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Heritability and Correlation of Drought Resistance Traits and Agronomic Traits in Peanut (*Arachis hypogaea* L.)

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Abstract: Several physio-morphological traits are related to pod yield of peanut. Improvement of these traits should lead to yield increase under drought conditions. The objective of this study was to evaluate (1) heritability of drought resistance traits, yield and yield components and (2) relationships among these traits. A cross of two parents (ICGV 98324 and KK 4) differing in physio-morphological traits was used in this study. Pot experiments of F₂ and F₃ populations were set up in the open field with rainout shelters. One hundred and twenty eight entries were subjected to water stress during 28 to 70 days after sowing and evaluation of the studied characters was conducted at appropriate time. Data were recorded for Root Dry Weight (RDW), Root Length (RL), Root Surface (RS), Root Volume (RV), Specific Leaf Area (SLA), SPAD Chlorophyll Meter Reading (SCMR), biomass, pod yield, pod number per plant, seed number per pod, 100-seed weight and Harvest Index (HI). Heritability estimates in broad sense for root characters and drought resistance traits were low to intermediate, ranging from 0.27 to 0.59. Similarly, low to intermediate heritability estimates in broad sense were found for pod yield and yield components, ranging from 0.20 to 0.57. Heritability estimates in narrow sense were much lower than in broad sense. The correlation coefficients among root characters were inter-related positively, whereas negative correlation coefficients were observed among physiological characters. Root characters were closely related to biomass production but they were not related to yield and yield components except for pod number per plant.

Key words: Breeding, inheritance, SLA, SPAD chlorophyll meter reading, water stress

INTRODUCTION

Peanut yield in rain-fed areas has been limited by drought stress because pod yield and other growth parameters have been severely affected (Pimratch *et al.*, 2008; Songsri *et al.*, 2008a; Awal and Ikeda, 2002; Nautiyal *et al.*, 2002; Reddy *et al.*, 2003; Nigam *et al.*, 2005). Yield losses have been estimated to be 56-85% (Nageswara Rao *et al.*, 1989), depending on crop growth stages when the crop was exposed to drought (Awal and Ikeda, 2002; Reddy *et al.*, 2003), drought intensity and drought duration (Nautiyal *et al.*, 2002; Nigam *et al.*, 2005). Even in irrigated areas, peanut is frequently exposed to drought because water supply is not sufficient.

The use of drought-resistant varieties is an important strategy to combat the drought problem. These varieties should be able to provide higher yield under drought conditions. Genetic variability for drought resistance has been reported in peanut (Erickson and Ketring, 1985; Upadhyaya, 2005; Songsri *et al.*, 2008a-c, 2009). However,

breeding for drought resistance based on pod yield is lacking behind due to significant genotype×environment interactions (Wright *et al.*, 1996).

Alternative breeding strategies using physiological traits as selection criteria have been proposed by some researchers. Rapid progress in drought resistance breeding has been achieved based on characters like Harvest Index (HI), Water Use Efficiency (WUE), Specific Leaf Area (SLA) and SPAD Chlorophyll Meter Reading (SCMR) (Nigam *et al.*, 2005). The SLA and SCMR have been found to be highly correlated with WUE (Nageswara Rao *et al.*, 2001; Sheshshayee *et al.*, 2006) and have been used as surrogate traits for WUE (Nigam *et al.*, 2005; Lal *et al.*, 2006; Sheshshayee *et al.*, 2006; Arunyanark *et al.*, 2008; Jongrungklang *et al.*, 2008; Pimratch *et al.*, 2008). Specific leaf area and SCMR have been found to be negatively correlated (Nageswara Rao *et al.*, 2001; Upadhyaya, 2005).

Earlier studies have indicated differential responses for relative water content in peanut (Painawadee *et al.*, 2009) and it was positively correlated with chlorophyll

content and grain yield in rice under drought conditions Pirdashti *et al.* (2009). Leaf water status is dependent on rooting density, root distribution, ability of roots to extract water, behavior of stomata closure and transpiration rate (Kramer, 1969, 1983; Gregory, 2006). Root systems play a crucial role in determining shoot water status and therefore effective water uptake is an important determinant of drought resistance (Huang *et al.*, 1997; Huang, 2000; Kashiwagi *et al.*, 2006). Larger root systems and deep growth of root systems into lower soil profile can take up more water to support plant growth and yield (Ludlow and Muchow, 1990; Turner *et al.*, 2001). Moreover, deep and prolific root systems have been associated with enhanced avoidance of terminal drought stress in chickpea (Ludlow and Muchow, 1990; Serraj *et al.*, 2004). Selections with more extensive root systems could extract more soil water from greater soil volumes than selections with limited root system. Many root characteristics have been shown to be under genetic control and quantitatively inherited (O'Toole and Bland, 1987). Genetic variation for root characters has been found among peanut genotypes (Ketring, 1984). Differences among peanut genotypes were observed and Virginia type possessed longer taproots and had faster root growth rates than Spanish type (Huang and Ketring, 1987; Maiti *et al.*, 2002).

Information on the heritability of RWC, SCMR, SLA, HI, biomass, pod yield, pod number per plant, seed number per pod, 100-seed weight and root traits and the phenotypic correlations among these traits will be useful for planning suitable breeding strategies for improving drought tolerance. The effective selection for traits under improvement depends on sufficient additive genetic variation of the traits that are expressed as heritability. Phenotypic relationships among traits are also important when simultaneous selection of multiple traits is to be carried out for high yield under drought stress conditions. Therefore, the present research was undertaken to estimate the (1) heritability of drought resistance traits, yield and yield components and (2) relationships among these traits.

