



Research Article

Postharvest Treatments for Improving the Quality of Fresh and Processed Costata Persimmon Fruits

¹Ahmed Mohamed Saied Hussein, ²Omaima Mohamed Hafez, ²Nagwa Selmy Zayed, ²Malaka Abd Elfatah Saleh and ¹Mohie Mostafa Kamil

¹Department of Food Technology, National Research Center, Dokki, Cairo, Egypt

²Department of Pomology, National Research Centre, Dokki, P.O. Box 12622, Cairo, Egypt

Abstract

Background and Objective: Kaki (Costata) is ripening differently than other fruits. Really, hard persimmons to ripen so expect to wait for several weeks for ripe. Persimmon contain relatively high content of dietary fibers, total and major phenolics, main minerals and trace elements make persimmon preferable for health. This study aimed to improve the quality of Kaki fruits through some postharvest treatments to remove astringency and ripening. Fruit leathers products provide attractive, colored and flavor, therefore, leather kaki of postharvest treated fruits were also prepared and evaluated to improve the quality of processed fruits. **Materials and Methods:** Postharvest pretreatment was carried out to improve the color quality of Kaki fruits by treating fruits separately with 0% (T1), 2% (T2), 3% (T3) calcium chloride solution and exposed to CO₂ (T4). Moisture, protein, fat, fiber, ash, total solids, total sugars, reducing and non reducing sugars, total flavonoid, antioxidant of the samples were determined. **Results:** Postharvest treatments caused a noticeable increase in color parameters of Kaki fruits compared to untreated fruits. Where, T2 and T3 had the highest lightness value (L = 37.61 and 37.23); followed with T4 (35.76). While, redness (a*) maximized in T4 (19.61), followed with T2 (17.18) and T3 (15.50). Also chemically, protein content was maximized in postharvest treated fruits by CO₂ (5.41%), followed with treated fruits with 3% Ca Cl₂ (2.50%). But, TSS of treated kaki fruit juice was decreased slightly, where they ranged between 18.4-18.6 compared to untreated fruits (20.1). Furthermore, post harvest Kaki fruits were evaluated to produce leather product. The obtained Hunter color parameter showed that the lowest color parameters was found in leather of untreated fruits, while the highest value was found in leather of treated fruits with CO₂. **Conclusion:** Sensory evaluation indicated that treated fruits not affected significantly in color, odor, appearance and overall acceptability of leather product.

Key words: Persimmon "Costata" cultivar, Kaki fruits, postharvest treatments, astringency removal and ripening, fruits leather

Received: September 07, 2018

Accepted: November 09, 2018

Published: December 15, 2018

Citation: Ahmed Mohamed Saied Hussein, Omaima Mohamed Hafez, Nagwa Selmy Zayed, Malaka Abd Elfatah Saleh and Mohie Mostafa Kamil, 2019. Postharvest treatments for improving the quality of fresh and processed costata persimmon fruits. Asian J. Crop Sci., 11: 1-7.

Corresponding Author: Ahmed Mohamed Saied Hussein, Department of Food Technology, National Research Center, Dokki, Cairo, Egypt
Tel: +201016426755

Copyright: © 2019 Ahmed Mohamed Saied Hussein *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Kaki (Costata) is ripening differently than other fruits. Really, hard persimmons to ripen so expect to wait for several weeks for ripe. Immature Kaki fruits have a high proanthocyanidin-type tannin content which it makes astringent and bitter. The astringent type persimmon fruit is not edible at harvest due to the presence of soluble tannins that cause an astringent taste in the mouth¹. The fruit becomes edible during postharvest ripening, when soluble tannins polymerize and become insoluble². A higher CO₂ concentration always produces more acetaldehyde and makes astringency removal faster. Also, temperature plays an important role in anaerobic metabolism; more acetaldehyde and ethanol accumulate at higher temperatures³. The eating quality of persimmon is considered best at the end of the pre-climacteric stage owing to presence of maximum sugars and desired orange color^{4,5}. One of the most important mineral elements determining fruit quality is Calcium. Postharvest Ca treatments used to increase Ca content of the cell wall which delaying senescence, resulting in firmer, higher quality fruit and with less susceptible to disease^{6,7}.

