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Tracheal Responsiveness to Histamine and Histamine (H₁) Receptor Blockade by Chlorpheniramine in an Animal Model of COPD

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Abstract: Airway Hyperresponsiveness (AHR) is the main feature of asthma which also exists in Chronic Obstructive Pulmonary Disease (COPD). However the mechanism of AHR is uncertain due to the complexity stimuli action used for measuring AHR. The mechanism of competitive antagonism blockade, which is measured as concentration ratio-1 (CR-1) is far simpler than that of agonists and depends only on drug delivery to the receptor sites and receptor affinity. Therefore, in this study we have examined the histamine (H₁) receptor blockade by chlorpheniramine on isolated tracheal chains of a model of COPD compared to control guinea pigs. Experimental models of COPD was induced in guinea pigs by exposing animals to cigarette smoke for three months. The responses of tracheal chains of COPD and control animals (for each group n=7) to cumulative concentrations of histamine (H) in the absence and presence of 10 nM chlorpheniramine were measured and the effective concentrations of H causing 50% of maximum response (EC₅₀ H) were obtained. The chlorpheniramine blockade (CR-1) was calculated by: (post chlorpheniramine EC₅₀ H/EC₅₀ H)-1. In addition, the contractility of tracheal chains due to 50 μM histamine concentration and hematocrit were also measured. The tracheal responses of COPD guinea pigs were significantly higher than those of control animals to histamine (EC₅₀ H for COPD and control animals were 19±2.45 and 34.28±2.06 μM, respectively, p<0.001). The histamine (H₁) receptor blockade by chlorpheniramine (CR-1) was also significantly higher in trachea of COPD compared to that of control animals (2.62±0.51 vs 0.44±0.06, p<0.001). There were significant correlations between tracheal response to EC₅₀ H and (CR-1) (r=-0.564, p<0.05). The hematocrit in COPD animal was significantly higher than control animals (p<0.001). However there were not significantly difference in contractility response of tracheal chains between COPD and control animals. The enhanced histamine (H₁) receptor blockade increased tracheal response to histamine in tracheal chains of COPD animals and significant correlation between these two phenomenon may indicate that increased drug delivery to the receptors could be a determinant factor for bronchial responsiveness to stimuli in COPD.

Key words: Tracheal responsiveness, histamine H₁ receptors blockade, COPD, guinea pig, chlorpheniramine

INTRODUCTION

Chronic Obstructive Pulmonary Disease (COPD) is a global health problem, reaching almost epidemic proportions in the developing world^[1]. The research into understanding of basic mechanisms of the disease and development of new treatment to prevent the progression of this condition presents a major challenge. By leading to a clearer understanding of the key events in the pathophysiology of COPD and enabling short term studies to develop appropriate strategies, animal models can provide a framework for the rational and safe design of expensive and long term clinical studies. In cigarette smokers it is believed that airflow obstruction is caused by parenchyma disease (emphysema) and/or by smoke-

induced distortion of the structure of the small airways^[2-4]. Recently a model of cigarette smoke-induced lung disease in which guinea pigs develop airflow obstruction and emphysematous lung destruction has been described^[5].

There are reports regarding airway hyperresponsiveness to different stimuli in animals exposed to cigarette smoke^[6-10], but little is known regarding the mechanism(s) of increased airways responsiveness in animal exposed to cigarette smoke as well as asthmatic patients. This is mainly due to complexity of action of stimuli using in measuring AHR.

The mechanism(s) action of a competitive antagonist measured as the degree of rightward shift or dose ratio or concentration ratio (DR or CR) is far simpler than that of

an agonist and depends only to concentration of antagonist at the receptor sites [I] and receptor affinity (K_a)^[11]. Thus, receptor blockade by a competitive antagonist could be inside the mechanism (s) of airway responsiveness. In previous studies we demonstrated enhanced blockade of different receptors by their antagonists in both asthmatic patients^[12-14] and sensitized animals^[15,16].

Therefore, in the present study tracheal responsiveness of guinea pigs exposed to cigarette smoke to histamine and histamine (H₁) receptor blocked by chlorpheniramine was studied.

