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Sweet Pepper Biomass Production and Partitioning as Affected by Different Shoot and Root-zone Conditions

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Abstract: Three greenhouse experiments on sweet pepper plants were carried out to investigate the effect of manipulating three root zone conditions namely volumetric Water Content (WC), Electrical Conductivity (EC) and minimum root zone temperature independently from each other on biomass production and partitioning of sweet pepper plants. Interactions of WC with fruit load and EC with climate were also investigated. Treatments were in Exp. I, two water content levels, High 80% (HWC) and low 50% (LWC) with an EC $3.0 \pm 0.5 \text{ mS cm}^{-1}$; in Exp. II, High EC (HEC) 6.5 mS cm^{-1} and low (LEC) 2.5 mS cm^{-1} both under normal (HET) or suppressed transpiration (LET) and in Exp. III, heated (18°C) or non heated nutrient solution (control). Under treatments of Exp. I, two fruit load treatments, Normal Fruit Load (NFL) and Manipulated Fruit Load (MFL) were applied. Dry matter production was not affected by reducing water content treatments in the root zone. While HEC reduced dry matter production under control condition (HET), manipulation of greenhouse climate (LET) mitigated this negative effect and dry matter production was not different than LEC treatment. In addition, raising nutrient solution minimum temperature increased significantly dry matter production. Total plant fresh weight followed the same pattern of dry matter production. No significant differences were observed in dry matter partitioning as affected by all treatments although there was a tendency for more dry matter partitioned to the stems due to lower fruit number in MFL or high incidence of blossom-end rot in HEC treatment. Partitioning between leaves and stems followed allometric relationship while partitioning to the fruits seemed to follow the principle of sink strength.

Key words: Sweet pepper, *Capsicum annuum* L., salinity, water content, root zone temperature, biomass production, partitioning, electrical conductivity

INTRODUCTION

The ongoing water scarcity worldwide necessitates the careful use of this resource. Due to a growing competition from other sectors and the rapid increase in population, agriculture will have to do less with fresh water and do more with marginal quality water. Reduction in irrigation regimes and/or using low quality water is some of those options to increase production efficiency. This may lead to unfavorable growth conditions such as temporarily drought and/or salinity build up resulting in reduction in total plant biomass production. Protected cultivation enables for manipulation of growth conditions, root and shoot conditions, which increase production efficiency. Many studies investigated the effect of different irrigation frequencies on the growth and

production of field crops^[1-4] as well as greenhouse crops^[5]. These studies reported negative effect of long irrigation intervals on general plant growth parameters and production. Salinity may also build up in soilless cultivation closed systems as a result of nutrient accumulation and/or of using poor quality water^[6,7]. Salinity stress commonly affects negatively plant growth of vegetables such as tomato^[8,9] and sweet pepper^[10-12]. Bruggink *et al.*^[13] mentioned that it should be possible to improve plant growth by adapting the salinity level in relation to the rate of transpiration, thus diminishing water deficits in the plant at high transpiration. The other way round was mentioned to be also possible where tomato crop growth and yield under salinity condition could be improved by manipulating greenhouse climate (transpiration rate)^[14-18]. However, there is little known

about the response of sweet pepper crop grown under different salinity levels to greenhouse climate manipulation^[11].

Another option for the grower to improve plant growth is to manipulate root zone temperature. In practice, potential transpiration increases suddenly at sunrise while water uptake may be delayed because of the low nutrient solution temperature since water uptake was reported to be improved by warming up the nutrient solution^[19-22]. However, these researchers have used 24 h/day heated nutrient solution. In countries like Egypt or regions like the Mediterranean, daytime temperature is high enough that there is no need to heat the nutrient solution during the day. On the other hand, during nighttime, nutrient solution temperature may drop as a result of a drop of greenhouse air temperature (commonly unheated plastic house). Therefore, heating the nutrient solution during nighttime may be enough.

