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Research Article

Molecular Detection of Polymorphism of Heat Shock Protein 70 (*Hsp70*) in the Semen of Iraqi Holstein Bulls

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Abstract

Background and Objective: *Hsp70* is a molecular chaperone and cell preservation against heat shock, considered as the ideal biological marker for measuring heat stress in animals, its polymorphism may affect stress tolerance bulls' fertility. The study aimed to identify polymorphism of *hsp70* gene in Holstein bulls born in Iraq. **Materials and Methods:** The study was carried out during 1st November, 2015 to 31st January, 2016, 29 Holstein bulls born in Iraq, 2.5-3.5 years old, back to Artificial Insemination Center of Abou-Ghareeb Baghdad, Iraq, were used. Semen was collected by using the artificial vagina. DNA extracted then PCR amplified, sequencing, BLAST analysis and multiple sequence alignment were carried out. **Results:** The results showed three haplotypes, haplotype G1, haplotype G2 and haplotype G3 as a compare with the same gene in GenBank, they all showed silent mutations in different positions except haplotype G2 showed a missense mutation in position 1451 as well as silent mutations. **Conclusion:** It is concluded that the Holstein bulls born in Iraq have new polymorphism of *Hsp70* gene as a compare with same gene in GenBank, because of mutations.

Key words: Molecular detection, heat shock protein 70 polymorphism, Holstein bulls, heat stress

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Heat shock protein 70 (*Hsp70*) is one of HSPs family and its size is 68-73 kDa¹. It is encoded by a single exon of the *Hsp70* gene, the open reading frame to this gene is 1926 bp approximately. The protein comprises 641 amino acids². The work of *Hsp70* is molecular chaperones and cell preservation against heat shock, able of denaturing proteins³, it's one of the most important roles is prevent aggregation of the partially folded proteins remodel folding pathways and regulate activity of proteins⁴. Expressed in response to heat stress and environmental stress⁵, *hsp70* gene was considered as the ideal biological marker for measuring heat stress in animals⁶. Polymorphism of *hsp70* gene explains the differences between individuals in the tolerance of stress conditions such as heat stress, polymorphism produced by genetic mutations which can be a useful to resist heat stress⁷. Nucleotide polymorphisms in the *hsp70* gene promoter region may affect stress tolerance and haplotypes of *Hsp70* were associate with bulls' fertility and heat tolerance^{8,9}, the difference in reproductive performance was also associated with polymorphism of *hsp70* gene¹⁰. The *Hsp70* is available in spermatozoa of mature bulls¹¹. Although the origin of the Holstein cattle is the cold regions (Europe), can be bred and preserve in tropical and sup-tropical environment like Iraq¹², *Hsp70* playing a pivotal function in environmental and heat stress adaptation¹³⁻¹⁴, because it is playing important roles in essential cellular processes and also related to stress¹⁵. The main objective of this study was to identify the manipulation of genotype (molecular changing) to capable with harsh environment (high temperature). Therefore, polymorphisms of the *hsp70* gene of the Holstein bulls born in Iraq were identified. Since there have been no studies and the lack of information dealing with this subject, this study aimed to identify polymorphism of *Hsp70* gene in Holstein bulls which are born in Iraq.

MATERIALS AND METHODS

Animals and semen collection: The study was carried out during 1st November, 2015 to 31th January, 2016. Twenty nine Holstein bulls born in Iraq of known fertility, 2.5-3.5 years old, back to Artificial Insemination Center of Abou-Ghareeb Baghdad, Iraq were used in the present study. Semen was collected from all bulls by using the artificial vagina method.

DNA extraction: The DNA was extracted from semen samples using Chelex-100® (Sigma Aldrich, USA) as already described

by Walsh *et al.*¹⁶ and modified by Manuja *et al.*¹⁷. Concentration and purity of DNA was estimated by using Nano-drop system (Nano Drop thermo scientific 200, USA). To determine the best purity, with an estimate 260/280 ratio, which should be 1.8, that indicate the purity of the DNA¹⁸.

