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Research Article Gut Endocrine Cells in the Stomach of Sunda Porcupines (*Hystrix javanica*)

¹Teguh Budipitojo, ²Yuda Heru Fibrianto and ³Guntari Titik Mulyani

¹Department of Anatomy,

²Department of Physiology,

³Department of Internal Medicine, Faculty of Veterinary Medicine, Gadjah Mada University, 55281 Yogyakarta, Indonesia

Abstract

Background and Objective: The regional distribution and relative frequency of gastrointestinal endocrine cells has been found to vary with the animal species and feeding habits. This study aimed to analyze the existence and distribution of immunoreactive endocrine cells to chromogranin, ghrelin, somatostatin, serotonin and gastrin in the gastric mucosa of sunda porcupine (Hystrix javanica) using immunohistochemistry. Materials and Methods: Five adult sunda porcupines were used without sexual distinction. The presences of endocrine cells were visualized by ABC immunohistochemical methods using five types of polyclonal antisera, specific for chromogranin, ghrelin, somatostatin, serotonin and gastrin. **Results:** In the stomach of sunda porcupine, endocrine cells immunoreactive for all of these markers were observed. The chromogranin-immunoreactive cells were found with greatest frequency, in all stomach regions. In all regions of the stomach, endocrine cells secreting ghrelin were also detected in weaker frequency compare with chromogranin. The somatostatin immunoreactive cells were detected rare in the gastric and fundic regions, serotonin cells were found rare in fundic but few in pyloric regions, while gastrin cells were detected abundance in pyloric region of stomach of the sunda porcupines. Gastrin positive cell was also detected in the duodenal part of sunda porcupine. The chromogranin-positive cells were the most numerous, being more prevalent in the stomach glands. The other endocrine cells were identified in smaller numbers, some of them located in all regions or in specific regions. The finding of these cell types in the mucosal stomach confirms their preferential location in the final portions of the principal regions of the stomach and suggests control by feedback of its functions. Conclusion: In conclusion, the present study show for the first time the immunolocalization of chromogranin, ghrelin and gastrin immunoreactive cells in the stomach of sunda porcupine. However, in contrast with previous finding of crested porcupine and other rodents, the present result found serotonin immunoreactive cells in the fundic and pyloric gland regions and somatostatin immunoreactive cells in the gastric and fundic gland regions of sunda porcupine stomach. The differences may was caused by feed types, feeding habits and geographical locations of the species.

Key words: Sunda porcupine, gut endocrine cells, chromogranin, ghrelin, gastrin

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Corresponding Author: Teguh Budipitojo, Department of Anatomy, Faculty of Veterinary Medicine, Gadjah Mada University, 55281 Yogyakarta, Indonesia

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Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Gastrointestinal endocrine cells were dispersed throughout the epithelia and gastric glands of the alimentary tract. These cells produce many biologically active polypeptides and amines, some of them hormones¹. These hormones have important functions in the overall regulation of the digestive process, such as secretion of intestinal and associated glands, gut motility, nutrient absorption and intestinal blood flow². The role of endocrine cells and their respective regulator peptides has attracted the interest of many investigators, who have described them in the gastrintestinal tract (GIT) of other vertebrates, such as amphibia³, fish⁴, birds⁵ and mammals⁶, seeking to establish the morphological role of these cells and respective peptides responsible for regulating numerous body functions.

The regional distribution and relative frequency of gastrointestinal endocrine cells has been found to vary with the animal species and feeding habits². Fourteen types of endocrine cells have been demonstrated in the gastrointestinal tract of Tree Shrew⁷ and Babirusa⁸ by immunohistochemistry. Six types of endocrine cells have been shown in the malayan pangolin, manis javanica⁹ and five in the barking deer¹⁰. These studies reveal interspecific differences and suggest correlation between endocrine cell distribution and feeding habits. This study aimed to analyze the existence and distribution of immunoreactive endocrine cells to chromogranin, ghrelin, somatostatin, serotonin and gastrin in the gastric mucosa of sunda porcupine (*Hystrix javanica*) using immunohistochemistry.

