

Asian Journal of Plant Sciences

ISSN 1682-3974





∂ OPEN ACCESS

Asian Journal of Plant Sciences

ISSN 1682-3974 DOI: 10.3923/ajps.2022.339.346



Research Article Agrophysiological Variation in Flax Affected by Folic Acid and Ag Nanoparticles Foliar Applications

¹Mervat Sh. Sadak, ²Bakry Ahmed Bakry and ²Mohamed Farouk El-Karamany

¹Department of Botany, Agricultural and Biological Research Institute, National Research Centre, 33 El Bohouth St., P.O. 12622, Dokki, Giza, Egypt ²Department of Field Crops Research, Agricultural and Biological Research Institute, National Research Centre, 33 El Bohouth St., P.O. 12622, Dokki, Giza, Egypt

Abstract

Background and Objective: Nanotechnology can be used to improve food security by increasing crop production. Nanoparticles are tiny particles that have the potential to boost the growth and yield of various plants including the flax plant. So, the present work was conducted to evaluate the efficiency of foliar application of folic acid, Ag nanoparticles on improving the growth, yield quantity and quality of flax plants under sandy soil conditions. **Materials and Methods:** Thus, two field experiments were carried out at the experimental station of the National Research Centre, Al-Nubaria district El-Behira Governorate, Egypt, in the 2019-2020 and 2020-2021 winter seasons. Foliar application of different concentrations of folic acid at rates of (50, 100 and 150 ppm) and Ag NPs (0.0, 20, 40 and 60 ppm) was carried out twice. **Results:** The results indicating that all the applied treatments showed highly significant variations in all the investigated growth parameters (shoot length, shoot fresh and dry weight, root length and fresh weight). There were significant increases in all biochemical parameters due to all concentrations used in all treatments. **Conclusion:** The promoting effect reached maximum at 100 mg L⁻¹ of folic acid and 40 mg L⁻¹ of Ag NPs foliar application and for all biochemical characters and yield quantity and quality.

Key words: Flax, folic acid, Ag-nanoparticles, productivity, quality, sandy soil, vegetable oils

Citation: Sadak, M.S., B.A. Bakry and M.F. El-Karamany, 2022. Agrophysiological variation in flax affected by folic acid and Ag nanoparticles foliar applications. Asian J. Plant Sci., 21: 339-346.

Corresponding Author: Mohamed Farouk El-Karamany, Department of Field Crops Research, Agricultural and Biological Research Institute, National Research Centre, 33 El Bohouth St., P.O. 12622, Dokki, Giza, Egypt

Copyright: © 2022 Mervat Sh. Sadak *et al.* This is an open access article distributed under the terms of the creative commons attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Flax (Linum usitatissimum L.) is one of the important plants, its seeds show a very high antioxidant activity and are increasingly proposed as an important source of oil and antioxidants. The flax plant has been grown as a dual-purpose plant (as a fibre and oil source) in many countries as Egypt^{1,2}. Regarding the national economy, flax plays a prominent role through its large possibilities of exportation and fabrication^{3,4}. The need for traditional edible oils increased due to population growth over the world and an increase in the demand for the plant. Thus it is required to find new renewable resources of vegetable oils. It can be mentioned that flax seeds produce a vegetable oil known as flaxseed or linseed oil. Oil quality is usually valued according to the content of Essential Fatty Acids (EFAs) as α-linolenic (omega-3) to linoleic (omega-6) unsaturated fatty acids, and it is among the biggest suppliers of omega-3⁵.

Folic acid (vitamin B₉) is one of the most prominent B-complex vitamins, it is a water-soluble vitamin. Folic acid is active in plants in its reduced form as tetra-hydro-folic acid (tetra-hydro-folate) and tetra-hydro-folic coenzymes⁶⁻⁸. These folic acid derivatives have different functions, in photosynthesis⁹, biochemical conversions of nitrogen, carbon, and sulfur as well as synthesis and catabolism of protein amino acids¹⁰ and nucleic acids^{11,12}. Emam *et al.*¹³ stated that vitamin treatments did not only stimulate oil production but also activated the antioxidative properties of flax seeds in terms of increasing the endogenous contents of glutathione, ascorbic acid and total phenols. However, the observed stimulation of oil production was found to be at the expense of carbohydrate and protein accumulation in vitamin-treated flax plants.

