

Asian Journal of Plant Sciences

ISSN 1682-3974





Effect of Gibberellic Acid Spray on Growth, Nutrient Uptake and Yield Attributes During Various Growth Stages of Black Cumin (*Nigella sativa* L.)

S.H. Shah, I. Ahmad and Samiullah Department of Botany, Aligarh Muslim University, Aligarh 202 002, India

Abstract: Field trials were conducted on black cumin (*Nigella sativa* L.), sprayed with either deionized water (control) or 10^{-5} M GA₃ at 40 (vegetative stage) or 60 (flowering stage) Days after Sowing (DAS) to characterize the effects of hormone treatment on the mentioned parameters and select the suitable growth stage for spray in order to achieve desired results. Shoot length/plant, leaf number/plant, leaf area/plant, branch number/plant, dry weight/plant and accumulations of N, P and K were assayed at 70 DAS. Capsule number/plant, seeds capsule⁻¹, 1000 seed weight, seed yield q ha⁻¹, harvest index and Seed Yield Merit (SYM) were analyzed at harvest (130 DAS). It was noted that growth, NPK accumulation and seed yield were maximal when spraying of GA₃ was carried out at 40 DAS. However, spraying at 60 DAS was not much effective in terms of the parameters studied. Moreover, there was a significant difference in spray treatments at various growth stages only when GA₃ was sprayed and not when water was sprayed.

Key words: Dry matter production, gibberellic acid, *Nigella sativa*, NPK uptake, yield

INTRODUCTION

Nigella sativa L. is an important herb, used extensively in medicinal as well as in food formulations (Babayan et al., 1978; Takruri and Dameh, 1998). The extract of its seeds shows anticancerous activity and has been found much effective against Dalton's lymphoma ascites cells (Salomi et al., 1989). Moreover, the black seed oil has also been reported to increase the flow of bile experimentally and is of great use in paralysis, particularly facial paralysis, back pain and rheumatism (Riaz et al., 1996; Saeed et al., 1996). However, the supply of this medicinal plant through collection from forests, is not keeping pace with its continuous increasing demand in the domestic as well as international markets. As such there is an excellent scope for the promotion and extension of its cultivation to a commercial level. In this context, the use of growth regulators, particularly Gibberellic acid (GA₃) is thought to be trendsetting, as it addresses the improper source-sink relationship and internal hormonal imbalance occurring during growth. GA₃ is an essential growth hormone known to be actively involved in various physiological activities such as growth, flowering and ion transport (Wareing and Phillips, 1981; Takei et al., 2002; Khan and Samiullah, 2003), besides stimulating leaf area expansion (Brock, 1993) and inducing elongation and osmoregulation in internodes (Azuma et al., 1997), in addition to increasing dry matter and biomass production (Bhaskar et al., 1997; Gupta and Dutta, 2001).

Given the potential beneficence of these effects, along with the fact that hormone requirement, relative concentrations and responses are different at different growth stages (Robert, 1999) therefore, the present study was aimed at characterizing the effect of GA₃ on *Nigella sativa* L., sprayed during various growth phases.

MATERIALS AND METHODS

Field experiments were carried out during 1999-2000 and 2000-2001 at the Agricultural Farm, Aligarh Muslim University, Aligarh, India (27° 53′ N, 78° 54′ E and 187.45 m altitude) to select the most appropriate growth stage of N. sativa for GA3 spray for augmenting productivity. The spraying of $10^{-5}\,\mathrm{M}\;\mathrm{GA_3}$ was performed at 40 (vegetative stage) and 60 (flowering stage) Days after Sowing (DAS) of crop growth. GA₃ (10⁻⁵ M) was sprayed at the rate of 600 L ha⁻¹ and the control set was applied with deionized water. The soil of the experimental plots was sandy loam and available N, P and K was 195.6, 21.7 and 229.6 kg ha⁻¹, respectively. Seeds were procured from the Regional Research Institute of Unani Medicine, Aligarh, India. They were surface sterilized with 0.01% mercuric chloride solution, followed repeated washings with double distilled water. The seeds were then sown in 24 m² plots and at the seedling stage at a distance of 30 cm between rows and 15 cm between plants in a row. Each treatment was replicated thrice.