MATERIALS AND METHODS

Animals and cigarette smoke exposure: Fourteen adult Dunkin-Hartley guinea pigs (400 to 500 g) of both sexes were divided into two groups of 7 experimental and 7 controls. Experimental animals were exposed to cigarette smoke as previously described^[17,18]. The animals were exposed to cigarette smoke in an awake, restrained state and spontaneously breathing in a smoking chamber which was a modification of that described by Simami *et al.*^[18]. Animals were placed in a plexiglas box with their heads secured in a compartment (15x12x7 cm). Twenty millilitre puffs of cigarette smoke was drawn out of cigarettes with syringes and then exhausted at a rate of two puffs per minute into the animal's head chamber. Exposure of animals to each cigarette lasted for 8-9 min, with a 10 min resting period between cigarettes. The animals were initially exposed to one commercial non-filter cigarette per day and this dose was increased to a maximum of 5 cigarettes per day over a 2-week period. In a pilot study it was observed that animals couldn't tolerate the exposure of cigarette smoke of more than 5 cigarettes per day. The exposure to the smoke of 5 cigarettes per day, 6 days per week, continued for 3 months. The guinea pigs exposed to cigarette smoke were called COPD animals. The control animals were not exposed to cigarette smoke and they were kept in the animal house under normal conditions for the same period of time. The study was approved by the ethical committee of Mashhad University of Medical Sciences.

Tissue preparations: Guinea pigs were killed by a blow on the neck and trachea were removed. Each trachea was cut into 10 rings (each containing 2-3 cartilaginous rings). All the rings were then cut open opposite the trachealis muscle and sutured together to form a tracheal chain^[19,20]. Tissue was then suspended in a 10 mL organ bath (organ bath 61300, BioScience Palmer-Washington, Sheerness, Kent, UK) containing Krebs-Henseleit

solution of the following composition (mmol/L): NaCl 120, NaHCO₃ 25, MgSO₄ 0.5, KH₂PO₄ 1.2, KCl 4.72, CaCl₂ 2.5 and dextrose 11.

The Krebs solution was maintained at 37°C and gassed with 95% O₂ and 5% CO₂. Tissue was suspended under isotonic tension of 1 g and allowed to equilibrate for at least 1 h while it was washed with Krebs solution every 15 min.

Measurement of tracheal response to histamine and histamine H₁ receptor blockade: In each experiment two cumulative Log Concentration-response Curves (LCRC) of histamine phosphate (MW=308 BDH Chemical Co. Ltd, UK) induced contraction of tracheal chain were obtained, one 10 min after producing 10 nM concentration of chlorpheniramine maleate (MW=391, Sigma Chemical Ltd, UK) in organ bath (adding 0.1 mL of 1 μM chlorpheniramine solution to organ bath= post chlorpheniramine histamine response curve) and the other 10 min after adding the same volume of saline (baseline histamine response curve).

Cumulative log concentration-response curve of tracheal chain to increasing concentrations of histamine (0.1 μM to 10 mM) was obtained with addition of consecutive concentrations every 2 min. To obtain the curve the percentage of contraction of the tracheal smooth muscle due to each concentration of histamine in proportion to the maximum contraction obtained, in baseline histamine response curve, was calculated and plotted against log concentration of histamine.

The effective concentration of histamine causing 50% of maximum response (EC₅₀ H) of baseline and post chlorpheniramine histamine response curve in each experiment was measured (expressed as EC₅₀ H and post chlorpheniramine EC₅₀ H, respectively). The tracheal response to histamine was considered as EC₅₀ H. The histamine H₁ receptor blockade by chlorpheniramine was assessed as concentration ratio minus one (CR-1) which was calculated by: (post chlorpheniramine EC₅₀ H/EC₅₀ H)-1.

The experiments for measuring post chlorpheniramine histamine response curve and baseline histamine response curve in each tracheal chain were performed randomly with 1 h resting period between each of the two experiments while washing the tissues every 10 min. Tracheal responses to histamine were tested on incubated tissues with 1.4 μM indomethacin 30 min prior and during obtaining LCRC in the presence of both saline and chlorpheniramine. In all experiments responses were recorded on a kymograph (ET8 G-Boulitt, Paris) and measured after fixation.

Pathological evaluation: The lungs of all COPD animals underwent normal pathological evaluation using hematoxin-eosin dye by a pathologist.