This study was intended to investigate different approaches of manipulating root zone conditions with or without shoot environment aiming at improving plant biomass production using sweet pepper crop as a test plant.

MATERIALS AND METHODS

Three Experiments I, II and III were carried out using sweet pepper crop (*Capsicum annuum* L.) and each focused on one of the factors in the root environment namely, Water Content (WC), Electrical Conductivity (EC) and minimum Root Zone Temperature (RZT), respectively. Plants in Exp. I and II were grown in Rockwool slabs in controlled climate glasshouses in The Netherlands and in Exp. III plants were grown in Nutrient Film Technique (NFT) in unheated plastic house in Egypt. Plants in Exp. I and II were trimmed to two first order branches while those in Exp. III were grown un-trimmed as a standard practices in both sites.

Exp. I consisted of two volumetric water content treatments, High (HWC) 80% and Low (LWC) 50%. WC and EC were monitored online by two dielectric water content sensors. Irrigation was triggered by signals from the sensors. EC in the two treatments was kept at $3.0 \pm 0.5 \text{ mS cm}^{-1}$ by switching the irrigation automatically from nutrient solution with EC 3.0 mS cm^{-1} to a nutrient solution with EC 1.0 mS cm^{-1} whenever the EC in the root zone was above 3.0 mS cm^{-1} .

Exp. II consisted of two EC levels in the root zone namely High EC (HEC) 6.5 mS cm^{-1} and Low EC (LEC) 2.5 mS cm^{-1} and both were under two different greenhouse climates. First greenhouse was controlled according to the standard Dutch practice and used as

control (HET) and the second greenhouse (LET) was set to 25% less ventilation opening compared to the control to have lower Vapor Pressure Deficit (VPD). In addition, if necessary, humidification was applied in the second greenhouse to suppress transpiration not to exceed $0.15 \text{ l plant}^{-1} \text{ h}^{-1}$.

Exp. III consisted of two treatments, unheated nutrient solution (control) and heated nutrient solution by setting minimum nutrient solution temperature to 18°C .

In addition, for the interest of studying the interactive effect of sink strength with water content in the root zone on dry matter production and partitioning, Exp I included two fruit manipulation treatments. Manipulated Fruit Load (MFL) by allowing only two flowers per plant per week to develop to fruits and Normal Fruit Load (NFL) by allowing all possible flowers to set and develop to fruits. Both of the two fruit treatments were under the two water treatments.

Crop measurements: In Exp. I, fresh and dry weights of all plant organs were measured in destructed samples which were carried out once a month for NFL and every two months for MFL treatment. Two plants per replicate were taken out in every sample (6 plants per treatment). Plants next to destructed samples were not included in any measurements. In Exp. II destructed samples were taken every two weeks starting from the generative stage and each consisted of three plants for each treatment for determining dry matter production. Destructed samples were taken once a month in Exp. III.

Exp. I and II were arranged in a split plot design with climate treatment as main plot and EC as the sub-main in Exp I meanwhile, Water content in the root as main plot and fruit load treatment as sub-main. Exp. III was arranged in a Complete Randomized Block Design.

RESULTS

Total plant fresh weight was slightly higher only during the warm periods in HWC and this resulted in 6% difference between the two water treatments (Fig. 1A). On the other hand, total plant fresh weight was significantly reduced by HET-HEC in the time that LET-HEC had the same fresh weight when compared to LEC under the two climate conditions (Fig. 1B). Total plant fresh weight increased also by heating up the nutrient solution (Fig. 1C).

