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Therapeutic Implications of Blockers of Advanced Glycation End Products (AGEs)-their Receptor (RAGE) System

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Abstract: Non-enzymatic modification of proteins by reducing sugars, a process that is also known as Maillard reaction, leads to the formation of Advanced Glycation End-products (AGEs) *in vivo*. There is a growing body of evidence that formation and accumulation of AGEs progress during normal aging and at an extremely accelerated rate under diabetes, thus being involved in the pathogenesis of various diseases such as diabetic vascular complications and neurodegenerative diseases. Recently, engagement of their receptor, RAGE, is shown to activate its down-stream signaling and subsequently evoke inflammatory responses in various types of cells. Therefore, inhibition of AGE formation or blockade of the RAGE signaling may be a promising target for therapeutic intervention in the AGE-RAGE-related devastating disorders. In this review, we discuss several types of blockers of the AGE-RAGE system and their therapeutic implications in diseases.

Key words: AGEs, RAGE, oxidative stress, PEDF, diabetic vascular complications

INTRODUCTION

The modification, aggregation and deposition of proteins are a prominent part of many pathological processes and can play a direct role in tissue damage. The pathological role of the non-enzymatic modification of proteins by reducing sugars, a process that is known as glycation (also called the Maillard reaction), has become increasingly evident in various types of diseases^[1,2]. It is now well established that early glycation products undergo progressive modification over time *in vivo* to the formation of irreversible cross-links, after which these molecules are termed Advanced Glycation End-products (AGEs). AGEs have been implicated in the development of many of the pathological sequelae of diabetes and aging, such as atherosclerosis and diabetic microangiopathy^[1,2]. Furthermore, there is a growing body of evidence that RAGE is a signal-transducing receptor for AGEs and that engagement of RAGE by AGEs evokes inflammatory responses in vascular wall cells as well^[3]. Recently, it has become clear that the AGE-RAGE system also has a role in neurodegenerative diseases such as Alzheimer's Disease (AD)^[4], Parkinson's disease^[5], Creutzfeldt-Jakob disease^[6] and amyotrophic lateral sclerosis^[7]. Thus, inhibition of AGE formation or blockade of the RAGE signaling may be a therapeutic approach. In this review, we discuss several types of blockers of the AGE-RAGE system (Table 1) and their therapeutic implications in diseases.

Table 1: Blockers of the AGE-RAGE system

1. Inhibitors of AGE formation
I. Guanidine structure
(i) Aminoguanidine
(ii) Metformin
II. Vitamin
(i) Pyridoxamine
(ii) Benfotiamine
III. Thiazolidine structure
(i) OPB-9195
IV. Piperazine structure
(i) Tenilsetam
2. AGE breakers
(i) PTB
(ii) ALT-711
3. Inhibitors of the AGE-RAGE signaling
(i) PEDF

Inhibitors of AGE formation

Guanidine structure: Guanidine compounds, such as aminoguanidine (AG) and metformin, can trap α -dicarbonyl compounds, thus preventing their further reactions with amino groups of proteins. This type of drug is one of the most studied inhibitors of AGEs both *in vitro* and *in vivo*.

AG (Pimagedine): AG is a prototype therapeutic agent for the prevention of AGE formation. The first report of intervention to prevent AGE formation by AG was the prevention of arterial wall protein cross-linking in diabetic animals^[8]. Since then, use of AG to prevent AGE formation *in vitro* and *in vivo* has given evidence of the involvement of AGEs in many disease processes and abnormal physiological states.

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AG was introduced as a hydrazine reagent for trapping reactive carbonyls formed during the Maillard reaction, especially Amadori intermediates, thus impeding their conversion into AGEs. AG reacts not only extensively with Amadori carbonyl groups of glycated proteins but also with dicarbonyl compounds such as methylglyoxal, glyoxal and 3-deoxyglucosone^[9].