Shoot length/plant, leaf number/plant, leaf area/plant, dry weight/plant and N, P, K uptake in plants were

recorded at 80 DAS. Leaf area was calculated gravimetrically. The leaf area of 10% of total leaves in each treatment was determined by outlining on graph paper and the dry weight of these leaves was recorded. The leaf area/plant was computed using the leaf dry weight/plant and dry weight of those leaves for which the area was estimated (Watson, 1958). Dry weight/plant was recorded by drying the plants at 80°C for 24 h. The concentration of N and P in the plant was estimated, adopting the methods of Lindner (1944) and Fiske and Subba Row (1925), respectively. Content of K was analysed by flame photometer. The uptake of N, P and K were the product of their concentration in the plant and dry weight of plant. At maturity (130 DAS), capsules/plant, seeds/capsule, 1000 seed weight, seed yield ha-1 and Harvest Index (HI) were determined. Twenty plants from each plot were removed and the capsule number plant⁻¹ was recorded. Random samples were taken from threshed seeds for determination of the 1000 seed weight. The seed yield from three randomly selected plants was noted after threshing the seeds. Harvest index was determined by dividing the seed yield by the biological yield. Moreover, SYM was obtained by using the method given by Imsande (1992). Analysis of variance was carried out on the data obtained and LSD (p = 0.05) was calculated (Gomez and Gomez, 1984).

RESULTS AND DISCUSSION

The effect of GA₃ administration on growth, nutrient uptake and yield was found to be significantly promotive, except for seeds/capsule, 1000 seed weight and HI (Table 1-3). It was also found that spraying of GA₃ at 40 DAS proved more stimulatory for the parameters studied in comparison to the 60 DAS spray. In context of the 40 DAS spray, promotion of growth in the crop may probably be because the spray was available at a stage when the plant was in the phase of maximum cell division

and expansion at apical, lateral and intercalary regions through meristems and hence it enhanced the key constituents of the building blocks (organs) of the infrastructure of the plant maximally. At the time of division, these decisive cells need more nutrients, which are made available by the efficient manipulation, absorption and utilization of the available nutrients, triggered by the GA₃ spray (Shah, 2004). This ultimately may be the cause for more plant height, leaf number, leaf area and branch number (Table 1). Moreover, the GA₃ induced wall extensibility (Huttly and Phillips, 1995) leading to cell expansion may be assigned as another possible reason for the elongation of internodes (increase in plant height) (Moore, 1989; Taiz and Zeiger, 1998) and expansion of leaf area (Sarma, 1985; Khan, 1996; Khan et al., 1998), which inturn manifests itself in the form of more dry matter as observed (Table 1). This stance is further supported by the strong positive correlation between maximum leaf area and dry matter (r = 0.925**). On the contrary, spraying at 60 DAS did not show such marked effects due to the inappropriate phase of stimulation, as by then the vegetative phase was already over.

Similarly, there was observed an increase in the nutrient uptake (Table 2) of the test plants due to the GA₃ spray, more prominent in the 40 DAS stage than in the 60 DAS stage. It is known that the application of GA₃ invokes the inherent genetic potential of the crop for maximum performance (Moore, 1989; Taiz and Zeiger, 1998). Also, as aforesaid, the spray of GA₃ during the active growth phase of the crop triggers judicious utilization of resources and results in a better source-sink relationship. Hence, the cause of increased nutrient uptake may be the increased assimilation of all nutrients effectively during the vegetative phase, under maximal uptake potential. Similar results have been obtained for GA₃ application on *N. sativa* and other crops by Stopkinska (1986), Khan *et al.* (1998), Mousa *et al.* (2001)