There was a minor difference between the treatments of water content concerning total plant dry matter production (Fig. 2A). Meanwhile, total plant dry weight was significantly reduced by HET-HEC. LET-HEC had almost the same dry matter production compared to LEC

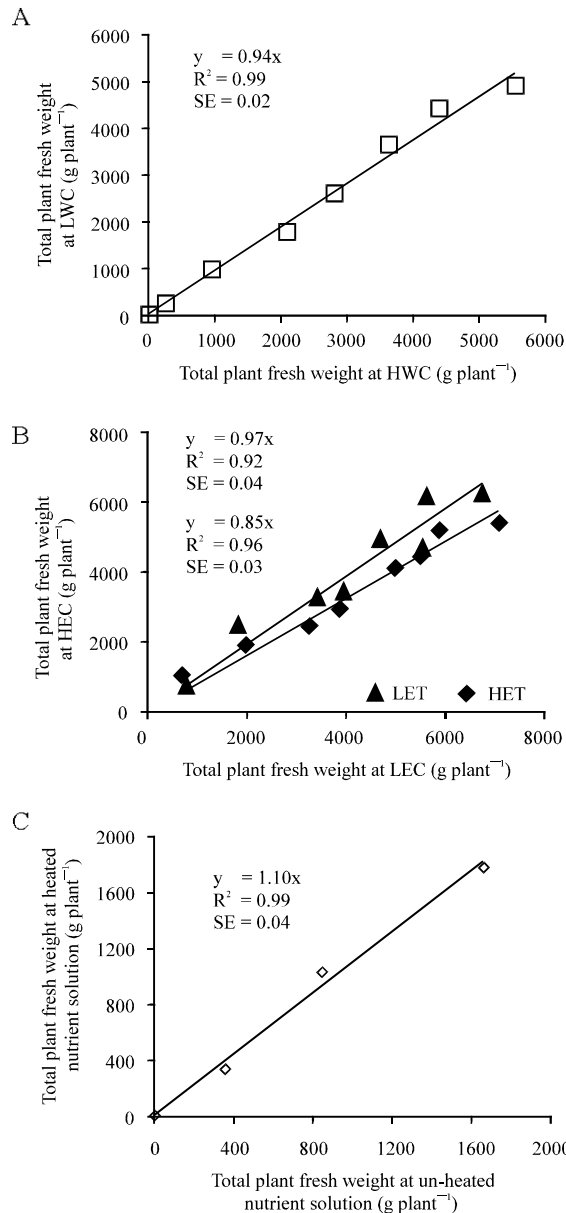


Fig. 1: Total plant fresh weight (g plant⁻¹) of sweet pepper plants grown in Low Water Content (LWC) versus High Water Content (HWC) in the root zone (A), High EC (HEC) versus Low EC (LEC) nutrient solution under the two climate conditions low transpiration (LET) and High transpiration (HET)(B) and heated nutrient solution versus un heated nutrient solution (C)

(Fig. 2B). It was clear that dry matter production responded positively to the heating treatment (Fig. 2C).

