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Research Article Mycorrhiza Modulates Morphology, Color and Duration of Flowers in Hyacinth

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Abstract

Background and Objective: Hyacinth, one of world-famous bulbous ornamental plants is widely planted in various habitats. The present work tried to evaluate whether and how inoculation with arbuscular mycorrhizal fungi (AMF) regulates flower morphology and flowering duration of hyacinth. **Materials and Methods:** Three AMF species, namely, *Diversispora spurca* (*D. spurca*), *Diversispora versiformis* (*D. versiformis*) and *Funneliformis mosseae* (*F. mosseae*) were inoculated into potted hyacinth (*Hyacinths orientalis* L. Anna Marie) plants for 130 days. Flower morphology, flowering duration, nutrient status, indole acetic acid (IAA), chlorophyll and anthocyanin concentration were measured. Data were analyzed by one-way analysis of variance (ANOVA) with SAS. Significances of treatments were compared by the Duncan's multiple range tests at p<0.05. **Results:** Root mycorrhizal colonization varied from 38.0-48.7%. Amongst three AMF species, only *F. mosseae* significantly (p<0.05) has increased flower biomass, opening flower number, flower stem height and floret diameter. *Diversispora spurca* and *F. mosseae*, respectively prolonged flower duration for 1.4 and 3.3 days. Mycorrhizal plants with *F. mosseae* had significantly (p<0.05) higher IAA concentration of flowers and roots, higher anthocyanin concentration of flowers, higher chlorophyll b and total chlorophyll levels of leaves and greater N, P and K levels of flowers and roots. *Diversispora spurca* colonized plants possessed significantly (p<0.05) higher flower IAA levels and leaf chlorophyll b and total chlorophyll concentrations. *Diversispora spurca* colonized in hyacinth, which is closely related with nutrient status, IAA levels and AMF species. Meanwhile, *F. mosseae* exhibited the best stimulated effects and can consider to be used in hyacinth cultivation.

Key words: Arbuscular mycorrhiza, auxin, chlorophyll, Funneliformis mosseae, hyacinth, nutrient

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Competing Interest: The authors have declared that no competing interest exists.

Data Availability: All relevant data are within the paper and its supporting information files.

INTRODUCTION

Soil arbuscular mycorrhizal fungi (AMF) can form mutualistic symbiosis with upto 80% of land's plants, arbuscular mycorrhizas. Arbuscular mycorrhizal symbiosis characterizes with mutual benefits: Host plants supply AMF with 3~20% of photosynthates and in return AMF provides water and nutrients to host plants by developed hyphal netmork^{1,2}. Such mycorrhizal plants generally reproduce more successfully³, effectively improve transplant survival and growth⁴ and confer stronger resistance to abiotic and biotic stresses⁵. As a result, inoculation with AMF is regarded as an effecter to improve plant growth and development of ornamental and crop plants^{6,7}.

Hyacinth (Hyacinthus orientalis L.) is one of worldfamous bulbous ornamental plants, which is widely applied in potted plants, cut-flower and nature plantation⁸. Particularly, hyacinth is well known for its gorgeous flower color and powerful fragrance. In floriculture, AMF shows positive effects on flower quality, flower growth and flowering time. Asrar et al.9 reported that mycorrhizal inoculation with Glomus deserticola enhanced both flower biomass and flower size in snapdragon (Antirhinum majus). Scagel¹⁰ reported that inoculation with AMF reduced flowering 7-8 days earlier in harlequin (Sparaxis tricolor). AMF increased the number of flowers in gerbera (Gerbera jamesonii)¹¹. These results indicate that mycorrhizas have potential effects on flower growth and flowering time of ornamental plants. However, the information regarding AMF effects on hyacinth plants is poorly known.

The purpose of the present study was to analyze the effects of different AMF species on flower quality, flowering duration, nutrient status, indole acetic acid (IAA), chlorophyll and anthocyanin concentration of potted hyacinth plants.

MATERIALS AND METHODS

Plant set-up: *Hyacinths orientalis* L. Anna Marie was used in this study. The seed balls of hyacinths were chosen with 16-17 cm of the cross-section diameter and sterilized with 75% alcohol solution for 10 min and placed into a 3-L plastic pot on October 23, 2015. The pot was supplied with 2.0 kg autoclaved (121°C, 0.11 MPa, 2 h) substrates of soil and sand (3:1, v/v).