MATERIALS AND METHODS

Five stomachs of adult sunda porcupines, *Hystrix javanica*, about 67 cm in length, were purchased from a merchant in Tawangmangu, Central java, Indonesia and were used as samples. Stomach tissues of *Hystrix javanica* were fixed for 24 h in Bouin's solution, dehydrate in ethanol, cleared in xylene and embbeded in parrafin.

Stomach sections were cut serially in 4-5 µm thickenesses and stained by hematoxylin and eosin (HE) for conventional histological evaluation. For immunohistochemical staining of avidin-biotin peroxidase complex (ABC) methods¹¹ tissue sections were deparaffinized in xylene and rehydrated in decreasing series of ethanol concentrations. After washing in Phosphate Buffered Saline (PBS), endogenous peroxidase activity was blocked by incubating the section in H_2O_2 in methanol for 15 min. Primary antibodies of porcine chromogranin (Lot 8808014; Incstar, Stilwater, OK, USA) at 1:5,000 dilutions, ghrelin (bs-0467R; Bioss Inc. Massachusetts, USA) at 1:1000 dilution, serotonin (Sero-2-3; J Nishitsutsuji, Japan) at 1:5,000 dilution, gastrin (GP-1304; Yanaihara Institute, Japan) at 1:10,000 dilution and somatostatin (20H2T; Incstar, Stilwater, OK, USA) at 1:5,000 dilution were applied overnight at 4°C. To prevent non spesific staining, sections were incubate with nomal goat serum before incubation with the primary antibodies. The secondary antibody was applied for 45 min at room temperature. The immunoreactive site were visualized by tris-HCL buffer containing 3,3 diaminobenzidine tetrahydrocloride. Section were counterstained with mayer hematoxylin and examined with a conventional light microscope and photomicrographs were taken with digital camera.

The specificity of the immunohistochemical staining was confirmed by the replacement of primary antibody with normal rabbit serum. The semiquantitatif number of positif endocrine cell were graded subjectively into 4 classes as rare, few, moderate and numerous.

RESULTS

The present study revealed five kinds of endocrine cells immunoreactive for chromogranin, ghrelin, somatostatin, serotonin and gastrin in the gastric mucosa of sunda porcupine (*Hystrix javanica*). The chromogranin-positive cells were the most numerous, being more prevalent in the stomach glands. The other endocrine cells were identified in smaller numbers, some of them located in all regions or in specific regions.

The chromogranin immunoreactive cells were found rare, few and numerous in the gastric, fundic and pyloric gland regions, respectively (Fig. 1a-c). In all regions of the stomach, endocrine cells secreting ghrelin were detected rare, a weaker frequency compare with chromogranin (Fig. 1d-f).

The somatostatin immunoreactive cells were detected rare in the gastric and fundic regions (Fig. 2a, b), serotonin immunoreactive cells were found rare in fundic but few in pyloric regions (Fig. 2c, d), while gastrin immunoreactive cells were detected numerous in pyloric region of stomach of the sunda porcupines (Fig. 2e). Gastrin positive cell was also detected in the duodenal part of sunda porcupine, individually (Fig. 1f).

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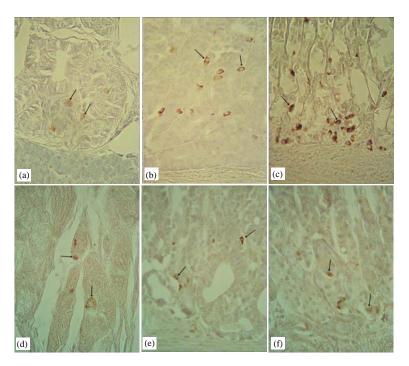


Fig. 1(a-f): Immonolocalization of chromogranin and ghrelin positive cells in the gastric, fundic and pyloric regions of the stomach of sunda porcupine (*Hystrix javanica*). As indicated by arrows, chromogranin cells were detected rare, few and numerous in the (a) Gastric, (b) Fundic, (c) Pyloric gland regions, respectively, while ghrelin cells were detected rare in (d) Gastric, (e) Fundic and (f) Pyloric gland regions of stomach of the sunda porcupines