Nowadays nanotechnology uses are increasing in various fields like information technology, energy, the medical sector, and agriculture. Nanotechnology has proved its ability to solve problems in agriculture and related industries. The use of nanotechnology in agriculture is expected to have many environmental benefits¹⁴. Many studies on NPs release have risen on how the release would affect ecosystem health and human safety^{15,16}. Increasing nanoparticles (ENPs) are exploited in most applications and products due to their physico chemistry. Pathway of ENPs entry into plants mainly via stomata or cuticle¹⁷, ENPs have a positive or negative effect. It has been reported that ENPs can play a role in enhancing energy harvesting for photosynthesis, electron transport, photoreduction activity of PSII, and oxygen evolution^{18,19}. Under certain conditions, plants can generate natural mineralized nanoparticles required for plant growth²⁰. Silver nanoparticle Ag NPs are the most commonly used manufactured nanoparticles in a wide range of commercial products²¹. Ag NPs have been linked to crop yield improvement in agriculture. Several studies have shown that suitable concentrations of Ag NPs have an important role in improving seed germination^{22,23} plant growth²⁴⁻²⁶, enhancing photosynthetic quantum efficiency and chlorophyll content^{24,27}, and increasing water and fertilizer use efficiency²⁸.

However, the present work was conducted to evaluate the efficiency of foliar application of folic acid, Ag nanoparticles on improving the growth, yield quantity and quality of flax plants.

MATERIALS AND METHODS

Study area: Two field experiments were carried out at the experimental station of the National Research Centre, Al-Nubaria district El-Behira Governorate-Egypt, in the 2019/2020 and 2020/2021 winter seasons. The soil of the experimental site was sandy. Mechanical, chemical and nutritional analysis of the experimental soil is reported in Table 1 according to Carter and Gregorich²⁹.

Methodology: The experimental design was a complete randomized block design with 3 replications, to investigate the effect of foliar application with 3 levels of folic acid at rates of (50, 100 and 150 mg L⁻¹) and Ag NPs (0.0, 20, 40 and 60 mg L⁻¹) on growth, seed yield and yield components of flax (Letwania-9) cultivar.

Flax seeds of Letwania-9 cultivar were sown in the mid of November, 2019/2020 and 2020/2021 winter seasons in rows 3.5 m long, and the distance between rows was 20 cm apart, plot area was 10.5 m² (3.0 m in width and 3.5 m in length). The seeding rate was 2000 seeds m⁻². Pre-sowing, 150 kg per fed of calcium super-phosphate (15.5% P₂O₅) were used. Nitrogen was applied after emergence in the form of ammonium nitrate 33.5% N at a rate of 75 kg per fed in 5 equal doses. Potassium sulfate (48% K₂O) was added at 2 equal doses of 50 kg per fed. Irrigation was carried out using the new sprinkler irrigation system where water was added every 5 days. Foliar application of different concentrations of Ag NPs (0.0, 20, 40 and 60 ppm) and folic acid at rates of (50, 100 and 150 ppm) was carried out twice, where plants were sprayed after 45 and 60 days from sowing. Water deficit (by skipping one irrigation after the 2 foliar applications) aiming to avert spray treatments washing. Plant samples were taken after 75 days from sowing for measurements of growth characters and some biochemical parameters.

Sand											
Course 2000-200 (μ %) Fine 200-20 (μ %)			Silt 20-0 (µ %)		Clay<2 (μ %)				Soil texture		
47.46		36.19		12.86			4.28				Sandy
Chemical an	alysis										
				Soluble o	cations (meq l	L ⁻¹)		Soluble	anions (meq	L ⁻¹)	
pH 1:2.5	EC (dSm ⁻¹)	CaCo ₃	OM (%)	Na ⁺	K+	Mg ⁺	Ca++	CO3-	HCO₃ [−]	CI-	SO ₄ -
7.60	0.13	5.3	0.06	0.57	0.13	0.92	1.0	0.0	1.25	0.48	0.89
Nutritional a	inalysis/Available r	outrients									
Macro eleme	ent (ppm)			Micro ele	ement (ppm)						
N	Р		K	Zn		Fe			Мn		Cu
52	12.0		75	0.14		1.4			0.3		0.00

52 12.0 75 0.14 Growth traits of plant height (cm), shoot fresh and dry weight (g)/plant, root length (cm), fresh and dry (g)/plant, were measured. At harvest, flax plants were harvested when signs of full maturity appeared, after removing capsules carefully, the following flax traits were measured, plant height (cm), fruiting zone length (cm), technical stem length (cm), number of fruiting branches/plant, number of capsules/plant, biological yield/plant (g), seed yield/plant (g), and 1000-seeds wt. (g), were recorded on random samples of ten guarded p l ants in each plot. Also, straw yield (ton per fed), seed yield (kg per fed) and biological yield (ton per fed) and oil yield (kg per fed) were studied. Some biochemical analysis of the seed yield was done such as oil (%), carbohydrates (%).

Table 1: Some physical and chemical characteristics of the experimental soil

Chemical analysis: Photosynthetic pigments contents (chlorophyll a, b and carotenoids) in fresh leaves were estimated using the method of Lichtenthaler and Buschmann³⁰. Indole acetic acid content was extracted and analyzed using the method of Hansen and Halkier³¹. Total phenol content was measured as described by William and Metzger³². Seed oil content was determined using Soxhlet apparatus and petroleum ether (40-60°C) according to Das *et al.*³³. Carbohydrates were determined calorimetrically according to the method of Pak and Simon³⁴.

