



Asian Journal of Plant Sciences

ISSN 1682-3974

science
alert

ANSI*net*
an open access publisher
<http://ansinet.com>

The Effects of Epicuticular Wax Cover on the Rate of Water Loss of *Sorghum bicolor* (L.) Moench

¹Mijitaba Hamissou and ²Dale E. Weibel

¹Department of Biology, Jacksonville State University, Jacksonville, Alabama, 36285 USA

²Department of Agronomy, Oklahoma State University, Stillwater, OK 74078 USA

Abstract: Experiments were conducted to determine the role of epicuticular wax cover on the rate of water loss of sorghum plants. Relative Water Content (RWC), water potential (Ψ), transpiration rate, stomatal conductance and stomatal density were measured in 3 near-isogenic lines of sorghum variety ROKY62, bloom (*BmBm*), sparse-bloom (*hh*) and bloomless (*bmbm*) under different water conditions. Under drought conditions, bloomless sorghum exhibited more negative Ψ values than the bloom type, -1.70 Mpa as compared to -1.43 Mpa for the bloomless and bloom lines, respectively. Under normal water conditions, similar trend was observed in Ψ , where the values were -1.62 and -1.21 Mpa for the bloomless and bloom lines, respectively. Data indicated that there is a positive relationship between wax cover and the internal water content of plants. The bloomless plants had higher stomatal conductance and transpiration rates than the bloom or sparse-bloom plants. Microscopic examinations revealed higher stomatal densities on the flag leaf than on the third leaf down in all 3 isogenic lines. We conclude that wax cover on sorghum leaves reduces the transpirational water loss and prevents a rapid decrease in plant water potential.

Key words: Sorghum, epicuticular wax, transpiration, stomatal conductance

INTRODUCTION

Sorghum bicolor (L.) Moench is one of the most important sources of carbohydrate in many arid countries. It is grown in the African sub-Sahara and in the semi-arid regions of India, Pakistan and Indonesia as source of staple food. These regions are characterized by scanty rainfall and high evaporation rates, making agricultural practices challenging. Sorghum is also grown in the semi-arid portion of the Great Plains of the United States.

The vegetative body of sorghum plants is normally covered with a powdery wax. The wax cover may range from a heavy covering termed "bloom" to a light covering known as "sparse bloom." While working with the World Collection of Sorghum in Coimbatore, Ayyangar and Ponnaiya^[1] discovered that an African variety from the group *sorghum elegans* did not exhibit visible wax covering. They termed the condition "bloomless" and determined through back crossing that wax cover on sorghum leaves is controlled by a single dominant gene *BmBm*. Peterson *et al.*^[2] reported the occurrence of two independent genes *bm₁bm₁* and *bm₂bm₂* controlling bloomlessness and three independent genes *h₁h₁*, *h₂h₂*

and *h₃h₃* controlling sparse-bloom condition. The presence or absence of wax cover in sorghum is believed to play important roles in the plant's drought resistance mechanism, in plant-insect interactions^[3], plant digestibility and yield^[4,5].

Wax belongs to a class of organic substances exuded from plant cells and that may appear in combination with cutin and suberin^[6]. Wax exudates are most often deposited on the cuticular layer enveloping the epidermis at the interface of the plant and the atmospheric environments, playing a defense mechanism against desiccation and insect herbivores. Wilkinson and Cummins^[7] analyzed the composition and roles of wax on sorghum and concluded that the presence of epicuticular wax might explain the resistance of sorghum to water loss. They determined that plant surface wax prevents water loss by reducing solar energy load on the plant through increased reflectance, an avoidance of reduced water potential and maintenance of a more complete stomatal control over transpiration^[8]. The objective of this research was to investigate some physiological parameters associated with water loss on three isogenic lines of sorghum, variety ROKY62, the bloom, the sparse-bloom and the bloomless.