MATERIALS AND METHODS

Plant materials: Two parental lines (ICGV 98324 and KK 4) were used in a cross to generate an F₁ hybrid. The parents were selected because they were different in many characters such as RWC and SCMR as evaluated in one of our studies (Painawadee *et al.*, 2009). ICGV 98324 is a drought resistant line from ICRISAT and it was identified as drought resistant because of high total biomass and pod yield under drought conditions (Nageswara Rao *et al.*, 1992; Nigam *et al.*, 2003, 2005). KK 4, a released variety in Thailand, is a Valencia bunch

type with erect growth habit, early maturity (100 days), small-seeded nature and high drought tolerance index for RDW (Painawadee *et al.*, 2009). The F₁ hybrid was further grown in a small plot for multiplication and generation advance. The F₂ and F₃ generations were used in the experiments.

Crop management and experimental designs: The experiment was conducted at the Field Crop Research Station of Khon Kaen University located in Khon Kaen Province, Thailand (latitude 16° 28' N, longitude 102° 48' E, 200 m above sea level). One hundred and twenty eight plants in the F₂ generation were first evaluated in an un-replicated trial during December, 2006 to April, 2007 and the 128 progenies in the F₃ generation were later evaluated in a randomized complete block design with four replications during June, 2007 to October, 2007. Crop management for both trials was identical.

The plants were grown in pots with 25 cm diameter and 70 cm height under open environment and removeable rainout shelters were available if necessary. Pots were filled with soil (Yt; fine-loamy, siliceous, isohypothermic, Oxic Paleustults). The soil properties are given in Table 1.

The soil was filled into the pots in four columns of the soil profile in order to make soil bulk density uniform. Three plastic tubes were installed to supply water to each soil column from the bottom to the column below the top soil column and the top soil column was surface irrigated as detailed by Songsri *et al.* (2009) and Painawadee *et al.* (2009).

Lime at the rate of 19.2 g pot⁻¹ was incorporated into the soil prior to soil filling and phosphorus fertilizer as triple superphosphate at the rate of 12.12 g P pot⁻¹ and potassium fertilizer as muriate of potash (KCl) at the rate 15.26 g K pot⁻¹ were applied immediately before planting.

Table 1: Soil properties for F₂ and F₃ generations experiment

Soil properties	Generations	
	F ₂	F ₃
Physical (%)		
Sand particle	71.00	55.00
Silt particle	20.00	29.00
Clay particle	9.00	16.00
Field capacity	10.00	10.00
1/3 available water	5.00	6.00
Permanent wilting point	3.00	4.00
Chemical		
pH (1:1 H ₂ O)	5.58	4.50
Organic matter (%)	0.47	0.36
Total nitrogen (%)	0.02	0.03
Available phosphorus (ppm)	7.00	2.00
Potassium (ppm)	23.50	29.50
Calcium (ppm)	216.50	215.50
Bulk density (g cm ⁻³)	1.48	1.47

Seeds were treated with captan (3a, 4, 7, 7a-tetrahydro-2-[(trichloromethyl)thio]-1H-isoindole-1, 3 (2H)-dione) at the rate of 5 g kg⁻¹ seed for fungal control and ethrel 48% at the rate of 2 ml L⁻¹ water to break seed dormancy. Rhizobium inoculation was done by applying a water-diluted commercial peat-based inoculum of Bradyrhizobium (mixture of strains THA, 201 and THA, 205; Department of Agriculture, Ministry of Agriculture and Cooperatives, Bangkok, Thailand) on the hills before planting.

The seeds were over planted and the seedlings were thinned to one plant per hill at 14 days after sowing (DAS). Gypsum (CaSO₄) at the rate of 9.58 g pot⁻¹ was applied at 40 DAS. The pots were kept weed free by regular manual weeding. Pests and diseases were controlled by weekly application of carbosulfan [2-3-dihydro-2,2-dimethylbenzofuran-7-yl (dibutylaminothio) methylcarbamate 20% w/v, water soluble concentrate] at 2.5 L ha⁻¹, methomyl [S-methyl-N-((methylcarbamoyl) oxy) thioacetimidate 40% soluble powder] at 1.0 kg ha⁻¹ and carboxin [5,6-dihydro-2-methyl-1,4-oxath-ine-3-carboxanilide 75% wettable powder] at 1.68 kg ha⁻¹.

Uniform irrigation at field capacity was supplied to the experiments from planting to 12 DAS. Withholding of irrigation was carried out after 12 DAS to allow soil water gradually get reduced until reaching at 1/3 available water

(28 DAS) and the soil moisture content was maintained at this level until 70 DAS. After 70 DAS, soil moisture level at field capacity was brought back until harvest.

The method of calculation for plant water use proposed by Songsri *et al.* (2008a-c, 2009) was followed. It was found that crop water requirement was identical to crop water loss through plant transpiration and soil evaporation. Therefore, crop water requirement is the product of evaporation (a pan) multiplied by crop coefficient for peanut. For each water level, soil moisture was controlled uniformly until harvest.

Weather parameters: Weather data were obtained from the nearest meteorological station. The maximum and minimum air temperatures ranged between 33.1 and 20°C in F₁ trial and 32.4 and 25.8°C in F₂ trial (Fig. 1). Daily pan evaporation ranged from 2.88 to 9.84 mm in F₁ trial and 0.6 to 8.9 mm in F₂ trial. The relative humidity values were 75.6% in F₁ trial and 89% in F₂ trial. The solar radiations and average sunshine hours were 18.9 MJ/m²/day and 10 h in F₁ trial and 13.9 MJ/m²/day and 6.4 h in F₂ trial, respectively.