Statistical analysis: Combined analysis of variance of complete randomized block design for the substances and their concentrations across the two seasons were performed using the method described by Lawal³⁵. since the trend was similar in both seasons the homogeneity test Bartlett's equation was applied and the combined analysis of the

two seasons was done according to the method³⁶. Means were compared by using the Least Significant Difference (LSD) at 5 and 1%.

Feddan = 4200 m^2 (Local area meter)

RESULTS

Effect on growth parameters: Folic acid (50, 100 and 150 mg L⁻¹) and silver nanoparticle Ag Nps (20, 40 and 60 mg L⁻¹) with all concentrations showed highly significant increases in growth parameters of flax plant such as shoot length, shoot fresh and dry weight, root length and fresh weight except root length treatment with 50 mg L⁻¹ folic acid and 20 mg L⁻¹ Ag NPs the increases were not significant as compared with control plants as shown in Table 2. Regarding treatment with folic acid 100 mg L⁻¹ was the most effective concentration on all the studied parameters compared with the other used concentrations. While 40 mg L⁻¹ was the most effective concentration of Ag NPs compared with the other Ag NPs concentrations. Folic acid with 100 mg L⁻¹ showed the highest significant increases in shoot length (68.67 cm), shoot fresh weight (4.37 g), shoot dry weight (0.59 g). While Ag NPs showed the highest increases in root length (13.67 cm) and root fresh weight (0.49 g) compared with other treatments.

Photosynthetic pigments, IAA and phenolics: Regarding photosynthetic pigments of flax plants grown under drought stress in sandy soil is represented in Table 3. Data show that folic acid foliar application at rates of (0, 50, 100, 150 mg L⁻¹) or Ag NPs foliar application at rates of (0, 20, 40, 60 mg L⁻¹) caused significant increases in chlorophyll a, chlorophyll b, carotenoids and total pigments, IAA and phenol contents of flax plants. Regarding foliar application with folic acid

Asian J. Plant Sci., 21 (2): 339-346, 2022

	Shoots	Roots				
Treatments (mg L ⁻¹)	 Length (cm)	Fresh weight (g)	Dry weight (g)	 Length (cm)	Fresh weight (g)	
Control	56.33	2.14	0.29	11.00	0.19	
Folic acid						
50	65.33	3.07	0.42	12.00	0.27	
100	68.67	4.37	0.59	13.33	0.48	
150	66.67	4.09	0.56	13.33	0.37	
Ag NPs						
20	64.00	2.95	0.40	11.67	0.36	
40	67.67	4.00	0.54	13.67	0.49	
60	63.33	3.29	0.45	11.67	0.49	
LSD _{0.05}	2.02	0.28	0.04	0.79	0.05	
LSD _{0.01}	3.90	0.53	0.07	1.53	0.09	

Table 2: Effect of folic acid and Ag NF	s foliar applications le	vels on growth parameters	of flax plants at 75 day	/s after sowing combined of two seasons
---	--------------------------	---------------------------	--------------------------	---

Table 3: Effect of Ag NPs and folic acid foliar applications levels on biochemical characters of flax plants at 75 days after sowing combined of two seasons

Photosynthetic pigments ($\mu q q^{-1}$ fresh weight)

Treatments (mg L ⁻¹)	Chlorophyll a	Chlorophyll b	Carotenoids	Total pigments	IAA (μg/100 g FW)	Phenolic (mg/100 g FW
Control	1009.7	529.3	249.9	1788.9	22.92	64.29
Folic acid						
50	1377.5	626.0	286.5	2290.0	25.68	84.79
100	1760.4	654.9	338.8	2754.1	47.73	106.68
150	1480.1	606.7	321.3	2408.2	41.58	86.90
AgNPs						
20	1177.2	620.2	277.0	2074.4	34.14	75.31
40	1454.3	659.4	319.9	2433.6	43.93	87.51
60	1409.8	681.9	282.5	2374.1	37.36	100.21
LSD _{0.05}	18.3	14.5	3.5	39.5	0.50	1.12
LSD _{0.01}	36.0	28.10	6.70	77.66	0.98	2.17

100 mg L⁻¹ gave the highest values of chlorophyll a, chlorophyll b, carotenoids, total pigment, IAA and phenolics contents compared with the other concentrations (50 and 150 mg L⁻¹). While, the highest promotive effect of Ag NPs was obtained from foliar treatment with 40 mg L⁻¹, in chlorophyll a, carotenoids, total pigments, IAA and phenolics. While 60 mg L⁻¹ gave the highest increases in chlorophyll b content of flax plants. Moreover, Folic acid with 100 mg L⁻¹ was the most effective treatment followed by 40 mg L⁻¹ Ag NPs compared with control plants and the other treated plants.