PCR amplification: Amplifications were performed in 25 µL reactions (Table 1). According to First BASE Laboratories/Malaysia, primer was used to amplify conserved region within the HSP70 promotor, *Hsp70*-F: ATGGCGAAAAACATGGCTATCGGC, *hsp70*-R: CTAATCCACCTCTCAATGGTGGGGCC, First BASE Laboratories/Malaysia, the PCR (A-2040-1 My Genie/Gradient Thermal Bioneer Korea) amplification cycling protocols of PCR amplification are summarized in Table 2. The PCR product was detected on 1.5% ethidium bromide stained agarose gel. PCR product size was 1926 bp (Fig. 1).

Sequencing: The PCR products purification, then sequenced in First BASE Laboratories/Malaysia. Nucleotide sequence alignments and comparisons were done using the Geneious version 10.1.3 software. BLAST analysis were carried out on website <http://www.ncbi.nlm.nih.gov>, Multiple Sequence Alignment were carried out on website <http://www.ebi.ac.uk/Tools/msa/clustalo/> and compared with same gene in Gene Bank accession number AY662497.1.

RESULTS AND DISCUSSION

Results of sequencing and the multiple sequence alignment were compared with heat shock protein *Hsp70* gene of the Holstein bulls in the gene bank. Three new haplotypes were detected and classified as three groups.

First group, the haplotype G1, is the closest haplotype to heat shock protein *hsp70* gene in the Holstein bulls in the

Table 1: Components of PCR reactions

First BASE laboratories (2016)	
Components	Amount (µL)
Water, nuclease free	9.5
2X PCR master mix	12.5
Forward primer (10 µM)	1.0
Reverse primer (10 µM)	1.0
DNA template (75 ng)	1.0
Total	25.0

Table 2: Cycling protocol of PCR amplification

Cycle steps	Temperature (°C)	Time	No. of cycles
Initial denaturation	95	5 min	1
Denaturation	94	30 sec	30
Annealing	61	30 sec	
Extension	72	2 min	
Final extension	72	10 min	1

