



Asian Journal of Crop Science

ISSN 1994-7879

science
alert
<http://www.scialert.net>

ANSI*net*
an open access publisher
<http://ansinet.com>



Research Article

Storage Root Nutrient and Yield Enhancement in Sweet Potato Variety VitAto Using Empty Fruit Bunch Compost and Hexaconazole

Borhan Bin Abdul Haya @ Yahya, Mohd Yusoff Abdullah and Nur Kharunisa Tajarudin

Faculty of Plantation and Agrotechnology, Universiti Teknologi Mara (UiTM), P.O. Box 40450, Shah Alam Selangor, Malaysia

Abstract

Background: The effectiveness of oil palm Empty Fruit Bunch (EFB) compost and hexaconazole (HEX) growth regulator in increasing the storage root yield of sweet potato variety. VitAto grown on sandy tin tailing soil was investigated. **Materials and Methods:** Four treatments were used namely, the recommended rate of inorganic fertilizer practice (control), EFB compost and the combination of EFB compost with 10 and 30 ppm HEX. The field experiment layout was a Randomized Complete Block Design (RCBD) with four replications. **Results:** At the maturity stage, the EFB compost with 30 ppm HEX treatment significantly increased the storage root number, fresh weight, dry mass production and harvest index by 125, 35.1, 16.9 and 15.2% higher than control treatment, respectively. At this stage also, this treatment significantly increased the storage root potassium (K) concentration (69.4%) and content (106.9%) higher than the control treatment, respectively. The result showed that the K nutrient was the main nutrient that can be efficiently supplied by EFB compost to the plant. Most nutrients, in particular K uptake were enhanced by the application of HEX. Both K nutrient and HEX at 30 ppm increased the yield primarily through an increase in the storage root number. The higher storage root number and greater proportion of assimilate translocation to the storage root contributed to an increase in the fresh weight and subsequent dry mass production. **Conclusion:** The combination treatment of EFB compost with 30 ppm HEX was better than other treatments in term of increasing the storage root nutrient concentrations, contents and most of yield parameters. The combination of EFB compost and PGR could be considered as an alternative practice to the application of inorganic fertilizer in VitAto cultivation on sandy soil.

Key words: VitAto, sweet potato, EFB compost, hexaconazole, storage root yield, potassium (K), sandy soil

Received: July 19, 2016

Accepted: August 20, 2016

Published: September 15, 2016

Citation: Borhan Bin Abdul Haya @ Yahya, Mohd Yusoff Abdullah and Nur Kharunisa Tajarudin, 2016. Storage root nutrient and yield enhancement in sweet potato variety. VitAto using empty fruit bunch compost and hexaconazole. Asian J. Crop Sci., 8: 87-95.

Corresponding Author: Borhan Bin Abdul Haya @ Yahya, Faculty of Plantation and Agrotechnology, Universiti Teknologi Mara (UiTM), P.O. Box 40450, Shah Alam Selangor, Malaysia

Copyright: © 2016 Borhan Bin Abdul Haya @ Yahya *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Sweet potato or scientifically known as *Ipomoea batatas* (L.) Lam. ranks 7th most important food crop after wheat, rice, maize, Irish potato, barley and cassava worldwide¹. In Malaysia, VitAto variety introduced by MARDI in 2007 is an orange-fleshed sweet potato that is high in β -carotene content and antioxidant compounds such as vitamin C, E and anthocyanin².

VitAto is widely grown on sandy soil in East coast states of Peninsular Malaysia. Yield has been generally low under this condition. The market price for storage roots are usually based on size and currently marketed as grades A, B and C for large, medium and small storage roots, respectively³. VitAto storage roots obtained by farmers are mostly from grades B and C and only a limited few from grade A. This low yield consequently contributes to a low income obtained by farmers.

Empty Fruit Bunches (EFB) is one of the waste or by-product derived from Fresh Fruit Bunch (FFB) after palm oil extraction process in the mill⁴. In 2014, the availability of EFB was abundant with the production volume reached 21 million tons in Malaysia⁵. The use of organic fertilizer or compost was not uncommon in sweet potato cultivation as reported in several studies⁶⁻⁸. However, there is a lack of information or fundamental aspect on how EFB can affect or influence sweet potato storage root yield and quality. The previous study of EFB on sweet potato was focused on the use as the mulching material, parameters studied were on the leaf growth and storage root yield⁹.

Hexaconazole (HEX) belongs to the group of fungicide called triazole compounds. These compounds are extremely effective at very low concentrations¹⁰. The use of HEX as a plant growth regulator has been shown to increase several physiological aspects of yield on root crops such as tuber root, fresh weight, dry mass production, increased in tuber number and length, pigment content, starch, protein and antioxidant of carrot¹¹, sweet potato¹² and white radish¹³. Other triazole compounds such as paclobutrazol were used in floral induction of mango in Malaysia¹⁴. However, HEX was used mostly as a fungicide but never as a growth regulator on any crops in Malaysia.