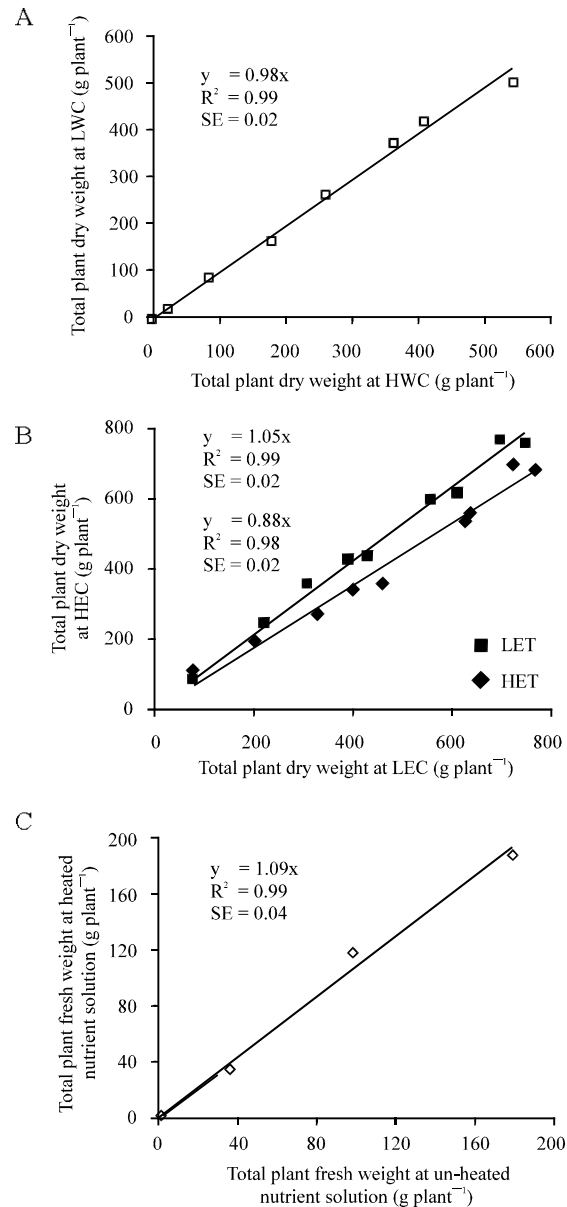


Fig. 2: Total plant dry weight (g plant⁻¹) of sweet pepper plants grown in Low Water Content (LWC) versus High Water Content (HWC) in the root zone (A), High EC (HEC) versus Low EC (LEC) nutrient solution under the two climate conditions low transpiration (LET) and High transpiration (HET)(B) and heated nutrient solution versus un heated nutrient solution (C)

In order to analyze how the reduction in dry matter production by HET-HEC was brought about, a validated

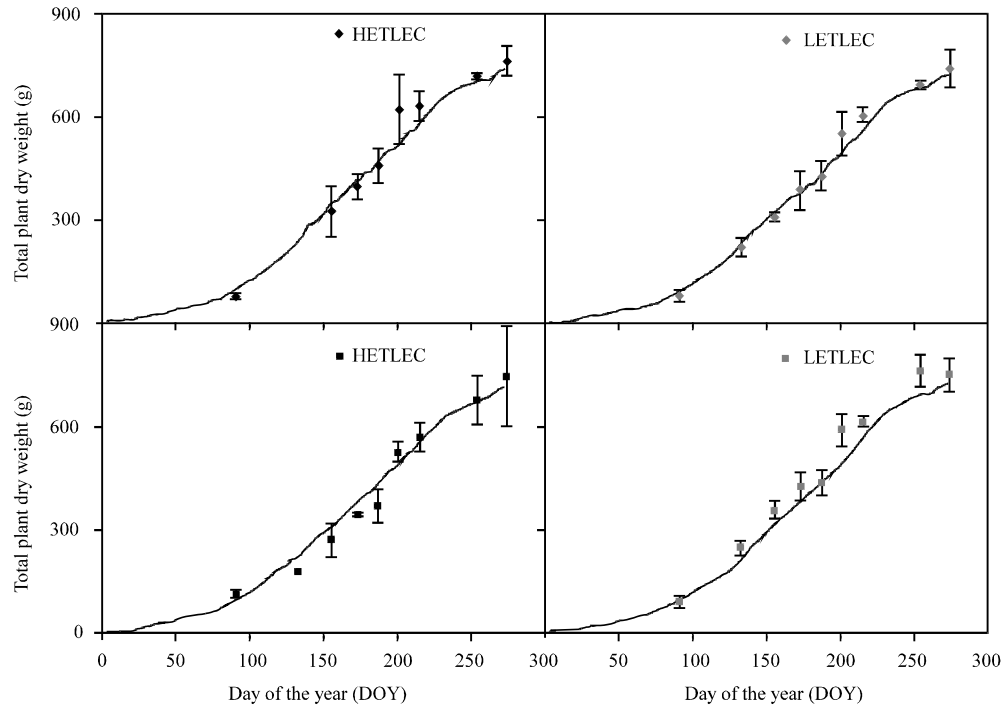


Fig. 3: Actual (symbols) and simulated (lines) total plant dry matter production (g plant^{-1}) of sweet pepper plants grown under the four combinations of EC levels and climate conditions. Error bars are twice standard error