Three AMF species, namely, *Diversispora spurca* (C.M. Pfeiff., C. Walker and Bloss) C. Walker and A. Schüβler, *D. versiformis* (P. Karst.) Oehl, G.A. Silva and Sieverd and *Funneliformis mosseae* (T.H. Nicolson and Gerd.) C. Walker

and A. Schüßler were used in this study. These AMF species were provided by the Bank of Glomeromycota in China (BGC) and propagated by pot culture in terms of identified spores and white clover under the host plant of *Trifolium repens* for 16 weeks. At the time of seedball planting, approximate 1100 spores of each AM fungus were inoculated into a pot as the AMF treatment. And the non-AMF control received same amount of autoclaved inoculums plus 2 mL inoculums filtrate (25 µm filter) to keep the similar microbial environment except the AM fungus. All treated plants were placed in a greenhouse with 768 μ mol m⁻² sec⁻¹ photosynthetic photon flux density and 18/10°C day/night temperature. During acclimation of hyacinth plant, the pots were weekly rotated to avoid position effect. After 130 days, the test plants were in the full-bloom flowering period (>50% blooming of flowers) and then harvested.

Experimental design: The experiment consisted of 4 treatments with absolutely randomized blocked design. The 4 treatments included the inoculation with *D. spurca*, *D. versiformis, F. mosseae* and non-AMF control. Each treatment had 9 replicates, leading to a total of 36 pots.

Determinations of variables: The 1 cm long fresh root segments were cut and stained with 0.05% (w/v) trypan for 3 min by the protocol of Phillips and Hayman¹². Root mycorrhizal colonization was estimated as the percentage of infected root lengths against total tested root lengths.

In the full-flowering period, flower morphological traits, including stem height, inflorescence length, floret diameter, opening floret number and total floret number were measured. The flowering duration was from the fist floret blooming to the last.

Leaf chlorophyll content was determined by the method described by Lichtenthaler and Wellburn¹³. Anthocyanin concentration in flowers was determined as per the protocol of Garcia-Viguera *et al.*¹⁴

The 0.2 g fresh samples of flowers, leaves and roots were used to determine IAA concentration in terms of the protocol of Chen *et al.*¹⁵ under the Enzyme-Linked Immunosorbent Assay (ELISA) conditions.

N contention of flowers, leaves and roots was determined by the protocol of Zhao *et al.*¹⁶. P and K concentrations in tissues were determined by the protocol of Wu and Zou¹⁷.

Statistical analysis: Data were analyzed by one-way variance of analysis (ANOVA) with SAS 8.1 software (SAS Institute Inc., Cary, NC, USA). Probabilities of significance were used to test

the significance among treatments. The significant differences between treatments were compared with the Duncan's multiple range (DMR) tests at p<0.05.

RESULTS AND DISCUSSION

Root mycorrhizal colonization and plant biomass production: No AMF structure (Fig. 1a) was observed in the non-inoculated hyacinth plants and the inoculated plants showed great root mycorrhizal colonization (Fig. 1b-d), which ranged from 38.0-48.7% (Table 1). Meanwhile, significantly (DMR, p<0.05) higher root mycorrhizal colonization was ranked as the trend of *F. mosseae>D. spurca>D. versiformis* (Table 1). This suggests different compatibility between AMF and hyacinth plants, which had been confirmed in previous studies¹⁸.

Compare to non-AMF treatment, inoculation with D. spurca, D. versiformis and F. mosseae did not significantly (p<0.05) alter fresh and dry weight of roots (Table 1). In regard to flowers, amongst three AMF species used, only F. mosseae-inoculated treatment significantly (p<0.05) increased fresh and dry weight of flowers by 27.34 and 12.54%, respectively as compared with non-AMF treatment. In these AMF species, D. spurca did not affect leaf fresh and dry biomass and *D. spurca* and *D. versiformis* heavily decreased leaf fresh and dry biomass, in comparison with non-AMF inoculation. The result demonstrated that only F. mosseae, other than D. spurca and D. versiformis, had a positive effect on plant biomass production of hyacinth. It is in agreement with earlier studies in zinnia (Zinnia elegans)¹⁹ and pot marigold (*Calendula officinalis*)²⁰. It suggests that different AMF play different role on plant growth, due to functional

diversity of AMF²¹. And, the compatablity of hyacinth plants with *F. mosseae* was greater than *D. spurca* and *D. versiformis.* As reported by the represent study, *F. mosseae* significantly promoted the growth of hyacinth, while *D. spurca* and *D. versiformis* were contrary. It is concluded that *F. mosseae* might elevate mineral nutrient absorption, increase IAA concentration and alter resource allocation. The results agreed with the research of Nadeem *et al.*²²