Data collection:

Plant water status: Relative Water Content (RWC) was recorded on each of the four leaflets of the tetrafoliate leaf

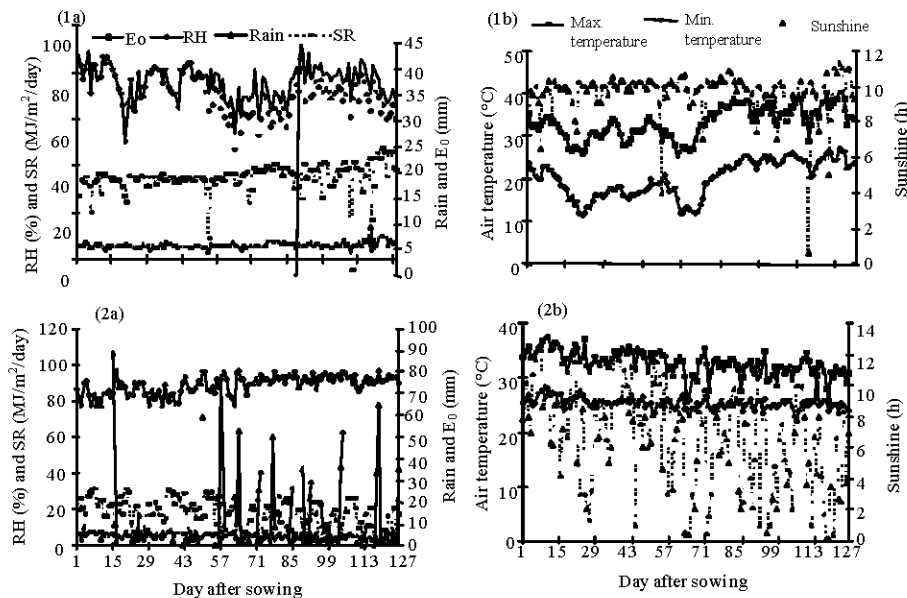


Fig. 1: Daily mean air temperature, rain fall, evaporation (E₀), relative humidity (RH), solar radiation (SR), maximum and minimum air temperature (max. and min. temp.) and sunshine (h) all season grow for experiment 1 (1a, b) and experiment 2 (2a, b)

at 70 DAS. Only one fully expanded second or third leaf from the apex of the main axis of each pot was used to record RWC. Leaves were detached and kept in sealable plastic bags in ice box and transported to the laboratory and leaf fresh weight was recorded. The leaf samples were then soaked in distilled water for 8 h and blotted for surface drying and water-saturated leaf weight was determined. The samples were oven-dried at 80°C until reaching constant weight and leaf dry weight could be determined. The RWC was calculated based on the formula suggested by González and González-Vilar (2001) as follows:

$$\text{RWC (\%)} = \frac{[\text{FW}-\text{DW}]/(\text{TW}-\text{DW}) \times 100}$$

where, FW is the sample fresh weight, TW is the sample turgid weight and DW is the sample dry weight.

Leaf parameters: SPAD Chlorophyll Meter Reading (SCMR), Specific Leaf Area (SLA) and Relative Water Content (RWC) were recorded at 70 days after sowing at 9.00-10.00 AM. The second leaf from terminal bud of the main stem of each plant was detached and kept in sealable plastic bags in ice box. The leaf samples were soon transported to a laboratory. Fresh weight was recorded soon after reaching the laboratory and SCMR was measured immediately by a Minolta handheld portable SCMR meter (SPAD- 502 Minolta, Tokyo, Japan), using four leaflets per sample. In recording the SCMR, care was taken to ensure that the SPAD meter sensor fully covered the leaf lamina and the interference from veins and midribs could be avoided. The same samples were further measured for leaf area, using a leaf area meter (LI 3100C Area meter, LI COR Inc., USA). The samples were then oven-dried at 80°C until reaching constant weight and leaf dry weight could be determined. The SLA was calculated using the following equation (Nageswara Rao *et al.*, 2001):

$$\text{SLA} = \frac{\text{Leaf area (cm}^2\text{)}}{\text{Leaf dry weight (g)}}$$

Root traits, yield and yield components: Biomass (roots and shoots included), root dry weight, shoot dry weight and harvest index were determined at harvest (120 DAS). Shoots were cut at crown level and the roots were washed in running tap water to remove the soil. The complete removal of the soil was done on a 3 mm sieve. Care was taken to recover the root system completely. The roots were thoroughly cleaned and straightened by repeated dipping and rising in buckets of clear water. Then, The root samples were put on a flatbed scanner (Epson Perfection V700 Photo, Epson Inc., Cheung Shwan,

Kowloon, HK) and scanned using WinRhizo Pro 2004a (Regent instruments, Inc., Quebec, QC) at 400 dpi (Gaur *et al.*, 2008; Songsri *et al.*, 2008a). Top and bottom lighting systems were used to eliminate shadows and maximize contrast. The captured grayscale image was analyzed with WinRhizo to measure Root Surface (RS), Root Length (RL) and Root Volume (RV). Root samples and above ground samples were oven dried at 80°C until constant weights were reached. After oven drying, Root Dry Weight (RDW) and shoot dry weight were recorded. Harvest index was calculated using the following relationship (Wright and Nageswara Rao, 1994):

$$\text{HI} = \frac{\text{Pod yield}}{\text{Pod yield} + \text{shoot and root dry weight}}$$

At harvest, pods were removed from shoots and immature pods were not included in any calculation of yield parameters. Pod yields were determined after air drying to approximately 8% moisture content. Pod number per plant seed number per pod and 100 seed weight were also recorded at final harvest.

Statistical analysis: The data of the F₃ generation were subjected to analysis of variance (Gomez and Gomez, 1984) and the significant variance of the progenies was further partitioned into genetic variance and error variance. Heritability estimates in broad sense (h_{bs}^2) were calculated using the following relationship (Singh *et al.*, 1993):

$$h_{bs}^2 = \frac{\sigma_g^2}{(\sigma_g^2 + \sigma_e^2)}$$

Where:

$$\sigma_e^2 = M_e, \sigma_g^2 = (M_g - M_e)/b,$$

σ_e^2 = Environmental variance

M_e = Mean square of error

σ_g^2 = Genotypic variance

M_g = Mean square of genotype

The Standard Error (SE) associated with broad sense heritability estimate was calculated as:

$$\text{SE}(h_{bs}^2) = (1-h^2) [1+(b-1) h^2] [2/(bf)]^{1/2}$$

Where:

b = Replication

f = Degree of freedom of error.