Change in yield and yield components: Table 4 and 5 show the effect of foliar application at different concentrations of folic acid or Ag NPS on yield parameters of flax plants grown under the newly reclaimed sandy soil. Data clearly show that foliar application with different levels increased significantly yield and its components such as plant height (cm), fruiting zone length (cm), technical stem length (cm), number of fruiting branches/plant, number of capsules/plant, biological yield/plant (g), seed yield/plant (g), straw yield (ton per fed), seed yield (kg per fed), biological yield (ton per fed), 1000 seeds weight (g), oil (%), carbohydrates (%) and oil yield (per fed). Folic acid with 100 and Ag NPs 40 mg L⁻¹ were the most effective treatments compared with the other used concentrations. The increase in yield components of flax in response to different treatments relative to untreated plants might result from the increased number of fruiting branches/plant, the number of capsules/plant and 1000 seed weight (g).

Rank correlation coefficients: Biological yield (ton per fed), straw yield (ton per fed) and/ or seed yield (kg per fed) of flax showed a very strong and positive correlation with other traits and yield components, namely, shoot length (cm), shoot fresh wt. (g), shoot dry wt. (g), root length (cm), root fresh wt. (g), chlorophyll a, chlorophyll b, total pigments, IAA, phenolic, technical stem length (cm), fruiting zone length (cm), plant height (cm), no. of fruiting branches/plant, no. of capsules/ plant, 1000 seeds wt. straw yield (ton per fed) and seed yield (kg per fed) showed significant and positive ($p \le 0.05$) correlation with fruiting zone length (cm),

Asian J. Plant Sci., 21 (2): 339-346, 2022

Table 4: Effect of Ag NPs and folic acid foliar applications on seed yield and yield components of flax plants grown under sandy soil conditions combined of two seasons

Treatments	Plant	Fruiting zone	Technical stem	No. of fruiting	No. of	Biological	Seed yield/
(mg L ⁻¹)	height (cm)	length (cm)	length (cm)	branches/plant	capsules/plant	yield/plant (g)	plant (g)
Control	66.00	13.33	52.67	18.00	33.00	0.87	2.83
Folic acid							
50	73.33	15.00	58.33	23.67	35.00	1.08	3.45
100	75.00	15.67	59.33	27.33	38.33	1.38	4.07
150	70.33	17.00	53.33	26.33	34.33	1.41	3.12
Ag NPs							
20	71.67	17.33	54.33	26.33	32.67	1.01	3.22
40	77.00	20.33	56.67	21.00	42.67	2.64	4.16
60	75.00	20.33	54.67	28.00	39.67	1.65	3.04
LSD _{0.05}	2.57	1.07	2.05	1.64	1.37	0.09	0.22
LSD _{0.01}	4.97	2.08	3.96	3.18	2.65	0.18	0.43

Table 5: Effect of Ag NPs and folic acid foliar applications on seed yield and yield components of flax plants grown under sandy soil conditions combined of two seasons

Treatments	Straw yield	Seed yield	Biological yield	1000-seeds	Oil	Carbohydrates	Oil yield
(mg L ⁻¹)	(ton per fed)	(kg per fed)	(ton per fed)	weight (g)	(%)	(%)	(kg per fed)
Control	1.68	347.92	2.03	4.79	32.79	29.43	114.08
Folic acid							
50	2.49	527.39	3.01	5.06	34.01	31.01	179.38
100	2.92	660.73	3.58	5.80	35.73	32.83	236.10
150	2.32	533.56	2.86	5.04	34.65	31.65	184.86
Ag NPs							
20	2.57	472.29	3.04	5.19	33.79	31.12	159.59
40	2.96	584.51	3.54	6.04	35.79	32.46	209.18
60	2.15	495.10	2.65	5.12	34.72	32.82	171.91
LSD _{0.05}	0.01	0.01	0.01	0.13	0.31	0.87	1.70
LSD _{0.01}	0.02	0.02	0.02	0.25	0.43	1.19	2.34

Table 6: Rank correlation coefficients between biological yield, straw yield and/ or seed yields per fed with other studied traits for flax

Traits	Straw yield (ton per fed)	Seed yield (kg per fed)	Biological yield (ton per fed
Shoot length (cm)	0.79**	0.77**	0.80**
Shoot fresh wt. (g)	0.67**	0.73**	0.69**
Shoot dry wt. (g)	0.67**	0.73**	0.69**
Root length (cm)	0.55**	0.61**	0.57**
Root fresh wt. (g)	0.67**	0.84**	0.71**
Chlorophyll a	0.74**	0.69**	0.75**
Chlorophyll b	0.75**	0.81**	0.77**
Carotenoids	0.73**	0.74**	0.74**
Total pigments	0.78**	0.75**	0.79**
IAA	0.67**	0.83**	0.71**
Phenolic	0.66**	0.65*	0.67**
Technical stem length (cm)	0.59**	0.63**	0.61**
Fruiting zone length (cm)	0.38*	0.33	0.38*
Plant height (cm)	0.68**	0.68**	0.69**
No. of fruiting branches/plant	0.33	0.49**	0.36
No. of capsules/plant	0.52**	0.57**	0.54**
Biological yield/plant (g)	0.56**	0.51**	0.56**
Seed yield/plant (g)	0.85**	0.79**	0.86**
Straw yield (ton per fed)	-	0.89**	1.00**
Seed yield (kg per fed)	0.89**	-	0.92**
Biological yield (ton per fed)	1.00**	0.92**	-
1000-seeds wt. (g)	0.85**	0.77**	0.85**