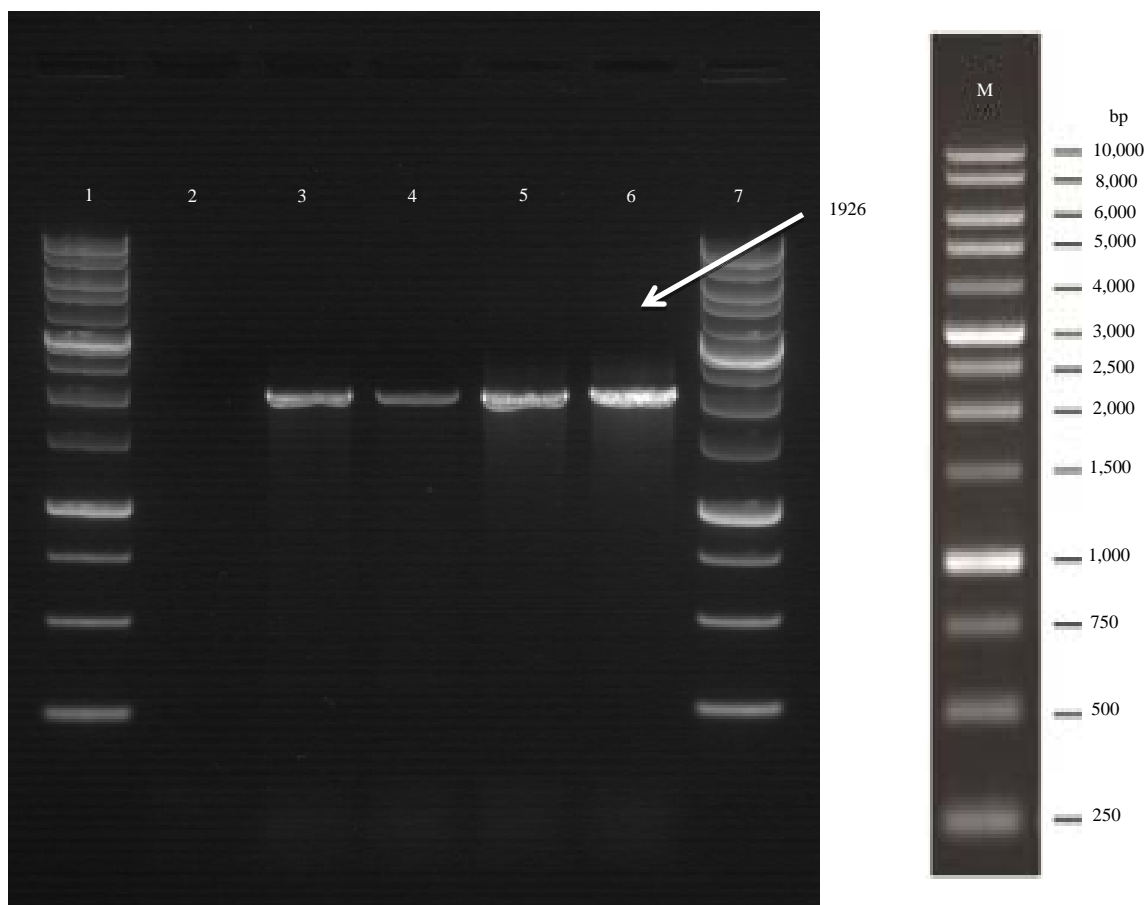


Fig. 1: Gel electrophoresis of PCR products of *Hsp70* gene

Sample 1 and 7: DNA marker, Sample 2: -ve is no-template control (water to replace DNA template), Sample 3-6: DNA template (*Hsp70* gene), A total 75 ng of DNA sample was used in one 25 μ L PCR reaction. only 3 μ L of PCR product was run on 1% TAE agarose of PCR product was run on 1% TAE agarose gel at 100 V, 60 min

gene bank. This haplotype was exposed to genetic mutations in nucleotides at positions, 6 (C<G), 174 (C<G), 282 (A<G) and 1339 (T<C). All mutations are silent that did not encode a new amino acid. Fifteen Bulls were found in this group.

Second group, the haplotype G2, in comparison with *Hsp70* gene in the Holstein bulls in the gene bank and first group. Nucleotides were substituted in eight different positions, 114 (G<A), 174 (C<G), 282 (A<G), 1339 (T<C), 1451 (C<A), 1590 (A<G), 1695 (C<T) and 1719 (G<T). These genetic mutations are all silent except the mutation at the position 1451 (missense mutation). It encoded for new amino acid (Aspartic acid instead of alanine). The change nucleotide cytosine (C) instead of adenine (A) led to a change code GCC to GAC which encode a new amino acid¹⁹. Six bulls were found in this group.

Third group, the haplotype G3, in comparison with *Hsp70* gene in the Holstein bulls in the gene bank and other groups, in this group nucleotides were substituted for

each position 114 (G<A), 174 (C<G), 282 (A<G), 1339 (T<C), 1590 (A<G), 1695 (C<T) and 1719 (G<T). All mutations are silent. Eight bulls were found in this group.

The results of the Multiple Sequence Alignment (MSA) of *hsp70* gene in the bulls (Fig. 2) showed that the convergence ratio of G1, G2 and G3 haplotypes were 99.73, 99.51 and 99.56%, respectively.

These results showed silent mutations in different positions of *hsp70* gene, although it does not encode a new amino acid²⁰ but it may influence the relevant protein through change in transcription and may impact the precision or efficiency of splicing of mRNA or control of transcripts^{21,22}. On the other hand the missense mutation that occurred in haplotype G2 may change protein properties due to differences between amino acids, aspartic and alanine²³.