Therefore, the objectives of this study were to investigate the effects of EFB compost, the combination of EFB compost with HEX on storage root yield components and nutrient concentrations and contents of VitAto in a comparison to the recommended inorganic fertilizer practice currently adopted by most VitAto farmers.

MATERIALS AND METHODS

Experimental site: The field experiment was conducted at MARDI Rawang station, Selangor (N3°16'17.89", E101°30'52.19") from March-July, 2013. The soil type was sandy tin tailing with 90.8% sand, 4.5% silt and 4.7% clay¹⁵. The monthly means for selected climatic factors were 24.1-28.6°C for temperature, 3.0-7.1 mm for rainfall and 76.2-83.4% for relative humidity during the experimental period as shown in Table 1.

Planting materials, treatments and experimental design:

VitAto stem cuttings obtained from MARDI Rawang station were used as planting materials. There were four treatments used in this study. The control treatment was the standard recommended fertilizer practice for the cultivation of VitAto on sandy soil¹⁶. The second treatment consisted of EFB compost (F-EFB) with Nitrogen (N), Phosphorus (P) and Potassium (K) nutrient contents equivalent to the inorganic fertilizer nutrient content in the control treatment. The other two treatments consisted of the EFB compost with 10 (third treatment) or 30 ppm HEX (fourth treatment) designated as F-EFB+10 ppm and F-EFB+30 ppm HEX, respectively. The experimental design was a Randomized Complete Block Design (RCBD) replicated four times. The EFB compost used was aGricare® Premium Compost (Myagri Eco-Biosciences Sdn. Bhd., Selangor, Malaysia) while the fungicide Anmi4.8SC (Advansia Sdn. Bhd., Selangor, Malaysia) was used as plant growth regulator with HEX as the main active ingredient. The formulation of this plant growth regulator was based on previous studies^{11,12,17}. These three treatments; EFB compost with 10 and 30 ppm HEX were used based on their potential to give high yield as indicated on findings of earlier preliminary study. The inorganic fertilizers for control and EFB compost were applied at 7, 28 and 35 days after planting (DAP) based on the VitAto production manual¹⁶. The application of HEX using drenching method was given at 20, 40 and 60 DAP.

Table 1: Selected climatic factors during the experimental period (February to June, 2013)

Year/month	Temperature (°C)		Rainfall (mm)	Mean relative humidity (%)
	Minimum	Maximum		
February	24.1	28.1	3.3	83.4
March	25.7	28.2	3.3	79.8
April	25.7	28.4	6.3	82.5
May	25.5	28.6	7.1	80.7
June	25.7	28.5	3.0	76.2

Source: Data were obtained from the Malaysian Meteorological Department in Kepong Station

Growth analysis procedure: The destructive growth analysis method by Hunt¹⁸ coupled with the sequential growth analysis technique for systematic sampling of plant samples by harvest was employed¹⁹. One plant sample per replicate from each treatment was harvested with a total of four samples per treatment at each harvest. The storage root samples were obtained at 30, 55, 77 and 99 DAP corresponding to the storage root initiation, early and middle bulking and maturity growth stages, respectively. The sample was used to measure yield components of storage root. All samples were oven dried at 72°C for 48 h for dry mass determination and harvest index. Dried storage root samples obtained from the final harvest (99 DAP) were also used for nutrient analysis.

Storage root yield components: The yield parameters measured consisted of storage root and pencil size root numbers, fresh weight, storage root dry mass and harvest index. The classification of storage root was based on

Wilson and Lowe²⁰ where, root diameter greater than 15 mm refer to the storage root and root between 5-15 mm refer to the pencil root. The pencil root was not considered as part of the yield but its diameter range was used to determine smaller storage root known as pencil size root. The difference in physical characteristics between both roots were identified and distinguished before the actual measurement and separation was carried out (Fig. 1). In addition, the storage root dry mass production and value of harvest index at the maturity stage were also used as yield component parameters. The storage root dry mass production was the sum of storage root and pencil size root. Harvest index was the ratio of storage root dry mass per total plant dry mass.

Storage root nutrients analysis: Nitrogen concentration analysis was based on Elementar Analysensysteme GmbH²¹ using vario MACRO cube CHNS elemental analyzer (Elementar Analysensysteme GmbH, Hanau, Germany). The P and K concentrations analysis were based on Campbell and Plank²²



Fig. 1(a-d): Classification on type of roots, (a) Fibrous root, (b) Pencil root, (c) Pencil size root and (d) Storage root

using ICP-OES, Optima 7300 DV (Perkin Elmer, Massachusetts, United states). The storage root nutrient content was computed by multiplying the value of nutrient concentration with the storage root dry mass.