Heritability estimates in narrow sense (h_n^2) were also calculated by parent offspring method using the data of F₃ on F₂ families as suggested in Smith and Kinman (1965):

$$h_{ig}^2 = b/2r_{op}$$

Where:

b = Regression coefficient or slope

r_{op} = Relationship of parents-offspring.

Linear regression coefficients (b) were calculated by regressing of F₃ progeny means (Y_i) on F₂ plants means (X_i). Standard errors (SE) for the slope of each regression were calculated as follows (Ibrahim and Quick, 2001):

$$SE = \left[\frac{Y_i^2 - (X_i Y_i) 2 / X_i^2}{(n - 2) X_i^2} \right] / X_i^2$$

where, n is number of families.

Simple correlation coefficients based on progeny means of F₃ generation were calculated to determine the relationship between yield and yield components with drought resistance traits. All calculations were accomplished using STATISTIX 8 software program.

RESULTS

Heritability estimates for root parameters and drought resistant traits: Broad sense heritability estimates for root parameters and drought resistant traits, in general, were much higher than those in narrow sense (Table 2). Heritability estimates in broad sense ranged from 0.27 for SCMR to 0.59 for root surface and root volume, whereas heritability estimates in narrow sense ranged from 0.00 for SLA to 0.13 for root dry weight.

Heritability estimates for yield and yield components: Heritability estimates in broad sense for yield and yield components were relatively higher than those in narrow sense (Table 3). Broad sense heritability estimates ranged from 0.20 for Harvest Index (HI) to 0.57 for pod number per plant, whereas narrow sense heritability estimates ranged from 0.13 for pod yield to 0.23 for pod number per plant and 100-seed weight.

Phenotypic correlation between drought resistant traits and drought resistant traits and yield components: All root parameters (root length, root dry weight, root surface and root volume) were positively and significantly associated with the high correlation coefficients between 0.67, p<0.01 to 0.98, p<0.01 (Table 4). In contrast to root parameters, all traits related to drought resistance (RWC, SLA and SCMR) were negatively and significantly associated and the correlation coefficients ranged between -0.17, p<0.05 to -0.51, p<0.01). Root characters were also positively and significantly correlated with biomass production and in lesser extent with pod number per plant (Table 5). Most drought resistance traits (RWC,

Table 2: Broad sense and narrow sense heritability and standard errors for root dry weight (RDW), root length (RL), root surface (RS), root volume (RV), specific leaf area (SLA), relative water content (RWC) and SPAD chlorophyll meter reading (SCMR)

Character	Heritability	
	Broad sense	Narrow sense
RDW	0.34±0.03	0.13±0.01
RL	0.58±0.04	0.10±0.00
RS	0.59±0.04	0.11±0.00
RV	0.59±0.03	0.12±0.00
SLA	0.57±0.04	0.00
RWC	0.46±0.04	0.01±0.00
SCMR	0.27±0.03	0.05±0.00

Table 3: Broad sense and narrow sense heritability and standard errors for biomass, pod yield, number pod per plant, number seed per pod, 100-seed weight and harvest index (HI)

Character	Heritability	
	Broad sense	Narrow sense
Biomass	0.30±0.03	0.16±0.00
Pod yield	0.47±0.04	0.13±0.00
Pod No. plant ⁻¹	0.57±0.04	0.23±0.01
Seed No. pod ⁻¹	0.24±0.03	0.19±0.02
100 seed weight	0.46±0.04	0.23±0.01
HI	0.20±0.02	0.21±0.01

Table 4: Correlation coefficients among drought resistance traits for peanut in F₃ generation

Drought resistance traits	Drought resistance traits					
	RDW	RL	RS	RV	RWC	SCMR
RL	0.67**					
RS	0.71**	0.98**				
RV	0.73**	0.92**	0.97**			
RWC	0.03	-0.05	-0.00	0.04		
SCMR	0.15	0.11	0.10	0.09	-0.51**	
SLA	-0.03	0.07	0.06	0.03	-0.17*	-0.40**

*, ** Significant at the p ≤ 0.05 and p ≤ 0.01, respectively

Table 5: Correlation coefficients between yield and yield components with drought resistance traits for peanut in F₃ generation

Drought resistance traits	Yield and yield components					
	HI	Biomass	Pod yield	Pod No. plant ⁻¹	Seed No. pod ⁻¹	100 seed weight
RDW	-0.20*	0.65**	0.13	0.10	0.02	0.04
RL	0.04	0.43**	0.04	0.29**	0.02	0.06
RS	0.00	0.42**	0.01	0.26**	0.02	0.05
RV	-0.04	0.39**	0.04	0.17*	0.01	0.02
SLA	0.02	0.02	0.02	-0.07	0.00	-0.03
RWC	0.04	0.07	0.04	0.11	0.05	0.03
SCMR	-0.01	0.10	0.01	0.20*	0.05	0.00

*, ** Significant at the p ≤ 0.05 and p ≤ 0.01, respectively

SCMR and SLA) were not correlated with pod yield and yield components except for SCMR with pod number per plant (0.20, p<0.05).