Flower morphology and flowering duration: Among three AMF species used, only *F. mosseae* significantly (DMR, p<0.05) increased height of flower stem, floret diameter and the number of opening flowers and the other two AMF species did not alter flower morphology (Table 2). In addition, F. mosseae and D. spurca inoculated plants showed 3.3 and 1.4 days longer flowering duration than non-inoculated plants and D. versiformis shortened 0.7 days of flower duration. Previous studies had shown that AMF could increase, reduce or have no effect on flower traits and flowering duration of ornamental plants²³⁻²⁶. Inoculation with *Glomus* intraradices increased flower stem height of Chrysanthemum *morifolium*²³ while had no effect on marigold²⁴. AMF also increased the number of flower in strawberry and Hypericum perforatum^{25,26}. Similarly, AMF inoculation prolonged or also shortened flowering duration^{27,28}. It is concluded that AMF-mediated flower morphology and flowering duration are strongly dependent on AMF species. As a whole, F. mosseae exhibited the best efficiency on flower than other two AMF species.

Chlorophyll concentration in leaf: Among three AMF species used, *D. versiformis* significantly (DMR, p<0.05) decreased

Table 1: Effects of AMF inoculation on root colonization and biomass production of hyacinth plants

	Fresh weight (g	FW/plant)		Dry weight (g D	Dry weight (g DW/plant)			
							colonization	
Treatments	Flowers	Leaves	Roots	Flowers	Leaves	Roots	(%)	
Non-AMF	34.27±3.31 ^b	14.67±1.12ª	47.25±4.48ª	3.27±0.19 ^b	0.53±0.04ª	2.51 ± 0.18^{ab}	0.0 ± 0.0^{d}	
D. spurca	32.27±3.17 ^b	12.94±1.14 ^{ab}	44.40±4.05ª	3.02 ± 0.13^{b}	0.51 ± 0.04^{ab}	2.27±0.11 ^b	42.4±3.8 ^b	
D. versiformis	30.80±2.57 ^b	10.83±0.80°	44.59±3.56ª	3.18±0.29 ^b	0.43±0.03°	2.45±0.17 ^{ab}	38.0±3.1°	
F. mosseae	43.64±3.89ª	11.53±1.10 ^{bc}	46.01±4.16ª	3.68±0.14ª	0.44±0.03 ^{bc}	2.68±0.15ª	48.7±4.7ª	

Data (Means \pm SD, n = 3) followed by different letters among treatments indicate significant differences (DMR, p<0.05) between treatments

Table 2: Effects of AMF inoculation on flower morphological traits and flowering duration of hyacinth plants in full-bloom pe	eriod
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	Flower stem	Inflorescence	Floret	Opening floret	Total floret	Flowering
Treatments	height (cm)	length (cm)	diameter (cm)	number/plant	number/plant	duration (day)
Non-AMF	20.0±1.6 ^b	11.1±1.0 ^{ab}	2.9±0.2 ^b	24±2 ^b	55 ± 5^{ab}	22.1±2.0 ^{bc}
D. spurca	21.2±1.9 ^{ab}	11.7±1.1 ^{ab}	3.1±0.2 ^{ab}	16±1°	55 ± 5^{ab}	23.5±0.6 ^b
D. versiformis	19.0±1.4 ^b	10.4±0.7 ^b	3.0 ± 0.2^{b}	12±1°	53±5 [⊾]	21.4±0.9°
F. mosseae	23.2±1.4ª	12.3±1.0ª	3.3±0.1ª	45±4ª	59±5ª	25.4±0.7ª

Data (Means \pm SD, n = 9) followed by different letters among treatments indicate significant differences (DMR, p<0.05) between treatments

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Fig. 1(a-d): Root mycorrhizal colonization of hyacinth plants by (a) Non-AMF, (b) *Diversispora spurca*, (c) *Diversispora versiformis* and (d) *Funneliformis mosseae*