Table 1: Dry matter of different plant organs (fraction of total plant dry weight) of sweet pepper plants under different root and shoot conditions

Experiment No.	Treatments	Leaves	Stems	Fruits
I	HWC-MFL	15.70	30.59	53.71
	HWC-NFL	14.18	27.24	58.58
	LWC-MFL	13.54	26.83	59.62
	LWC-NFL	12.54	25.85	61.25
LSD 5%		N.S	N.S	N.S
II	HET-LEC	11.19ab	21.87	66.9
	HET-HEC	11.39ab	22.40	66.2
	LET-LEC	9.44b	19.33	71.2
	LET-HEC	12.82a	20.99	66.2
LSD 5%		2.93	N.S	N.S
III	Heated nutrient solution	15.84	20.75	63.41
	Un-heated nutrient solution	20.1	25.63	55.27
LSD 5%		N.S.	N.S.	N.S.

NS = non significant

explanatory crop growth model developed by Gijzen^[23] was used. Simulation results show that the model can predict plant dry matter production very well under different conditions (Fig. 3). There were also some differences among the treatments in their simulation results. These differences among the treatments were brought about mainly by differences in Leaf Area Index (LAI). LAI was introduced as an input in the model. The smaller LAI in HET-HEC was the main reason for the lowest dry matter production by the crop. As there was

no provision in the model for the effect of EC, resulting differences in calculated production were caused by differences in climate and LAI. Within a greenhouse, (high and low EC) differences can only come from LAI. Modeled differences were slightly smaller than real ones (about 5%). Hence, some other effect (stomatal conductance) plays a role. Indeed a sensitivity analysis of the model revealed that a reduction of 15% in maximum stomatal conductance (g_{max}) may result in the 5% remaining of the observed reduction in total dry matter production.

Dry matter partitioning: Dry Matter (DM) partitioning to the fruits was the highest fraction and averaged more than 60% in all experiments meanwhile leaves had the lowest fraction of plant dry weight (Table 1). Although differences in partitioning to different parts were non significant almost in all treatments, plants under HEC had a tendency for higher dry matter partitioning to the stems compared to plants under LEC treatments. Similarly, plants with Manipulated Fruit Load (MFL) tended to partition more assimilates to the stems compared to NFL plants. However, the partitioning relationship between leaves and stems was linear all the time for all the treatments in all experiments (Fig. 4).

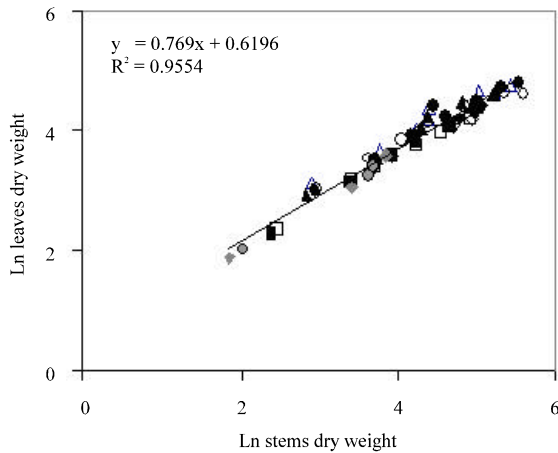


Fig. 4: Relationship between dry weights of leaves and stems (transformed to natural logarithm) for all treatments in all experiments. Triangles are low EC and circles are high EC, blank is high transpiration and black is low transpiration. Squares are normal fruit load and diamonds are manipulated fruit load, blank is low water content and black is high water content. Gray circles are heated nutrient solution and gray diamonds are unheated nutrient solution