Statistical analysis: The data was analyzed using one way analysis of variance (ANOVA) followed by Tukey's pairwise comparisons for separation of means. The relationship between variables was determined using Pearson correlation. The $p \leq 0.05$ were used to determine levels of significance.

RESULTS AND DISCUSSION

Yield components

General trend: The storage root number and fresh weight were continuously increased from the storage root initiation to maturity stages (Fig. 2a, b). The pencil size root number and fresh weight were generally increased from the storage root initiation to the early bulking stage (55 DAP) before declining thereafter (Fig. 2c, d). At the early bulking stage, the storage root number for F-EFB was 200% significantly higher than the

control while at the mid bulking (77 DAP) through maturity stages, both F-EFB and F-EFB+30 ppm HEX were 167% and 125% significantly higher than the control treatment, respectively. The storage root fresh weight for F-EFB+10 ppm and F-EFB+30 ppm HEX were 87 and 35% significantly higher than control at the mid bulking (77 DAP) and maturity stages, respectively. However, there were no significant differences between treatments at the earlier growth stages.

The pencil size root number was not significantly affected by treatment application throughout the entire growing period, however, at the mid bulking stage (77 DAP), the pencil size root for the control was higher than other treatments. At the early bulking stage (55 DAP), the pencil size root fresh weight for the F-EFB treatment was significantly higher than the control and at the mid bulking stage (77 DAP), the F-EFB+10 ppm was significantly higher than F-EFB+30 ppm while at the maturity stage, the control treatment was significantly higher than F-EFB+10 ppm HEX. The storage root dry mass production and harvest index for F-EFB+30 ppm HEX were 17 and 16% significantly higher than control at the maturity stage, respectively (Fig. 3a, b).

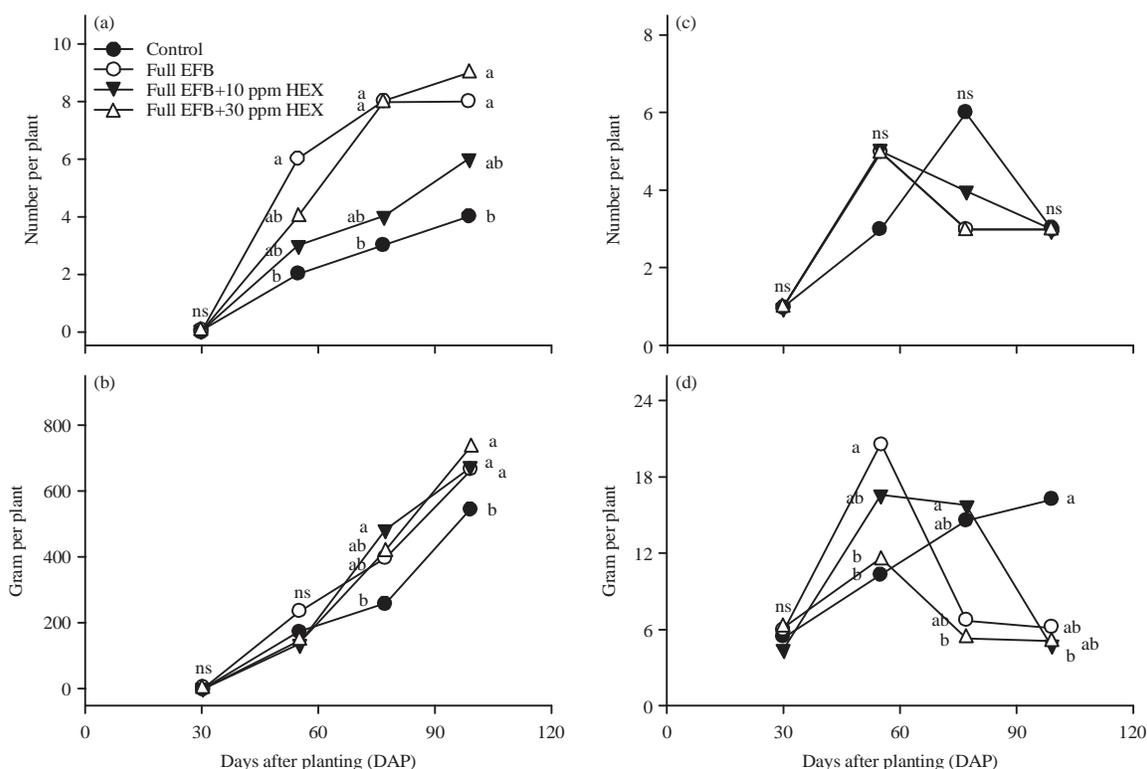


Fig. 2(a-d): Yield components, (a) Storage root number, (b) Storage root fresh weight, (c) Pencil size root number, (d) Pencil size root fresh weight as affected by various EFB compost and HEX combination treatments throughout the entire growing period, different letters follow different treatments at specific harvest time indicate a significant difference at $p \leq 0.05$ while ns indicate non-significant and the values are the means of four replicates