Table 1: Effect of 10⁻⁵ M GA₃ spray at different growth stages of black cumin (*Nigella sativa* L.) on shoot length/plant, leaf number/plant, leaf area/plant, dry weight/plant and branch number/plant at 70 DAS (average of 1999-2000 and 2000-2001)

weight plant and branch number/plant at 70 DAS (average of 1999-2000 and 2000-2001)								
Spray treatment	Spray stages (days after sowing)	Shoot length (cm)	Leaf No.	Leaf area (cm²)	Drey weight (g)	Branch No.		
Water	40 (vegetative)	46.51	29.10	295.10	1.78	7.55		
	60 (flowering)	41.12	25.01	285.23	1.30	6.64		
GA_3	40 (vegetative)	78.51	42.15	430.16	2.85	10.90		
	60 (flowering)	61.41	35.94	370.25	2.24	9.05		
LSD at 5%		12.05	5.120	22.160	0.16	1.61		

Table 2: Effect of 10⁻⁵ M GA₃ spray at different growth stages of black cumin (Nigella sativa L.) on N, P and K uptake at 70 DAS (average of 1999-2000 and 2000-2001)

Spray treatment	Spray stages (days after sowing)	N uptake (mg/plant)	P uptake (mg/plant)	K uptake (mg/plant)
Water	40 (vegetative)	34.45	3.73	38.73
	60 (flowering)	33.30	3.41	34.42
GA_3	40 (vegetative)	60.41	6.01	64.50
	60 (flowering)	48.18	4.37	47.50
LSD at 5%		10.12	0.63	12.12

Table 3: Effect of 10⁻⁵ M GA₃ spray at different growth stages of black cumin (*Nigella sativa* L.) on capsule number/plant, seed number/capsule, 1000 seed weight, seed yield q ha⁻¹, harvest index and seed yield merit (SYM) at harvest 130 DAS (average of 1999-2000 and 2000-2001)

Spray	Spray stages	Capsule	0 1/ 1	1000 seed	seed yield	Harvest	07774
treatment	(days after sowing)	number/plant	Seeds/capsule	weight (g)	(q ha ⁻¹)	Indes (%)	SYM
Water	40 (vegetative)	16.50	52.01	2.50	8.31	30.21	251.04
	60 (flowering)	15.26	52.17	2.45	8.07	30.01	242.18
GA_3	40 (vegetative)	23.75	55.21	2.55	11.12	31.36	348.72
	60 (flowering)	19.60	54.91	2.50	9.92	31.45	311.98
LSD at 5%		3.210	NS^1	NS^1	1.03	NS^1	14.210

¹Not significant

and Hashmi (2003). Further the nutrient uptake did not show marked increase during the 60 DAS stage, which may have been because, as the plant ages, there is reduced influx of nutrient by the old and suberized roots (Barbar, 1980). Moreover, the translocation of nutrient to sinks (seeds) during their formation and subsequent development of fruit can also be reasonably considered to be responsible for nutrient depletion at later stages of growth.

With regard to various yield attributes taken under study, the treatment with GA3 proved more effective at the 40 DAS stage in increasing the number of capsules/plant, seed yield and seed yield merit, than at the 60 DAS stage. However, either treatment did not show much effect on HI, number of seeds/capsule or the 1000 seed weight, which may be attributed to the dilution with growth effect, arising due to the voluminous sink (Table 3). As per Thorne (1966), vegetative growth before flowering has considerable influence on the yield of the crop. Synchronously, in our research, spraying at a phase that coincides with intense vegetative growth resulted in higher demand for absorption of nutrients and water to bring the best out of the crop, in the form of enhanced number of capsules and subsequently, seed yield and seed yield merit. In other words, the ample availability of nutrients, efficient re-directed mobilization of metabolites and the modification of the rate or duration of dry mass accumulation at the critical growing phase induced by the hormone treatment (Davies, 1995; Naidu and Swamy, 1995), coupled with the delayed senescence and prevented abortion of plant organs under GA₃ administration (Noden and Leopold, 1978), Saks and Vanstaden (1992) can be said to be responsible for the observed increase in yield of the crop. Such findings gain support from those of Saran and Mehta (1985), Khan et al. (1998) and Mousa et al. (2001).