DISCUSSION

The progress in breeding for drought resistance in peanut based on pod yield has been slow because of high G×E interactions (Wright *et al.*, 1996). The use of surrogate traits related to drought resistance has been

suggested by many authors (Nageswara Rao *et al.*, 2001; Nigam *et al.*, 2005) as the inheritance of these characters must be simpler than pod yield. The information on the heritability and the relationships among characters is important for plant breeders to formulate appropriate breeding strategies to achieve breeding goals. The aims of this study were to understand whether heritability estimates for characters under investigation were sufficient for further improvement of these characters and to explore whether physiological characters and other drought related characters could be used as surrogate traits for pod yield under drought conditions.

For drought resistance and physiological characters, the heritability estimates in broad sense were higher than those in narrow sense. Similar lower narrow sense heritability estimates were also observed for pod yield and its related traits. Higher broad-sense heritability estimates could be due to the inclusion of non-heritable genetic variance and variance due to G×E interactions (Falconer, 1996; Brown and Caligari, 2008). Low heritability estimates observed in this study indicate the difficulty in improving these characters.

The heritability estimates in narrow sense in this study may be under estimated because of high intra plant variation of individual plants in the F₂ generation. Single plants in the F₂ generation might be not appropriate for evaluation of narrow sense heritability estimates and the heritability estimates may be improved if more advanced generations were used with appropriate replicated trials. Root systems themselves are exceedingly complex structures, typically being composed of thousands of individual root axes. Higher narrow-sense heritability estimates for crop growth rate, reproductive duration, partitioning and yield were obtained based on F₄ on F₃ rather than based on F₃ on F₂ (Ntare and Williams, 1998).

Heritability estimates in broad sense and narrow sense for yield and yield components followed a similar pattern to those for root parameters and traits related to drought resistance. However, narrow sense heritability estimates for yield and yield components were somewhat higher than those for root parameters and traits related to drought resistance. Better improvement for these traits would be expected especially for characters with higher narrow sense heritability estimates such as pod number per plant (0.23), seed number per pod (0.19), 100-seed weight (0.23) and harvest index (0.21).

With regard to the heritability estimates, drought resistance and physiological characters were not superior to yield and yield components and therefore, the use of drought resistance and physiological characters as surrogate traits for pod yield and yield components in

this population will not be effective. The disappointing results observed in this study could be due to the low genetic variability for these characters in the parental materials. However, the use of more diverse germplasm as parental materials should increase selection efficiency and these surrogate traits are still worth-exploring in peanut.

To the best of our knowledge so far, the report on the heritability for root characters in peanut has not been available in the open literature and the direct comparison of the results is not possible. In peanut, Songsri *et al.* (2008b) found rather high broad sense heritability estimates of 0.73-0.98 for biomass, pod yield, HI, SCMR and SCMR in the F_{4.7} and F_{4.8} generations. For other legumes, however, broad-sense heritability estimates of 0.51-0.55 for root area, 0.47-0.50 for root length and 0.51-0.61 for root mass in common bean under limited soil phosphorus supply has been reported (Araújo *et al.*, 2005). In clover, broad sense heritability estimates for root characters were between 0.2 and 0.4 (Caradus and Woodfield, 1990).

The relationships of root parameters indicated that it was not necessary to evaluate all parameters and evaluation of the most convenient and less expensive characters would be sufficient. In this case, root dry weight is recommended. Ketring (1984) also found positive correlation between root volume and root dry weight in peanut and suggested to select for extensive rooting traits to develop more drought tolerant peanut cultivars. Huang and Ketring (1987) also obtained highly positive linear correlation coefficients for root dry weight with root volume and total dry weight in peanut. High inter-relationships among root characters were also observed in pea (McPhee, 2005) and common bean (Araújo *et al.*, 2005). They also suggested that the high correlation between root mass and root area justifies screening genotypes based solely on root mass. However, negative correlation between root length and root dry weight were found in rice (Sasmal, 2008). This could be due to the fact that rice is normally grown under water-flooded conditions.

In contrast to root parameters, all traits related to drought resistance (RWC, SLA and SCMR) were negatively and significantly associated and the correlation coefficients ranged between -0.17, $p \leq 0.05$ to -0.51, $p \leq 0.01$). The results were in agreement with those reported under non-stressed conditions (Nageswara Rao *et al.*, 2001; Upadhyaya, 2005) and end of season drought conditions (Nigam and Aruna, 2008). More recently, Songsri *et al.* (2009) found consistent relationships between SLA and SCMR under different water regimes. The significant interrelationships between

SLA and SCMR suggested that SCMR could be used as a reliable and rapid measure to identify genotypes with low SLA in peanut. Nageswara Rao *et al.* (2001) reported that SCMR could be used as a reliable and rapid measure to identify genotypes with low SLA or high Specific Leaf Nitrogen (SLN) which are surrogate measures of Transpiration Efficiency (TE) in peanut. In cowpea studied under mid-season drought, the high RWC of leaves was maintained in some of the genotypes by stomata closure and a reduction of leaf area. Drought avoidance by maintaining high leaf water content was negatively associated with SLA (Anyia and Herzog, 2004). Jongrungklang *et al.* (2008) found that the more severe the drought stresses the more was the increase in the SCMR. In fact, plant water status is related to level of soil moisture. Therefore, the results in this study showed the negative and significant relationship between SCMR and RWC. The water loss from cells might effect the concentration of chlorophyll content. During the stress period in rice, SCMR increased with stress but declined rapidly within 3 days after re-irrigation (Duy Nang, 2004). Contrastingly, in rice under drought stress conditions the correlation between SCMR with RWC was positive and significant and SCMR decreased with drought stress compared to control (Pirdashti *et al.*, 2009).

However, drought resistance traits and root traits were not associated. This could be largely due to different times of evaluation between the two groups of characters. RWC, SCMR and SLA were evaluated at 70 DAS, whereas RDW, RS and RV were evaluated at harvest because of limited number of samples and evaluation of root traits being highly destructive. Furthermore, Ketring and Reid (1993) found that groundnut was able to establish both a deep and laterally spreading root system fairly early during the growing cycle, providing adaptation to drought occurrence during and later in the season. Root growth estimations at early vegetative growth stages may be of limited use considering the growth stage \times genotype interaction for root growth in chickpea (Canci *et al.*, 2004; Krishnamurthy *et al.*, 1996).