Persimmon (*Diospyros kaki* L.) is the main and leading that consumed in the Egyptian market. It is not left to ripe on the trees and harvested at mature stage to ripened for commercial production and marketing⁸. Persimmon is relatively high content of dietary fibers, total and major phenolics, main minerals and trace elements make persimmon preferable for healthy⁹. It is also a good source of fiber and vitamins, mainly A and C. It is mainly eaten fresh, but can be frozen, canned or dried and can be stored for up to 6 month in modified or controlled atmospheres¹⁰. The dried persimmon portions could be used as ingredient in products such as muesli, snacks and breakfast cereals¹¹.

Fruit leather, also called a fruit sheet, is a dehydrated fruit based confectionery dietary product which is often eaten as snack, dessert or after rehydration. It is chewy and flavorful, low in fat and high in fiber and carbohydrates; it is also light weight and easily for storing and packaging. Fruit leathers made from fresh fruit pulp or a mixture of fruit juice concentrates and other ingredients after a complex operation that involves a dehydration step^{12,13}. Fruit pulp-based fruit leathers are nutritious and sensorially acceptable to customers. They contain substantial quantities of dietary fibers, carbohydrates, minerals, vitamins and antioxidants that remain in finished product¹⁴.

Most fresh fruits have a short harvest season and are sensitive to deterioration; therefore, making fruit leather from fresh fruits is an effective method to preserve fruits¹³. Fruit

leathers are manufactured by dehydrating a fruit puree into a leather like sheet. Fruit leathers are often considered as a health food, therefore there are large numbers of fruit leather products available on the market, i.e., apricot leather and mango leather.

Basically, fruit pulps are mixed with appropriate quantities of sugar, pectin, acid and color and then dried into sheet-shaped products. Gujral and Brar¹⁴ added sugars and pectin to mango leathers. The sugar gave the product a sweeter taste and increased the solids content; then pectin was used to thicken the pulp, modify the flexible texture and ensure the retention of the shapes of the dried product. Furthermore, they also prepared mango leather with the addition of potassium meta bisulphite to get better sensory qualities and the results were satisfactory for consumers. Various additives can be used, such as glucose syrup, sodium metabisulphite and ascorbic acid, depending on the types of fruit leather¹⁵⁻¹⁷.

Fresh fruits are known to be excellent sources of vitamins, minerals, fibers, carbohydrates and other bioactive compounds. The objectives of this study were to improve the quality of Kaki fruits through some postharvest treatments to remove astringency and ripening. Fruit leathers provide attractive, colored and flavorsome products for people, therefore, leather kaki fruits were also prepared and evaluated to improve the quality of fresh and processed fruits.

MATERIALS AND METHODS

Plant sample: Kaki fruits (Costata cultivar) an astringent persimmon cultivar were harvested at maturity stage and coloring (two thirds of the surface) at mid of October 2015, from productive trees 12 years old planted at 4×4 m distant and grow in a loamy soil under flood irrigation system, in a private orchard located at Qalyubia Governorate Egypt. The productive trees were similar in growth vigor had a good physical condition and received similarly the recommended horticultural practices. The soil had 2.04% organic matter, 8.4 pH, 0.32 dsn⁻¹ EC, 1.6% CaCO₃, 2.8% P, 47.2% K, 1000% Ca, 114% Mg, 15.8% Na, 7.6 ppm Fe, 3.4 ppm Mn, 1.4 ppm Zn and 1.7 ppm Cu.

Post harvest treatments: Four hundred and twenty mature fruits (60 kg) "greenish-yellow color" were selected free from visual defects "undamaged, uniformed in shape, free from blemishes and pathogen infection" were divided into 4 treatments, every treatment contained 105 fruits (15 kg) and washed separately with tap water, air dried, placed into plastic baskets. The four treatments were: T₁ untreated fruits as a

control. The T₂ and T₃ fruits were immersed for 5 min in 20 L tanks containing 2 and 3% (w/v) calcium chloride solution, respectively. The T₄ fruits were exposed with CO₂ treatment¹⁸. All treatments were stored at 20°C for 2 days, then immediately transported to the laboratory of food technology, National Research Centre (NRC)-Dokki Giza-Egypt.