Statistical analysis: The data of tracheal response to histamine (EC₅₀ H), histamine H₁ receptor blockade (CR-1) maximum response, muscle contractility and hematocrit were quoted as mean±SEM. In comparing all values between COPD and control guinea pigs unpaired “t” were employed. Tracheal response to histamine (EC₅₀ H), was related to (CR-1), using the least square regression. Significance was accepted at p<0.05.

RESULTS

Histology: The following pathological changes were observed in the lung of animals exposed to cigarette smoke:

- 1) Increased intra-alveolar septum in all specimens (infiltration of leukocytes).
- 2) Increased lymphatic tissue in the lung parenchyma of all specimens.
- 3) The destruction of alveolar wall and existence of emphysema in the lungs of most animals.
- 4) Intra-alveolar bleeding in most animal’s lungs.

Tracheal response to histamine: The mean value of EC₅₀ H in tracheal chains of COPD animals (19±2.49 μM, range 8-25 μM) was significantly lower than in control animals (34.28 ±2.06 μM, range 28-45 μM, p<0.001), (Fig. 1a and Table 1). The most responsive trachea of COPD animals was 5.62 times more sensitive to histamine than the least responsive trachea from control animals.

Chlorpheniramine blockade (CR-1): The rightward shift of the post chlorpheniramine histamine response curve compared to the baseline histamine response curve in tracheal chains of COPD animals was greater than that of control animals (Fig. 2). Mean CR-1 in tracheal chains of COPD animals (2.62±0.51, range 0.3-4.8) was 6 times greater than in control animals (0.44±0.06, range 0.2-0.7, p<0.001), (Fig. 1b and Table 1). The value of CR-1 of the most sensitive trachea of COPD animals was 24 times greater than that for the least sensitive trachea from control animals.

Relationship between bronchial response to histamine and chlorpheniramine blockade: There was a significant negative correlation between tracheal response to histamine (EC₅₀ H) and histamine H₁ receptor blockade by chlorpheniramine (r=-0.564, p<0.05; Fig. 3).

Table 1: Values of tracheal response to histamine (EC₅₀ H), histamine (H₁) receptor blockade by chlorpheniramine (CR-1), hematocrit and contractility response of tracheal chains (contraction response to 50 μM concentration of histamine) and in tracheal chains of control and COPD, (for each group, n=7) animals and statistical differences between the two groups

Tracheal response	Control	COPD	Sta. Dif.
EC ₅₀ (μmol)	34.28±2.06	19.00±2.45	p<0.001
(CR-1)	0.44±0.06	2.62±0.51	p<0.001
Contractility	0.18±0.03	0.20±0.03	NS
Hematocrit	21.80±0.91	34.57±2.12	p<0.001

Values are quoted as mean±SEM. Sta. Dif.; Statistical difference

Contractility: The mean value of the post chlorpheniramine maximum response in tracheal chains of control and COPD animals (99.71±2.85 and 86.28±11.67, respectively) were not significantly lower than maximum response in the presence of saline (100±0.00% for both cases). The mean value of contractility in tracheal chains of COPD animals (0.20±0.04 g, range 0.07-0.36 g) also was not significantly higher than control animals (0.18±0.03 g, range 0.07-0.33).

Hematocrit: The mean value of hematocrit in COPD animals (34.57±2.125%, range 25-44) was significantly higher than control animals (21.8±0.91%, range 20-25, p<0.001).

DISCUSSION

This study showed increased tracheal response to histamine in guinea pig exposed to cigarette smoke (COPD) compared to control animals. The histological findings in animal exposed to cigarette smoke showed increased intra-alveolar septum (infiltration of leukocytes), increased lymphatic tissue, destruction alveolar walls and intra-alveolar bleeding. Therefore, the pathological changes in the lungs of these animals are similar to those of COPD patients^[21-23]. The pathological changes in the lungs of animals exposed to cigarette smoke were also similar to those of previous studies^[5,24,25]. The hematocrit in COPD animals were also significantly higher than control group. The pathological changes in the lung of the guinea pigs exposed to cigarette smoke and increased hematocrit in these animals confirmed the induction of COPD in the experimental group of animals. Tracheal responsiveness to histamine seen in COPD guinea pigs was similar to the results of our previous study in asthmatic patients^[13].