DISCUSSION

The non significant difference in dry matter production under water content treatments in the root environment can be explained on the basis that plants were not under any degree of water stress. As yield is the main fraction of biomass production and it was not affected by the treatments^[24], difference in total plant dry weight was not expected. In addition, it was reported that if the canopy is already complete, mild water stress should have little or no effect on biomass production rate^[25]. On the other hand, under different EC conditions, the improvement in plant total dry matter production under LET-HEC compared to HET-HEC maybe due to the improvement in plant water relations which reflected on net photosynthesis. With severe water stress, stomatal opening and photosynthesis per unit leaf area are reduced^[25]. Plant growth and production is a function of the relationship between dry matter production (photosynthesis) and water content (related to water uptake and transpiration) in plant tissue^[26,27]. The two components of this relationship, dry matter and water, are interdependent^[28] and very much affected by conditions during growth such as solar radiation and EC in the root zone (or the irrigation water). Reduction in plant dry matter production has been reported to be caused by high

EC in the root zone^[10-12,29]. This reduction was also noticed in the data of this study. However, as with other crops such as tomato^[14,16-18], reducing vapor pressure deficit of the greenhouse resulted in mitigating the negative effect of HEC. Since VPD of the greenhouse and EC of the root zone affect the same state variable of plant water status, this mitigation is brought about by improving plant water relations. Zabari *et al.*^[11] mentioned that leaf conductance and area were the causes of the reduction in dry matter production caused by HEC. The data of this study show that both of these two parameters are affected by the imposed conditions (HEC and VPD). However, the main accurate measurable factor that was more affected is leaf area. The small reduction in g_{max} calculated by the sensitivity analysis of the model show that actual stomatal conductance is even smaller to be measured with a fair degree of accuracy by any current existing devices. Prediction of leaf area development is still a weak feature of crop growth models^[30] and that is the reason for giving it as an input to crop growth models as in many other studies^[31,32].

Dry matter production was also improved by heating the nutrient solution. This was reported earlier for many crops^[33] as well as for pepper^[34]. The latter explained this increment based on a higher photosynthesis. Under the same greenhouse conditions, photosynthesis is a function of leaf area and stomatal conductance. Leaf area was found to be higher under the heating treatment in this study. In addition, data in literature indicate that stomatal conductance increased when root zone temperature increased^[35].

Partitioning between leaves and stems follows always an allometric relationship^[36]. The latter found the allometric relationship to be constant under different shoot conditions of temperature and plant densities. The same was also observed here under all the experimental treatments of root zone environment. On the other hand, partitioning to the fruits always follows sink strength of generative parts^[31,36,37]. The sink strength of generative parts depends on number of fruits and sink strength of each fruit. The sink strength of individual fruit depending mainly on age and temperature^[36]. Since EC may affect both the number of growing fruits on the plant and the incidence of BER, sink strength may also be affected by EC. This may or may not, depending on the degree of EC effect, influence dry matter partitioning. EC does not affect DM partitioning in tomato with exception of very high level of EC ($EC = 17 \text{ mS cm}^{-1}$)^[38] and this is what have been observed also in sweet pepper during this work. The tendency for more allocation of dry matter to the stems in HEC and MFL treatments can be due to the lower fruit sink strength, compared to that observed in the LEC and

NFL treatments, respectively. Indeed higher partitioning to the stems with the lower number of fruits has been observed in earlier study^[39].

CONCLUSIONS

It can be concluded that total dry matter production is not affected by reducing water content in the root zone to 50% if the water potential can be controlled. Under conditions of over-abundant supply of water, effect of changing the EC in the root zone does not appear until a threshold of transpiration is reached. Limiting the minimum nutrient solution temperature from falling below a certain degree, has a positive advantage on plant fresh and dry matter production. Root zone conditions do not seem to have a direct effect on dry matter partitioning. The effect of root environment on dry matter partitioning may come through the effect on fruit sink strength.