Preparing kaki leather: Fruit was rinsed, chopped, mixed, homogenized with 12% water and 0.03% citric acid; then cooked at low heating through uncovered kettle and stirred during cooking. Simmer and stir were continued until the fruit purée has thickened. The purée should be very smooth. Puree was poured into line baking sheet. Baking sheet was placed in the oven at low temperature (40°C). The fruit leather is ready when it is no longer sticky, but has a smooth surface. After which, the fruit leather was peeled up from the plastic wrap and stored in the refrigerator.

Chemical analysis: Moisture, protein, fat, fiber, ash, total solids, total sugars, reducing and non reducing sugars of the samples were determined according to the methods of AOAC¹⁹. Total soluble solids (TSS) were determined using a Hand refractometer (ATAGO, Japan) and expressed as Brix value. Acidity was measured according to the method of AOAC¹⁹ and expressed as citric acid (%). Brix/acid ratio was calculated by dividing the total soluble solids on the total acidity values for each sample. The pH was measured using Hanna pH-meter HI 9021 m Germany. Also, vitamin C content was determined according to AOAC¹⁹ using 2, 6 dichlorophenol-endo-phenol. Viscosity was measured using HAAKE viscometers (Haake, Mess-Technik GmbH. Co., Germany), thermostatic bath was used to control working temperature within 25°C. Viscosity result was determined in centipoise (cP) unit according to the method of Ibarz *et al.*²⁰.

Determination of hunter color parameters: Color parameters (L*, a* and b*) of jam samples were determined using a spectro-colorimeter (Tristimulus Colour Machine) with the CIE lab color scale (Hunter, Lab Scan XE-Reston VA, USA) in the reflection mode. The instrument was standardized with white tile of Hunter Lab Color Standard (LX No.16379): X = 72.26, Y = 81.94 and Z = 88.14 (L* = 92.46; a* = -0.86; b* = -0.16) according to Sapers and Douglas²¹.

Total phenolics and flavonoids contents: Total phenolics content of sheet samples were determined using the method of Folin-Ciocalteu²². Results were expressed as gallic acid equivalent (mg GAE/g dry weight). Flavonoids contents of

sheet samples were determined using AlCl₃ method²³ and expressed as catechine equivalents (mg CAT/g sample dry weight).

Antioxidant activity: Antioxidant activity was determined using DPPH radical-scavenging assay and assay as reported by Grzegorzczak *et al.*²⁴. Various concentrations of ethanol and ethanol extracts of tested samples (50, 100, 150 and 200 µg mL⁻¹) were added to 4 mL of 0.1 mM DPPH solution in methanol and the mixture was shaken vigorously. After incubation for 30 min at room temperature the absorbance was recorded at 517 nm. TBHQ used as a reference in the same concentration range of tested extract. A control solution, without a tested compound was prepared in the same manner as the assay mixture. All analyses were carried out in triplicate. The degree of decolorization indicates the radical-scavenging efficiency of the extract. The antioxidant activity of tested samples was calculated as an inhibitory effect (I%) of the DPPH radical formation as follows:

$$\text{Inhibition (\%)} = 100 \times \frac{A_{517}(\text{control}) - A_{517}(\text{sample})}{A_{517}(\text{control})}$$

The EC₅₀ value was defined as the concentration (µg mL⁻¹) of the compound required to scavenge the DPPH radical by 50%.

Sensory evaluation: Sensory evaluation of sheet samples was carried out through evaluating taste, odor, color, mouth feel, appearance and overall acceptability as described by Hussein and Shedeed²⁵.

Statistical analysis: Statistical analysis of tested samples was carried out using SPSS program to calculate standard deviations, one way analysis of variance (ANOVA), with multiple ranges least significant difference (LSD) test (p<0.05).