Root characters were also positively and significantly correlated with biomass production and in lesser extent with pods number per plant. This could indicate that larger plants had larger root system and also had high number of pods. Although RDW was not significantly correlated with pod yield and pod number per plant, it was negatively and significantly correlated with harvest index. The results might suggest that the effect of roots contributed indirectly to pod yield through biomass production and subsequently through pod number per plant, but large root systems had low contribution to partitioning efficiency as indicated by negative correlation

with harvest index. Similarly, McPhee (2005) observed positive correlation between total root characters and biomass and the present authors also found inter-relationships among root characters. However, Passioura (1983) hypothesized that yield could be increased by decreasing roots as they represent a high energy. Siddique *et al.* (1990) found that wheat genotypes with high HI would have lower root/shoot ratios, indicating less investment in roots. In fact, the turnover of roots can be relatively rapid, with a half life of 30-40 days in peanut (Krauss and Deacon, 1994). Therefore, even if the root/shoot ratio at a given point in time in many species is only between 10 and 40%, a complete turnover of roots in about 40 days would bring the root/shoot ratio close to 100% over the entire life cycle.

Most drought resistance traits (RWC, SCMR and SLA) were not correlated with pod yield and yield components except for SCMR with pod number per plant (0.20, $p \leq 0.05$). More greenish plants yielded more pods than did plants with lighter color under non-stress conditions. According to Wuruna *et al.* (2009), number of pod per plant was significantly correlated with SCMR at 60 days after emergence, but the correlation was not significant under 1/3 available water (severe drought stress). SCMR is an indicator of the photo-synthetically active light-transmittance characteristics of the leaf, which is dependent on the unit amount of chlorophyll per unit leaf area (chlorophyll density) (Richardson *et al.*, 2002). Significant and positive correlation between SCMR and chlorophyll content was observed and SCMR was also closely related with chlorophyll density (Arunyanark *et al.*, 2008, 2009).

CONCLUSION

Heritability and correlation information provides a guideline for selection of characters in breeding population. In this study, broad sense heritability estimates for most characters were not high. Narrow sense heritability estimates were much lower than broad sense heritability estimates and they were expected to be underestimated because of high intra-plant variation in the F_2 population. Based on heritability estimates, selection for RWC, SCMR, root parameters and other traits, characters related to drought resistance and agronomic traits in this population may be difficult in early segregating generation and evaluation should be carried out in more advanced generations. As there were high inter-correlations among root parameters, evaluation of root dry weight alone is sufficient because it was more simple, economical and less time-consuming.

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REFERENCES

- Anyia, A.O. and H. Herzog, 2004. Water-use efficiency, leaf area and leaf gas exchange of cowpeas under mid-season drought. *Eur. J. Agron.*, 20: 327-437.
- Araújo, A.P., I.F. Antunes and M.G. Teixeira, 2005. Inheritance of root traits and phosphorus uptake in common bean (*Phaseolus vulgaris* L.) under limited soil phosphorus supply. *Euphytica*, 145: 33-40.
- Arunyanark, A., S. Jogloy, C. Akkasaeng, N. Vorasoot, T. Kesmla, R.C. Nageswara Rao, G.C. Wright and A. Patanothai, 2008. Chlorophyll stability is an indicator of drought tolerance in peanut. *J. Agron. Crop Sci.*, 194: 113-125.
- Arunyanark, A., S. Jogloy, N. Vorasoot, C. Akkasaeng, T. Kesmla and A. Patanothai, 2009. Stability of relationship between chlorophyll density and soil plant analysis development chlorophyll meter readings in peanut across different drought stress conditions. *Asian J. Plant Sci.*, 8: 102-110.
- Awal, M.A. and T. Ikeda, 2002. Recovery strategy following the imposition of episodic soil moisture deficit stands of peanut (*Arachis hypogaea* L.). *J. Agron. Crop Sci.*, 188: 185-192.
- Brown, J. and P. Caligari, 2008. An Introduction to Plant Breeding. 1st Edn., Wiley-Blackwell, Oxford, UK., ISBN : 978-1-4051-3344-9 pp: 66-115.
- Canci, H., M. Cagirgan and C. Toker, 2004. Genotypic variations for root and shoot growth at seedling stage in chickpea mutants. *Int. Chickpea Pigeonpea Newsletter*, 11: 11-12.
- Caradus, J.R. and D.R. Woodfield, 1990. Estimates of heritability for and relationships between, root and shoot characters of white clover. I. Replicated clonal material. *Euphytica*, 46: 203-209
- Duy-Nang, N., 2004. Nitrogen uptake, leaf nitrogen, chlorophyll content and leaf color of rice (*Oryza sativa* L.) as affected by drought stress. M.Sc. Thesis, Univers. Philip., Los Banos.
- Erickson, P.I. and D.L. Ketring, 1985. Evaluation of peanut genotypes for resistance to water stress *in situ*. *Crop Sci.*, 25: 870-876.
- Falconer, D.S., 1996. Introduction to Quantitative Genetics. 2nd Edn., Longman Group, New York.
- Gaur, P.M., L. Krishmanan and J. Kashiwagi, 2008. Improving drought-avoidance root traits in chickpea (*Cicer arietinum* L.)—current status of research at ICRISAT. *Plant Prod. Sci.*, 11: 3-11.
- Gomez, K.A. and A.A. Gomez, 1984. Statistical Procedures for Agricultural Research. 2nd Edn., John Wiley and Sons Inc., New York, ISBN-10: 0471870927.
- González, L. and M. González-Vilar, 2001. Determination of Relative Water Content. In: Handbook of Plant Ecophysiology Techniques, Roger, M.J.R. (Ed.). Springer, Netherlands, ISBN: 978-0-7923-7053-6, pp: 207-212.
- Gregory, P.J., 2006. The Functioning Root System: Plant Roots Growth, Activity and Interaction with Soils. Blackwell Publishing Ltd., Oxford, UK., ISBN-10: 1405119063.
- Huang, M. and D.L. Ketring, 1987. Root growth characteristics of peanut genotypes. *Jour. Agric. China.*, 36: 41-52.
- Huang, B., R.R. Duncan and R.N. Carrow, 1997. Drought-resistance mechanisms of seven warm-season turfgrasses under surface soil drying: II. Root aspects. *Crop Sci.*, 37: 1863-1869.
- Huang, B., 2000. Role of Root Morphological and Physiological Characteristics in Drought Resistance of Plants. In: Plant-Environment Interactions, Wilkinson, R.F. (Ed.). Marcel Inc., New York, ISBN: 13978-0824703776, pp: 39-64.
- Ibrahim, A. and J.S. Quick, 2001. Heritability of heat tolerance in winter and spring wheat. *Crop Sci.*, 41: 1401-1405.
- Jongrunklang, N., B. Toomsan, N. Vorasoot, S. Jogloy, T. Kesmla and A. Patanothai, 2008. Identification of peanut genotypes with high water use efficiency under drought stress conditions from peanut germplasm of diverse origins. *Asian J. Plant Sci.*, 7: 628-638.
- Kashiwagi, J., L. Krishnamurthy, J.H. Crouch and R. Serraj, 2006. Variability of root length density and its contributions to seed yield in chickpea (*Cicer arietinum* L.) under terminal drought stress. *Field Crops Res.*, 95: 171-181.
- Ketring, D.L., 1984. Root diversity among peanut genotypes. *Crop Sci.*, 24: 229-232.
- Ketring, D.L. and J.L. Reid, 1993. Growth of peanut roots under field conditions. *Agron. J.*, 85: 80-85.
- Kramer, P.J., 1969. Plant and Soil Water Relationships: A Modern Synthesis. 1st Edn., Tata McGraw Hill Publishing Company Ltd., New Delhi, ISBN: 0-07-099399-8, pp: 296-345.