MATERIALS AND METHODS

Experiments were conducted at the Oklahoma State University, Stillwater, OK (USA), College of Agriculture's greenhouse facilities to measure several physiological parameters related to plant-water relation of sorghum. The experimental design consisted of 6 treatments in form of rows. Seeds of three isogenic lines of sorghum, variety ROKY62, bloom (carrying the gene *BmBm*), sparse-bloom (carrying the gene *h₁h₁*) and bloomless (carrying the gene *bmbm*) were sown in rows, 1.0 meter long separated by 0.75 m. Germinated seedlings were thinned to allow 10 cm spacing between consecutive plants within rows. The total number of plants per row was 11, but one plant at each extremity of the rows was excluded from the test. The remaining 9 plants per row were labeled and used for all data collection. The 6 treatments were replicated 3 times and randomized. The experimental rows were watered every other day and fertilized once during the experimental period. At boot stage and continuing for 2 weeks, drought stress was imposed by withholding water. Control treatments continued to receive water every other day until the experiment was terminated.

Seven days following the onset of imposed drought and continuing for 7 consecutive days, wax exudation on leaves were recorded daily on control and drought imposed plants by visual observations. The amount of surface wax deposited was ranked on all the test plants. Because of the lack of published guideline for visual observations, we set and followed the following scale: 1 = glossy leaf surface, no wax even when leaves are rubbed against; 2 = glossy but light scattering, trace of wax only after rubbing the leaves; 3 = the amount of surface covered by wax equal to the amount of surface not covered; 4 = plant body mostly covered by wax and 5 = flaky or powdery wax over the entire plant body.

Two sets of leaf discs were taken every day from each of the test plants. One set was used for water potential determination as described by Kanematsu^[6] and the second set was used for Relative Water Content (RWC) determination as described by Turner^[9]. Water potential data were statistically analyzed using the analysis of variance (ANOVA) for a Randomized Block Design^[10].

Stomatal density was determined according to modified techniques of Sinclair and Dun^[5], Witham *et al.*^[11]. The modification involved replacing the liquid plastic with finger nail polish. Both ad-axial and ab-axial surfaces of the leaves were investigated microscopically and the numbers of stomata per view field were recorded. All measurements were performed on the flag leaves and the third leaves down. The data were analyzed statistically using ANOVA for a Randomized Block Design.

Stomatal conductance and transpiration rates were determined over the 7 days on all experimental plants, using a steady state porometer Li-Cor 6000, according to Kanematsu^[6]. Measurements were performed on the flag leaves and the third leaves down. The data were statistically analyzed using an analysis of variance (ANOVA) for a Randomized Block Design.

RESULTS

Wax deposition on the plant surface was determined using our grading scale described above. Wax covering was higher in the bloom plants than in the sparse bloom plants. We observed more intense wax deposition under imposed drought conditions than under well-watered conditions (Fig. 1a). The occurrence of wax increased linearly as the number of days without water increased, in both bloom and sparse-bloom lines (Fig. 1b). Over the entire experimental period, no surface wax was detectable with our scaling method on the bloomless plants under drought or well-watered conditions.

Plants Relative Water Content (RWC) were calculated as percent of [(fresh weight-dry weight)/(turgid weight-dry weight)]. Under imposed drought, the bloom and sparse-bloom lines had higher relative water content than the bloomless line (Fig. 2b). Under well-watered conditions however, Fig. 2a, the RWC values of the 3 isogenic lines were similar, with daily averages of 91.9, 90.3 and 91.4% for the bloom, the sparse-bloom and the bloomless, respectively. As the number of days without water was prolonged, the RWC values of the bloomless line decreased at faster rates than those of the bloom and sparse-bloom lines. At day 7 of imposed drought, the RWC values were 76.0, 60.2 and 53.3% for the bloom (waxy), the sparse-bloom and the bloomless (non-waxy), respectively.