RESULTS AND DISCUSSION

Chemical properties of postharvest treatment kaki fruits ("Costata" cv.) were determined and compared with untreated fruits sample. Table 1 showed that the chemical composition of treated kaki fruits was affected slightly compared to non-treated sample, where moisture content of untreated or treated fruits ranged between 18.10-23.88%. Ash content was slightly higher in treated fruits with CO₂ followed with untreated fruits and treated fruits with 3% CaCl₂. Also, protein content was slightly higher in post-harvest treated fruits with CO₂ followed with treated fruits with 3, 2% CaCl₂

Table 1: Effect of postharvest treatments of kaki fruits on their physicochemical properties of juice

Physicochemical properties	Untreated fruits (T1)	Fruits immersed in CaCl ₂ /5 min		Fruits exposed to CO ₂ (T4)
		2% (w/v) (T2)	3% (w/v) (T3)	
Moisture (%)	20.63±0.25	18.10±0.34	23.88±0.45	22.31±0.32
Ash (%)	0.55±0.09	0.18±0.01	0.42±0.03	0.87±0.05
Protein (%)	0.36±0.02	0.60±0.04	2.50±0.21	5.41±0.18
Fiber (%)	36.36±0.65	29.77±0.52	83.31±0.33	60.09±0.29
pH	6.10±0.11	6.11±0.250	6.06±0.30	6.09±0.15
Acidity (%)	0.28±0.01	0.19±0.01	0.96±0.02	0.21±0.01
TSS (%)	20.1±0.09	18.6±0.110	18.6±0.070	18.4±0.080

Table 2: Effect of calcium chloride and CO₂ postharvest treatments on the hunter color parameters of Kaki fruits

Samples	L	a	b	a/b	ΔE	Satu-ration	Hue
Rawfruits (T1)	26.12±.015	13.94±0.13	30.85±0.25	0.45±0.01	42.76±0.13	33.85±0.15	85.31±0.67
Fruits were immersed for 5 min in 20 L tanks containing CaCl₂ solution							
2% w/v (T2)	37.61±0.19	17.18±0.15	48.24±.32	0.36±0.05	63.54±0.22	51.21±0.32	88.81±0.52
3% w/v (T3)	37.23±0.24	15.50±.0.19	46.58±0.17	0.33±0.09	61.61±0.35	49.09±0.39	86.31±0.70
Fruits exposed							
CO ₂ (T4)	35.76±0.29	19.61±0.22	50.40±0.22	0.39±0.03	64.83±0.28	54.08±0.44	87.58±0.46

and untreated fruits. The lowest fiber content was found in untreated fruits (36.36%), while it was higher in treated fruits with 2% CaCl₂ (27.77%) and CO₂ (83.81 and 60.09%). The obtained results were agreed with Baltacioglu and Artik²⁶, where they stated that the ash, crude fiber, moisture, protein and total fat contents of the persimmon fruits were similar in astringent and non-astringent species.

On the other hand, pH of untreated fruits not affected with post-harvest treatment, where pH ranged between 6.06-6.11 in untreated or treated fruits. But the acidity was higher in treated fruits with 3% CaCl₂. Also, total soluble solids (TSS) of treated kaki fruit juice decreased slightly compared to untreated fruits.

Fruits color is one of the most important sensory properties that affected on the consumer preference. Postharvest pretreatment was carried out to improve the color quality of Kaki fruits by immersing fruits separately in tanks containing 2% (T2), 3% (T3) calcium chloride solution for 5 min and exposed to CO₂ (T4). The effect of previous postharvest treatment on the color parameters (L*, a* and b*) of Kaki fruits were evaluated and compared with raw fruits (T1). Table 2 showed that CaCl₂ or CO₂ treatments improved color parameters of fruits, where T2 and T3 had the highest lightness value followed with T4. Also, fruits redness (a*) maximized in T4 followed with T2 and T3. The same trend was also noticed in yellowness (b*), saturation and hue values of fruits. It could be concluded that CaCl₂ or CO₂ post-harvest treatments caused a noticeable increase in color parameters of Kaki fruits compared to untreated fruits.

Post harvest Kaki fruits were processed to produce kaki fruit leather. The amount of the obtained leather product was referred to fruits weight and expressed as a yield. Data in

Table 3: Effect of calcium chloride and CO₂ postharvest treatments on the yield of kaki leather

Samples	Fruits weight (g)	Kaki fruit leather weight (g)	Yield (%)
Untreated fruits (T1)	1400	418	29.86
Fruits were immersed for 5 min in 20 L tanks containing CaCl₂ solution			
2% (w/v) (T2)	1400	471	33.64
3% (w/v) (T3)	1400	499	35.64
Fruits exposed to CO ₂ (T4)	1400	503	35.93

Table 3 indicated that the kaki leather yield of untreated fruits was lower than kaki leather of treated fruits with 2%, 3% CaCl₂ and CO₂.