- Kramer, P.J., 1983. Water Relations of Plants. 1st Edn., Academic Press, New York, ISBN: 0-12-425040-8, pp: 187-261.
- Krauss, U. and J.W. Deacon, 1994. Root turnover of groundnut (*Arachis hypogaea* L.) in soil tubes. Plant and Soil, 166: 259-270.
- Krishnamurthy, L., O. Ito and C. Johansen, 1996. Genotypic Differences in Root Growth Dynamics and its Implications for Drought Resistance in Chickpea. In: Dynamics of Roots and Nitrogen in Cropping Systems of the Semi-Arid Tropics. Ito, O., C. Johansen, J.J. Adu Gyamfi, K. Katayama, J.V.D.K. Kumar Rao and T.J. Rego (Eds.). JIRCAS Agriculture Series No. 3. Japan International Research Center for Agricultural Sciences, Tsukuba, pp: 235-250.
- Lal, C., K. Hariprasanna, A.L. Rathnakumar, H.K. Gor and B.M. Chikani, 2006. Gene action for surrogate traits of water-use efficiency and harvest index in peanut (*Arachis hypogaea*). Ann. Applied Biol., 148: 165-172.
- Ludlow, M.M. and R.C. Muchow, 1990. A critical evaluation of traits for improving crop yield in water-limited environments. Adv. Agron., 43: 107-153.
- Maiti, R.K., P. Wesche-Ebeling, A. Núñez-Gonzalez and E. Sánchez-Arreola, 2002. Root System and Mineral Nutrition. In: The Peanut (*Chis hypogaea* op, Maiti, R.K. and P. Wesche-Ebeling (Eds.). Science Publishers Inc., Enfield, NH, USA., ISBN: 157808-232, pp: 125-146.
- McPhee, K., 2005. Variation for seedling root architecture in the core collection of pea germplasm. Crop Sci., 45: 1758-1763.
- Nageswara Rao, R.C., J.H. Williams and M. Singh, 1989. Genotypic sensitivity to drought and yield potential of peanut. Agron. J., 81: 887-893.
- Nageswara Rao, R.C., L.J. Reddy, V.K. Mehan, S.N. Nigam and D. McDonald, 1992. Drought research on groundnut at ICRISAT. Proceedings of the International Work Shop, Groundnut-a Global Perspective, Nov. 25-29, ICRISAT Center Andhra Pradesh, India, pp: 455-455.
- Nageswara Rao, R.C., H.S. Talwar and G.C. Wright, 2001. Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using chlorophyll meter. J. Agron. Crop Sci., 189: 175-182.
- Nautiyal, P.C., R.C. Nageswara Rao and Y.C. Joshi, 2002. Moisture-deficit-induced changes in leaf-water content, leaf carbon exchange rate and biomass production in groundnut cultivars differing in specific leaf area. Field Crops Res., 74: 67-79.
- Nigam, S.N., M.S. Basu and A.W. Cruickshank, 2003. Hybridization and description of the trait-based and empirical selection programs. Proceedings of the International Workshop on Breeding for Drought-Resistant Peanuts, Feb. 25-27, ICRISAT Centre andhra Pradesh, India, ACIAR, Canberra, Australia, pp: 15-17.
- Nigam, S.N., S. Chandra, K. Rupa Sridevi, Manohar Bhukta and A.G.S. Reddy *et al.*, 2005. Efficiency of physiological trait-based and empirical selection approaches for drought tolerance in groundnut. Ann. Applied Biol., 146: 433-439.
- Nigam, S.N. and R. Aruna, 2008. Stability of soil plant analytical development (SPAD) chlorophyll meter reading (SCMR) and specific leaf area (SLA) and their association across varying soil moisture stress conditions in groundnut (*Arachis hypogaea* L.). Euphytica, 160: 111-117.
- Ntare, B.R. and J.H. Williams, 1998. Heritability of components of a simple physiological model for yield in groundnut under semiarid rainfed conditions. Field Crops Res., 58: 25-33.
- O'Toole, J.C. and W.C. Bland, 1987. Genotypic variation in crop plant root systems. Adv. Agron., : 91-145.
- Painawadee, M., S. Jogloy, T. Kesimal, C. Akkasaeng and A. Patanothai, 2009. Identification of traits related to drought resistance in peanut (*Arachis hypogaea* L.). Asian J. Plant Sci., 8: 120-128.
- Passioura, J.B., 1983. Roots and drought resistance. Agric. Water Manage., 7: 256-280.
- Pimratch, S., S. Jogloy, N. Vorasoot, B. Toomsan, A. Patanothai and C.C. Holbrook, 2008. Relationship between biomass production and nitrogen fixation under drought stress conditions in peanut genotypes with different levels of drought resistance. J. Agron. Crop Sci., 194: 15-25.
- Pirdashti, H., Z.T. Sarvestani and M.A. Bahmanyar, 2009. Comparison of physiological responses among four contrast rice cultivars under drought stress conditions. PWASET, 37: 52-53.
- Reddy, T.Y., V.R. Reddy and V. Anbumozhi, 2003. Physiological responses of groundnut (*Arachis hypogaea* L.) to drought stress and its amelioration: A critical review. Plant Growth Regul., 41: 75-88.
- Richardson, A.D., S.P. Duigan and G.P. Berlyn, 2002. An evaluation of noninvasive methods to estimate foliar chlorophyll content. New Phytologist, 153: 185-194.
- Sasmal, B., 2008. Relationship of root and shoot characters in parent, F1 and F2 populations of rice. J. Agron. Crop Sci., 159: 260-263.