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REFERENCES

1. Doorenbos, J. and A.H. Kassam, 1979. Yield response to water. FAO Irrigation and Drainage Paper, pp: 33.
2. Imtiyaz, M., N.P. Mgadla, B. Chepete and S.K. Manase, 2000. Response of six vegetable crops to irrigation schedules. Agric. Water Manag., 45: 331-342.
3. Jaimez, R.E., F. Rada and C. Garcia-Nunez, 1999. The effect of irrigation frequency on water and carbon relations in three cultivars of sweet pepper (*Capsicum chinense jacq*), in a tropical semiarid region. Sci. Hort., 81: 301-308.
4. Pandey, R.K., J.W. Maranville and A. Admou, 2000. Deficit irrigation and nitrogen effects on maize in a Sahelian environment. I. Grain yield and yield components. Agric. Water Manag., 46: 1-13.
5. Chartzoulakis, K. and N. Drosos, 1997. Water requirements of greenhouse grown sweet pepper under drip irrigation. Acta Hort., 449: 175-180.
6. Bohme, M., 1994. Effects of closed systems in substrate culture for vegetable production in greenhouses. Acta Hort., 396: 45-54.
7. Bohme, M., 1995. Evaluation of organic, synthetic and mineral substrates for hydroponically grown cucumber. Acta Hort., 401: 209-217.
8. Hayward, H.E. and E.M. Long, 1943. Some effects of sodium salts on the growth of tomato. Plant Physiol., 18: 556-569.
9. Sanchez Conde, M.P. and P. Azuara, 1979. Effect of balanced solutions with different osmotic pressure on tomato plant. J. Plant Nutr., 1: 297-307.
10. Pitacco, A., O. Lain and C. Giulivo, 1990. Gas exchange and architecture of sweet pepper as affected by water stress. First Congress of the European Society of Agronomy, Colmar, France.
11. Zabri, A.W., S.W. Burrage and K.S. Chartzoulakis, 1997. The effects of Vapour Pressure Deficit (VPD) and enrichment with CO₂ on water relations, photosynthesis, stomatal conductance and plant growth of sweet pepper (*Capsicum annuum* L.) grown by NFT. Acta Hort., 449: 561-567.
12. Chartzoulakis, K.S. and G. Klapaki, 1998. Effects of NaCl salinity on growth and yield of two pepper cultivars. Acta Hort., 511: 143-149.
13. Bruggink, G.T., H.E. Schouwink and E.A.J.M. Coolen, 1987. Effects of different day and night osmotic pressure of the nutrient solution on growth, water potentials and osmotic potentials of young tomato plants in soilless culture. Soilless Cultures, 3: 8-19.
14. De Kreij, C., 1992. Tomaat. Hoge Luchtvochtigheid geeft minder neusrot. Groenten+Fruit/Glasgroenten No. 20: 33.
15. De Kreij, C. and A. Huys, 1993. Paprika. Luchtbevochtiging is zinloos. Groenten+Fruit/Glasgroenten No. 1: 19.
16. Stanghellini, C., W.Th.M Van Meurs, L. Simonse and J. Van Gaalen, 1998. Combined effect of climate and concentration of the nutrient solution on a greenhouse tomato crop. I: Vegetative growth. Acta Hort., 458: 221-229.
17. Stanghellini, C., W.Th.M. Van Meurs, F. Corver, E. Van Dulleman and L. Simonse, 1998. Combined effect of climate and concentration of the nutrient solution on a greenhouse tomato crop. II: Yield quality and quantity. Acta Hort., 458: 231-237.
18. Li, Y., 2000. Analysis of greenhouse tomato production in relation to salinity and shoot environment. Ph.D Thesis, Wageningen Agric. Univ., pp: 95.