Color quality of the obtained fruits leather were evaluated and compared with leather of untreated fruit. Table 4 showed that the lowest lightness was found in leather of untreated fruits (control sample), while L* was higher in leather of post-harvest treated fruits with CO₂ (T4), followed with treated fruits with calcium chloride (T2 and T3). The same trends were found in redness and yellowness of the obtained leather, where a* and b* declined in control sample and was higher in T4, T2 and T3. Results concluded that the lowest Hunter color parameter were found in leather of untreated fruits and the highest color parameter were found in leather of post-harvest fruits with CO₂ (T4). This result could be due to the higher polyphenolic and flavonoid compounds and their antioxidant activities in treated fruits.

Bioactive compounds particularly phenolics and flavonoids are major interests in persimmon fruit. Table 5 compared these total phenolics, total flavonoids and their antioxidant activity in leather of untreated and treated kaki fruits. The obtained results showed that the total phenolics and antioxidant activity of leather affected slightly during preparing leather using untreated or treated fruits. The total phenolic compounds of Kaki leather of treated fruits with CO₂

Table 4: Effect of postharvest treatments of kaki fruits on color parameter of leather product

Leather	L*	a*	b*	a/b	ΔE	Saturation	Hue
Untreated fruits (T1)	35.38±0.15	9.88±0.06	19.14±0.14	0.52±0.07	41.42±0.40	21.54±0.14	87.00±0.70
Fruits immersed for 5 min in 20 L tanks containing CaCl₂ solution							
2% (w/v) (T2)	38.59±0.17	12.77±0.19	23.45±0.11	0.54±0.05	46.93±0.35	26.70±0.19	87.56±0.65
3% (w/v) (T3)	36.91±0.21	11.67±0.09	23.11±0.13	0.50±0.04	45.08±0.28	25.89±0.11	87.52±0.59
Fruits exposed to CO ₂ (T4)	40.13±0.25	14.18±0.12	28.31±0.16	0.50±0.02	51.12±0.22	31.66±0.17	87.98±0.50

Table 5: Total phenolics, total flavonoids and antioxidant activity (DPPH) in fruits leather of untreated and postharvest treated kaki fruits

Phytochem and activities	Kaki leather of untreated fruits	Kaki leather of fruits immersed in CaCl ₂ / 5 min		
		2% w/v	3% w/v	Kaki leather of fruits exposed to CO ₂
Total phenolics (mg g ⁻¹)	0.3594	0.3336	0.3675	0.4330
Total flavonoids (mg g ⁻¹)	ND	ND	ND	ND
DPPH (Trolox mg g ⁻¹)	0.0824	0.0905	0.0839	0.1090

ND: Not detected

Table 6: Effect of preparing kaki leather of untreated and post-harvest treated fruits on its phenolic compounds

Compound (μg g ⁻¹)	Fresh fruits				Kaki leather of fruits			
	Untreated	Immersed in CaCl ₂ /5 min		Exposed to CO ₂	Untreated	Immersed in CaCl ₂ / 5 min		Exposed to CO ₂
		2%	3%			2%	3%	
Gallic	1556.01	1565.02	2113.97	1665.29	3993.99	4244.75	4071.06	3633.55
Protocatechuic	23.94	19.88	26.66	18.78	31.04	33.71	36.63	35.61
Gentisic	4.50	6.57	5.50	6.01	14.83	19.61	9.96	14.26
Chlorogenic	1.36	0.76	1.16	1.98	2.31	3.26	3.37	3.48
Vanillic	1.90	2.50	4.39	3.63	17.37	18.65	20.52	16.19
Ferulic	0.72	0.23	7.13	0.79	ND	ND	ND	ND
Sinapic	1.47	ND	5.23	ND	9.91	2.18	4.95	9.22
Rutin	3.07	2.82	7.16	2.78	7.26	9.37	12.64	9.47
Myricetin	1.90	0.47	3.98	3.27	1.40	6.26	1.42	4.20
Cinnamic	0.21	1.08	0.10	0.30	0.54	0.65	0.77	0.62
Quercetin	0.68	0.70	0.65	0.36	0.40	1.19	1.57	0.66
Kaempferol	0.95	0.90	0.55	0.32	ND	ND	1.74	0.78