These mutations may effect on protein folding and stability²⁴, protein function²⁵ and protein-protein interactions²⁶.

```
AY662497.1 ATGGCGAAAAACATGGCTATCGGCATCGACCTGGGCACCACCTACTCCTGCGTAGGGGTG 60
-----TACTCCTGCGTAGGGGTG 18
-----TACTCCTGCGTAGGGGTG 18
-----TACTCCTGCGTAGGGGTG 18
*****

AY662497.1 TTCCAGCACGGCAAGGTGGAGATCATCGCCAACGACCAGGGCAACCGCACCACCCCCAGC 120
G1 TTCCAGCACGGCAAGGTGGAGATCATCGCCAACGACCAGGGCAACCGCACCACCCCCAGC 78
G2 TTCCAGCACGGCAAGGTGGAGATCATCGCCAACGACCAGGGCAACCGCACCACCCCCAGC 78
G3 TTCCAGCACGGCAAGGTGGAGATCATCGCCAACGACCAGGGCAACCGCACCACCCCCAGC 78
*****

AY662497.1 TACGTGGCCTTACCAGTACCGAGCGGCTCATCGGCGGATGCGGCCAAGAGCCAGGTGGCG 180
G1 TACGTGGCCTTACCAGTACCGAGCGGCTCATCGGCGGATGCGGCCAAGAGCCAGGTGGCG 138
G2 TACGTGGCCTTACCAGTACCGAGCGGCTCATCGGAGATGCGGCCAAGAGCCAGGTGGCG 138
G3 TACGTGGCCTTACCAGTACCGAGCGGCTCATCGGAGATGCGGCCAAGAGCCAGGTGGCG 138
*****

AY662497.1 CTGAACCCGCGAGAACACGGTGTTCGACGCGAAGCGCGCTGATCGGCCGCAAGTTCGGAGAC 240
G1 CTGAACCCGCGAGAACACGGTGTTCGACGCGAAGCGCGCTGATCGGCCGCAAGTTCGGAGAC 198
G2 CTGAACCCGCGAGAACACGGTGTTCGACGCGAAGCGCGCTGATCGGCCGCAAGTTCGGAGAC 198
G3 CTGAACCCGCGAGAACACGGTGTTCGACGCGAAGCGCGCTGATCGGCCGCAAGTTCGGAGAC 198
*****

AY662497.1 CCGGTGGTGCAGTCGGACATGAAGCACTGGCCTTTCGCGTCAACGACGGAGACAAG 300
G1 CCGGTGGTGCAGTCGGACATGAAGCACTGGCCTTTCGCGTCAACGACGGAGACAAG 258
G2 CCGGTGGTGCAGTCGGACATGAAGCACTGGCCTTTCGCGTCAACGACGGAGACAAG 258
G3 CCGGTGGTGCAGTCGGACATGAAGCACTGGCCTTTCGCGTCAACGACGGAGACAAG 258
*****

AY662497.1 CCTAAGGTGCAGGTGAGCTACAAGGGGAGACCAAGGCGTTCTACCCGGAGGAGATCTCG 360
G1 CCTAAGGTGCAGGTGAGCTACAAGGGGAGACCAAGGCGTTCTACCCGGAGGAGATCTCG 318
G2 CCTAAGGTGCAGGTGAGCTACAAGGGGAGACCAAGGCGTTCTACCCGGAGGAGATCTCG 318
G3 CCTAAGGTGCAGGTGAGCTACAAGGGGAGACCAAGGCGTTCTACCCGGAGGAGATCTCG 318
*****

AY662497.1 TCGATGGTGTGACCAAGATGAAGGAGATCGCCGAGGCGTACCTGGGCCACCCGGTGACC 420