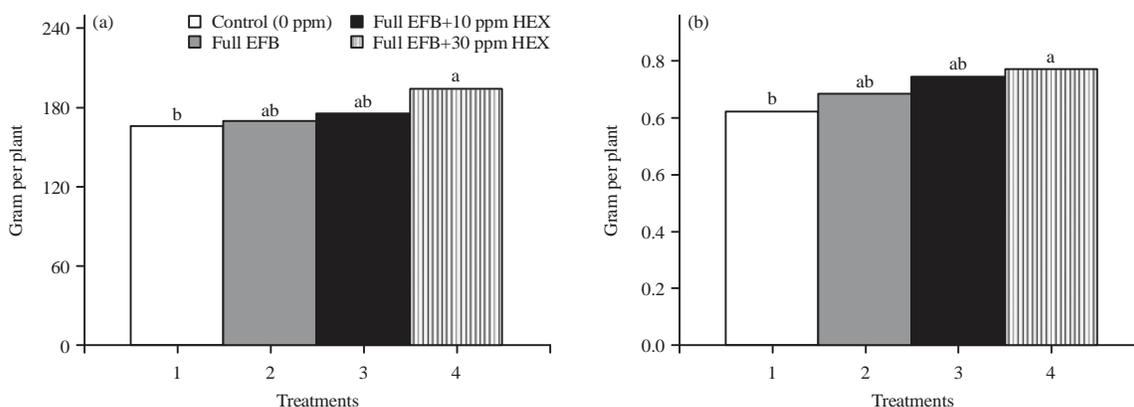


Fig. 3(a-b): Yield components (a) Storage root dry mass and (b) Harvest index as affected by various EFB compost and HEX combination treatments at the maturity stage, different letters follow different treatments at specific harvest time indicate a significant difference at $p \leq 0.05$ while ns indicate non-significant and the values are the means of four replicates

Table 2: Correlation coefficients of various parameters related to yield components as affected by various EFB compost and HEX combination treatments during the entire growing period

Parameters	No. of storage root	Storage root fresh weight	No. of pencil size root	Pencil size root fresh weight	Storage root dry mass production
Storage root fresh weight	0.50***				
No. of pencil size root	-0.05 ^{ns}	-0.34*			
Pencil size root fresh weight	-0.14 ^{ns}	-0.24 ^{ns}	0.72***		
Storage root dry mass production	0.51***	0.97***	0.02 ^{ns}	0.001 ^{ns}	
Harvest index	0.51***	0.93***	-0.17 ^{ns}	-0.27 ^{ns}	0.90***

*, **, ***Significant at $p \leq 0.05$, $p \leq 0.01$ and $p \leq 0.001$, respectively, ^{ns}No significant correlation and the values are the means of four replicates

Table 3: Nitrogen, phosphorus and potassium concentrations of storage root as affected by various EFB compost and HEX combination treatments at the maturity stage

Treatment	Nutrient concentration (%)			
	Control	F-EFB	F-EFB+10 ppm	F-EFB+30 ppm
Storage root				
Nitrogen	1.89 ^a	1.94 ^a	1.80 ^a	1.86 ^a
Phosphorus	0.18 ^a	0.16 ^a	0.13 ^a	0.18 ^a
Potassium	0.62 ^c	0.97 ^{ab}	0.77 ^{bc}	1.05 ^a

Different letters follow different treatments within row indicate a significant difference at $p \leq 0.05$ and the values are the means of four replicates

Table 4: Nitrogen, phosphorus and potassium contents of storage root as affected by various EFB compost and HEX combination treatments at the maturity stage

Treatment	Nutrient content (g)			
	Control	F-EFB	F-EFB+10 ppm	F-EFB+30 ppm
Storage root				
Nitrogen	3.33 ^a	3.21 ^a	3.24 ^a	3.35 ^a
Phosphorus	0.32 ^a	0.21 ^a	0.22 ^a	0.30 ^a
Potassium	1.01 ^b	1.68 ^{ab}	1.29 ^b	2.09 ^a

Different letters follow different treatments within row indicate a significant difference at $p \leq 0.05$ and the values are the means of four replicates

Correlation studies of the various parameters related to the yield components: Most yield component parameters shows significant positive correlation except with pencil size

root number and fresh weight (Table 2). However, the pencil size root parameters were positively correlated to each others.