Fig. 2: Effects of AMF inoculation on leaf chlorophyll a, chlorophyll b, total chlorophyll and flower anthocyanin concentration in hyacinth plants. Data (Means \pm SD, n = 3) followed by different letters above bars showed significant differences (DMR, p<0.05) between treatments

leaf chlorophyll a concentration and *D. spurca* and *F. mosseae* considerably increased leaf chlorophyll b and total chlorophyll concentrations than non-AMF treatment (Fig. 2). *D. spurca* significantly (p<0.05) increased the content of chlorophyll b by 39.63% and *F. mosseae* significantly increased the content of total chlorophyll by 15.38%. The higher chlorophyll concentration in AM plants suggested higher photosynthetic capacity or greater N level⁹. The same result was reported by Lu *et al.*²⁹ in *Morus alba* plants.

Anthocyanin concentration in flowers: AMF inoculation with *D. spurca* and *D. versiformis* did not affect flower anthocyanin concentration as compared with non-AMF inoculation (Fig. 2). However, inoculation with *F. mosseae* possessed 81.1% significantly (DMR, p<0.05) higher flower anthocyanin concentration than non-AMF treatment. Such result is in accordance with the AMF-induced flower biomass production and morphology under *F. mosseae* conditions. *F. mosseae* increased the anthocyanin concentration that may be connected with the high N concentration in flowers,

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Fig. 3: Effects of AMF inoculation on IAA content of flowers, leaves and roots in hyacinth plants. Data (Means \pm SD, n = 3) followed by different letters above bars showed significant differences (DMR, p<0.05) between treatments

	Table 3: Effects of AMF inoculation on N, P and K contents of flowers, leaves and roots in hyacinth r	olants
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	Flowers (mg DW/plant)			Leaves (mg DW/plant)			Roots (mg DW/plant)		
Treatments	 N	Р	K	 N	P	К	N	P	К
Non-AMF	68.09±7.92 ^b	7.90±0.11 ^ь	54.91±1.71 ^b	25.78±2.28ª	2.88±0.26ª	26.35±0.88ª	34.94±1.51 ^₅	4.65±0.55ª	31.00±0.70 ^b
D. spurca	65.22±7.76 ^b	7.88±0.24 ^b	56.80±0.30 ^b	22.19±1.55 [⊾]	2.07±0.21 ^b	22.21±2.64 ^b	35.87±3.85 ^b	4.49±0.25ª	30.28±3.02 ^b
D. versiformis	70.58±4.46 ^b	7.84±0.17 ^b	62.09±5.35 ^b	18.06±1.79°	1.76±0.21 ^b	18.14±1.51°	34.27±1.58 ^b	4.33±0.09ª	29.38±1.11 ^b
F. mosseae	96.84±6.67ª	11.12±0.13ª	74.10±5.95ª	18.73±1.86 ^{bc}	1.74±0.18 ^b	17.50±2.08°	43.45±1.57ª	4.94±0.39ª	40.69±3.94ª

Data (Means \pm SD, n = 3) followed by different letters among treatments indicate significant differences (DMR, p<0.05) between treatments

because higher N concentration influenced anthocyanin production in *Hibiscus sabdariffa* plants³⁰.

IAA levels of tissues: In this study, compared with non-AMF treatment, three AMF species did not provide any effect on leaf IAA levels (Fig. 3). In roots, only F. mosseae exhibited 20.02% significantly (DMR, p<0.05) higher IAA concentration than non-AMF treatment, in flowers, D. spurca and F. mosseae markedly increased IAA levels. Changes in these IAA levels were coincident with changes in flower morphology, in this study. This implied that AMF-induced IAA changes are closely related with AMF-induced flower morphological changes in hyacinth. It is well documented that IAA plays an important role in plant morphogenesis, such as stem elongation, pollen tube growth, flower and floral transition^{31,32}. Studies in the past also proved that inoculation with AMF significantly increased flower IAA concentration in onion³³ and soybean³⁴ and root IAA content in trifoliate orange³⁵.