G1 TCGATGGTGTGACCAAGATGAAGGAGATCGCCGAGGCGTACCTGGGCCACCCGGTGACC 378
G2 TCGATGGTGTGACCAAGATGAAGGAGATCGCCGAGGCGTACCTGGGCCACCCGGTGACC 378
G3 TCGATGGTGTGACCAAGATGAAGGAGATCGCCGAGGCGTACCTGGGCCACCCGGTGACC 378
*****

AY662497.1 AACGCGGTGATCACCGTGCCGGCCTACTTCAACGACTCGCAGCGGCAGGCCACCAAGGAC 480
G1 AACGCGGTGATCACCGTGCCGGCCTACTTCAACGACTCGCAGCGGCAGGCCACCAAGGAC 438
G2 AACGCGGTGATCACCGTGCCGGCCTACTTCAACGACTCGCAGCGGCAGGCCACCAAGGAC 438
G3 AACGCGGTGATCACCGTGCCGGCCTACTTCAACGACTCGCAGCGGCAGGCCACCAAGGAC 438
*****

AY662497.1 GCGGGGGTGATCGCGGGGCTGAACGTGCTGAGGATCATCAACGAGCCCACGGCCGCCGCC 540
G1 GCGGGGGTGATCGCGGGGCTGAACGTGCTGAGGATCATCAACGAGCCCACGGCCGCCGCC 498
G2 GCGGGGGTGATCGCGGGGCTGAACGTGCTGAGGATCATCAACGAGCCCACGGCCGCCGCC 498
G3 GCGGGGGTGATCGCGGGGCTGAACGTGCTGAGGATCATCAACGAGCCCACGGCCGCCGCC 498
*****

AY662497.1 ATCGCCTACGGCCTGGACAGGACGGGCAAGGGGGAGCGCAACGTGCTCATCTTTGATCTG 600
G1 ATCGCCTACGGCCTGGACAGGACGGGCAAGGGGGAGCGCAACGTGCTCATCTTTGATCTG 558
G2 ATCGCCTACGGCCTGGACAGGACGGGCAAGGGGGAGCGCAACGTGCTCATCTTTGATCTG 558
G3 ATCGCCTACGGCCTGGACAGGACGGGCAAGGGGGAGCGCAACGTGCTCATCTTTGATCTG 558
*****

AY662497.1 GGAGGGGGCACGTTTCAGCTGTCCATCTGACGATCGACGACGGCATCTTCGAGGTGAAG 660
G1 GGAGGGGGCACGTTTCAGCTGTCCATCTGACGATCGACGACGGCATCTTCGAGGTGAAG 618
G2 GGAGGGGGCACGTTTCAGCTGTCCATCTGACGATCGACGACGGCATCTTCGAGGTGAAG 618
G3 GGAGGGGGCACGTTTCAGCTGTCCATCTGACGATCGACGACGGCATCTTCGAGGTGAAG 618
*****
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Fig. 2: Continue

AY662497.1	GCCACGGCCGGGACACGCACCTGGGCGGGGAGGACTTCGACAACAGGCTGGTGAACCAC	720
G1	GCCACGGCCGGGACACGCACCTGGGCGGGGAGGACTTCGACAACAGGCTGGTGAACCAC	678
G2	GCCACGGCCGGGACACGCACCTGGGCGGGGAGGACTTCGACAACAGGCTGGTGAACCAC	678
G3	GCCACGGCCGGGACACGCACCTGGGCGGGGAGGACTTCGACAACAGGCTGGTGAACCAC	678

AY662497.1	TTCTGGAGGAGTTCAAGAGGAAGCACAAGAAGGACATCAGCCAGAACAAGCGGGCCGTG	780