Nutrient concentrations and contents in the storage root:

The F-EFB+30 ppm HEX had 69 and 36% significantly higher storage root K concentration than the control and F-EFB+10 ppm HEX treatments, respectively (Table 3). At the maturity stage, the K nutrient content of storage root was significantly higher in F-EFB+30 ppm treatment as compared to control and F-EFB+10 ppm HEX treatments. However, N and P contents were not significantly different in all treatments (Table 4).

Correlation studies of the various parameters related to storage root nutrient concentrations and contents with yield components at the maturity stage:

The storage root K concentration was significantly correlated with storage root number ($r = 0.83^{***}$) while K nutrient content of the storage root was significantly correlated with both storage root number and fresh weight ($r = 0.82^{***}$ and 0.67^{**} , respectively). However, both N and P concentrations and contents of storage root did not show any significant correlation with any of yield component parameters (Table 5).

Table 5: Correlation coefficient of various parameters related to yield components and storage root nutrient concentrations and contents as affected by various EFB compost and HEX combination treatments at the maturity stage

Storage root	Parameters					
	No. of storage root	Storage root fresh weight	No. of Pencil size root	Pencil size root fresh weight	Storage root dry mass	Harvest index
Nitrogen						
Concentration	0.10 ^{ns}	-0.23 ^{ns}	0.10 ^{ns}	-0.40 ^{ns}	-0.56 ^{ns}	0.04 ^{ns}
Content	-0.30 ^{ns}	-0.03 ^{ns}	0.46 ^{ns}	-0.50 ^{ns}	0.48 ^{ns}	0.04 ^{ns}
Phosphorus						
Concentration	0.55 ^{ns}	-0.18 ^{ns}	-0.15 ^{ns}	0.25 ^{ns}	0.00 ^{ns}	-0.25 ^{ns}
Content	0.48 ^{ns}	-0.20 ^{ns}	-0.15 ^{ns}	0.01 ^{ns}	0.34 ^{ns}	-0.22 ^{ns}
Potassium						
Concentration	0.83 ^{***}	0.53 ^{ns}	-0.10 ^{ns}	-0.40 ^{ns}	0.24 ^{ns}	0.38 ^{ns}
Content	0.82 ^{***}	0.67 ^{**}	-0.08 ^{ns}	-0.50 ^{ns}	0.58 [*]	0.45 ^{ns}

*, **, ***Significant at $p \leq 0.05$, $p \leq 0.01$ and $p \leq 0.001$, respectively, ^{ns}No significant correlation and the values are the means of four replicates

Yield components

Response of yield components to treatments application:

In general, the storage root number, fresh weight, dry mass production and harvest index responded positively to the application of F-EFB and the combination treatments of EFB and HEX. Among these treatments, the F-EFB+30 ppm HEX was considered the best treatment since, it significantly increased all yield component parameters higher than the control. This study indicated that the application of EFB compost alone was able to increase yield components comparable to the application of recommended inorganic fertilization. The use of organic fertilizer as main sources of nutrients that lead to either better or comparable yield with inorganic fertilizer was reported in many sweet potato studies^{6,7,23}. Similar applications on other crops were also reported such as corn applied with corn straw or farmyard manure²⁴ and cassava applied with cattle manure²⁵. The hexaconazole application was also reported to increase storage root number, fresh weight and dry mass production of several crops such as carrot¹¹ and sweet potato^{12,13}. Therefore, a combination of EFB compost with HEX was likely to further enhance the yield of VitAto as indicated by higher storage root number, fresh weight, dry mass production and harvest index.

Relationship among yield components: Storage root number is a yield component parameter that represents the sink size²⁶. It acts as a storage organ that accumulates or stores assimilate or dry mass produced. The higher the storage root number the greater would be the sink capacity. Higher dry mass production and subsequent allocation are essential to ensure that all storage roots can be sufficiently filled. Storage root cells must be fully formed and expanded before they are able to store assimilates²⁷. Treatments that can increase storage root numbers can also increase other yield components as shown particularly in the F-EFB+30 ppm HEX treatment. To

maximize marketable yields, it is important to attain as high number of storage roots as possible²⁸. As shown in the result, storage root number was significantly correlated with storage root dry mass production. This finding was in agreement with the results obtained by Bourke²⁹ and Bhagsari³⁰. In this study, the storage root numbers also significantly correlated with storage root fresh weight similar to the finding of study by Bhagsari³⁰. The higher storage root fresh weight contributed to higher storage root dry mass production as shown in correlation studies which was also observed by several researchers^{30,31}. It showed that the higher dry mass partitioning to storage root led to the improvement of storage root size. The close relationship of the harvest index with the storage root yield found in this study was in agreement with other reported studies^{30,31}.

