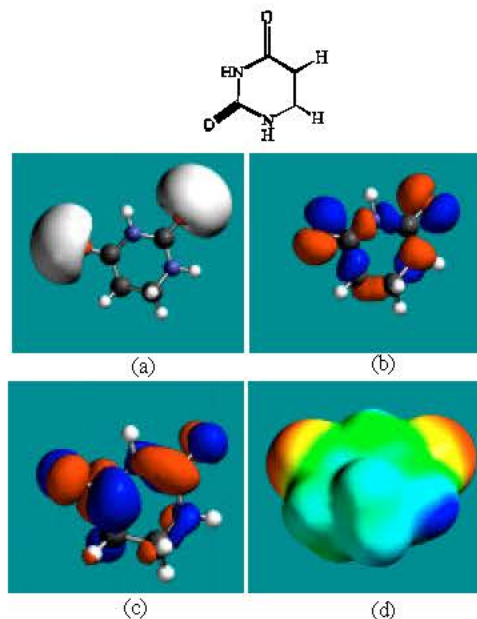


Fig. 6: Structure of DHU giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

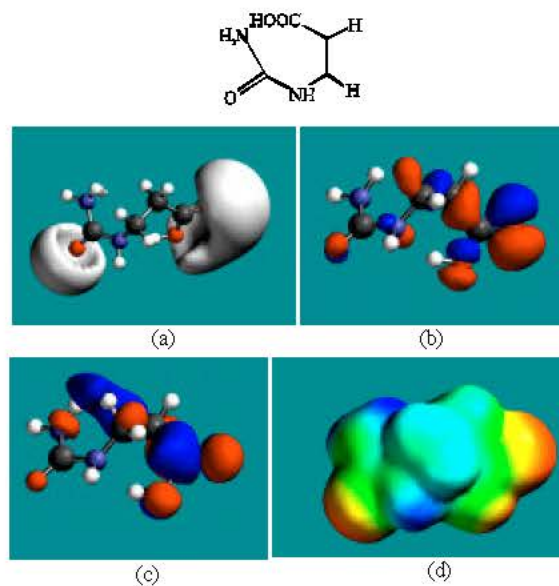


Fig. 7: Structure of BUPA giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

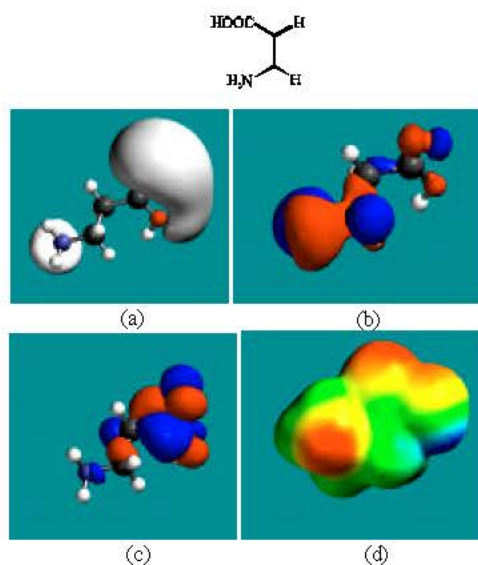


Fig. 8: Structure of BALA giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

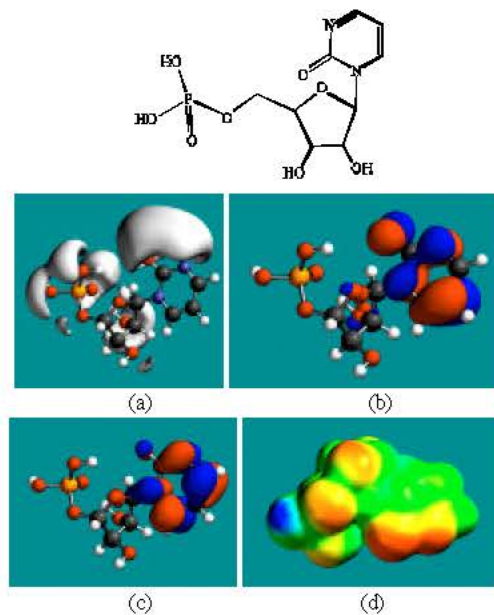


Fig. 9: Structure of ZEB-P giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