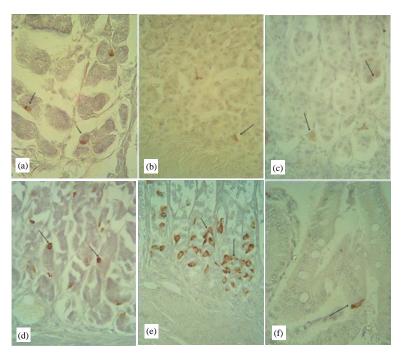


Fig. 2(a-f): Immonolocalization of somatostatin, serotonin and gastrin positive cells in the stomach of sunda porcupine (*Hystrix javanica*). As indicated by arrows, somatostatin cells were detected rare in the (a) Gastric, (b) Fundic gland regions, serotonin cells were found rare in (c) Fundic but few in (d) Pyloric gland regions, while gastrin cells were detected numerous in (e) Pyloric gland region of stomach of the sunda porcupines and (f) Gastrin positive cell was also detected in the duodenal part of sunda porcupine

DISCUSSION

Chromogranin has been used as a marker of endocrine cells as it is found in a wide spectrum of gut endocrine cells in mammals¹² and was detected in all of stomach gland regions of babirusa⁸ and malayan pangolin⁹. In agreement with these previous findings, the present study found the presences of chromogranin immunoreactive cells in all gastric gland regions of sunda porcupine.

Ghrelin is a peptide hormone produced in the gastrointestinal tract which functions as a neuropeptide in the central nervous system¹³. Besides regulating appetite, ghrelin also plays a significant role in regulating the distribution and rate of use of energy¹⁴. Ghrelin immunoreactive cells are found mainly in the stomach¹⁵. In rodents, ghrelin immunoreactive cells were observed in all regions of the gastrointestinal tract and were most abundance in the mucosal layer of fundic gland region¹⁶. In agreement with these previous findings, the present study found ghrelin immunoreactive cells in all gastric gland regions of sunda porcupine without any differences on the density between gland regions.

Serotonin is secreted by the enterocromafin cells, widely distributed in the nervous system and in the gastro-enteropancreatic endocrine cells¹⁷, where it is used to regulate intestinal movements¹⁸. Serotonin immunoreactive cells were detected in the whole gastrointestinal tract of rat¹⁹, mouse²⁰ and porcupine²¹. In the crested porcupine, serotonin immunoreactive cells were detected throughout the whole gastrointestinal tract and showed the highest frequencies in the cardiac region of stomach and duodenum^{21,22}. In contrast with previous finding in crested porcupine and other rodents, the present result found serotonin immunoreactive cells in the fundic and pyloric gland regions of sunda porcupine stomach but not in the gastric gland region.

The somatostatin is a neuroendocrine hormone which was first isolated from the hypothalamus of the sheep²³ and plays important inhibition roles on the secretion of other neuroendocrine hormones²⁴. Somatostatin immunoreactive cells were found predominantly in the pylorus gland region of manchurian chipmunk²⁵ and gerbil²⁶ stomach. In contrast with previous finding, somatostatin immunoreactive cells were not detected in the stomach of the crested porcupine²². In the present study, however, somatostatin immunoreactive cells were found in the gastric and fundic gland regions of sunda porcupine stomach but not in the pyloric gland region.

Gastrin is a peptide hormone produced by G cells of the pyloric antrum of the stomach and duodenum²⁷. This hormone acts by increasing the force of antrum contractions and the constriction of the pyloric sphincter²⁸. In agreement

with ostrich²⁹ and malayan pangolin⁹, the present study found gastrin immunoreactive cells in the gastric glands of the pyloric gland region and duodenum of sunda porcupine (*Hystrix javanica*).

CONCLUSION

The present study show for the 1st time the immunolocalization of chromogranin, ghrelin and gastrin immunoreactive cells in the stomach of sunda porcupine. However, in contrast with previous finding of crested porcupine and other rodents, the present result found serotonin immunoreactive cells in the fundic and pyloric gland regions and somatostatin immunoreactive cells in the gastric and fundic gland regions of sunda porcupine stomach. The differences may caused by feed types, feeding habits and geographical locations of the species.

