

Asian Journal of Plant Sciences

ISSN 1682-3974





Calcium Oxalate Crystals in Vegetative and Floral Organs of *Galanthus* sp. (Amaryllidaceae)

Nuran Ekici and Feruzan Dane Department of Biology, Faculty of Science and Arts, Trakya University, Edirne, Turkey

Abstract: In this study Ca-oxalate crystals in vegetative and floral organs of *Galanthus plicatus* L., *Galanthus gracilis* L., *Galanthus elwesii* L. were investigated. The crystals were detected in all vegetative organs but they were only detected in tepal and ovary of floral organs. They were not observed in mature anthers and old adventative roots. Calcium oxalate crystals were of the raphide type.

Key words: Galanthus, amaryllidaceae, Ca-oxalate, generative, vegetative

INTRODUCTION

Ca-oxalate crystals are inclusions (biominerals) that are often seen in higher plants (Esau, 1976; Franceschi and Horner, 1980) and in fungi (Arnott, 1995). The cells in a plant tissue that produce the crystals are generally referred to as crystal idioblasts (Foster, 1956; Arnott, 1982; Monje and Baran, 2002). Calcium oxalate occurs in two hydration states in plants, as the monohydrate (whewellite) or as the dihydrate (weddellite; Frey-Wyssling, 1981; Arnott, 1982). A number of crystal habits have been found for both hydration states: raphids, prisms, styloids, druses and crystal sand (Franceschi and Horner, 1980; Arnott, 1982). Both the chemical nature and the morphology of these crystals, as well as their localization within the plant body, could be specific for a given species (Monje and Baran, 2002). So they are used in plant taxonomy (Lersten, 1974). The Ca-oxalate crystals in the epiderma, mesophyll, cortex, phloem, xylem parenchyma and pericarp of various plants were observed by plant anatomists (Esau, 1976). Presence of the crystals in various tissues of laticifers (Dehgan and Craig, 1978), corm (Sunell and Healy, 1979) and seeds (Ilarslan et al., 1997) is known. In recent years it is also notable that these crystals were present in transitory floral organs such as stamens, gynoecia and petals (Meric and Dane, 2004). They are quite prevalent in floral organs of many taxa including Dilleniaceae, Liliaceae, Palmae, Malvaceae, Cunoniaceae, Euphorbiaceae (Tilton and Horner, 1980), Solanaceae (Horner and Wagner, 1992), Leguminosae (Buss and Lersten, 1972). The roles of Ca-oxalate crystals in plant growth and development remained unclear

(Prychid and Rudall, 1999). Recently, detailed description of the calcium oxalate raphide crystal content of Leucojum aestivum L. in vegetative and generative tissues in different phases of its life cycle were studied (Ekici and Dane, 2007). In this study Ca-oxalate crystals of some Galanthus species which is sister genus with Leucojum were researched. The aim of this study is to contribute to the solutions of the problems about Ca-oxalate crystals and taxonomical problems of Amaryllidaceae.

MATERIALS AND METHODS

Studies of Ca-oxalate crystal types were made on plants from herbarium materials of Galanthus plicatus L. (EDTU 2055), Galanthus gracilis L. (EDTU 4723), Galanthus elwesii L. (EDTU 3101). Vouchers were kept in the herbarium of Biology Department of Trakya University (EDTU). Clearing-squash technique which developed by Herr (1971) was used to observe Ca-oxalate crystals in different tissues of Galanthus sp. Dry herbarium materials of Galanthus sp. were hydrated in warm water and then they were first pretreated in lactic acid (85%) for 24 h at room temperature. After this pretreatment materials were transferred directly to a recently developed clearing fluid composed of lactic acid (85%), chloral hydrate, phenol, clove oil and xylene (2:2:2:1, by weight) for 48 h at room temperature. Then they were taken on slides with a small amount of clearing fluid by means of a Pasteur pipette for microscopic examination (Herr, 1971). Some of the slides were coloured with 2% aceto-orcein. All preparations were examined under Olympus BH-2 microscope. Photos were taken with Olympus C-5060 digital camera.

RESULTS AND DISCUSSION

In this study Ca-oxalate crystals in vegetative and floral organs of Galanthus plicatus L., Galanthus gracilis L., Galanthus elwesii L. were investigated. Crystals were observed in three species of Galanthus in vegetative organs as young adventative roots, bulb membrane (Fig. 1a-c), flower stalk (Fig. 2a-c) and leaf cells (Fig. 3a-c). They were

only observed in mature anthers and ovary (Fig. 4a-c) and tepal cells (Fig. 5a-c) of floral organs. Crystal content was dense in bulb membrane, flower stalks, leaves, tepals, ovary and young adventative roots, respectively. Calcium oxalate crystals were of the raphide type.