were higher than Kaki leather of untreated or treated fruits with CaCl₂. Also, antioxidant activity (DPPH) of kaki leather of treated fruits with CO₂ was slightly higher than kaki leather of untreated or treated fruits with CaCl₂. But, Novillo *et al.*²⁷ found that submitted fruit to destringency treatment with CO₂, lowered the values of both soluble polyphenols content and total antioxidant capacity. It was noticed also that total flavonoids compounds not detected in kaki leather of untreated or treated fruits.

Phenolic compounds of post-harvest treated kaki fruits were determined and evaluated to produce fruits leather. About 12 phenolic compounds were detected in fresh kaki fruits. Table 6 showed that gallic was the main phenolic compound in untreated kaki fruits followed with protocatechuic, gentisic, rutin, vanillic and myricetin. Also, gallic was maximized in treated kaki fruits with 3% CaCl₂ to reach 2113.97 μg g⁻¹ followed with treated fruits with CO₂. Also, Kaki leather of untreated or treated fruits characterized with its higher percentage of phenolic compounds. This result could be due to concentration of Kaki fruits in leather products. While, ferulic was not detected in kaki leather of untreated or treated fruits. These results agreed with

Table 7: Effect of preparing kaki leather of untreated and post-harvest treated fruits on total tannins and carotenoids

Compound (μg g ⁻¹)	Untreated	immersed in CaCl ₂ /5 min		Exposed to CO ₂
		2%	3%	
Fresh fruits				
Total tannins (μg g ⁻¹)	40.892	36.464	47.016	38.545
Total carotenoids (μg g ⁻¹)	14.591	15.385	15.076	19.400
Leather				
Total Tannins (μg g ⁻¹)	225.473	211.101	184.014	159.875
Total carotenoids (μg g ⁻¹)	107.426	73.540	112.077	104.433

Yaqub *et al.*²⁸ who stated that major phenolic compounds that present in persimmon are ferulic acid, p-coumaric acid and gallic acid.

Persimmon fruit can be used in the manufacturing of products with functional characteristics because of its bioactive properties. It is an excellent source of ascorbic acid, tannins and carotenoids, having healthy aspects owing to their antioxidant and other health protecting activities²⁹. Therefore, total tannins and carotenoids of postharvest treated kaki fruits and its leather products were evaluated. Table 7 showed that post harvest treated fruits affected on total tannins where untreated fruits was 40.892 μg g⁻¹, while

Table 8: Sensory evaluation of kaki leather

Sensory properties	Kaki leather				LSD at 0.05
	Untreated fruits	Fruits immersed in CaCl ₂ /5 min		Fruits exposed to CO ₂	
		2% w/v	3% w/v		
Taste (20)	16.29±2.82 ^b	13.81±3.77 ^c	18.11±2.20 ^a	14.88±1.13 ^c	2.55
Color (20)	18.13±1.01	18.11±1.06	18.25±0.33	18.02±0.19	NS
Odor (20)	17.17±2.17	18.35±2.38	17.14±2.26	18.24±3.15	NS
Texture (20)	18.71±0.68 ^b	17.82±1.20 ^c	19.65±1.03 ^a	17.35±0.96 ^c	1.04
Appearance (20)	19.15±0.61	18.33±1.09	18.61±1.19	17.65±0.95	ND
Overall acceptability (100)	89.45±0.85	86.42±0.96	91.76±0.80	86.14±0.75	ND

treated fruits with CaCl₂ decreased to 36.464 µg g⁻¹. This result could be due to the effect of CO₂ treatment to remove astringency based on the in solubilization of tannins by the acetaldehyde generated during anaerobic respiration, which was triggered when fruit was exposed to a high CO₂ atmosphere³⁰. Also, total carotenoids of untreated fruits (14.591 µg g⁻¹) slightly increased to be ranged between 15.076-19.400 µg g⁻¹ in CaCl₂ and CO₂. This result could be due to carotenoid contents rapidly increase as green mature fruit changes to soft mature persimmon³¹.