SIGNIFICANT STATEMENT

The present study show for the first time the immunolocalization of chromogranin, ghrelin and gastrin immunoreactive cells in the stomach of porcupine, especially sunda porcupine (*Hystrix javanica*) which is endemic to Indonesia. In the previous report on crested procupine (*Hystrix cristata*) which is endemic to Italy, Sicily, North Africa and sub-Saharan Africa. Timurkaan *et al.*²¹ and Yaman *et al.*²² showed the presence of serotonin and somatostatin in the gastrointestinal tract. However, in contrast with previous finding of crested porcupine and other rodents, the present result found serotonin immunoreactive cells in the fundic and pyloric gland regions and somatostatin immunoreactive cells in the gastric and fundic gland regions of sunda porcupine stomach.

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REFERENCES

- Schonhoff, S.E., M. Giel-Moloney and A.B. Leiter, 2004. Minireview: Development and differentiation of gut endocrine cells. Endocrinology, 145: 2639-2644.
- Gunawardene, A.R., B.M. Corfe and C.A. Staton, 2011. Classification and functions of enteroendocrine cells of the lower gastrointestinal tract. Int. J. Exp. Pathol., 92: 219-231.

- Xie, Z., D. Guo, C. Lu, B. Li and L. Shan, 2012. Argentaffine cells in the digestive tract of hibernating and non-hibernating in Chinese fire-bellied newt (*Cynops orientalis*). Int. J. Morphol., 30: 1389-1394.
- Vieira-Lopes, D.A., N.L. Pinheiro, A. Sales, A. Ventura, F.G. Araujo, I.D. Gomes and A.A. Nascimento, 2013. Immunohistochemical study of the digestive tract of *Oligosarcus hepsetus*. World J. Gastroenterol., 19: 1919-1929.
- Yang, J., L. Zhang, X. Li, L. Zhang, X. Liu and K. Peng, 2012. An immunohistochemical study on the distribution of endocrine cells in the digestive tract of gray goose (*Anser anser*). Turk. J. Vet. Anim. Sci., 36: 373-379.
- Furness, J.B., L.R. Rivera, H.J. Cho, D.M. Bravo and B. Callaghan, 2013. The gut as a sensory organ. Nat. Rev. Gastroenterol. Hepatol., 10: 729-740.
- Yamada, J., M. Tauchi, W. Rerkamnuaychoke, H. Endo and N. Chungsamarnyart *et al.*, 1999. Immunohistochemical survey of the gut endocrine cells in the common tree shrew (*Tupaia belangerl*). J. Vet. Med. Sci., 61: 761-767.
- Agungpriyono, S., A.A. Macdonald, K.Y. Leus, N. Kitamura and I.K. Adnyane *et al.*, 2000. Immunohistochemical study on the distribution of endocrine cells in the gastrointestinal tract of the babirusa, *Babyrousa babyrussa* (Suidae). Anat. Histol. Embryol., 29: 173-178.
- Nisa, C., N. Kitamura, M. Sasaki, S. Agungpriyono and C. Choliq et al., 2005. Immunohistochemical study on the distribution and relative frequency of endocrine cells in the stomach of the Malayan Pangolin, *Manis javanica*. Anat. Histol. Embryol., 34: 373-378.
- Adnyane, I.K.M., A.B. Zuki, M.M. Noordin and S. Agungpriyono, 2011. Immunohistochemical study of endocrine cells in the gastrointestinal tract of the barking deer, *Muntiacus muntjak*. Anatomia Histol. Embryol., 40: 365-374.
- 11. Dabbs, D.J., 2014. Diagnostic Immunohistochemistry. 3rd Edn., Churchill Livingstone Elsevier, Philadelphia, PA., USA.
- El-Salhy, M., O.H. Gilja, D. Gundersen, J.G. Hatlebakk and T. Hausken, 2014. Duodenal chromogranin a cell density as a biomarker for the diagnosis of irritable bowel syndrome. Gastroenterol. Res. Pract. 10.1155/2014/462856.
- 13. Burger, K.S. and L.A. Berner, 2014. A functional neuroimaging review of obesity, appetitive hormones and ingestive behavior. Physiol. Behav., 136: 121-127.
- Dickson, S.L., E. Egecioglu, S. Landgren, K.P. Skibicka, J.A. Engel and E. Jerlhag, 2011. The role of the central ghrelin system in reward from food and chemical drugs. Mol. Cell. Endocrinol., 340: 80-87.
- 15. Hamza, S., I. Vulkova, M. Gulubova and D. Sivrev, 2014. Characteristics of ghrelinpositive cells of the stomach in the rat. Trak. J. Sci., 12: 141-143.