In addition several other studies showed AHR in guinea pigs exposed to cigarette smoke to different stimuli^[6-10]. However, the results of the present study also demonstrated an increased histamine H₁ receptor blockade by chlorpheniramine (CR-1) in COPD animals. The increased receptor blockade by a competitive

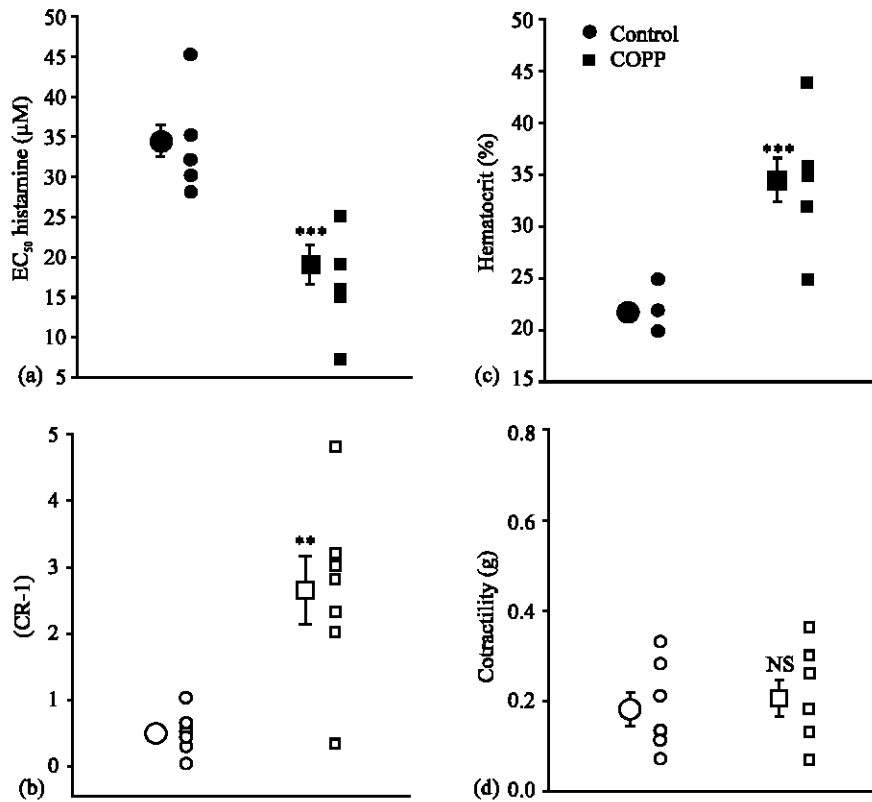


Fig. 1: Individual values and mean±SEM (big symbols with bars) of tracheal response to histamine (EC₅₀ H) (a), histamine (H₁) receptor blockade by chlorpheniramine (CR-1) (b), hematocrit (c) and contractility response of tracheal chains (contraction response to 50 µM concentration of histamine) (d), in tracheal chains of control (circle symbols) and COPD (square symbols) (for each group, n=7). Statistical differences between values in COPD with those of control animals: NS; Non-significant, *, p<0.05, **, p<0.01, ***, p<0.001

antagonist was similar to the results of our previous *in vitro* results indicating increased muscarinic and histamine (H₁) receptors by atropine^[16] and chlorpheniramine^[15] in sensitized guinea pigs, respectively. The consistent increase in antagonist blockade in the present study and our previous studies in sensitized animals^[15,16] indicated that the cause of enhanced receptor blocked is increase of either receptor affinity (K_a) or drug delivery to the receptors ([I]) or both of these factors^[1]. The fact that two receptor systems showed enhanced competitive antagonist blockade in sensitized animals^[15,16] and one receptor system in cigarette exposed animals in *in vitro* studies suggests that the abnormality lies with [I] rather than K_a. This conclusion is also supported by *in vitro* experiments, which predict that receptor affinity for a given antagonist shows little variation between species and tissues^[26]. We therefore suggest that this enhanced antagonist blockade may be caused by epithelial damage leading to increased

epithelial permeability and accessibility of ligands to the receptor sites.

In fact, the increased airway epithelial permeability to different agents has been demonstrated in animals exposed to cigarette smoke^[18,27] and in smokers^[28]. The existence of airway inflammation in animal exposed to cigarette smoke^[17,29,30], smokers^[28,31] and COPD^[32,33] is well documented. Thus airway inflammation can cause epithelial damage; and this, in turn, can result in better access of ligands to the active sites in the airways, causing increased receptor blockade by competitive antagonists.