Diabetic retinopathy is a microvascular complication of diabetes characterized by loss of pericytes, microaneurysm formation and acellular capillaries in the retinal microvessels^[10]. Treatment of diabetic rats for 26 weeks with AG prevented a 2.6 fold accumulation of AGE products at branching sites of pre-capillary arterioles and thereby prevented abnormal endothelial cell proliferation and significantly diminished pericyte dropout^[11]. In the same study, treatment for 75 weeks with AG decreased the number of acellular capillaries and microaneurysms as well^[11]. AG inhibited Vascular Endothelial Growth Factor (VEGF) expression in rats with long-term galactosemia^[12]. Furthermore, AGE infusion resulted in significant leukostasis and blood-retinal barrier dysfunction, one of the characteristic features of background retinopathy, by inducing retinal VEGF expression in non-diabetic healthy mice^[13,14]. These observations suggest that AG could also exert beneficial effects on early diabetic retinopathy by suppressing AGE-induced retinal VEGF overexpression. A multicenter clinical trial revealed that Pimagedine slowed the progression of diabetic retinopathy, although it was terminated early due to safety concerns^[15,16].

AG has also been shown to inhibit the accumulation of renal AGEs and to retard the development of microalbuminuria and mesangial expansion in streptozotocin-induced diabetic rats^[17]. AG inhibited the development of diabetic nephropathy in Otsuka Long Evans Tokushima Fatty (OLETF) rats, a model of type 2 diabetes as well^[18]. Recently, AG was found to decrease the expression of pro-sclerotic growth factors such as transforming growth factor- β (TGF- β) and platelet-derived growth factor-BB and connective tissue growth factor and thus to prevent the development of glomerulosclerosis and tubulointerstitial fibrosis in experimental diabetic nephropathy^[19,20]. AG treatment also prevented albuminuria in diabetic hypertensive rats without affecting blood pressure^[21].

Double-blinded, placebo-controlled, randomized clinical trials of Pimagedine (ACTION; A Clinical Trial in Overt Nephropathy) were designed to evaluate the safety and efficacy of AG in retarding the rate of progression of renal disease in patients with overt diabetic nephropathy. Pimagedine therapy reduced the 24 h total urinary proteinuria and prevented the decrease in glomerular

filtration rate in patients with diabetes^[22]. However, the effects of Pimagedine on serum creatinine doubling were found not to be significant; serum creatinine doubled in 26% of the placebo-treated patients and in 20% of those who received Pimagedine ($p = 0.099$). This study is noteworthy in providing the first clinical proof of the concept that inhibiting AGE formation can result in a clinically important attenuation of the serious complication of diabetes. Reported side effects of AG in clinical therapy were gastrointestinal disturbance, abnormalities in liver function tests, flu-like symptoms and a rare vasculitis^[23].

Metformin: Metformin (dimethylbiguanide) was introduced into clinical practice in 1957 as an oral anti-hyperglycemic agent for the management of type 2 diabetes^[24]. Metformin is a guanidine compound that is structurally related to AG, thus suggesting that it may also have a potential effect on the inhibition of glycation reactions. Several groups reported inhibitory effects of metformin on protein glycation^[25-27].

A number of studies have shown that metformin is beneficial in reducing diabetes-associated vascular risks beyond the benefits expected from its anti-hyperglycemic effect. Chronic metformin treatment prevented functional and structural alterations of the diabetic myocardium associated with glycation^[28]. Metformin has also been shown to inhibit AGE formation in peripheral nerves and improved their function in diabetic animals^[29]. Blockade of glycation reaction might provide one possible mechanism to explain the beneficial effects of metformin on diabetic vascular complications^[30].

Vitamin: Recently, vitamin B complexes such as pyridoxamine and thiamine pyrophosphate have been found to inhibit the formation of AGEs both *in vitro* and *in vivo*.

Pyridoxamine (PM) (Pyridorin): PM, originally described as a post-Amadori inhibitor (so-called Amadorins) of AGE formation, also inhibits the formation of advanced lipoxidation end-products (ALEs) on protein during lipid peroxidation reaction^[31-34].