Simultaneously, it is assumed that seed yield and harvest index frequently do not provide satisfactory measures of plant yield efficiency, because a large plant may have high yield and low harvest index and a small plant may have vice-versa. Seed yield merit which combines the merits of these two traits (Imsande, 1992), was also found to be influenced by the GA₃ spray in our study. It would hence be safe to conclude that spray of

 GA_3 at the vegetative phase (40 DAS) holds promising potential for the productivity and overall performance of N. sativa L.

ACKNOWLEDGMENT

The authors are sincerely grateful to Dr. H. Hussain for reading the manuscript and useful comments.

REFERENCES

Azuma, T., S. Ueno, N. Uchida and T. Yasuda, 1997. Gibberellin induced elongation and osmoregulation in internodes of floating rice. Physiol. Plant., 99: 517-522.

Babayan, V.K., D. Koottungal and G.A. Halaby, 1978. Proximate analysis, fatty acid and amino acid composition of *N. sativa* L. seeds. J. Food Sci., 43: 1314-1315.

Barbar, S.A., 1980. Soil-plant interactions in the phosphorus nutrition of plants. In: The Role of Phosphorus in Agriculture. (Eds. Khasawneh, F.E., E.C. Sample and E.J. Kamprath). ASA-CSSA-SSSA, Madison, WI., USA., pp. 591-615.

Bhaskar, S., K. Vasontha and H.C. Srivastava, 1997. Influence of growth regulators on production of herbage and oil in patchouli (*Pogostemon patchouli*). Ind. Perfurm., 41: 98-101.

Brock, T.G., 1993. Combined effects of hormones and light during growth promotion in primary leaves of *Phaseolus vulgaris*. Can. J. Bot., 71: 501-50.

Davies, P.J., 1995. The plant hormones: their nature, occurrence and functions. In Plant Hormones. (Ed. Davies, P.T.) (Kluwer Academic Publishers: Dordrecht), pp. 1-12.

Fiske, C.H. and R.Y. Subba, 1925. The colorimetric determination of phosphorus. J. Biol. Chem., 66: 375-400.

Gomez, K.A. and A.A. Gomez, 1984. Statistical Procedure for Agricultural Research. J. Wiley and Sons, New York.

Gupta, V.N. and S.K. Datta, 2001. Influence of gibberellic acid on growth and flowering in chrysanthemum (*Chrysanthemum morifolium* Ramat) cv. Jayanti. Ind. J. Plant Physiol., 6: 420-422.