However, our previous studies^[12-14] also showed enhanced blockade when pharmacological antagonists were administered by i.v. injection in asthmatic patients and this cannot be due to increased epithelial permeability. The strongest possibility is increased tissue permeability due to airway inflammation. If the permeability of a physical barrier of some kind close to the

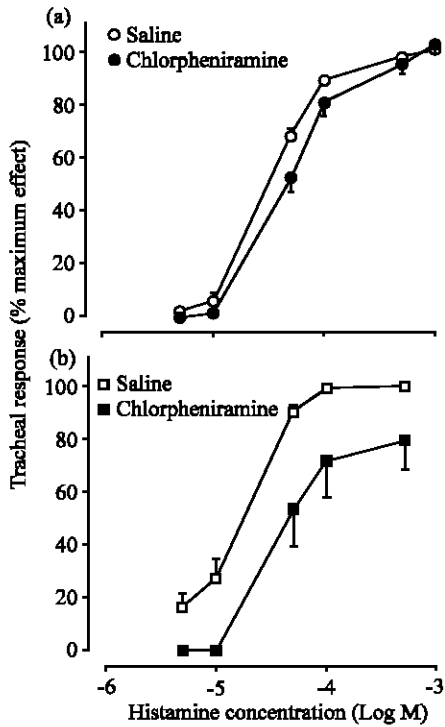


Fig. 2: Cumulative log concentration-response curves of histamine induced contraction of isolated trachea in the presence of saline and chlorpheniramine of control (a) and COPD animals (b), (for each group, n=7)

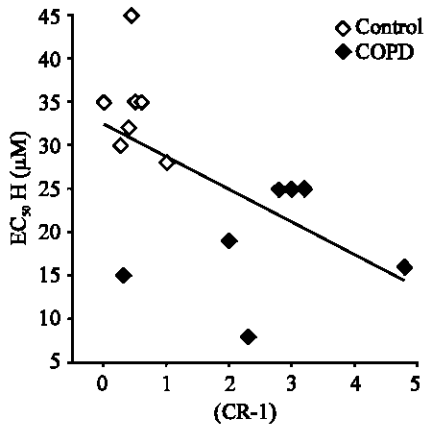


Fig. 3: Correlation between tracheal response to histamine (EC₅₀ H); and muscarinic receptor blockade by chlorpheniramine (CR-1) in all control and COPD guinea pigs (n=14): r=-0.542, p<0.01

receptor was increased in COPD and asthma, it could increase diffusion of antagonist ligands administered by either way and would explain the variation in (CR-1) produced by both routes of administration. In addition,

the results of the present *in vitro* study cannot be fully explained by increased epithelial permeability because the barrier role of epithelium against ligands diffusion^[34] may be appreciated only when perfused tracheal or bronchial tubes are exposed to ligands from the mucosal sides but not in the model used in this study, using tracheal rings. A stripe of tracheal smooth muscle is covered by epithelium only from one side and easily exposed to chemicals from the remaining three sides; this mostly excludes the barrier function of the epithelium. The increased chlorpheniramine blockade in tracheal chains of COPD guinea pigs exposed to cigarette smoke shown in this study, as well as increased chlorpheniramine, atropine and propranolol blockade in our previous *in vivo* studies in asthmatic patients^[12-14] and enhanced atropine and chlorpheniramine blockade in tracheal chains of sensitized animal studies^[15,16], is perhaps due to higher concentration of antagonists at the receptor sites achieved by an increased epithelial and tissue permeability leading to an increase in [I]. In addition, destruction of lung parenchyma is also documented in exposed animals to cigarette smoke^[17]. This can support an increased tissue permeability and better accessibility of ligands to the receptor sites.

The results of this study also showed significant correlations between histamine H₁ receptor blockade by chlorpheniramine (CR-1) and tracheal response to histamine. The significant correlation between (CR-1) and tracheal response to histamine in the present study, as well as blockade by other antagonists and agonist responsiveness in previous studies^[12-16], indicates that bronchial hyperresponsiveness to different stimuli in COPD and asthma, at least in part, is due to increased bronchial epithelial and tissue permeability which is perhaps due to airway inflammation in these diseases. If this is true, the treatment of these diseases should be focused on preventing this increased permeability.