Stitt *et al.*^[35] recently reported that PM prevented the development of pericyte loss and formation of acellular capillaries in diabetic rats. PM also inhibited the progression of renal disease and decreased hyperlipidemia and apparent redox imbalances in diabetic rats^[36]. Further, PM was found to inhibit AGE/ALE formation and hyperlipidemia and protected against renal and vascular pathology in non-diabetic obese rats as well^[37]. Phase II trials are ongoing to evaluate the efficacy

of Pyridorin in inhibiting the progression of albuminuria in patients with early stage diabetic kidney disease. The trials are also monitoring plasma triglyceride and cholesterol levels and several other parameters relevant to diabetes and kidney function.

Benfotiamine: Benfotiamine, a lipid soluble compound of thiamine, was found to be a potent inhibitor of glycation^[38]. Hammes and Brownlee *et al.*^[39] have recently found that benfotiamine inhibited the three major biochemical pathways (AGE formation, protein kinase C activation and the polyol pathway activation) as well as hyperglycemia-associated NF- κ B activation. They showed that benfotiamine prevented experimental diabetic retinopathy by activating the pentose phosphate pathway enzyme transketolase in the retinas, which converts glyceraldehyde-3-phosphate and fructose-6-phosphate into pentose-5-phosphates and other sugars^[39].

Thiazolidine structure: Synthetic compounds with thiazolidinedione structure have recently been found to be an effective inhibitor of AGE formation.

OPB-9195: OPB-9195 ((\pm)-2-isopropylidenehydrazono-4-oxo-thiazolidin-5-ylacetamide) is known to trap carbonyl intermediates of advanced glycation with an activity about one order of magnitude more potent than that of AG^[40].

OPB-9195 prevented the progression of diabetic nephropathy by lowering serum concentrations of AGEs and their deposition of glomeruli in OLETF rats^[40]. OPB-9195 was found to retard the progression of diabetic nephropathy in these animals by blocking type IV collagen production and suppressing overproduction of two growth factors, TGF- β and VEGF^[40]. OPB-9195 also prevented the progression of diabetic nephropathy in RAGE-overexpressed mice^[41].

OPB-9195 has been shown to exert beneficial effects on diabetic neuropathy as well; it improved tibial motor nerve conduction velocity and restored the decrease in sciatic nerve Na⁺-K⁺-ATPase activity in diabetic rats, which was in parallel with suppression of oxidative stress-induced DNA damage^[42]. These observations suggest that reactive oxygen species production induced by AGE-RAGE interactions might be involved in endoneurial vascular dysfunction and nerve injury in diabetic neuropathy. Recently, long-term administration of OPB-9195 has been found to reduce blood pressure and oxidative damage in stroke-prone spontaneously hypertensive rats, suggesting a pathological role for AGEs in hypertension^[43].

Piperazine structure: Tenilsetam, (\pm)-3-(2-thienyl)-2-piperazinone, a cognition-enhancing and possible anti-dementia drug was also shown to have the ability to inhibit AGE formation.

Tenilsetam: Tenilsetam was first introduced as an anti-ischemic and then as a cognition-enhancing drug^[44,45]. Now it is used for treatment of patients suffering from AD. Tenilsetam also inhibits reducing sugar-induced polymerization and cross-linking of proteins *in vitro*^[46]. Since aggregation of amyloid β (A β), the major component of senile plaques in AD, is significantly accelerated by protein cross-linking *via* AGEs, tenilsetam could exert beneficial effects on AD, at least in part, by attenuating the AGE-induced A β formation^[47].

AGE breakers: AGEs cause proteins that are normally flexible to become rigid. The cells, tissues and blood vessels exposed to AGEs become stiff and increasingly dysfunctional. In diabetic patients, the extent of protein cross-linking is accelerated due to prolonged diabetic exposure. AGE breakers are one the novel types of AGE inhibitors that could eliminate the AGE products both *in vitro* and *in vivo*.

N-phenacylthiazolium bromide (PTB) and ALT-711: Novel AGE-protein cross-link breakers such as PTB and its stable derivative ALT-711, which are able to selectively cleave and break the established glucose-derived AGE-protein cross-links were reported^[48,49].