Plants water potential (Ψ) values were more negative under imposed drought than under well watered conditions (Table 1). Under drought conditions, the Ψ values of the flag leaves were -1.43 and -1.53 Mpa in the

Table 1: Averages water potential (Mpa) of ROKY 62 bloom, sparse-bloom and bloomless sorghum lines under drought and well watered conditions

Leaf	Variety	Water potential (Mpa)	
		Droughted	Undroughted
Flag leaf	ROKY62 Bloom	-1.43a	-1.21a
	Sparse bloom	-1.53ab	-1.40a
	Bloomless	-1.70b	-1.62b
Third leaf	Bloom	-1.42a	-1.20a
	Sparse bloom	-1.49a	-1.36a
	Bloomless	-1.66b	-1.57ab

Numbers, within column, followed by the same letter(s) are not statistically different at 5% probability level

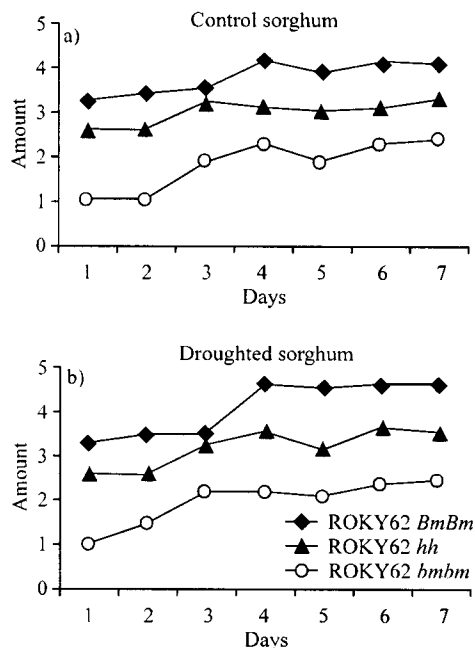


Fig. 1: Progression of wax cover on sorghum leaves over 7 days, under well watered conditions (1a) and under imposed drought (1b.) Plants were water stressed for seven days and the amount of wax deposition recorded on the scale of 1 to 5 with 1 having no visible wax and 5 being very waxy for 7 more consecutive days

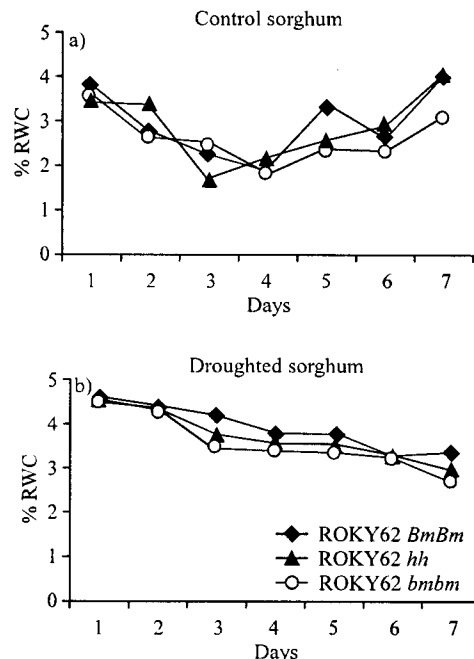


Fig. 2: Changes in relative water content of sorghum over 7 days, under well watered conditions (2a) and under imposed drought (2b.) Plants were water stressed for seven days and the change in RWC were determined over 7 days for 7 more consecutive days

Table 2: Average stomatal density (number of stomata cm^{-2}) of the flag leaf and the third leaf down of ROKY62 sorghum isogenic lines

Leaf	Variety	ad-axial Surface ^[1]	ab-axial Surface ^[2]	Ratio ^[1,2]
Flag	Bloom	6.89	20.11	1:29b
	Sparse bloom	10.11	17.81	1:18a
	Bloomless	9.10	22.97	1:25a
Third	Bloom	9.88	15.97	1:10a
	Sparse bloom	5.97	13.44	1:22a
	Bloomless	10.92	17.70	1:16a

Numbers followed by the same letter are not significant at 5% probability level

Table 3: Stomatal conductance and transpiration rates of the flag leaf and third leaf down of ROKY62