*Significant at 0.05 and **Significant at 0.00, combined data of 2 seasons

seed yield (kg per fed) showed very strong and positive $(p \le 0.01)$ correlation with no. of fruiting branches/plant.

On the contrary, biological yield (ton per fed) and or straw yield (ton per fed) showed negative correlations with number

of fruiting branches/plant, seed yield (kg per fed) also showed negative correlation with fruiting zone length Table 6. This indicates the importance of these traits in the yield and yield component of flax plants under sandy soil conditions.

DISCUSSION

The promotive effect of different treatments of either folic acid or Ag NPs on the growth and development of flax plants illustrate their important role in biochemical and physiological processes in plant cells. Folic acid treatments enhance agronomic performance and the other growth traits of the flax plant (Table 2). Dawood et al.³⁷ and Al-Maliky et al.³⁸ confirmed the positive role of folic acid on the growth traits of the faba bean plant. These increases in growth traits caused by folic acid could be related to its content of the most prominent of B complex vitamins besides its essential biochemical function in amino acid metabolism and nucleic acid synthesis¹², increasing cell division and expansion, biosynthesis of bioregulators as IAA and chlorophyll³⁹ as well as, nutrient absorption⁴⁰. The role of folic in such activities was demonstrated by an increase in the total vegetative growth as well as an increase in-plant weight and dry matter accumulation.

These results of Ag NPs effect on flax plant are in good harmony with those obtained by Latif *et al.*⁴¹, El-Batal *et al.*⁴² and Jurkow *et al.*⁴³ they found that treatment different plants with Ag NPs have a significant impact on growth parameters. Shelar and Chavan²³ stated that appropriate concentrations of Ag NPs play an important role in enhancing seed germination and plant growth. This enhancing effect of silver nanoparticles with different levels could be due to its effect in blocking ethylene signalling in flax plants⁴⁴. Moreover, the stimulating effect of Ag NPs on the growth and physiological attributes of the plant depends on the size and shape of the nanoparticle.

Exogenous treatment of folic acids caused significant increases in various constituents of photosynthetic pigment (Table 3). These increases may be ascribed with the function of folic acid as a central cofactor for one-carbon transfer reactions which are involved in many cellular reactions such as synthesis of chlorophyll and the photorespiration cycle⁴⁵. Moreover, folic coenzymes play a role in the biosynthesis of glycine, and an increased level of free glycine starts the development of plant porphyrins and their derivatives, chlorophylls¹⁰. In this regard, Dawood *et al.*³⁷ and Al-Maliky *et al.*³⁸ stated that foliar treatment of folic acid increased chlorophyll content in the faba bean plant.

Table 3 showed the significant increases in all photosynthetic pigment constituents of flax plants treated with Ag NPs with different concentrations. Those results incongruent with those of Farghaly and Nafady⁴⁶ and Latif *et al.*⁴¹ they concluded that Ag NPs improved photosynthesis and attributed these effects to the change of

nitrogen metabolism. Moreover, the increased contents of photosynthetic pigments constituents could increase the rate of photosynthesis, due to which there was more production of the photosynthesis process, which in turn increased the weight and growth and productivity of plant as it was observed in our study.

Foliar treatment of folic acid caused significant increases in yield and its components of flax plants grown under the sandy soil. These increases might be due ascribed to the increase in nutrient uptake and/or assimilation due to vitamin application⁴⁷. Stakhova *et al.*⁴⁸ stated that folic treatment promoted the formation of the dependent amino acids and increases the yield and quality of the seeds of pea (*Pisum sativum* L.) and barley (*Hordeum vulgare* L.). Moreover, strawberries, Li *et al.*⁴⁹ stated that folic acid is one of the most important micronutrients and has many forms, but the only folic acid form has cofactor activity. In this respect, a few pieces of literature reported that exogenous folic acid has a positive effect on the growth, yield and quality of some plants such as flax¹³, winter wheat⁵⁰ white button mushroom⁵¹.