On the other hand, total tannins and carotenoids were concentrated in leather products, where total tannins ranged between 159.875-225.473 µg g⁻¹ in leather products of untreated and treated. Also total carotenoids of leather products were concentrated and ranged between 73.540-112.077 µg g⁻¹ in leather products of untreated and treated fruits.

Sensory properties were evaluated in leather products of Kaki fruits before and after post-harvest treatment. Table 8 showed that, the highest taste and texture were found in kaki leather of post-harvest treated fruits with 3% CaCl₂. Furthermore, leather products of post-harvest treated fruits not affected significantly in color, odor, appearance and overall acceptability. From the obtained sensory evaluation it could be recommended to use post harvest kaki fruits to produce kaki leather.

CONCLUSION

The purpose of this study was to improve the quality of kaki fruits and evaluate its leather product. The obtained results concluded that treated fruits with CaCl₂ or CO₂ improved color parameters of fresh Kaki fruits and its leather product. Also, kaki leather of treated fruits characterized with its higher total phenolics compounds and antioxidant activity compared to leather of untreated fruits. Furthermore, sensory evaluation of treated fruits or its leather product not affected significantly in color, odor, appearance and overall acceptability.

SIGNIFICANCE STATEMENT

This study discovers a new leather kaki fruits product contain high concentration of several bioactive compounds. This product is characterized by its high healthy, nutritive value, good sensory and texture properties. This also opens the way for innovative research in the same field to produce more functional food products able to alleviate symptoms of several diseases.

ACKNOWLEDGEMENT

Our special thanks to the late Dr. Mamdouh Mohamed Nageib, Pomology Professor, National Research Center (NRC), for the support and supplying us with fruits used in this work.

REFERENCES

1. Kays, S.J., 1991. Postharvest Physiology of Perishable Plant Products. Van Nostrand Reinhold, New York, USA., Page: 1532.
2. Matsuo, T. and S. Ito, 1982. A model experiment for de-astringency of persimmon fruit with high carbon dioxide treatment: *In vitro* gelation of kaki-tannin by reacting with acetaldehyde. Agric. Biol. Chem., 46: 683-689.
3. Hribal, J., M. Zavrtnik, M. Simcic and R. Vidrih, 2000. Changes during storing and astringency removal of persimmon fruit *Diospyros kaki* L. Acta Alimentaria, 29: 123-136.
4. Zhang, Y.U., C. K. Song, C.Q. Jun, Z.S. Long and R.Y. Ping, 2003. Effects of Acetylsalicylic Acid (ASA) and ethylene treatments on ripening and softening of postharvest Kiwifruit. Acta Bot. Sinica, 45: 1447-1452.
5. Arnal, L., C. Besada, P. Navarro and A. Salvador, 2008. Effect of controlled atmospheres on maintaining quality of persimmon fruit cv. "Rojo Brillante". J. Food Sci., 73: S26-S30.
6. Hafez, O.M., H.A. Hamouda and M.A. Abd-El-Mageed, 2010. Effect of calcium and some antioxidants treatments on storability of Le Conte pear fruits and its volatile components. Selcuk Tarim Bilimleri Dergisi, 24: 87-100.
7. Sultan, M., S. Ibrahim, O.M. Hafez and M.A. Saleh, 2016. Cellulose sulfate active packaging material with treatments on orange shelf life. Int. J. Pharm Tech Res., 9: 79-90.