- Serraj, R., L. Krishnamurthy, J. Kashiwagi, J. Kumar, S. Chandra and J.H. Crouch, 2004. Variation in root traits of chickpea (*Cicer arietinum* L.) grown under terminal drought. *Field Crops Res.*, 88: 115-127.
- Sheshshayee, M.S., H. Bindumadhava, N.R. Rachaputi, T.G. Prasad, M. Udayakumar, G.C. Wright and S.N. Nigam, 2006. Leaf chlorophyll concentration relates to transpiration efficiency in peanut. *Ann. Applied Biol.*, 148: 7-15.
- Siddique, K.H.M., R.K. Belford and D. Tennant, 1990. Root:shoot of old and modern, tall and semi-dwarf wheat in mediterranean environment. *Plant and Soil*, 121: 89-98.
- Singh, M., S. Ceccarelli and J. Hamblin, 1993. Estimation of heritability from varietal trials data. *Theor. Applied Genet.*, 86: 437-441.
- Smith, J.D. and M.L. Kinman, 1965. The use of parent-offspring regression as an estimator of heritability. *Crop Sci.*, 5: 595-596.
- Songsri, P., S. Jogloy, N. Vorasoot, C. Akkasaeng, A. Patanothai and C.C. Holbrook, 2008a. Root distribution of drought-resistant peanut genotypes in response to drought. *J. Agron. Crop Sci.*, 194: 92-103.
- Songsri, P., S. Jogloy, T. Kesmla, N. Vorasoot, C. Akkasaeng, A. Patanothai and C.C. Holbrook, 2008b. Response of reproductive characters of drought resistant peanut genotypes to drought. *Asian J. Plant Sci.*, 7: 427-439.
- Songsri, P., S. Jogloy, T. Kesmla, N. Vorasoot, C. Akkasaeng, A. Patanothai and C.C. Holbrook, 2008c. Heritability of drought resistance traits and correlation of drought resistance and agronomic traits in peanut. *Crop Sci.*, 48: 2245-2253.
- Songsri, P., S. Jogloy, C.C. Holbrook, T. Kesmla, N. Vorasoot, C. Akkasaeng and A. Patanothai, 2009. Association of root, specific leaf area and SPAD chlorophyll meter reading to water use efficiency of peanut under different available soil water. *Agric. Water Manage.*, 96: 790-798.
- Turner, N.C., G.C. Wright and K.H.M. Siddique, 2001. Adaptation of grain legumes (pulses) to water limited environments. *Adv. Agron.*, 71: 193-231.
- Upadhyaya, H.D., 2005. Variability for drought resistance related traits in the mini core collection of peanut. *Crop Sci.*, 45: 1432-1440.
- Wright, G.C. and R.C. Nageswara Rao, 1994. Groundnut Water Relations. In: *The Groundnut Crop: A Scientific Basis for Improvement*, Smartt, J. (Ed.). Chapman and Hall, London, ISBN: 978-0412408205, pp: 281-325.
- Wright, G.C., R.C. Nageswar-Rao and M.S. Basu, 1996. A Physiological Approach to the Understanding of Genotype by Environment Interaction: A Case Study on Improvement of Drought Adaptation in Groundnut. In: *Plant Adaptation and Crop Improvement*, Cooper, M. and G.L. Hammer (Ed.). CAB International, Wallingford, UK., ISBN: 0-85199-108-4.
- Wunna, H., S. Jogloy, B. Toomsan and J. Sanitchon, 2009. Response to early drought for traits related to nitrogen fixation and their correlation to yield and drought tolerance traits in peanut (*Arachis hypogaea* L.). *Asian J. Plant Sci.*, 8: 138-145.