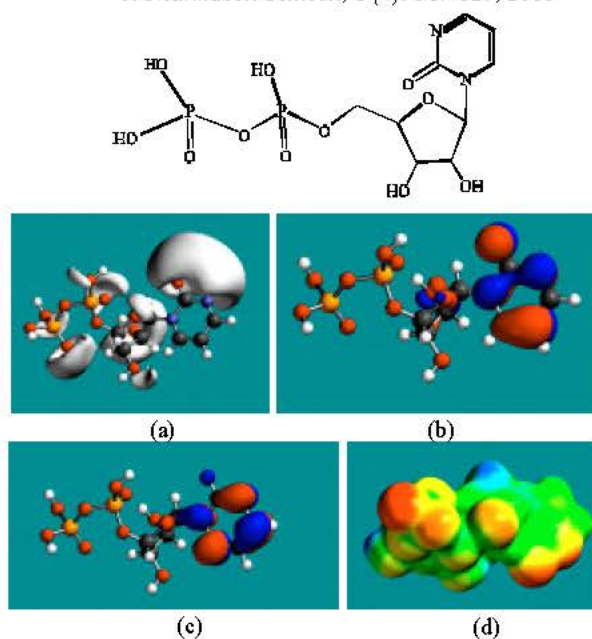


Fig. 10: Structure of ZEB-PP giving in: (a) the electrostatic potential (greyish envelope denotes negative electrostatic potential), (b) the HOMOs, (where red indicates HOMOs with high electron density) (c) the LUMOs (where blue indicates LUMOs) and in (d) the density of electrostatic potential on the molecular surface (where red indicates negative, blue indicates positive and green indicates neutral)

In the case of ZEB, uridine and uracil, the electrostatic potential is found to be more negative around hydroxyl and carbonyl and ethereal oxygen atoms indicating that the positions may be subject to electrophilic attack. In the case of 2-PMDN and DHU, the electrostatic potential is found to be more negative around oxygen and nitrogen atoms indicating that the positions may be subject to electrophilic attack. In the case of BUPA and BALA, the electrostatic potential is found to be more negative around carbonyl and hydroxyl oxygen atoms and the amino nitrogen atoms, indicating that the positions may be subject to electrophilic attack. In the case of ZEB-P, ZEB-PP and ZEB-PPP, the electrostatic potential is found to be more negative around hydroxyl and carbonyl and ethereal and phosphate oxygen atoms indicating that the positions may be subject to electrophilic attack. This is discussed more fully later when nature of surface charge densities are considered.

In the case of ZEB, both HOMOs with high electron density and LUMOs are found to be centered on the non-hydrogen atoms of the pyrimidine ring. In the case of uridine, both HOMOs with high electron density are found to be centered on most of the non-hydrogen atoms of the pyrimidinone ring and some non-hydrogen atoms of the ribose ring whereas LUMOs are found to be centered on the non-hydrogen atoms of pyrimidinone ring.

The presence of both electron-rich (red) and electron-deficient (blue) regions on the surface of ZEB and its metabolites (Fig. 2d and 10d) indicate that they can be subject to electrophilic and nucleophilic attacks. The metabolites DHU and BUPA that have somewhat greater abundance of electron-deficient regions than ZEB and other metabolites, indicating that they may be more subject to nucleophilic attack such as that glutathione and nucleobases in DNA. Reaction with glutathione would cause glutathione depletion, thus inducing cellular toxicity and oxidation of nucleobases would cause DNA damage. However, the two metabolites are found to have the larger LUMO-HOMO energy differences so that they would be less labile. This means that the rate of such adverse reactions may not be significant.

Conclusions

Zebularine is a pyrimidinone ribinucleoside that is a potent inhibitor of DNA methyltransferases. Based on both *in vitro* and *in vivo* activity in mammalian cells, ZEB has been proposed for clinical evaluation as an oral antitumour agent. Molecular modelling analyses show that ZEB and all its metabolites have kinetic stability. Thus, although the presence of electron-deficient regions on the molecular surfaces of ZEB and its metabolites indicate that they may react with cellular glutathione and may cause oxidation of nucleobases in DNA, in actual fact the rates of such adverse reactions are not expected to be significant.

Abbreviations

ZEB:	Zebularine, 2(1H)-pyrimidinone riboside
CDA:	Cytidine deaminase
DHU:	Dihydrouracil
THU:	Tetrahydrouridine
2-PMDN:	2-pyrimidinone
BUPA:	β -ureidipropionic acid
BALA:	β -alanine
LUMO:	Lowest unoccupied molecular orbital
HOMO:	Highest occupied molecular orbital

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