G1	TTCTGGAGGAGTTCAAGAGGAAGCACAAGAAGGACATCAGCCAGAACAAGCGGGCCGTG	738
G2	TTCTGGAGGAGTTCAAGAGGAAGCACAAGAAGGACATCAGCCAGAACAAGCGGGCCGTG	738
G3	TTCTGGAGGAGTTCAAGAGGAAGCACAAGAAGGACATCAGCCAGAACAAGCGGGCCGTG	738

AY662497.1	AGGCGGCTGCGCACCGCATGCGAGCGGGCCAAGAGAACCTTGTCTCCAGCACCCAGGCC	840
G1	AGGCGGCTGCGCACCGCATGCGAGCGGGCCAAGAGAACCTTGTCTCCAGCACCCAGGCC	798
G2	AGGCGGCTGCGCACCGCATGCGAGCGGGCCAAGAGAACCTTGTCTCCAGCACCCAGGCC	798
G3	AGGCGGCTGCGCACCGCATGCGAGCGGGCCAAGAGAACCTTGTCTCCAGCACCCAGGCC	798

AY662497.1	AGCCTGGAGATCGACTCCCTGTTTCGAGGGCATCGACTTCTACACGTCCATCACCAGGGCG	900
G1	AGCCTGGAGATCGACTCCCTGTTTCGAGGGCATCGACTTCTACACGTCCATCACCAGGGCG	858
G2	AGCCTGGAGATCGACTCCCTGTTTCGAGGGCATCGACTTCTACACGTCCATCACCAGGGCG	858
G3	AGCCTGGAGATCGACTCCCTGTTTCGAGGGCATCGACTTCTACACGTCCATCACCAGGGCG	858

AY662497.1	CGGTTTCGAGGAGCTGTGCTCCGACCTGTTCCGGAGCACCCCTGGAGCCCGTGGAGAAGGCG	960
G1	CGGTTTCGAGGAGCTGTGCTCCGACCTGTTCCGGAGCACCCCTGGAGCCCGTGGAGAAGGCG	918
G2	CGGTTTCGAGGAGCTGTGCTCCGACCTGTTCCGGAGCACCCCTGGAGCCCGTGGAGAAGGCG	918
G3	CGGTTTCGAGGAGCTGTGCTCCGACCTGTTCCGGAGCACCCCTGGAGCCCGTGGAGAAGGCG	918

AY662497.1	CTACGCGACGCCAAGCTGGACAAGGCGCAGATCCACGACCTGGTCTGGTGGGGGGCTCC	1020
G1	CTACGCGACGCCAAGCTGGACAAGGCGCAGATCCACGACCTGGTCTGGTGGGGGGCTCC	978
G2	CTACGCGACGCCAAGCTGGACAAGGCGCAGATCCACGACCTGGTCTGGTGGGGGGCTCC	978
G3	CTACGCGACGCCAAGCTGGACAAGGCGCAGATCCACGACCTGGTCTGGTGGGGGGCTCC	978

AY662497.1	ACCCGCATCCCCAAGGTGCAGAAGCTGCTGCAGGACTTCTTCAACGGGCGCGACCTCAAC	1080
G1	ACCCGCATCCCCAAGGTGCAGAAGCTGCTGCAGGACTTCTTCAACGGGCGCGACCTCAAC	1038
G2	ACCCGCATCCCCAAGGTGCAGAAGCTGCTGCAGGACTTCTTCAACGGGCGCGACCTCAAC	1038
G3	ACCCGCATCCCCAAGGTGCAGAAGCTGCTGCAGGACTTCTTCAACGGGCGCGACCTCAAC	1038

AY662497.1	AAGAGCATCAACCCCGACGAGGCGGTGGCGTACGGGGCGGCGGTGCAGGCGGCCATCCTG	1140
G1	AAGAGCATCAACCCCGACGAGGCGGTGGCGTACGGGGCGGCGGTGCAGGCGGCCATCCTG	1098