Delayed intervention with ALT-711 attenuated the decrease in large artery compliance in diabetic rats^[49]. Further, ALT-711 treatment was found to decrease renal AGE levels and reduce albumin excretion rate in diabetic animals^[50,51]. A recent clinical trial has shown that patients who received ALT-711 experienced statistically significant reduction in arterial pulse pressure and an increase in large artery compliance compared to patients who received placebo^[52].

Inhibitors of the AGE-RAGE signaling

Pigment Epithelium-Derived Factor (PEDF): Compared to the strategies of preventing AGE formation or breaking preformed AGEs, the manipulation of the AGE-RAGE signaling pathways as a therapeutic option in diabetic vascular complications remains much less developed. However, recently, we have found that PEDF, one of the superfamily of serine protease inhibitors with potent neuronal differentiating activity in human retinoblastoma

cells^[53], inhibited the AGE-induced pericyte apoptosis, the earliest histopathological hallmark of early diabetic retinopathy, through its anti-oxidative properties^[54]. Furthermore, PEDF was found to prevent the AGE-elicited chemokine production and angiogenesis, another vascular derangements in diabetic retinopathy^[55-57]. Since the levels of vitreal PEDF are decreased in angiogenic eye diseases such as proliferative diabetic retinopathy^[58], substitution of PEDF may disrupt inappropriate retinal cell responses to AGEs, thus being a promising strategy for treatment of patients with diabetic retinopathy.

There is a growing body of evidence that the AGE-RAGE system is implicated in the pathogenesis of various diseases such as diabetic vascular complications. Therefore, inhibition of AGE formation or blockade of the RAGE signaling may be a promising therapeutic target for the treatment of AGE-related disorders. Effectiveness of several types of AGE inhibitors or RAGE-signal blockers described here should be confirmed by multicenter, randomized, double-blinded clinical trials.

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REFERENCES

1. Vlassara, H., R. Bucala and L. Striker, 1994. Pathogenic effects of advanced glycosylation: Biochemical, biologic and clinical implications for diabetes and aging. *Lab. Invest.*, 70: 138-151.
2. Brownlee, M., 1995. Advanced protein glycosylation in diabetes and aging. *Ann. Rev. Med.*, 46: 223-234.
3. Naka, Y., L.G. Bucciarelli, T. Wendt, L.K. Lee, L.L. Rong, R. Ramasamy, S.F. Yan and A.M. Schmidt, 2004. RAGE axis, animal models and novel insights into the vascular complications of diabetes. *Arterioscler. Thromb. Vasc. Biol.*, 24: 1342-1349.
4. Sasaki, N., R. Fukatsu, K. Tsuzuki, Y. Hayashi, T. Yoshida, N. Fujii, T. Koike, I. Wakayama, R. Yanagihara, R. Garruto, N. Amano and Z. Makita, 1998. Advanced glycation end products in Alzheimer's disease and other neurodegenerative diseases. *Am. J. Pathol.*, 153: 1149-1155.
5. Castellani R., M.A. Smith, P.J. Richey and G. Petty, 1996. Glycoxidation and oxidative stress in Parkinson disease and diffuse Lewy body disease. *Brain Res.*, 737: 195-200.
6. Sasaki, N., M. Takeuchi, H. Choei, S. Kikuchi, Y. Hayashi, N. Nakano, H. Ikeda, S. Yamagishi, T. Kitamoto, T. Saito and Z. Makita, 2002. Advanced Glycation End Products (AGE) and their receptor (RAGE) in the brain of patients with Creutzfeldt-Jakob disease with prion plaques. *Neurosci. Lett.*, 326: 117-120.
7. Kikuchi, S., K. Shinpo, A. Ogata, S. Tsuji, M. Takeuchi, Z. Makita and K. Tashiro, 2002. Detection of N-(carboxymethyl)lysine (CML) and non-CML advanced glycation end products in the anterior horn of amyotrophic lateral sclerosis spinal cord. *Amyotroph. Lateral Scler. Other Motor Neuron Disord.*, 3: 63-68.
8. Brownlee, M., H. Vlassara, A. Kooney, P. Ulrich and A. Cerami, 1986. Aminoguanidine prevents diabetes-induced arterial wall protein cross-linking. *Science*, 232: 1629-1632.
9. Thornalley, P.J., A. Yurek-George and O.K. Argirov, 2000. Kinetics and mechanism of the reaction of aminoguanidine with the α -oxoaldehydes glyoxal, methylglyoxal and 3-deoxyglucosone under physiological conditions. *Biochem. Pharmacol.*, 60: 55-65.
10. Mandarino, L.J., 1992. Current hypotheses for the biochemical basis of diabetic retinopathy. *Diabetes Care*, 15: 1892-1901.
11. Hammes, H.P., S. Martin, K. Federlin, K. Geisen and M. Brownlee, 1991. Aminoguanidine treatment inhibits the development of experimental diabetic retinopathy. *Proc. Natl. Acad. Sci., USA.*, 88: 11555-11558.
12. Frank, R.N., R. Amin, A. Kennedy and T.C. Hohman, 1997. An aldose reductase inhibitor and aminoguanidine prevent vascular endothelial growth factor expression in rats with long-term galactosemia. *Arch. Ophthalmol.*, 115: 1036-1047.
13. Stitt, A.W., T. Bhaduri, C.B. McMullen, T.A. Gardiner and D.B. Archer, 2000. Advanced glycation end products induce blood-retinal barrier dysfunction in normoglycemic rats. *Mol. Cell Biol. Res. Commun.*, 3: 380-388.
14. Moore, T.C., J.E. Moore, Y. Kaji, N. Frizzell, T. Usui, V. Poulaki, I.L. Campbell, A.W. Stitt, T.A. Gardiner, D.B. Archer and A.P. Adamis, 2003. The role of advanced glycation end products in retinal microvascular leukostasis. *Invest. Ophthalmol. Vis. Sci.*, 44: 4457-4464.
15. Vasan, S., P.G. Foiles and H.W. Founds, 2001. Therapeutic potential of AGE inhibitors and breakers of AGE protein cross-links. *Expert Opin. Invest. Drugs*, 10: 1977-1987.