Physical and chemical conditions such as temperature, pressure, pH and ion concentration, may affect crystal growth, location and properties (Franceschi

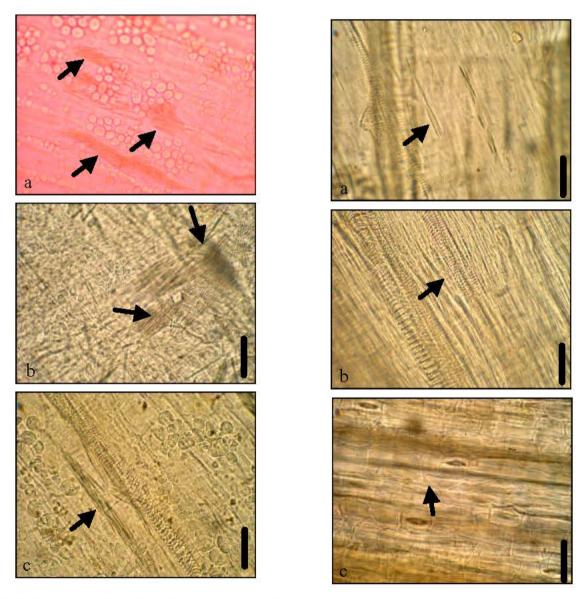


Fig. 1: Ca-Oxalate crystals in the bulb membrane cells of $\it Galanthus\,$ sp. (a) $\it G.\,$ elwesii (Coloured with 2% aceto-orcein); (b) $\it G.\,$ gracilis and (c) $\it G.\,$ plicatus. Scale bars = 40 μm

Fig. 2: Ca-oxalate crystals in the flower stalk cells of Galanthus sp. (a) G. elwesii; (b) G. gracilis and
(c) G. plicatus. Scale bars = 40 μm

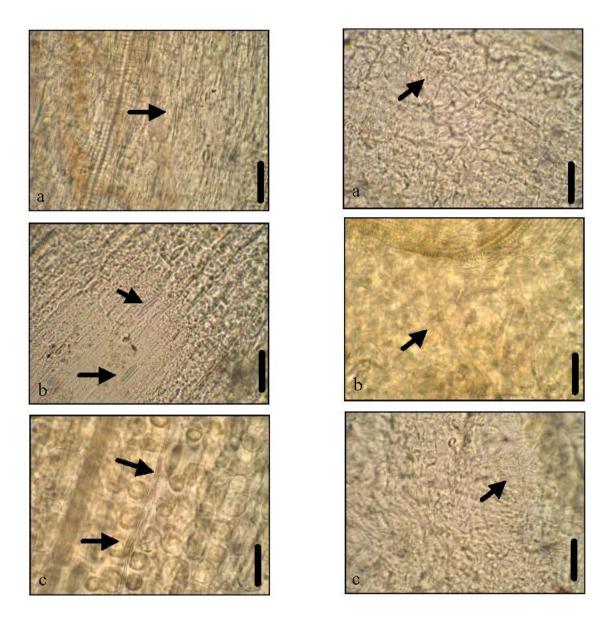


Fig. 3: Ca-Oxalate crystals in the leaf cells of Galanthus sp. (a) G. elwesii; (b) G. gracilis and (c) G. plicatus. Scale bars = 40 μm

Fig. 4: Ca-Oxalate crystals in the ovary cells of Galanthus sp. (a), G. elwesii (b) G. gracilis and (c) G. plicatus. Scale bars = 40 µm

and Horner, 1980), however it is considered that crystal formation within the cell is under genetic control (Ilarslan et al., 2001).

Although some species of monocotyledones have different crystal types in adjacent cells, a particular taxon can have specific crystal shape (Prychid and Rudall, 1999). According to Watson and Dallwitz (2005) mezophyll cells of leaves of Amaryllidaceae plants contain raphide crystals. In this study, raphide crystals were

observed in all vegetative and generative tissues except anthers and old adventative roots of Galanthus sp. at their flowering phases like L. aestivum which sister genus with Galanthus (Ekici and Dane, 2007). Their existence in meristematic cells has been demonstrated in several studies (Dane et al., 2000; Ekici and Dane, 2007; Horner and Wagner, 1980; Kausch and Horner, 1983; Seago and Marsh, 1989) as they were seen in young adventative roots of Galanthus sp. These crystals were also seen in



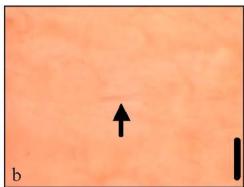




Fig. 5: Ca-oxalate crystals in the tepal cells of Galanthus sp. (a) G. elwesii; (b) G. gracilis (Coloured with 2% aceto-orcein) and (c) G. plicatus. Scale bars = 20 μm

ovary wall of *Triteleia* (Themidaceae) (Berg, 2003) which is related taxon to Amaryllidaceae and *Galanthus nivalis* (Amaryllidaceae) (Chudzik and Sniezko, 2003), *L. aestivum* (Amaryllidaceae) (Ekici and Dane, 2007) like *G. plicatus*, *G. gracilis* and *G. elwesii*.

In conclusion, this study was determined CaOx crystals in G. plicatus, G. gracilis and G. elwesii, being members of Amaryllidaceae in floral stages of its

development. Although crystals were seen in connective tissue of anthers of L. aestivum in pollen mitosis phase they were not seen in mature anthers of Galanthus species and L. aestivum. In further studies we will focus on crystal content of these taxa during their life cycles.