Nutrient status in tissues: Compared to non-AMF, plants inoculated with *F. mosseae* had 42.22, 40.76 and 34.95% significantly (DMR, p<0.05) higher N, P and K concentrations in flowers and the other AMF species had no significant changes in flower N, P and K levels (Table 3). Conversely, in

leaves, the AMF plants possessed lower N, P and K content than non-AMF plants. In roots, only *F. mosseae*-inoculated hyacinth plants exhibited relatively higher N and K content than other treatments. Therefore, in this study, *F. mosseae* had the best positive effect on N, P and K assimilating in flowers and roots. The similar results were shown in pepper³⁶, pelargonium³⁷ and *Miscanthus sacchariflorus*³⁸ plants. N nutrition may help to maintain chlorophyll synthesis and plant relative growth rate^{39,40}. AMF played a vital role in enhanced P and K acquisition^{41,42}. Meanwhile, K is involved in a wide range of function in plants as a carrier ion in hormonal stressed signals^{37,43}. As a result, AMF-modulated N, P and K changes are associated with AMF-modulated biomass production changes in hyacinth plants.

CONCLUSION

In this study, mycorrhizal inoculation could heavily affect biomass production, chlorophyll, anthocyanin and IAA synthesis and nutrient acquisition of hyacinth plants. Meanwhile, only *F. mosseae* had strongly positive compatibility with hyacinth plants. AMF-modulated biomass production may be associated with AMF-modulated nutrient changes. AMF-induced flower morphological changes are due to AMF-induced IAA changes.

SIGNIFICANCE STATEMENTS

This study evaluated the effects of three AMF species, namely, *Diversispora spurca, Diversispora versiformis* and *Funneliformis mosseae* in flower morphology, color and duration of hyacinth. It found that only *F. mosseae* had the positive stimulated effects on morphology, color and lasting time of flowers, which is involved in IAA levels and N, P and K concentrations. This study provides new pathway to modulate flowering phase and flower quality of hyacinth plants through utilizing AMF as a stimulator in floriculture.

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REFERENCES

- 1. Ahulu, E.M., H. Andoh and M. Nonaka, 2007. Host-related variability in arbuscular mycorrhizal fungal structures in roots of *Hedera rhombea, Rubus parvifolius* and *Rosa multiflora* under controlled conditions. Mycorrhiza, 17: 93-101.
- Smith, S.E. and D.J. Read, 2008. Mycorrhizal Symbiosis. 3rd Edn., Academic Press, London, UK., ISBN-13: 978-0123705266, Pages: 800.
- Koide, R.T., 2010. Mycorrhizal Symbiosis and Plant Reproduction. In: Arbuscular Mycorrhizas: Physiology and Function, Koltai, H. and Y. Kapulnik (Eds.). Springer, New York, USA., ISBN-13: 9789048194896, pp: 297-320.
- 4. Vosatka, M., 1995. Influence of inoculation with arbuscular mycorrhizal fungi on the growth and mycorrhizal infection of transplanted onion. Agric. Ecosyst. Environ., 53: 151-159.
- 5. Petit, E. and W.D. Gubler, 2006. Influence of *Glomus intraradices* on black foot disease caused by *Cylindrocarpon macrodidymum* on *Vitis rupestris* under controlled conditions. Plant Dis., 90: 1481-1484.
- Vosatka, M. and J. Albrechtova, 2008. Theoretical Aspects and Practical Uses of Mycorrhizal Technology in Floriculture and Horticulture. In: Floriculture, Ornamental and Plant Biotechnology, Da Silva, J.A.T. (Ed.). Global Science Books Ltd., Middlesex, UK., pp: 466-479.
- 7. Koltai, H., 2010. Mycorrhiza in floriculture: Difficulties and opportunities. Symbiosis, 52: 55-63.
- 8. Yi, Y.B., K.S. Lee and C.H. Chung, 2002. Protein variation and efficient *in vitro* culture of scale segments from *Hyacinthus orientalis* L. cv. Carnegie. Sci. Hortic., 92: 367-374.