8. El-Zaabalawy, H.T., 2007. Trials to improve marketing characteristics and prolonging storage life of persimmon and mango fruits. Ph.D. Thesis, Fruit Science, Faculty of Agriculture, Zagazig University, Egypt.
9. Luo, Z., 2006. Extending shelf-life of persimmon (*Diospyros kaki* L.) fruit by hot air treatment. Eur. Food Res. Technol., 222: 149-154.
10. Telis, V.R.N., A.L. Gabas, F.C. Menegalli and J. Telis-Romero, 2000. Water sorption thermodynamic properties applied to persimmon skin and pulp. Thermochim. Acta, 343: 49-56.
11. Carcel, J.A., J.V. Garcia-Perez, E. Riera and A. Mulet, 2007. Influence of high-intensity ultrasound on drying kinetics of persimmon. Dry. Technol., 25: 185-193.
12. Huang, X. and F.H. Hsieh, 2005. Physical properties, sensory attributes and consumer preference of pear fruit leather. J. Food Sci., 70: E177-E186.
13. Maskan, A., S. Kaya and M. Maskan, 2002. Hot air and sun drying of grape leather (pestil). J. Food Eng., 54: 81-88.
14. Gujral, H.S. and S.S. Brar, 2003. Effect of hydrocolloids on the dehydration kinetics, color and texture of mango leather. Int. J. Food Propert., 6: 269-279.
15. Demarchi, S.M., N.A.Q. Ruiz, A. Concellon and S.A. Giner, 2013. Effect of temperature on hot-air drying rate and on retention of antioxidant capacity in apple leathers. Food Bioprod. Proces., 91: 310-318.
16. Ruiz, N.A.Q., S.M. Demarchi, J.F. Massolo, L.M. Rodoni and S.A. Giner, 2012. Evaluation of quality during storage of apple leather. LWT-Food Sci. Technol., 47: 485-492.
17. Sharma, S.K., S.P. Chaudhary, V.K. Rao, V.K. Yadav and T.S. Bisht, 2013. Standardization of technology for preparation and storage of wild apricot fruit bar. J. Food Sci. Technol., 50: 784-790.
18. Kato, K., 1990. Astringency removal and ripening in persimmons treated with ethanol and ethylene. HortScience, 253: 205-207.
19. AOAC., 1995. Official Methods of Analysis, Vitamins and Other Nutrients. Method No. 967.21. In: Ascorbic Acid in Vitamin Preparation and Juices, AOAC. (Ed.), AOAC International, USA., pp: 16-17.
20. Ibarz, A., C. Gonzalez and S. Esplugas, 1994. Rheology of clarified fruit juices. III: Orange juices. J. Food Eng., 21: 485-494.
21. Sapers, G.M. and F.W. Douglas, Jr., 1987. Measurement of enzymatic browning at cut surfaces and in juice of raw apple and pear fruits. J. Food Sci., 52: 1258-1285.
22. Singleton, V.L., R. Orthofer and R.M. Lamuela-Raventos, 1999. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods Enzymol., 299: 152-178.
23. Lamaison, J.L.C. and A. Carnet, 1990. Teneurs en principaux flavonoids des fleurs de *Crataegus monogyna* Jacq et de *Crataegus laevigata* (Poiret D. C) en fonction de la vegetation. Pharm. Acta. Helv., 65: 315-320.
24. Grzegorzczuk, I., A. Matkowski and H. Wysokinska, 2007. Antioxidant activity of extracts from *in vitro* cultures of *Salvia officinalis* L. Food Chem., 104: 536-541.
25. Hussein, A.M.S. and N.A. Shedeed, 2011. Production of good quality drinks from some Egyptian berry fruits varieties. Model. Meas. Control, 72: 26-37.
26. Baltacioglu, H. and N. Artik, 2013. Study of postharvest changes in the chemical composition of persimmon by HPLC. Turk. J. Agric. For., 37: 568-574.
27. Novillo, P., C. Besada, L. Tian, A. Bermejo and A. Salvador, 2015. Nutritional composition of ten persimmon cultivars in the "ready-to-eat crisp" stage. Food Nutr. Sci., 6: 1296-1306.
28. Yaqub, S., U. Farooq, A. Shafi, K. Akram, M.A. Murtaza, T. Kausar and F. Siddique, 2016. Chemistry and functionality of bioactive compounds present in persimmon. J. Chem., Vol. 2016. 10.1155/2016/3424025
29. Karaman, S., O.S. Toker, F. Yuksel, M. Cam, A. Kayacier and M. Dogan, 2014. Physicochemical, bioactive and sensory properties of persimmon-based ice cream: Technique for order preference by similarity to ideal solution to determine optimum concentration. J. Dairy Sci., 97: 97-110.
30. Novillo, P., A. Salvador, E. Llorca, I. Hernando and C. Besada, 2014. Effect of CO₂ deastringency treatment on flesh disorders induced by mechanical damage in persimmon. Biochemical and microstructural studies. Food Chem., 145: 454-463.
31. Britton, G., 1995. Structure and properties of carotenoids in relation to function. FASEB. J., 9: 1551-1558.