16. Thornalley, P.J., 2003. Use of aminoguanidine (Pimagedine) to prevent the formation of advanced glycation endproducts. *Arch. Biochem. Biophys.*, 419: 31-40.
17. Soulis-Liparota, T., M. Cooper, D. Papazoglou, B. Clarke and G. Jerums, 1991. Retardation by aminoguanidine of development of albuminuria, mesangial expansion and tissue fluorescence in streptozocin-induced diabetic rat. *Diabetes*, 40: 1328.
18. Yamauchi, A., I. Takei, Z. Makita, S. Nakamoto, N. Ohashi, H. Kiguchi, T. Ishii, T. Koike and T. Saruta, 1997. Effects of aminoguanidine on serum advanced glycation endproducts, urinary albumin excretion, mesangial expansion and glomerular basement membrane thickening in Otsuka Long-Evans Tokushima fatty rats. *Diabetes Res. Clin. Pract.*, 34: 127-133.
19. Kelly, D.J., R.E. Gilbert, A.J. Cox, T. Soulis, G. Jerums and M.E. Cooper, 2001. Aminoguanidine ameliorates overexpression of pro-sclerotic growth factors and collagen deposition in experimental diabetic nephropathy. *J. Am. Soc. Nephrol.*, 12: 2098-2107.
20. Twigg, S.N., Z. Cao, S.V. McLennan, W.C. Burns, G. Brammar, J.M. Forbes and M.E. Cooper, 2002. Renal connective tissue growth factor induction in experimental diabetes is prevented by aminoguanidine. *Endocrinology*, 143: 4907-4915.
21. Edelstein, D. and M. Brownlee, 1992. Aminoguanidine ameliorates albuminuria in diabetic hypertensive rats. *Diabetologia*, 35: 96-97.
22. Bolton, W.K., D.C. Cattran, M.E. Williams, S.G. Adler, G.B. Appel, K. Cartwright, P.G. Foiles, B.I. Freedman, P. Raskin, R.E. Ratner, B.S. Spinowitz, F.C. Whittier, J.P. Wuerth and Action I Investigator Group, 2004. Randomized trial of an inhibitor of formation of advanced glycation end products in diabetic nephropathy. *Am. J. Nephrol.*, 24: 32-40.
23. Freedman, B.I., J.P. Wuerth, K. Cartwright, R.P. Bain, S. Dippe, K. Hershon, A.D. Mooradian and B.S. Spinowitz, 1999. Design and baseline characteristics for the aminoguanidine clinical trial in overt type 2 diabetic nephropathy (Action II). *Control Clin. Trials*, 20: 493-510.
24. Bailey, C.J. and R.C. Turner, 1996. Metformin. *N. Engl. J. Med.*, 334: 574-579.
25. Tanaka, Y., H. Iwamoto, T. Onuma and R. Kawamori, 1997. Inhibitory effect of metformin on formation of advanced glycation end products. *Curr. Ther. Res.*, 58: 693-697.
26. Ruggiero-Lopez, D., M. Lecomte, G. Moinet, G. Patereau, M. Lagarde and N. Wiernsperger, 1999. Reaction of metformin with dicarbonyl compounds. Possible implication in the inhibition of advanced glycation end product formation. *Biochem. Pharmacol.*, 58: 1765-1773.
27. Rahbar, S., R. Natarajan, K. Yerneni, S. Scott, N. Gonzales and J.L. Nadler, 2000. Evidence that pioglitazone, metformin and pentoxifylline are inhibitors of glycation. *Clin. Chem. Acta.*, 301: 65-77.
28. Regan, T.J., G.N. Jyothirmayi, C. Laharm and A. Jain, 2001. Left ventricular diastolic dysfunction in diabetic or hypertensive subjects: Role of collagen alterations. *Adv. Exp. Med. Biol.*, 498: 127-132.
29. Tanaka, Y., H. Uchino, T. Shimizu, H. Yoshii, M. Niwa, C. Ohmura, N. Mitsuhashi, T. Onuma and R. Kawamori, 1999. Effect of metformin on advanced glycation endproduct formation and peripheral nerve function in streptozotocin-induced diabetic rats. *Eur. J. Pharmacol.*, 376: 17-22.
30. Beisswenger, P. and D. Ruggiero-Lopez, 2003. Metformin inhibition of glycation processes. *Diabetic Metab.*, 29: 6S95-103.
31. Onorato, J.M., A.J. Jenkins, S.R. Thorpe and J.W. Baynes, 2000. Pyridoxamine, an inhibitor of advanced glycation reactions, also inhibits advanced lipoxidation reactions. Mechanism of action of pyridoxamine. *J. Biol. Chem.*, 275: 21177-21184.
32. Price, D.L., P.M. Rhettt, S.R. Thorpe and J.W. Baynes, 2001. Chelating activity of advance glycation end-product inhibitors. *J. Biol. Chem.*, 276: 48967-48972.
33. Voziyani, P.A., T.O. Metz, J.W. Baynes and B.G. Hudson, 2002. A post-Amadori inhibitor pyridoxamine also inhibits chemical modification of proteins by scavenging carbonyl intermediates of carbohydrate and lipid degradation. *J. Biol. Chem.*, 277: 3397-3403.
34. Metz, T.O., N.L. Alderson, S.R. Thorpe and J.W. Baynes, 2003. Pyridoxamine, an inhibitor of advanced glycation and lipoxidation reactions: a novel therapy for treatment of diabetic complications. *Arch. Biochem. Biophys.*, 419: 41-49.
35. Stitt, A., T.A. Gardiner, N.L. Alderson, P. Canning, N. Frizzell, N. Duffy, C. Boyle, A.S. Januszewski, M. Chachich, J.W. Baynes, S.R. Thorpe and N.L. Anderson, 2002. The AGE inhibitor pyridoxamine inhibits development of retinopathy in experimental diabetes. *Diabetes*, 51: 2826-2832.