- Asrar, A.A., G.M. Abdel-Fattah and K.M. Elhindi, 2012. Improving growth, flower yield and water relations of snapdragon (*Antirhinum majus* L.) plants grown under well-watered and water-stress conditions using arbuscular mycorrhizal fungi. Photosynthetica, 50: 305-316.
- 10. Scagel, C.F., 2004. Inoculation with vesicular-arbuscular mycorrhizal fungi and rhizobacteria alters nutrient allocation and flowering of harlequin flower. HortTechnology, 14: 39-48.
- 11. Deljou, M.J.N., A. Marouf and H.J. Hamedan, 2014. Effect of inoculation with Arbuscular Mycorrhizal Fungi (AMF) on gerbera cut flower (*Gerbera jamesonii*) production in soilless cultivation. Acta Hortic., 1034: 417-422.
- 12. Phillips, J.M. and D.S. Hayman, 1970. Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. Trans. Br. Mycol. Soc., 55: 158-161.
- 13. Lichtenthaler, H.K. and R.R. Wellburn, 1983. Determinations of total carotenoids and chlorophylls *a* and *b* of leaf extracts in different solvents. Biochem. Soc. Trans., 11: 591-592.
- 14. Garcia-Viguera, C., P. Zafrilla and F.A. Tomas-Barberan, 1998. The use of acetone as an extraction solvent for anthocyanins from strawberry fruit. Phytochem. Anal., 9: 274-277.
- Chen, Q., W.B. Qi, R.J. Reiter, W. Wei and B.M. Wang, 2009. Exogenously applied melatonin stimulates root growth and raises endogenous indoleacetic acid in roots of etiolated seedlings of *Brassica juncea*. J. Plant Physiol., 166: 324-328.
- Zhao, X., Y. Zhou, S. Wang, G. Xing, W. Shi, R. Xu and Z. Zhu, 2012. Nitrogen balance in a highly fertilized rice-wheat double-cropping system in Southern China. Soil Sci. Soc. Am. J., 76: 1068-1078.
- 17. Wu, Q.S. and Y.N. Zou, 2009. Mycorrhizal influence on nutrient u ptake of citrus exposed to drought stress. Philipp. Agric. Scientist, 92: 33-38.
- Davoodian, N., J. Bosworth and N. Rajakaruna, 2012. Mycorrhizal colonization of *Hypericum perforatum* L. (Hypericaceae) from serpentine and granite outcrops on the Deer Isles, Maine. Northeastern Naturalist, 19: 517-526.
- Long, L.K., Q. Yao, Y.H. Huang, R.H. Yang, J. Guo and H.H. Zhu, 2010. Effects of arbuscular mycorrhizal fungi on zinnia and the different colonization between *Gigaspora* and *Glomus*. World J. Microbiol. Biotechnol., 26: 1527-1531.
- Hristozkova, M., M. Geneva, I. Stancheva, M. Boychinova and E. Djonova, 2016. Contribution of arbuscular mycorrhizal fungi in attenuation of heavy metal impact on *Calendula officinalis* development. Applied Soil Ecol., 101: 57-63.
- 21. Wu, Q.S., G.H. Li and Y.N. Zou, 2010. Roles of arbuscular mycorrhizal fungi on growth and nutrient acquisition of peach (*Prunus persica* L. Batsch) seedlings. J. Anim. Plant Sci., 21:746-750.
- 22. Nadeem, S.M., M. Ahmad, Z.A. Zahir, A. Javaid and M. Ashraf, 2014. The role of mycorrhizae and Plant Growth Promoting Rhizobacteria (PGPR) in improving crop productivity under stressful environments. Biotechnol. Adv., 32: 429-448.