G2	AAGAGCATCAACCCCGACGAGGCGGTGGCGTACGGGGCGGCGGTGCAGGCGGCCATCCTG	1098
G3	AAGAGCATCAACCCCGACGAGGCGGTGGCGTACGGGGCGGCGGTGCAGGCGGCCATCCTG	1098

AY662497.1	ATGGGGGACAAGTCGGAGAACGTGCAGGACCTGCTGTTGCTGGACGTGGCTCCCCTGTCG	1200
G1	ATGGGGGACAAGTCGGAGAACGTGCAGGACCTGCTGTTGCTGGACGTGGCTCCCCTGTCG	1158
G2	ATGGGGGACAAGTCGGAGAACGTGCAGGACCTGCTGTTGCTGGACGTGGCTCCCCTGTCG	1158
G3	ATGGGGGACAAGTCGGAGAACGTGCAGGACCTGCTGTTGCTGGACGTGGCTCCCCTGTCG	1158

AY662497.1	CTGGGACTGGAGACGGCCGGAGGCGTGATGACCGCCCTGATCAAGCGCAACTCCACCATC	1260
G1	CTGGGACTGGAGACGGCCGGAGGCGTGATGACCGCCCTGATCAAGCGCAACTCCACCATC	1218
G2	CTGGGACTGGAGACGGCCGGAGGCGTGATGACCGCCCTGATCAAGCGCAACTCCACCATC	1218
G3	CTGGGACTGGAGACGGCCGGAGGCGTGATGACCGCCCTGATCAAGCGCAACTCCACCATC	1218

AY662497.1	CCCACGAAGCAGACGCAGATCTTACCACCTACTCGGACAACCAGCCGGGCGTGCTGATC	1320
G1	CCCACGAAGCAGACGCAGATCTTACCACCTACTCGGACAACCAGCCGGGCGTGCTGATC	1278
G2	CCCACGAAGCAGACGCAGATCTTACCACCTACTCGGACAACCAGCCGGGCGTGCTGATC	1278
G3	CCCACGAAGCAGACGCAGATCTTACCACCTACTCGGACAACCAGCCGGGCGTGCTGATC	1278

AY662497.1	CAGGTGTACGAGGGCGAGAGGGCCATGACGCGGGACAACAACCTGCTGGGGCGCTTCGAG	1380
G1	CAGGTGTACGAGGGCGAGAGGGCCATGACGCGGGACAACAACCTGCTGGGGCGCTTCGAG	1338
G2	CAGGTGTACGAGGGCGAGAGGGCCATGACGCGGGACAACAACCTGCTGGGGCGCTTCGAG	1338
G3	CAGGTGTACGAGGGCGAGAGGGCCATGACGCGGGACAACAACCTGCTGGGGCGCTTCGAG	1338

Fig. 2: Continue

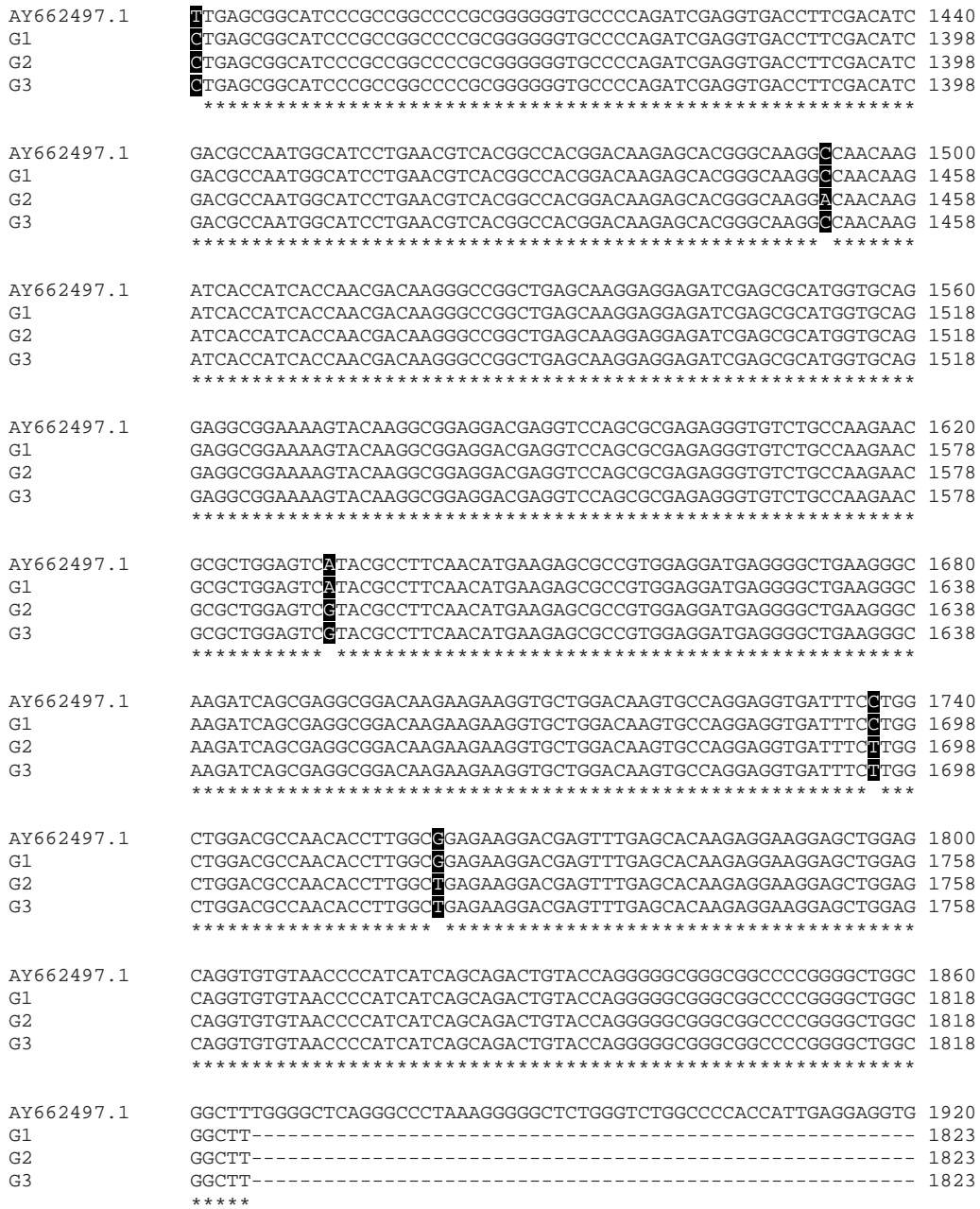


Fig. 2: Identified *Hsp70* gene in Iraqi Holstein bulls and the same gene in GenBank
CLUSTAL Omega (1.2.4) multiple sequence alignment

The result agree with Li *et al.*²⁷, Sodhi *et al.*²⁸, Kerekoppa *et al.*²⁹ and Bhat *et al.*³ on the possibility of polymorphism of *hsp70* gene in cattle.

While the results indicate that the Iraqi Holstein bulls have new haplotypes of *hsp70* in comparison with that recorded in GenBank. The haplotypes of this gene identified from Holstein cattle in Japan and Poland were identical accession number AY662497.1^{30,31}. Similar results were found