36. Degenhardt, T.P., N.L. Alderson, D.D. Arrington, R.J. Beattie, J.M. Basgen, M.W. Steffes, S.R. Thorpe and J.W. Baynes, 2002. Pyridoxamine inhibits early renal disease and dyslipidemia in the streptozotocin-diabetic rat. *Kidney Intl.*, 61: 939-950.
37. Alderson, N.L., M.E. Chachich, N.N. Youssef, R.J. Beattie, M. Nachtigal, S.R. Thorpe and J.W. Baynes, 2003. The AGE inhibitor pyridoxamine inhibits lipemia and development of renal and vascular disease in Zucker obese rats. *Kidney Intl.*, 63: 2123-2133.
38. Stracke, H., H.P. Hammes, D. Werkmann, K. Mavrikakis, I. Bitsch, M. Netzel, J. Geyer, W. Kopcke, C. Sauerland, R.G. Bretzel and K.F. Federlin, 2001. Efficacy of benfotiamine versus thiamine on function and glycation products of peripheral nerves in diabetic rats. *Exp. Clin. Endocrinol. Diabetes*, 109: 330-336.
39. Hammes, H.P., X. Du, D. Edelstein, T. Taguchi, T. Matsumura, Q. Ju, J. Lin, A. Bierhaus, P. Nawroth, D. Hannak, M. Neumaier, R. Bergfeld, I. Giardino and M. Brownlee, 2003. Benfotiamine blocks three major pathways of hyperglycemic damage and prevents experimental diabetic retinopathy. *Natl. Med.*, 9: 294-299.
40. Tsuchida, K., Z. Makita, S. Yamagishi, T. Atsumi, H. Miyoshi, S. Obara, M. Ishida, S. Ishikawa, K. Yasumura and T. Koike, 1999. Suppression of transforming growth factor beta and vascular endothelial growth factor in diabetic nephropathy in rats by a novel advanced glycation end product inhibitor, OPB-9195. *Diabetologia*, 42: 579-588.
41. Yamamoto, Y., I. Kato, T. Doi, H. Yonekura, S. Ohashi, M. Takeuchi, T. Watanabe, S. Yamagishi, S. Sakurai, S. Takasawa, H. Okamoto and H. Yamamoto, 2001. Development and prevention of advanced diabetic nephropathy in RAGE-overexpressing mice. *J. Clin. Invest.*, 108: 261-268.
42. Wada, R., Y. Nishizawa, N. Yagihashi, M. Takeuchi, Y. Ishikawa, K. Yasumura, M. Nakano and S. Yagihashi, 2001. Effects of OPB-9195, anti-glycation agent, on experimental diabetic neuropathy. *Eur. J. Clin. Invest.*, 31: 513-520.
43. Mizutani, K., K. Ikeda, K. Tsuda and Y. Yamori, 2002. Inhibitor for advanced glycation end products formation attenuates hypertension and oxidative damage in genetic hypertensive rats. *J. Hypertens.*, 20: 1607-1614.