Foliar treatment of flax plants with Ag NPs showed significant increases in yield and its components (Table 4 and 5). Silver is an excellent growth simulator⁵². Similar results of enhancing the role of Ag NPs treatments were obtained on mung bean by Najafi and Jamei⁵³ and Razzaq *et al.*⁵⁴ on the wheat plant. These increases in yield and yield components might be attributed to the increases in growth parameters, photosynthetic pigments, and IAA of treated flax plants.

CONCLUSION

This study showed the effect of foliar treatment of folic acid and Ag NPs on flax plants under sandy soil. Different concentrations increased plant growth, photosynthetic pigments, IAA, phenolics contents and yield quantity and quality. Among different concentrations, 100 and 40 mg L⁻¹ were the most effective treatments of folic acid and Ag NPs, respectively.

SIGNIFICANCE STATEMENT

This study discovers the possible synergistic effect of folic acid and Ag NPs foliar treatments that can be beneficial for increased plant growth, photosynthetic pigments, IAA, phenolics contents and yield quantity and quality. This study will help the researchers and farmers to maximize the flax productivity under sandy soil conditions by using 100 mg L⁻¹ of folic acid and 40 mg L⁻¹ of Ag NPs foliar application and for all biochemical characters and yield quantity and quality.

REFERENCES

- 1. El-Hariri, D.M., M.S. Hassanein and A.H.H. El-Sweify, 2004. Evaluation of same flax genotypes, straw yield, yield components and technological characters. J. Nat. Fibers, 1:1-12.
- 2. Bakry, A.B., T.A. Elewa and O.A.M. Ali, 2012. Effect of Fe foliar application on yield and quality traits of some flax varieties grown under newly reclaimed sandy soil. Aust. J. Basic Appl. Sci., 6: 532-536.
- 3. Khalifa, R.K.M., F.M. Manal, A.B. Bakry and M.S. Zeidan, 2011. Response of some flax varieties to micronutrients foliar application under newly reclaimed sandy soil. Aust. J. Basic Appl. Sci., 5: 1328-1334.
- Bakry, A.B., M.S. Sadak, M.M.S.A. Allah, T.M.A. El-Razik and M.G. Dawood, 2016. Maximizing the performance, productivity and quality traits of two flax cultivars by using some bio-fertilizers under newly reclaimed sandy soil. Res. J. Pharmaceut. Biol. Chem. Sci., 7: 429-441.
- Johnson, M., S. Ostlund, G. Fransson, B. Kadesjö and C. Gillberg, 2009. Omega-3/omega-6 fatty acids for attention deficit hyperactivity disorder: A randomized placebocontrolled trial in children and adolescents. J. Attention Disord., 12: 394-401.
- Nelson, D.L. and M.M. Cox, 2005. Lehninger: Principles of Biochemistry. 4th Edn., W.H. Freeman and Company, New York, USA, ISBN-13: 9780716743392, Pages: 1119
- Burguieres, E., P. McCue, Y.I. Kwon and K. Shetty, 2007. Effect of vitamin C and folic acid on seed vigour response and phenolic-linked antioxidant activity. Bioresour. Technol., 98: 1393-1404.
- 8. Bailey, S.W. and J.E. Ayling, 2009. The extremely slow and variable activity of dihydrofolate reductase in human liver and its implications for high folic acid intake. Proc. Nat. Acad. Sci., 106: 15424-15429.
- Grunert, R.R., A. Braune, E. Schnackenberg, W. Schloot and H. Krause, 2002. Genetic differences in enzymes of folic acid metabolism in patients with lip-jaw-palate clefts and their relatives. Mund Kiefer und Gesichtschirurgie, 6: 131-133.
- 10. Metzler, D.E. and C.M. Metzler, 2003. Biochemistry: The Chemical Reactions of Living Cells. Vol. 2. Academic Press, United States, ISBN-13: 978-0124925410, Pages: 1973.
- 11. Litwack, G., 2008. Folic Acid and Folates, Volume 79. 1st Edn., Academic Press, United States, ISBN-13: 9780123742322, Pages: 480.
- 12. El-Moghazy, T.F.A. and E.A.E.A.E. Al-Azzony, 2019. Effect of ascorbic, folic acids and hibiscus extract on geranium (*Pelargonium graveolens*). Am. J. Plant Biol., 4: 46-56.
- 13. Emam, M.M., A.H. El-Sweify and N.M. Helal, 2011. Efficiencies of some vitamins in improving yield and quality of flax plant. Afr. J. Agric. Res., 6: 4362-4369.
- 14. Abobatta, W.F., 2018. Nanotechnology application in agriculture. Acta Sci. Agric., 2: 99-102.