- 16. Sakata, I. and T. Sakai, 2010. Ghrelin cells in the gastrointestinal tract. Int. J. Peptides. 10.1155/2010/945056
- 17. Richter, G., F. Stockmann, J.M. Conlon and W. Creutzfeldt, 1986. Serotonin release into blood after food and pentagastrin: Studies in healthy subjects and in patients with metastatic carcinoid tumors. Gastroenterology, 91: 612-618.
- 18. Bornstein, J.C., 2012. Serotonin in the gut: What does it do? Front. Neurosci., Vol. 6. 10.3389/fnins.2012.00016.
- 19. Singh, P., S.R. Dutta and D. Guha, 2015. Gastric mucosal protection by aegle marmelos against gastric mucosal damage: Role of enterochromaffin cell and serotonin. Saudi J. Gastroenterol., 21: 35-42.
- 20. Yano, J.M., K. Yu, G.P. Donaldson, G.G. Shastri and P. Ann *et al.*, 2015. Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis. Cell, 161: 264-276.
- 21. Timurkaan, S., M. Karan and A. Aydin, 2005. Immunohistochemical study of the distribution of serotonin in the gastrointestinal tract of the porcupines (*Hystrix cristata*). Rev. Vet. Med., 156: 533-536.
- 22. Yaman, M., B.G. Tarakci, A. Bayrakdar, O. Atalar and O. Dabak, 2007. Immunohistochemical study of gastrointestinal endocrine cells in the porcupine (*Hystrix cristata*). Rev. Med. Vet., 158: 196-200.
- 23. Brazeau, P., W. Vale, R. Burgus, N. Ling, M. Butcher, J. Rivier and R. Guillemin, 1973. Hypothalamic polypeptide that inhibits the secretion of immunoreactive pituitary growth hormone. Science, 179: 77-79.
- 24. Wolin, E.M., 2012. The expanding role of somatostatin analogs in the management of neuroendocrine tumors. Gastrointest Cancer Res., 5: 161-168.
- 25. Lee, H.S., S.K. Ku and J.H. Lee, 1998. Localization of endocrine cells in the gastrointestinal tract of the Manchurian chipmunk, *Tamias sibiricus barberi*. Korean J. Biol. Sci., 2: 395-401.
- Lee, H.J., H.S. Lee, S.K. Ku, K.D. Park and K.S. Kim, 2000. Immunohistochemical study of the gastrointestinal endocrine cells in the Mongolian Gerbils, *Meriones unguiculatus*. Korean J. Vet. Res., 40: 653-660.
- 27. Kasajima, A., F. Fujishima, T. Morikawa, S. Kawasaki and S. Konosu-Fukaya *et al.*, 2015. G-cell hyperplasia of the stomach induces ECL-cell proliferation in the pyloric glands in a paracrinal manner. Pathol. Int., 65: 259-263.
- Rau, T.T., A. Sonst, A. Rogler, G. Burnat and H. Neumann *et al.*, 2013. Gastrin mediated down regulation of ghrelin and its pathophysiological role in atrophic gastritis. J. Physiol. Pharmacol., 64: 719-725.
- 29. Duritis, I., A. Mugurevics and L. Mancevica, 2013. The distribution of gastrin, somatostatin and glucagon immunoreactive (IR) cells in ostrich stomach during the pre and post hatching period. Anat. Histol. Embryol., 42: 362-368.