The present *in vitro* study indicated higher chlorpheniramine blockade at histamine H₁ receptors than tracheal response to histamine in tracheal chains of COPD guinea pigs compared to control animals (5.9 times for chlorpheniramine blockade and 1.7 times for histamine response).

In addition the contractility response of tracheal chains to histamine in COPD group was similar to the control animals. These findings suggested that mainly the prejunctional mechanisms were responsible for increased tracheal responsiveness in COPD animals^[35]. These mechanisms include: epithelial damage, neural control, inflammatory process and metabolism or absorption which affect antagonism blockade more than response to an agonist.

In conclusion, this study demonstrated enhanced histamine H₁ receptor blockade by chlorpheniramine in tracheal chains of COPD guinea pigs exposed to cigarette smoke. The cause of this enhanced antagonist blockade at the tracheal chains is perhaps increased epithelial and tissue permeability due to airway inflammation. The increased epithelial and tissue permeability of the airways appears, at least in part, is responsible for airway hyperresponsiveness.

REFERENCES

1. Dawkins, P.A. and R.A. Stockley, 2001. Animal models of chronic obstructive pulmonary disease. *Thorax*, 56: 972-977.
2. Bosken, C.H., B.R. Wiggs, P.D. Pare and J.C. Hogg, 1990. Small airway dimensions in smokers with obstruction to air flow. *American Review of Respiratory Diseases*, 42: 563-570.
3. Matsuba, K., J.L. Wright, B.R. Wiggs, P.D. Pare and J.C. Hogg, 1989. The change in airway structure associated with reduced forced expiratory volume in one second (FEV₁). *European Respiratory J.*, 2: 834-839.
4. Wright, J.L., L.M. Lawson, P.D. Pare, S. Kennedy, B. Wiggs and J.C. Hogg, 1984. The detection of small airways disease. *American Review of Respiratory Diseases*, 129: 898-994.
5. Wright, J.L. and A. Chung, 1990. Cigarette smoke causes physiological and morphological changes of emphysema in the guinea pig. *American Review of Respiratory Diseases*, 142: 1422-1426.
6. Dusser, D.J., T.D. Djokic and D.B. Borson, 1989. Cigarette smoke induces bronchoconstrictor hyperresponsiveness to substance P and inactivates airways neutral endopeptidase in the guinea pig. *J. Clinical Investigation*, 84: 900-906.
7. Han-Pin, K. and L. Ling-Chung, 1995. Sensory neuropeptides modulate cigarette smoke-induced decrease in neutral endopeptidase activity in guinea pig airways. *Life Sci.*, 57: 2187-2196.
8. Hulbert, W.M., T. Mclean and J.C. Hogg, 1985. The effect of acute airway inflammation on bronchial reactivity in guinea pig. *American Review of Respiratory Diseases*, 132: 7-11.
9. James, A.L., P. Dirks and H. Ohtaka, 1987. Airway responsiveness to intravenous and inhaled acetylcholine in the guinea pig after cigarette smoke exposure. *American Review of Respiratory Diseases*, 136: 158-1162.
10. Lee, L.Y., Y.P. Lou, J.L. Hong and J.M. Lundberg, 1995. Cigarette smoke-induced bronchoconstriction and release of tachykinins in guinea pig lungs. *Respiratory Physiology*, 99: 173-181.
11. Arunlakshana, O. and H.O. Schild, 1959. Some quantitative uses of drug antagonists. *British J. Pharmacol.*, 14: 48-58.
12. Boskabady, M.H. and P.D. Snashall, 1992. Enhanced muscarinic receptor blockade with atropine in the asthmatic tracheobronchial tree: Evidence for increased drug delivery. *American Review of Respiratory Diseases*, 145: 756-761.
13. Boskabady, M.H. and P.D. Snashall, 1997. Enhanced histamine H₁ receptor blockade with chlorpheniramine in the asthmatic tracheobronchial tree: Further evidence for increased drug delivery in asthma. *Med. JIRI.*, 11: 115-122.
14. Boskabady, M.H. and P.D. Snashall, 2000. Bronchial responsiveness to beta-adrenergic stimulation and enhanced beta-blockade in asthma. *Respirology*, 5: 111-118.
15. Boskabady, M.H., M. Harati and S. Adel Kardan, 1998. Enhanced chlorpheniramine blockade in isolated tracheal chains of asthmatic guinea-pigs. *Med. JIRI.*, 12: 265-271.
16. Boskabady, M.H. and S. Adel-Kardan, 1999. Increased muscarinic receptor blockade by atropine in tracheal chains of ovalbumine-sensitized guinea pigs. *Pharmacology*, 58: 300-308.
17. Sansores, R.H., R.T. Abboud, C. Becerril, M. Montano, C. Ramos, B. Vanda and M.L. Selman, 1997. Effect of exposure of guinea pigs to cigarette smoke on elastolytic activity of pulmonary macrophages. *Chest*, 112: 214-19.
18. Simani, A.S., S. Inoue and J.C. Hogg, 1974. Penetration of the respiratory epithelium of guinea pigs following exposure to cigarette smoke. *Laboratory Investigation*, 31: 75-81.
19. Holroyde, M.C., 1986. The influence of epithelium on the responsiveness of guinea-pig isolated trachea. *British J. Pharmacol.*, 87: 501-507.
20. McCaig, D., 1987. Comparison of autonomic responses in the trachea isolated from normal and albumin-sensitive guinea pig. *British J. Pharmacol.*, 92: 809-816.
21. Jeffery, P.K., 1997. Pathology of asthma and COPD: A synopsis. *European Respiratory Review*, 7: 111-118.
22. Rennard, S., 1997. Pathophysiological mechanisms of COPD. *European Respiratory Review*, 7: 206-210.
23. Saetta, M., W. Timens, P.K. Jeffery, D.S. Postma and N.M. Siafakas, 1998. Management of Chronic Obstructive Pulmonary Disease. *European Respiratory Monograph* 7.
24. Selman, M., M. Montano and B. Ramos, 1996. Tobacco smoke-induced lung emphysema in guinea pigs is associated with increased interstitial collagenase. *American J. Physiol.*, 271: L734-L743.