- 23. Vaingankar, J.D. and B.F. Rodrigues, 2012. Screening for efficient AM (arbuscular mycorrhizal) fungal bioinoculants for two commercially important ornamental flowering plant species of Asteraceae. Biol. Agric. Hortic., 28: 167-176.
- 24. Linderman, R.G. and E.A. Davis, 2004. Varied response of marigold (*Tagetes* spp.) genotypes to inoculation with different arbuscular mycorrhizal fungi. Sci. Hortic., 99: 67-78.
- 25. Bona, E., G. Lingua, P. Manassero, S. Cantamessa and F. Marsano *et al.*, 2015. AM fungi and PGP pseudomonads increase flowering, fruit production and vitamin content in strawberry grown at low nitrogen and phosphorus levels. Mycorrhiza, 25: 181-193.
- Lazzara, S., M. Militello, A. Carrubba, E. Napoli and S. Saia, 2017. Arbuscular mycorrhizal fungi altered the hypericin, pseudohypericin and hyperforin content in flowers of *Hypericum perforatum* grown under contrasting P availability in a highly organic substrate. Mycorrhiza, 27: 345-354.
- 27. Jin, Z., J. Li and Y. Li, 2015. Interactive effects of arbuscular mycorrhizal fungi and copper stress on flowering phenology and reproduction of *Elsholtzia splendens*. PLoS ONE, Vol. 10. 10.1371/journal.pone.0145793.
- Sohn, B.K., K.Y. Kim, S.J. Chung, W.S. Kim and S.M. Park *et al.*, 2003. Effect of the different timing of AMF inoculation on plant growth and flower quality of chrysanthemum. Scient. Hortic., 98: 173-183.
- 29. Lu, N., X. Zhou, M. Cui, M. Yu, J. Zhou, Y. Qin and Y. Li, 2015. Colonization with arbuscular mycorrhizal fungi promotes the growth of *Morus alba* L. seedlings under greenhouse conditions. Forests, 6: 734-747.
- 30. Zheng, S. and Y. Guo, 1998. Effects of staple nutrient on cell growth and anthocyanin production in suspension culture of *Hibiscus sabdariffa* L. Guihaia, 18: 70-74.
- Razem, F.A., K. Baron and R.D. Hill, 2006. Turning on gibberellin and abscisic acid signaling. Curr. Opin. Plant Biol., 9: 454-459.
- Alabadi, D., M.A. Blazquez, J. Carbonell, C. Ferrandiz and M.A. Perez-Amador, 2009. Instructive roles for hormones in plant development. Int. J. Dev. Biol., 53: 1597-1608.
- Torelli, A., A. Trotta, L. Acerbi, G. Arcidiacono, G. Berta and C. Branca, 2000. IAA and ZR content in leek (*Allium porrum*L.), as influenced by P nutrition and arbuscular mycorrhizae, in relation to plant development. Plant Soil, 226: 29-35.

- Meixner, C., J. Ludwig-Muller, O. Miersch, P. Gresshoff, C. Staehelin and H. Vierheilig, 2005. Lack of mycorrhizal autoregulation and phytohormonal changes in the supernodulating soybean mutant *nts1007*. Planta, 222: 709-715.
- 35. Wu, Q.S., C.Y. Liu, D.J. Zhang, Y.N. Zou, X.H. He and Q.H. Wu, 2016. Mycorrhiza alters the profile of root hairs in trifoliate orange. Mycorrhiza, 26: 237-247.
- 36. Turkmen, O., S. Sensoy, S. Demir and C. Erdinc, 2008. Effects of two different AMF species on growth and nutrient content of pepper seedlings grown under moderate salt stress. Afr. J. Biotechnol., 7: 392-396.
- Perner, H., D. Schwarz, C. Bruns, P. Mader and G. Eckhard, 2007. Effect of arbuscular mycorrhizal colonization and two levels of compost supply on nutrient uptake and flowering of pelargonium plants. Mycorrhiza, 17: 469-474.
- Sarkar, A., T. Asaeda, Q. Wang and M.H. Rashid, 2015. Arbuscular mycorrhizal influences on growth, nutrient uptake and use efficiency of *Miscanthus sacchariflorus* growing on nutrient-deficient river bank soil. Flora-Morphol. Distrib. Funct. Ecol. Plants, 212: 46-54.
- 39. Evelin, H., R. Kapoor and B. Giri, 2009. Arbuscular mycorrhizal fungi in alleviation of salt stress: A review. Ann. Bot., 104: 1263-1280.
- 40. Peng, Y., K.J. Niklas and S. Sun, 2010. The relationship between relative growth rate and whole-plant C:N:P stoichiometry in plant seedlings grown under nutrient-enriched conditions. J. Plant Ecol., 4: 147-156.
- Sarkar, A., T. Islam, G. Biswas, S. Alam, M. Hossain and N. Talukder, 2012. Screening for phosphate solubilizing bacteria inhabiting the rhizoplane of rice grown in acidic soil in Bangladesh. Acta Microbiol. Immunol. Hung., 59: 199-213.
- 42. Alloush, G.A. and R.B. Clark, 2001. Maize response to phosphate rock and arbuscular mycorrhizal fungi in acidic soil. Commun. Soil Sci. Plant Anal., 32: 231-254.
- 43. Peuke, A.D., W.D. Jeschke and W. Hartung, 2002. Flows of elements, ions and abscisic acid in *Ricinus communis* and site of nitrate reduction under potassium limitation. J. Exp. Bot., 53: 241-250.