- Hashmi, R.S., 2003. Studies on Physiomorphological Response of *Mentha arvensis* L. to nitrogen, phosphorus, gibberellic acid and kinetin application. Ph.D Thesis. Aligarh Muslim University, Aligarh, India
- Huttly, A.K. and A.L. Phillips, 1995. Gibberellin regulated plant genes. Physiol. Plant., 95: 310-317.
- Imsande, J., 1992. Agronomic characteristics that identify high yield, high protein content in soybean genotypes. Agron. J., 84: 409-414.
- Khan, N.A., 1996. Effect of gibberellic acid on carbonic anhydrase, photosynthesis, growth and yield of mustard. Biol. Plant., 38: 145-147.
- Khan, N.A., H.R. Ansari and Samiullah, 1998. Effect of gibberellic acid spray during ontogeny of mustard on growth, nutrient uptake and yield characteristics. J. Agron. Crop Sci., 181: 61-63.
- Khan, N.A. and Samiullah, 2003. Comparative effect of modes of gibberellic acid application on photosynthetic rate, biomass distribution and productivity of rape seed mustard. Physiol. Mol. Biol. Plants, 9: 141-145.
- Lindner, R.C., 1944. Rapid analytical methods for some of the more common inorganic constituents of plant tissues. Plant Physiol., 19: 70-89.
- Moore, T.C., 1989. Biochemistry and Physiology of Plant Hormones. Springer Verlag, New York.
- Mousa, G.T., I.H. El-Sallami and E.F. Ali, 2001. Response of *Nigella sativa* L. to foliar application of gibberellic acid, benzyladenine, iron and zinc. Assiut J. Agric. Sci., 32: 141-156.
- Naidu, C.V. and P.M. Swamy, 1995. Effect of gibberellic acid on growth biomass production and associated physiological parameters in some selected tree species. Ind. J. Plant Physiol., 38: 15-17.
- Noden, L.D. and A.C. Leopold, 1978. Phytohormones and the Endogenous Regulation of Senescence and Abscission. In: Phytohormones and Related Compounds: A Comprehensive Treatise. Phytohormones and the Development of Higher Plants, Eds., Letham, D.S., P.B. Goodwin and T.J.V. Higgins, Elsevier, Amsterdam, 2: 329-370.
- Riaz, M., M. Syed and F.M. Chaudhary, 1996. Chemistry of the medicinal plants of the genus Nigella. Hamdard Med., 39: 40-45.
- Robert, E.C., 1999. Introduction: Nature, Occurrence and Functioning of Plant Hormones. Biochemistry and Molecular Biology of Plant Hormones, Hooykaas, M.A. Hall and K.R. Libbenga (Eds.), pp. 3-22.

- Saeed, A., M.A. Rizvi and L. Ahmad, 1996. Cultivation of medicinal herbs at Madinat Al-Hikmah. Hamdard Med., 39: 23-26.
- Saks, Y. and J. Vanstaden, 1992. The role of gibberellic acid in the senescence of carnation flowers. J. Plant Physiol., 139: 484-488.
- Salomi, M.J., K.R. Panikkar, M. Kesavan, S.R. Donanana and K. Rajgopalan, 1989. Anticancer activity of *Nigella sativa* L. Ancient Sci. Life, 8: 262-266.
- Saran, B. and A.S. Mehta, 1985. Effects of growth regulators on the growth and yield of mustard. Geobios, 12: 24-25.
- Sarma, C.M., 1985. Interactions of gibberellic acid with kinetin and (2-chloroethyl) trimethyl ammonium chloride on the expansion growth of bean leaf disks. Proc. Nat. Acad. Sci. India, 55: 74-78.
- Shah, S.H., 2004. Morphophysiological response of black cumin (*Nigella sativa* L.) to nitrogen, gibberellic acid and kinetin application. Ph.D Thesis, Aligarh Muslim University, Aligarh, India.
- Stopkinska, J.D., 1986. Studies on interaction of growth regulators with potassium ions in some physiological process in bean (*Phaseolus vulgaris* L.). III. The effect of growth regulators on growth, chlorophyll levels and transpiration depending on potassium concentration. Acta Soc. Bot. Pol., 55: 577-588.
- Taiz, L. and E. Zeiger, 1998. Plant Physiology. 2nd Edn. Sinauer Associates Inc., Publishers, Sunderland, Massachusetts, USA.
- Takei, K., T. Takahashi, T. Sugiyama, T. Yamaya and H. Sokakibara, 2002. Multiple roots communicating nitrogen availability from roots to shoots: A signal transduction pathway mediated by cytokinin. J. Exp. Bot., 53: 971-977.
- Takruri, H.R.H. and M.A.F. Dameh, 1998. Study of the nutritional value of black cumin seeds (*N. sativa L.*) J. Sci. Food Agric., 76: 404-410.
- Thorne, G.N., 1966. Physiological Aspects of Grain Yield in Cereals. In: The Growth of Cereals and Grasses. (Eds. Milthorpe, F.L. and J.D. Ivins), pp. 88-103.
- Wareing, P.F. and I.D.J. Phillips, 1981. The control of growth and differentiation in plants. Pergamon Press, New York.
- Watson, D.I., 1958. The dependence of net assimilation rate on leaf area index. Ann. Bot., 22: 37-54.