25. Wright, J.L. and A. Churg, 1995. Smoke-induced lung emphysema in guinea pig is associated with morphometric evidence of collagen breakdown and repair. *American J. Physiol.*, 268: L17-L20.
26. Bowman, W.C. and M.J. Rand, 1980. *Textbook of Pharmacology* (2nd Edn.). Blackwell Scientific Publications, Oxford, pp: 39, 25.
27. Burns, A.R., S.P. Hosford and L.A. Dunn, 1989. Respiratory epithelial permeability after cigarette smoke exposure in guinea pigs. *J. Apply Physiol.*, 66: 2109-2116.
28. Taylor, R.G., J.E. Agnew, R.A. Francis, D. Pavia and S.W. Clarke, 1988. Respiratory epithelial permeabilities unrelated to bronchial reactivity and small airway function in young smokers and nonsmokers. *European Respiratory J.*, 1: 319-323.
29. Matsumoto, K., H. Aizawa and H. Inoue, 1998. Eosinophilic airway inflammation induced by repeated exposure to cigarette smoke. *European Respiratory J.*, 12: 387-394.
30. Nisikawa, M., N. Kakemisu and T. Ito, 1999. Superoxide mediates cigarette smoke-induced infiltration of neutrophils into the airways through nuclear factor-B activation and IL-8 mRNA expression in guinea pigs *in vivo*. *American J. Respiratory Cellular and Molecular Biol.*, 20: 189-198.
31. Finkelsteine, R., R.S. Fraser and H. Ghezzeo, 1995. Alveolar inflammation and its relation to emphysema in smokers. *American J. Respiratory and Critical Care Medicine*, 152: 1666-1672.
32. Cosio, M.G., J. Majo and M.G. Cosio, 2002. Inflammation of the airways and lung parenchyma in COPD: Role of T cells. *Chest*, 121: 160s-165s.
33. Pettersen, C.A. and K.B. Adler, 2002. Airways inflammation and COPD. *Chest*, 121: 142s-150s.
34. Mitchell, H.W., K.F. Willet and M.P. Sparrow, 1989. Perfused bronchial segment and bronchial strip: Narrowing vs isometric force by mediators. *J. Apply Physiol.*, 66: 2704-2709.
35. Sterk, P.G. and E.H. Bel, 1989. Bronchial hyperresponsiveness: The need for a distinction between hypersensitivity and excessive airway narrowing. *European Respiratory J.*, 2: 267-274.