Storage root versus pencil size root: The number and fresh weight of smaller storage root or also known as pencil size root were higher for the control compared to other treatments. The maximum production of pencil size roots for F-EFB and other combination treatments were observed at 55 DAP while for the control it was at 77 DAP. The F-EFB and the combination treatments attained their maximum production of new and smaller storage roots much earlier than the control treatment which seemed to continuously increase even beyond the bulking stage. Similar pattern was also observed in the fresh weight of pencil size root in which F-EFB and the combination treatments attained their maximum values at 55 DAP and gradually decreased thereafter. However, the fresh weight continued to increase to the maturity stage for the control treatment. This study suggested that when the sweet potato plant had reached the bulking stage, the production of new and smaller storage roots were reduced and most of the dry mass produced would be used for the growth of existing storage root. Similar observations were also reported elsewhere^{27,32}.

Relationship between storage root nutrient concentrations and contents with yield components at the maturity stage:

Storage root K concentration had significant correlation with storage root number while storage root K nutrient content showed significant correlation with storage root numbers, fresh weight and dry mass production. The K nutrient generally increased the yield through an increase in the storage root numbers²⁹. The K nutrient facilitated greater amount of photosynthate translocation from the source leaves to the storage root through an increase in the photosynthetic efficiency which enhanced the storage root size^{7,29,33}. The application of K fertilizer can increase the sink strength of storage root to attract greater photosynthate at all stages of growth, particularly during the maturity stage³⁴. In contrast, K deficiency could have caused significant reduction in several physiological characteristics such as total biomass production and root yield³⁵. The treatment that influences an increase in K concentration and content in plant are likely to increase the storage root yield at the harvestable stage as shown by F-EFB+30 ppm HEX treatment. Both EFB compost and HEX had something in common that capable of promoting the reproductive growth and at the same time reduced the growth of the vegetative parts. The EFB compost can effectively provide adequate K nutrients to the plant which in turn enhance assimilates translocation from leaves to the storage roots. The HEX is one of the triazole compounds that was reported to be capable altering nutrient uptake and plant nutrition apart from increasing the greater assimilate translocation from shoot to root as stated by Gomathinayagam *et al.*¹⁷. The HEX application¹² and K fertilization³³ were shown to increase root to shoot ratio of sweet potato.

The N and P concentrations and contents had non-significant correlation with any of the storage root yield components. Phosphorus nutrient was not considered important nutrient for sweet potato⁷ and the reduction in P might not significantly affect the yield markedly³⁶. This crop was reported to tolerate soil with low or deficient in P nutrient³⁷. This study also indicated that N was not a critical nutrient at the maturity stage. However, N would be the most needed nutrient during the vegetative stage which can promote in the production of assimilates through enhancement in leaf photosynthetic rates. The higher total dry mass production is one of the factors that can influence the storage root yield³². The total dry mass must be high to ensure high storage root dry mass. This role probably efficiently carried out by N rather than P or K nutrients. Bourke²⁹ stated that N influenced the storage root yield as a consequent to the lengthening the leaf area duration which in turn increased the

storage root weight. The second factor that can influence storage root yield is the efficiency of assimilates translocation to the storage root as described by Belehu³². This task is fulfilled by K nutrient as reported by previous studies^{7,29,33,34}. Therefore, N is important nutrient during the vegetative stage while K is an important nutrient during the storage root bulking and maturity stage³⁶ where most of the assimilates was allocated to the storage root.

CONCLUSION

Insufficient of available K for plant growth during storage root development could be the key problem of smaller and lower grades of storage root under recommended fertilization practice using inorganic fertilizer. In contrast, the EFB compost that efficiently supply the K nutrient to the plant and HEX especially at 30 ppm concentration increased the sink strength of storage root to attract more assimilate from the source. Consequently, they increased the sink size with appropriate number of storage root. In conclusion, the better treatment that can increase storage root yield and quality of VitAto is the combination of EFB compost and 30 ppm HEX treatments. These combination treatments could be used as an alternative practice to the application of inorganic fertilizer in VitAto cultivation on sandy soil.

ACKNOWLEDGMENT

The researchers would like to thank MARDI and specifically MARDI Rawang station for providing research plot, other logistics and field assistance during the entire experimental period.