44. Saletu, B., H.V. Semlitsch, P. Anderer, F. Resch, O. Presslich and P. Schuster, 1989. Psychophysiological research in psychiatry and neuropsychopharmacology. II. The investigation of antihypoxidotic/nootropic drugs (tenilsetam and co-dergocrine-mesylate) in elderlies with the Viennese Psychophysiological Test-System (VPTS). *Methods Find Exp. Clin. Pharmacol.*, 11: 43-55.
45. Munch, G., Y. Taneli, E. Schraven, U. Schindler, R. Schinzel, D. Palm and P. Riederer, 1994. The cognition-enhancing drug tenilsetam is an inhibitor of protein crosslinking by advanced glycosylation. *J. Neural. Transm. Park. Dis. Dement. Sect.*, 8: 193-208.
46. Munch, G., S. Mayer, J. Michaelis, A.R. Hipkiss, P. Riederer, R. Muller, A. Neumann, R. Schinzel and A.M. Cunningham, 1997. Influence of advanced glycation end-products and AGE-inhibitors on nucleation-dependent polymerization of beta-amyloid peptide. *Biochem. Biophys. Acta*, 1360: 17-29.
47. Shoda, H., S. Miyata, B.F. Liu, H. Yamada, T. Ohara, K. Suzuki, M. Oimomi and M. Kasuga, 1997. Inhibitory effects of tenilsetam on the Maillard reaction. *Endocrinology*, 138: 1886-1892.
48. Vasan, S., X. Zhang, A. Kapurniotu, A. Kapurniotu, J. Bernhagen, S. Teichberg, J. Basgen, D. Wagle, D. Shih, I. Terlecky, R. Bucala, A. Cerami, J. Egan and P. Ulrich, 1996. An agent cleaving glucose-derived protein crosslinks *in vitro* and *in vivo*. *Nature*, 382: 275-278.
49. Wolfenbuttel, B.H., C.M. Boulanger, F.R. Crijns, M.S. Huijberts, P. Poitevin, G.N. Swennen, S. Vasan, J.I. Egan, P. Ulrich, A. Cerami and B.I. Levy, 1998. Breakers of advanced glycation end products restore large artery properties in experimental diabetes. *Proc. Natl. Acad. Sci. USA.*, 95: 4630-4634.
50. Oldfield, M.D., L.A. Bach, J.M. Forbes, D. Nikolic-Paterson, A. McRobert, V. Thallas, R.C. Atkins, T. Osicka, G. Jerums and M.E. Cooper, 2001. Advanced glycation end products cause epithelial-myofibroblast transdifferentiation via the receptor for Advanced Glycation End-products (RAGE). *J. Clin. Invest.*, 108: 1853-1863.
51. Jerums, G., S. Panagiotopoulos, J. Forbes, T. Osicka and M.E. Cooper, 2003. Evolving concepts in advanced glycation, diabetic nephropathy and diabetic vascular disease. *Arch. Biochem. Biophys.*, 419: 55-62.
52. Kass, D.A., E.P. Shapiro, M. Kawaguchi, A.R. Capriotti, A. Scuteri, R.C. deGroot and E.G. Lakatta, 2001. Improved arterial compliance by a novel advanced glycation end-product crosslink breaker. *Circulation*, 104: 1464-1470.