in Italian⁶ and Chinese Holstein³². However, polymorphism of crossing Indian cattle with Holstein disagreed with this result³³.

There is no study in line with the results of this study on the location of mutations and perhaps the reason is that this study is the first of its kind on the *hsp70* gene polymorphism in the Holstein bulls in Iraq.

These results might indicate that the Iraqi Holstein bulls have adapted significantly to resist heat stress conditions through changes in the polymorphism of the *Hsp70* gene, that's because the mutations in *Hsp70* gene could be used as marker for heat stress tolerances²⁷ and assisted selection for resistant to heat stress in the breeding³⁴ but this needs to be further studied to determine the association between *Hsp70* gene polymorphism in semen and its characteristics in Holstein bulls born in Iraq.

CONCLUSION

This study showed three new haplotypes of the *Hsp70* gene in the Holstein bulls born in Iraq as a result of mutations in different positions, where Holstein can modify their genotypes of *hsp70* gene to adapting surrounding environmental conditions, even it was originated in different environments. These changes did not affect the animal fitness.

SIGNIFICANCE STATEMENTS

This study discovers that the *Hsp70* gene can manipulate by mutations (silent or missense) according to the environment conditions. This study will help the researcher to uncover the polymorphisms of *hsp70* gene in Holstein bulls born in Iraq and helps to study the relationship between *hsp70* gene polymorphism and semen quality as a guide to the selection of bulls.

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