- 15. Meng, H., T. Xia, S. George and A.E. Nel, 2009. A predictive toxicological paradigm for the safety assessment of nanomaterials. ACS Nano, 3: 1620-1627.
- Klaine, S.J., P.J.J. Alvarez, G.E. Batley, T.F. Fernandes and R.D. Handy *et al.*, 2008. Nanomaterials in the environment: Behavior, fate, bioavailability and effects. Environ. Toxicol. Chem., 27: 1825-1851.
- 17. Eichert, T., A. Kurtz, U. Steiner and H.E. Goldbach, 2008. Size exclusion limits and lateral heterogeneity of the stomatal foliar uptake pathway for aqueous solutes and water-suspended nanoparticles. Physiol. Plant., 134: 151-160.
- 18. Lei, Z., S. Mingyu, L. Chao, C. Liang and H. Hao *et al.*, 2007. Effects of nanoanatase TiO_2 on photosynthesis of spinach chloroplasts under different light illumination. Biol. Trace Elem. Res., 119: 68-76.
- 19. Giraldo, J.P., M.P. Landry, S.M. Faltermeier, T.P. McNicholas and N.M. Iverson *et al.*, 2014. Plant nanobionics approach to augment photosynthesis and biochemical sensing. Nat. Mater, 13: 400-408.
- Siddiqui, M.H. and M.H. Al-Whaibi, 2014. Role of nano-SiO₂ in germination of tomato (*Lycopersicum esculentum* seeds Mill.). Saudi J. Biol. Sci., 21: 13-17.
- Zhang, X.F., Z.G. Liu, W. Shen and S. Gurunathan, 2016. Silver nanoparticles: Synthesis, characterization, properties, applications and therapeutic approaches. Int. J. Mol. sci., Vol. 17. 10.3390/ijms17091534.
- 22. Barrena, R., E. Casals, J. Colon, X. Font, A. Sanchez and V. Puntes, 2009. Evaluation of the ecotoxicity of model nanoparticles. Chemosphere, 75: 850-857.
- 23. Shelar, G.B. and A.M. Chavan, 2015. Myco-synthesis of silver nanoparticles from *Trichoderma harzianum* and its impact on germination status of oil seed. Biolife, 3: 109-113.
- 24. Sharma, P., D. Bhatt, M.G.H. Zaidi, P.P. Saradhi, P.K. Khanna and S. Arora, 2012. Silver nanoparticle-mediated enhancement in growth and antioxidant status of *Brassica juncea*. Appl. Biochem. Biotechnol., 167: 2225-2233.
- 25. Kaveh, R., Y.S. Li, S. Ranjbar, R. Tehrani, C.L. Brueck and B. Van Aken, 2013. Changes in *Arabidopsis thaliana* gene expression in response to silver nanoparticles and silver ions. Environ. Sci. Technol., 47: 10637-10644.
- Vannini, C., G. Domingo, E. Onelli, B. Prinsi, M. Marsoni, L. Espen and M. Bracale, 2013. Morphological and proteomic responses of *Eruca sativa* exposed to silver nanoparticles or silver nitrate. PLoS ONE, Vol. 8. 10.1371/journal.pone.0068752.
- 27. Hatami, M. and M. Ghorbanpour, 2013. Effect of nanosilver on physiological performance of pelargonium plants exposed to dark storage. J. Hortic. Res., 21: 15-20.
- 28. Lu, C., C. Zhang, J. Wen, G. Wu and M. Tao, 2002. Research of the effect of nanometer materials on germination and growth enhancement of *Glycine max* and its mechanism. Soybean Sci., 21: 168-171.

- 29. Carter M.R. and E.G. Gregorich, 2007. Soil Sampling and Methods of Analysis. 2nd Edn., Taylor and Francis Group, Pages: 1264.
- Lichtenthaler, H.K. and C. Buschmann, 2001. Chlorophylls and Carotenoids: Measurement and Characterization by UV-VIS Spectroscopy. In: Current Protocols in Food Analytical Chemistry, Wrolstad, R.E., T.E. Acree, H. An, E.A. Decker and M.H. Penner *et al.* (Eds.)., John Wiley and Sons, New York, USA, pp: F4.3.1-F4.3.8.
- 31. Hansen, B.G. and B.A. Halkier, 2005. New insight into the biosynthesis and regulation of indole compounds in *Arabidopsis thaliana*. Planta, 221: 603-606.
- 32. Finch-Savage, W.E. and G. Leubner-Metzger, 2006. Seed dormancy and the control of germination. New Phytol., 171: 501-523.
- Das, M., S.K. Das and S.H. Suthar, 2002. Composition of seed and characteristics of oil from karingda [*Citrullus lanatus* (Thumb) Mansf]. Int. J. Food Sci. Technol., 37: 893-896.
- 34. Chow, P.S. and S.M. Landhausser, 2004. A method for routine measurements of total sugar and starch content in woody plant tissues. Tree Physiol., 24: 1129-1136.
- 35. Lawal, B., 2014. Applied Statistical Methods in Agriculture, Health and Life Sciences. 1st Edn., Springer International Publishing, Switzerland, ISBN-13: 978-3-319-37834-3, Pages: 799.
- de Smith, M.J., 2018. Statistical Analysis: A Comprehensive Handbook of Statistical Concepts, Techniques and Software Tools. 1st Edn., The Winchelsea Press, United Kingdom, ISBN-13: 978-1-912556-08-3, Pages: 99.
- Dawood, M.G., M.E. El-Awadi, K.M.G. El-Din and M.S.S.M.A. Shalaby, 2019. Improving performance and quality of faba bean plant via folic acid and α-tocopherol application. World Rural Obs., 11: 20-28.
- 38. Al-Maliky, A.W.A., A.N. Jerry and F.I. Obead, 2019. The effects of foliar spraying of folic acid and cysteine on growth, chemical composition of leaves and green yield of faba bean (*Vicia faba* L.). Basrah J. Agric. Sci., 32: 223-229.
- Jabrin, S., S. Ravanel, B. Gambonnet, R. Douce and F. Rébeillé, 2003. One-carbon metabolism in plants. Regulation of tetrahydrofolate synthesis during germination and seedling development. Plant Physiol., 131: 1431-1439.
- 40. Kilic, S. and H.T. Aca, 2016. Role of exogenous folic acid in alleviation of morphological and anatomical inhibition on salinity-induced stress in barley. Ital. J. Agron., 11: 246-251.
- 41. Latif, H.H., M. Ghareib and M.A. Tahon, 2017. Phytosynthesis of silver nanoparticles using leaf extracts from *Ocimum basilicum* and *Mangifira indica* and their effect on some biochemical attributes of *Triticum aestivum*. Gesunde Pflanzen, 69: 39-46.