Leaf	Variety	Conductance ($\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$)		Transpiration ($\text{mmol H}_2\text{O m}^{-2} \text{ s}^{-1}$)	
		Droughted	Undroughted	Droughted	Undroughted
Flag	Bloom	0.12	0.14	4.13	1.62
	Sparse bloom	0.13	0.14	4.72	1.64
	Bloomless	0.15	0.16	5.05	4.51
Third	Bloom	0.13	0.15	4.10	2.08
	Sparse bloom	0.15	0.16	4.60	2.18
	Bloomless	0.16	0.18	5.20	2.19

Each number is a mean of 81 individual measurements

bloom and sparse-bloom, respectively, as compared to -1.21 and -1.40 Mpa under well-watered conditions. The bloomless line had Ψ values of -1.70 Mpa under drought conditions as compared to -1.62 under well-watered conditions. Similar trends were observed with the third leaves down under drought conditions with Ψ values of -1.42 and -1.49 Mpa as compared to -1.20 and -1.36 Mpa for the bloom and the sparse-bloom lines, respectively. For the bloomless line, the values were -1.66 and -1.57 Mpa under droughted and well-watered conditions, respectively. Statistical analyses at 5% probability level indicated a significant difference exists between the water potential of the bloom and the bloomless line on both flag leaf and the third leaf down.

Stomatal densities of the flag leaf and the third leaf down of all test plants were determined microscopically and the stomatal ratios of ad-axial to ab-axial surfaces were calculated and presented in Table 2. Higher stomatal densities were observed on the lower surface of sorghum leaf than on the upper surface, with an average of ratio of 2.2 to 1. Statistical analyses were performed on the ratio

between ad-axial and ab-axial stomatal density to help understand the phenomenon of leaf rolling. The flag leaf ratio of the bloom lines statistically surpassed the sparse-bloom and the bloomless lines, explaining the physiological importance of the flag leaf to the emergence of the panicle, seed setting and maturity. Most of the stomatal aperture were closed under drought conditions but remained opened under normal water conditions.

Two other parameters of photosynthetic activities, the stomatal conductance and transpiration rates, were determined using a steady state porometer, Li-Cor 6000. Measurements were performed on the flag leaf and third leaf down. The transpiration rates of the flag leaves were higher than those of the third leaves down under drought conditions (Table 3). In the bloom and sparse-bloom lines, respectively, flag leaf photosynthetic values of 4.13 and 4.72 mmol H₂O m⁻² s⁻¹ were observed as compared to 4.10 and 4.60 mmol H₂O m⁻² s⁻¹ for their third leaves down. The stomatal conductance values did not exhibit a conclusive trend, but values of the third leaves were higher than those of the flag leaves.

DISCUSSION

Plant surface wax is an anatomical feature under the control of a family of genes among which the *BmBm*, *bmbm*, *HH*, *h₁h₁*, *h₂h₂* and *h₃h₃*. The phenotypic expressions of these genes appear to be enhanced by plant age as well as the plant water status as supported by this research. Wax deposition on sorghum leaf surface was recorded over the growing cycle of the plant. Present data indicated that wax cover reaches its maximum 25 to 30 days after germination, corresponding to boot stage or panicle emergence, when water balance is crucial for setting healthy and viable progeny seeds. The amount of wax cover not only increased with plant age but also with increasing water stress. The leaf rolling observed in sorghum plants was longitudinal to the main axis of the leaves, so as to expose the ab-axial surfaces high in wax cover but also high in stomatal density. It was demonstrated that wax plays an important role in preserving water status by reducing the solar energy load on the plant surface through reflectance and reduction the solar energy load on the leaf surface^[12]. When water becomes a limiting factor for growth and reproduction, leaf rolling and exposing the high stomatal density surface but high in wax cover to reflect solar radiation appears to be one physiological strategy used by plants to maintain a steady water status. This is of course a disadvantage for the bloomless sorghum lacking the epicuticular wax cover.

Present data agreed with published data of Wilkinson and Cummins^[7], who observed a relationship between the

presence of wax on the plant surface and water loss in sorghum. Under limiting water conditions, higher RWC and less negative Ψ values were recorded in the bloom and sparse-bloom sorghum than in the bloomless. This is an indication that preserving plant internal water to avoid a more reduced water potential is among one of the effects of the epicuticular wax deposit on plant body. Under well-watered conditions, there was no difference in RWC of the isogenic lines. The Ψ values were less negative when plants have some amount of wax cover on their leaf surface, indicating that wax cover plays an important role in preventing water loss.