19. Maher, M.J., 1978. The effect of root zone warming on tomatoes grown in nutrient solution at two air temperature. *Acta Hortic.*, 82: 113-120.
20. Kageyama, Y., 1980. Studies on the production of double truss fruiting tomatoes in water culture. III. Effect of solution temperature on the yield and uptake of nutrients and water. *Scientific Reports of the Fac. Agric. Okayam Univ.*, 56: 29-35.
21. Park, K.W., M.H. Chiang, J.H. Won and K.H. Jang, 1995. The effect of nutrient solution temperature on the absorption of water and minerals in Chinese leafy vegetables. *J. Kor. Soc. Hort. Sci.*, 36: 309-316.
22. Park, K.W., M.H. Chiang, J.H. Won and K.H. Jang, 1995. The growth pattern of Chinese leaf vegetables by nutrient solution temperature. *J. Korean Soc. Hortic. Sci.*, 36: 608-613.
23. Gijzen, H., 1992. Simulation of photosynthesis and dry matter production of greenhouse crops. CABO-DLO, The Netherlands. pp: 69.
24. Heuvelink, E. and A.M.R. Abdel-Mawgoud, 2001. Telen zonder drain is mogelijk. *Groenten en fruit*, pp: 22-23.
25. Hsiao, T.C., 1993. Growth and productivity of crops in relation to water status. *Acta Hortic.*, 335: 137-148.
26. Tyree, M.T. and M.G. Jarvis, 1982. Water in tissues and cells. In: *Encyclopedia of Plant Physiology: Physiological plant Ecology II, Water relations and Carbon assimilation (NS)*, 12B. (Lange, O.L., P.S. Nobel, C.B. Osmand and H. Ziegler (Eds.)). Springer Verlag, Berlin/New York, pp: 36-77.
27. Cosgrove, D.J., 1986. Update on cell walls: How do plant cell walls extend?. *Plant Physiol.*, 102: 1-6.
28. Frensch, J. and T.C. Hsiao, 1994. Transient responses of cell turgor and growth of maize roots as affected by changes in water potential. *Plant. Physiol.*, 104: 247-254.
29. Sonneveld, C., 1979. Effects of salinity on the growth and mineral composition of sweet pepper and eggplant grown under glass. *Acta Hortic.*, 89: 71-78.
30. Marcelis, L.F.M., E. Heuvelink and J. Goudriaan, 1998. Modelling biomass production and yield of horticultural crops: A review. *Sci. Hortic.*, 74: 83-111.
31. Marcelis, L.F.M., 1994. Fruit growth and dry matter partitioning in cucumber. Ph.D. Thesis, Agric. Univ. Wageningen, pp: 173.
32. Koning, A.De, 1994. Development and dry matter distribution in glasshouse tomato: A quantitative approach. Ph.D. Thesis, Wageningen Agric. Univ., pp: 240.
33. Cooper, A., 1973. Root temperature and plant growth: A review. Commonwealth Agricultural Bureau, England.
34. Gosselin, A. and M.J. Trudel, 1986. Root-zone temperature effects on pepper. *J. Am. Soc. Hortic. Sci.*, 111:2, 220-224.
35. Abdelhafeez, A.T., H. Harssema, G. Veir and K. Verkerk, 1971. Effects of soil and air temperature on growth, development and water use of tomatoes. *Neth. J. Agric. Sci.*, 19: 67-75.
36. Heuvelink, E. and L.F.M. Marcelis, 1999. Assimilate partitioning in fruit-bearing plants: environmental influences and source/sink balance. In: *Book of Abstracts of the 6th International Conference on Assimilate Transport and Partitioning (ICATP)*: New Castel, Australia, pp: 247.
37. Heuvelink, E., 1996. Tomato growth and yield: quantitative analysis and synthesis. Ph.D. Thesis, Wageningen Agric. Univ., pp: 326.
38. Ehret, D.L. and L.C. Ho, 1986. The effect of salinity on dry matter partitioning and fruit growth in tomatoes in nutrient film culture. *J. Hortic. Sci.*, 61: 361-367.
39. Nielsen, T.H. and B. Veierskov, 1988. Distribution of dry matter in sweet pepper plants (*Capsicum annuum* L.) during the juvenile and generative growth phases. *Sci. Hortic.*, 25: 179-187.