REFERENCES

1. Mohammad, N., S. Mansooreh, H.H. Khankahdani and E. Naseri, 2014. Feasibility sweet potato (*Ipomoea batatas* L.) growing in Southern regions of Iran (Minab) climate. *Scientia Agriculturae*, 5: 67-72.
2. Tan, S.L., A. Zaharah, A.M. Abdul Aziz, Y. Mohsin, A.K. ZainalAbidin and I. Abd Hamid, 2010. VitAto-variety keledak kaya dengan β -carotina [Vitato-a β -carotene rich sweetpotato variety]. *Buletin Teknologi Tanaman* 7: 23-33.
3. Rosly, A.R., 2010. Potensi tanaman VitAto Semerak [VitAto-potential crop in Semerak]. *Berita Transformasi Pertanian*.
4. Soom, R.M., W.H.W. Hasamudin, A.M. Top and K. Hassan, 2006. Thermal properties of oil palm fibre, cellulose and its derivatives. *J. Oil Palm Res.*, 18: 272-277.

5. MPOB., 2015. Fresh Fruit Bunch (FFB) received by mill for the month of December 2014. <http://bepi.mpob.gov.my/index.php/statistics/sectoral-status/127-sectoral-status-2014/674-ffb-received-by-mill-2014.html>
6. Mukhtar, A.A., B. Tanimu, U.L. Arunah and B.A. Babaji, 2010. Evaluation of the agronomic characters of sweet potato varieties grown at varying levels of organic and inorganic fertilizer. *World J. Agric. Sci.*, 6: 370-373.
7. Kareem, I., 2013. Growth, yield and phosphorus uptake of sweet potato (*Ipomoea batatas*) under the influence phosphorus fertilizers. *Res. J. Chem. Environ. Sci.*, 1: 50-55.
8. Agyarko, K., H.K. Dapaah, S. Buah and K.A. Frimpong, 2014. Sweet potato (*Ipomoea batatas*) yield parameters, soil chemical properties and cost benefit ratios following incorporation of poultry manure and inorganic NPK fertilizers in low nutrient Ghanaian soils. *Int. J. Plant Soil Sci.*, 3: 129-138.
9. Ravoof, A.A., 1988. Crop Production in Sand with Oil Palm Empty Fruit Bunch Mulching. In: *Technology for Rural Development: Proceedings of the Second Asia Conference*, Kuala Lumpur, Malaysia, December 4-7, 1985, Radhakrishna, S., M.M. Singh and C.K. John (Eds.). World Scientific, Singapore, ISBN: 9971502852, pp: 657-670.
10. Newman, J.P., 2014. Crop Production and Management. In: *Container Nursery Production and Business Management Manual*, Newman, J.P. (Ed.). University of California, Division of Agriculture and Natural Resources, Oakland, CA., USA., ISBN-13: 9781601078421, pp: 59-131.
11. Gopi, R., C.A. Jaleel, R. Sairam, G.M.A. Lakshmanan, M. Gomathinayagam and R. Panneerselvam, 2007. Differential effects of hexaconazole and paclobutrazol on biomass, electrolyte leakage, lipid peroxidation and antioxidant potential of *Daucus carota* L. *Colloids Surf. B: Biointerfaces*, 60: 180-186.
12. Sivakumar, T., A. Sundaramanickam and R. Panneerselvam, 2009. Changes in growth and pigment content in sweet potato by Triadimefon and Hexaconazole. *J. Phytol.*, 1: 333-341.
13. Sridharan, R., S. Raja and P. Sakthivel, 2015. Effect of triazole compounds on induced changes in growth biomass and biochemical content of white radish (*Raphanus sativus* L.). *J. Plant Stress Physiol.*, 1: 43-48.
14. Hartinee, A., M. Zabedah, H.M. Azhar, S.M. Asrul and T.M.T. Ab Malik, 2014. Effects of girdling technique on flowering of mango cv. Chok anan. *Trans. Malaysian Soc. Plant Physiol.*, 22: 101-104.
15. Vimala, P., 2005. Getting Started. In: *Organic Vegetable Cultivation in Malaysia*, Aini, Z., A. Sivapragasam, P. Vimala and M.N.M. Roff (Eds.). Malaysian Agricultural Research and Development Institute, Serdang, Malaysia, ISBN: 9789679364712, pp: 207-208.
16. MARDI., 2008. VitAto. Brochure, Unit Pengeluaran Bahan Tanaman, Biji Benih dan Baka Ternakan, Serdang, Malaysia.
17. Gomathinayagam, M., C.A. Jaleel, M.M. Azooz and R. Panneerselvam, 2009. Superoxide dismutase and ascorbate peroxidase profile changes with triazole applications in *Manihot esculenta* Crantz. *Global J. Mol. Sci.*, 4: 23-28.
18. Hunt, R., 1990. *Basic Growth Analysis: Plant Growth Analysis for Beginners*. 1st Edn., Unwin Hyman, London, UK., ISBN-13: 9780044453727, Pages: 112.
19. Jolliffe, P.A., G.W. Eaton and J.L. Doust, 1982. Sequential analysis of plant growth. *New Phytol.*, 92: 287-296.
20. Wilson, L.A. and S.B. Lowe, 1973. The anatomy of the root system in West Indian sweet potato (*Ipomoea batatas* (L.) Lam.) cultivars. *Ann. Bot.*, 37: 633-643.
21. Elementar Analysen systeme GmbH, 2005. Operating the vario EL: Sample preparation. In: *Vario EL ///CHNOS Elemental Analyser, Operating Instructions for Instruments Starting with Serial No. 11054041*, Elementar Analysensysteme GmbH, Hanau, Germany, May 2005, pp: 6.1-6.5.
22. Campbell, C.R. and C.O. Plank, 1998. Preparation of Plant Tissue for Laboratory Analysis. In: *Handbook of Reference Methods for Plant Analysis*, Kalra, Y.P. (Ed). Chapter 3, CRC Press, Boca Raton, FL., USA., ISBN-13: 9781420049398, pp: 37-49.
23. Taraken, I.T. and R. Ratsch, 2009. Sweetpotato Cultivation on Composted Mounds in the Highlands of Papua New Guinea. In: *Soil Fertility in Sweetpotato-based Cropping Systems in the Highlands of Papua New Guinea*, Kirchhof, G., (Ed.). Australian Centre for International Agricultural Research, Canberra, ISBN: 9781921531804, pp: 24-32.
24. Song, Z., H. Gao, P. Zhu, C. Peng and A. Deng *et al.*, 2015. Organic amendments increase corn yield by enhancing soil resilience to climate change. *Crop J.*, 3: 110-117.
25. Mathias, L. and V.H. Kabambe, 2015. Potential to increase cassava yields through cattle manure and fertilizer application: Results from Bunda College, Central Malawi. *Afr. J. Plant Sci.*, 9: 228-234.
26. Marschner, H., 1995. *Mineral Nutrition of Higher Plants*. 2nd Edn., Academic Press Ltd., London, UK., ISBN-13: 978-0124735439, Pages: 889.
27. Lebot, V., 2009. *Tropical Root and Tuber Crops: Cassava, Sweet Potato, Yams and Aroids*. CABI, Cambridge, MA., USA., ISBN-13: 9781845936211, Pages: 433.
28. Meyers, S.L., R.A. Arancibia, M.W. Shankle, J. Main, B. Gajanayake and K.R. Reddy, 2014. *Sweet Potato Storage Root Initiation*. Mississippi State University Extension Service, Mississippi, U.S., Pages: 3.
29. Bourke, R.M., 1985. Influence of nitrogen and potassium fertilizer on growth of sweet potato (*Ipomoea batatas*) in Papua New Guinea. *Field Crops Res.*, 12: 363-375.
30. Bhagsari, A.S., 1990. Photosynthetic evaluation of sweetpotato germplasm. *J. Am. Soc. Hortic. Sci.*, 115: 634-639.
31. Bhagsari, A.S. and D.A. Ashley, 1990. Relationship of photosynthesis and harvest index to sweet potato yield. *J. Am. Soc. Hortic. Sci.*, 115: 288-293.