53. Tombran-Tink, J., C.G. Chader and L.V. Johnson, 1991. PEDF: Pigment epithelium-derived factor with potent neuronal differentiative activity. *Exp. Eye Res.*, 53: 411-414.
54. Yamagishi, S., Y. Inagaki, S. Amano, T. Okamoto, M. Takeuchi and Z. Makita, 2002. Pigment epithelium-derived factor protects cultured retinal pericytes from advanced glycation end product-induced injury through its antioxidative properties. *Biochem. Biophys. Res. Commun.*, 296: 877-882.
55. Inagaki, Y., S. Yamagishi, T. Okamoto, M. Takeuchi and S. Amano, 2003. Pigment epithelium-derived factor prevents advanced glycation end products-induced monocyte chemoattractant protein-1 production in microvascular endothelial cells by suppressing intracellular reactive oxygen species generation. *Diabetologia*, 46: 284-287.
56. Yamagishi, S., S. Amano, Y. Inagaki, T. Okamoto, M. Takeuchi and H. Inoue, 2003. Pigment epithelium-derived factor inhibits leptin-induced angiogenesis by suppressing vascular endothelial growth factor gene expression through anti-oxidative properties. *Microvasc. Res.*, 65: 186-190.
57. Yamagishi, S., M. Takeuchi, Y. Inagaki, K. Nakamura and T. Imaizumi, 2003. Role of advanced glycation end products (AGEs) and their receptor (RAGE) in the pathogenesis of diabetic microangiopathy. *Intl. J. Clin. Pharmacol. Res.*, 23: 129-134.
58. Spranger, J., M. Osterhoff, M. Reimann, M. Mohlig, M. Ristow, M.K. Francis, V. Cristofalo, H.P. Hammes, G. Smith, M. Boulton and A.F. Pfeiffer, 2002. Loss of the antiangiogenic pigment epithelium-derived factor in patients with angiogenic eye disease. *Diabetes*, 50: 2641-2645.