REFERENCES

Arnott, H.J., 1982. Three Systems of Biomineralization in Plants with Comments on the Associated Organic Matrix. In: Nancollas, G.H. (Ed.) Biological Mineralization and Demineralization, Springer Verlag, Berlin, pp. 199-218.

Arnott, H.J., 1995. Calcium Oxalate in Fungi, in Calcium Oxalate in Biological Systems. Khan, S. (Ed.), CRC Press, Boca Raton.

Berg, R.Y., 2003. Development of ovule, embryo sac and endosperm in *Triteleia* (Themidaceae) relative to taxonomy. Am. J. Bot., 90: 937-948.

Buss, P.A. and N.R. Lersten, 1972. Crystals in tapetal cells of the Leguminosae. Bot. J. Linn. Soc., 65: 81-85.

Chudzik, B. and R. Sniezko, 2003. Calcium ion presence as a trait of receptivity in tenuinucellar ovules of *Galanthus nivalis* L. Acta. Biol. Cracoviensia Series Botanica, 45: 133-141.

Dane, F., C. Meriç and G. Huseyinova, 2000. Some Ultrastructural observations on calcium oxalate raphide crystal idioblasts and meristematic cells of the adventive root tips of *Sternbergia lutea* (L.) Ker-Gawl. ex Sprengel (*Amaryllidaceae*). Turk. J. Bot., 24: 71-80.

Dehgan, B. and M.E. Craig, 1978. Types of laticifers and crystals in *Jatropha* and their taxonomic implication. Am. J. Bot., 65: 345-352.

Ekici, N. and F. Dane, 2007. Calcium oxalate crystals during development of male and female gametophyte in *Leucojum aestivum* L. (Amaryllidaceae). Phytologia Balcanica, (Supplement) (In Press).

Esau, K., 1976. Anatomy of Seed Plant. California.

Foster, A.S., 1956. Plant idioblasts: Remarkable examples of cell specialization. Protoplasma, 46: 184-193.

Franceschi, V.R. and H.T. Horner, 1980. Calcium oxalate crystals in plants. Bot. Rev., 46: 361-427.

Frey-Wyssling, A., 1981. Crystallography of the two hydrates of crystalline calcium oxalate in plants. Am. J. Bot., 68: 130-141.

Herr, J.M., 1971. A new clearing-squash technique for the study of ovule development in angiosperms. Am. J. Bot., 58: 785-790.

Horner, H.T. and B.L. Wagner, 1980. Association of druse crystals with the developing stomium of *Capsicum annuum* (Solanaceae) anthers. Am. J. Bot., 67: 1347-1360.

- Horner, H.T. and B.L. Wagner, 1992. Association of four different calcium crystals in the anther connective tissue and hypodermal stomium of *Capsicum annuum* (Solanaceae) during microsporogenesis. Am. J. Bot., 67: 1347-1360.
- Ilarslan, H., R.G. Palmer, J. Imsande and H.T. Horner, 1997.

 Quantitative determination of calcium oxalate and oxalate in developing seeds of soybean (*Leguminosae*). Am. J. Bot., 84: 1042-1046.
- Ilarslan, H., R.G. Palmer and H.T. Horner, 2001. Calcium oxalate crystals in developing seeds of soybean. Ann. Bot., 88: 243-257.
- Kausch, A.P. and H.T. Horner, 1983. The development of mucilaginous raphide crystal idioblasts in young leaves of *Typha angustifolia* L. (Typhaceae). Am. J. Bot., 70: 691-705.
- Lersten, N.R., 1974. Morphology and distribution of colleters and crystals in relation to the taxonomy and bacterial leaf nodule symbiosis of *Psychotria* (*Rubiaceae*). Am. J. Bot., 61: 973-981.
- Meric, C. and F. Dane, 2004. Calcium oxalate crystals in floral organs of *Helianthus annuus* L. (Asteraceae). Acta Biol. Szegediensis, 48: 19-23.

- Monje, P.V. and E.J., Baran, 2002. Characterization of Calcium Oxalates Generated as Biominerals in Cactil. Plant Physiol., 128: 707-713.
- Prychid, C.J. and P.J. Rudall, 1999. Calcium oxalate crystals in monocotyledons: A review of their structure and systematics. Ann. Bot., 84: 725-739.
- Seago, J.L. and L. Marsh, 1989. Adventitious root development in *Typha glauca* with emphasis on the cortex. Am. J. Bot., 76: 909-923.
- Sunell, L.A. and P.L. Healy, 1979. Distribution of calcium oxalate crystal idioblasts in corm of taro (*Colocasia esculenta*). Am. J. Bot., 66: 1029-1032.
- Tilton, V.R. and H.T. Horner, 1980. Calcium oxalate raphide crystals and crystalliferous idioblasts in the carpels of *Ornithogalum caudatum*. Ann. Bot., 46: 533-539.
- Watson, L. and M.J. Dallwitz, 2005. The Families of Flowering Plants. http://delta-intkey.com/angio/www/amaryllidaceae. html.