32. Belehu, T., 2003. Agronomical and physiological factors affecting growth, development and yield of sweet potato in Ethiopia. Ph.D. Thesis, University of Pretoria.
33. George, M.S., G. Lu and W. Zhou, 2002. Genotypic variation for potassium uptake and utilization efficiency in sweet potato (*Ipomoea batatas* L.). *Field Crops Res.*, 77: 7-15.
34. Liu, H., C. Shi, H. Zhang, Z. Wang and S. Chai, 2013. Effects of potassium on yield, photosynthate distribution, enzymes' activity and ABA content in storage roots of sweet potato (*Ipomoea batatas* Lam.). *Aust. J. Crop Sci.*, 7: 735-743.
35. Tang, Z.H., A.J. Zhang, M. Wei, X.G. Chen, Z.H. Liu, H.M. Li and Y.F. Ding, 2015. Physiological response to potassium deficiency in three sweet potato (*Ipomoea batatas* [L.] Lam.) genotypes differing in potassium utilization efficiency. *Acta Physiologiae Plantarum*, Vol. 37. 10.1007/s11738-015-1901-0
36. FAO., 2005. Crop Response to Fertilizers. In: *Fertilizer Use by Crop in Ghana*, FAO (Ed.). Food and Agriculture Organization, Rome, pp: 39.
37. Djazuli, M. and T. Tadano, 1990. Comparison of tolerance to low phosphorus soils between sweet potato and potato. *J. Fac. Agric. Hokkaido Univ.*, 64: 190-200.