- El-Batal, A.I., F.A.E.L. Gharib, S.M. Ghazi, A.Z. Hegazi and A.G.M.A.E. Hafz, 2016. Physiological responses of two varieties of common bean (*Phaseolus vulgaris* L.) to foliar application of silver nanoparticles. Nanomater. Nanotechnol., Vol. 6. 10.5772/62202.
- Jurkow, R., R. Pokluda, A. Sękara and A. Kalisz, 2020. Impact of foliar application of some metal nanoparticles on antioxidant system in oakleaf lettuce seedlings. BMC Plant Biol., Vol. 20. 10.1186/s12870-020-02490-5.
- 44. Rezvani, N., A. Sorooshzadeh and N. Farhadi, 2012. Effect of nano-silver on growth of saffron in flooding stress. World Acad. Sci. Eng. Technol., 1: 517-522.
- 45. Hanson, A.D. and S. Roje, 2001. One-carbon metabolism in higher plants. Annu. Rev. Plant Physiol. Plant Mol. Biol., 52: 119-137.
- 46. Farghaly, F.A. and N.A. Nafady, 2015. Green synthesis of silver nanoparticles using leaf extract of *Rosmarinus officinalis* and its effect on tomato and wheat plants. J. Agric. Sci., 7: 277-287.
- 47. Abbas, Z., A.K. Tiwari and P. Kumar, 2018. Emerging Trends of Plant Physiology for Sustainable Crop Production. 1st Edn., Apple Academic Press, Boca Raton, ISBN-13: 9781315101224, Pages: 414.
- Stakhova, L.N., L.F. Stakhov and V.G. Ladygin, 2000. Effects of exogenous folic acid on the yield and amino acid content of the seed of *Pisum sativum* L. and *Hordeum vulgare* L. Appl. Biochem. Microbiol., 36: 85-89.
- Li, D., L. Li, Z. Luo, W. Mou, L. Mao and T. Ying, 2015. Comparative transcriptome analysis reveals the influence of abscisic acid on the metabolism of pigments, ascorbic acid and folic acid during strawberry fruit ripening. PLoS ONE, Vol. 10. 10.1371/journal.pone.0130037.
- 50. Vician, M. and P. Kovacik, 2013. The effect of folic application of mg-titanit fertilizer on phytomass, chlorophyll production and the harvest of winter wheat. Mendelnet, 3: 162-168.
- 51. Dahmardeh, M., Z. Poodineh and B.A. Fakheri, 2015. Effects of humic and folic acid on quantity and quality related traits of button mushroom (*Agaricus bisporus*). Biol. Forum Int. J., 7:823-828.
- 52. Sharon, M., A.K. Choudhary and R. Kumar, 2010. Nanotechnology in agricultural disease and food safety. J. Phytol., 2: 83-92.
- 53. Najafi, S. and R. Jamei, 2014. Effect of silver nanoparticles and Pb (NO₃)₂ on the yield and chemical composition of mung bean (*Vigna radiate*). J. Stress Physiol. Biochem., 10: 316-325.
- 54. Razzaq, A., R. Ammara, H.M. Jhanzab, T. Mahmood, A. Hafeez and S. Hussain, 2016. A novel nanomaterial to enhance growth and yield of wheat. J. Nanosci. Technol., 2: 55-58.