Transpiration rate was measured as the amount of water vapor exiting the stomatal openings/leaf area/second. The bloomless line had higher transpiration rate than the bloom line or the sparse-bloom line, indicating that the lack of epicuticular wax may favor desiccation under severe water stress. This is in agreement with published research by Jordan *et al.*^[13] who found that plants with lower amount of wax had a higher transpiration rates but were more digestible. It is evident that plants having the dominant alleles for surface wax can be important breeding materials for the improvement of food supply in many parts of the world.

Sorghum plants possess genetic and morphological mechanisms of adaptation to low rainfall environments. These adaptation mechanisms play important roles for the third world inhabitants who rely heavily on sorghum for food. The role of epicuticular wax in preventing water loss was investigated under greenhouse conditions. Based on this research, we conclude that wax cover on sorghum leaves reduces the transpirational water loss by preventing rapid decrease water potential and relative water content during periods of water stress. We found the existence of an inverse relationship between the amount of plant epicuticular wax and its rate of water loss, which led us to conclude that wax cover on the sorghum leaf surface is a better trade than the stomatal density in controlling water loss.

ACKNOWLEDGMENTS

Special thanks to Dr. Roland Dute, Department of Biology, Auburn University, Auburn, Alabama, USA and Dr. James Rayburn, Department of Biology, Jacksonville State University, Jacksonville, Alabama, USA for reviewing this manuscript.

REFERENCES

1. Ayyangar, G.N.R. and B.W.X. Ponnaiya, 1941. The occurrence and inheritance of a bloomless sorghum. *Current Sci.*, 10:408-409.

2. Peterson, G.C., K. Suksayretrup and D.E. Weibel, 1982. Inheritance of some bloomless and sparse bloom mutants in sorghum. *Crop Sci.*, 22: 63-67.
3. Ross, W.M. 1972. Effect of bloomless (*bmbm*) on yield in Combine Kafir-60. *Sorghum Newslett.*, 15: 121.
4. Cummins, D.G. and J.W.Jr. Dobson, 1972. Digestibility of bloom and bloomless sorghum leaves determined by a modified *in vitro* technique. *Agron. J.*, 64: 682-683.
5. Sinclair, C.B. and D.B. Dunn, 1961. Surface printing of plant leaves for phylogenic studies. *Stain Technol.*, 3: 299-304.
6. Kanemasu, T.E., 1975. Measurement of stomatal aperture and diffusive resistance. College of Agri. Res. Center., Washington State University Bull., 809: 1-2.
7. Wilkinson, R.E. and D.G. Cummins, 1981. Epicuticular fatty acids, fatty alcohol and alkane content of sorghum 'Redbine 60' leaves. *Crop Sci.*, 21: 397-400.
8. Esau, K., 1977. *Anatomy of Seed Plants*. 2nd Ed. John Wiley and Son Pub. Inc. New York
9. Turner, C.C., 1981. Techniques and Experimental Approaches for the Measurement of Plant Water Status. *Plant and Soil*, 58: 339-366.
10. SAS., 1989. SAS Institute, Inc. SAS User's Guide: Statistics, 1989 Edn. SAS.
11. Withman, F.M., D.F. Blaydes and R.M. Devlin, 1971. Stomates. In: *Experiments in Plant Physiology*. Van Nostrand Reinhold Co., pp: 103-107.
12. Blum, A., 1975. Effect of the *Bm* gene on epicuticular wax and water relations of *Sorghum bicolor*. (L.) Moench. *Isr. J. Bot.*, 24:50-51.
13. Jordan, W.R., P.J. Shouse, A. Blum, F.R. Miller and R.L. Monk, 1984. Environmental physiology of sorghum. II. Epicuticular wax load and cuticular transpiration. *Crop Sci.*, 24: 1168-1173.