

International Journal of Pharmacology

ISSN 1811-7775





Antigenotoxic Effect of Genistein and Gingerol on Genotoxicity Induced by Norethandrolone and Oxandrolone in Cultured Human Lymphocytes

Tanveer Beg, Yasir Hasan Siddique, Gulshan Ara, Jyoti Gupta and Mohammad Afzal Human Genetics and Toxicology Laboratory, Department of Zoology, Aligarh Muslim University, Aligarh (UP), 202002, India

Abstract: Norethandrolone and Oxandrolone were studied for their genotoxic effect on human lymphocyte chromosomes using chromosomal aberrations (CAs) and sister chromatid exchanges (SCEs) as parameters and subsequently Genistein and Gingerol were used as antigenotoxic agents to ameliorate the genotoxicity induced by the steroids. This experiment was aimed at finding the dosage at which these two steroids are genotoxic enough to cause chromosome damage. They were studied at 5, 10, 20, 30 and 40 μ M, respectively and were found to be significantly genotoxic at 30 and 40 μ M. Genistein and Gingerol proved to be equally effective in reducing genotoxic damage at appropriate doses. The results suggest a strong genotoxic effect of both steroids *in vitro* in human lymphocytes and also a significant antigenotoxic action of Genistein and Gingerol against steroid induced genotoxic damage.

Key words: Norethandrolone, oxandrolone, genistein, gingerol, androgens, genotoxicity, chromosomal aberrations, sister chromatid exchanges, human lymphocytes

INTRODUCTION

Genistein is an anticarcinogenic isoflavone found in soybean seeds in the form of glycosides. It has been demonstrated that genistein has a great number of biological activities in animal cells including weak estrogenic effect, inhibition of different kinases (including tyrosine kinase), antioxidant properties and modulation of cell proliferation and transformation (Muster et al., 1997). Epidemiological and animal model studies have shown a relationship between a diet high in soyfoods and a gamut of beneficial effects with regard to health, including reduced incidence of breast and prostate cancers, cardiovascular disease and post-menopausal ailments (Dixon and Ferreira, 2002). Several studies suggest that genistein exerts a protective effect against lipid peroxidation of low and high density lipoproteins (LDL and HDL) (Patel et al., 2001; Ferretti et al., 2004). Genistein is synthesized from the flavonone naringenin in plants through a ring migration reaction catalyzed by the cytochrome P450 enzyme isoflavone synthase (IFS). IFS genes have recently been cloned from many plant species and now genistein can be produced in nonlegumes through recombinant DNA technique (Dixon and Ferreira, 2002).

The key constituents of Ginger (*Zingiber officinale*) include volatile oils, oleoresin (Gingerol, shogaol), phenol

(Gingerol, zingerone), vitamins and minerals. Ginger has medicinal purposes (Huang *et al.*, 1990). It has carminative, diaphoretic and antispasmodic properties, reduces clotting, cholesterol and blood pressure, is anti-inflammatory, helpful in nausea and indigestion.

Naturally occurring anabolic steroids are synthesized in the testis, ovary and adrenal gland from cholesterol via pregnenolone. Synthetic anabolic steroids are based on the principal male hormone testosterone (Haynes and Murad, 1985). They bind to receptors in reproductive tissue, muscles and fat (Mooradian et al., 1987). Norethandrolone and Oxandrolone are oral anabolicandrogenic steroids (AAS), synthesized and approved under the brand names Nilevar and Anavar, respectively. They have moderate progestagenic activity and share liver toxicity issues common to 17-alkylated steroids. overdosage produce Acute can nausea gastrointestinal upset. Chronic use of Norethandrolone can lead to excessive androgens: menstrual irregularities and virilization in women and impotence, premature cardiovascular disease and prostatic hypertrophy in men (International Programme on Chemical Safety, 1993).

Precocious prostatic cancer has been described after long term anabolic steroid abuse (Roberts and Essenhigh, 1986). Cases where hepatic cancers have been associated with anabolic steroid abuse have been reported (Overly *et al.*, 1984). Also, androgen ingestion by a

pregnant mother can cause virilization of a female fetus (Dewhurst and Gordon, 1984). Oxandrolone is used in the treatment of anemia and hereditary angioedema, which causes episodes of swelling of the face, extremities, genitals, bowel wall and throat (Gannon, 1994) and also prescribed for a number of medical disorders causing involuntary weight loss, in order to promote muscle regrowth and in treating cases of osteoporosis. The only legitimate therapeutic indications for such anabolic steroids are in the replacement of male sex steroids in men who have androgen deficiency, e.g., due to loss of both testes, in the treatment of certain rare forms of aplastic anemia which are or may be responsive to anabolic androgens and in certain countries to counteract catabolic states, e.g., after major trauma (International Programme on Chemical Safety, 1993). The steroids have been reported to be genotoxic in human lymphocytes in vitro and in mouse bone-marrow cells (Dhillon et al., 1994; Hundal et al., 1997; Joosten et al., 2004; Siddique and Afzal, 2004a, b, c; Siddique and Afzal, 2005a; Siddique et al., 2005a,b, 2006a). The genotoxic effects of steroids can be reduced by the use of antioxidants and natural plant products (Ahmad et al., 2004; Siddique and Afzal, 2005b; Siddique et al., 2005b, 2006b, 2007a, b, c; Beg et al., 2007; Siddique et al., 2008a, b). The above studies are concerned with the estrogens and synthetic progestins. The present study deals with the androgens that are used in the treatment of various hormonal diseases and for boosting muscle growth in patients and also in body-building (International Programme on Chemical Safety, 1993).

The use of these two steroids by humans and their ability to act as genotoxic agents, as suggested by earlier studies was an ample reason for us to examine their genotoxic potential using cultured human lymphocytes. In this study, we primarily analyse the antigenotoxic effect of Genistein and Gingerol separately on the frequency of CAs and SCEs induced by Norethandrolone and Oxandrolone respectively, *in vitro* in human lymphocytes.

MATERIALS AND METHODS

Time and place of study: The study was conducted from April-September, 2007, in the Human Genetics and Toxicology Laboratory, Department of Zoology, AMU, Aligarh (UP), India.

Chemicals and reagents: Norethandrolone; Oxandrolone; Genistein (Sigma); Gingerol (Sigma); RPMI-1640 (Gibco); Fetal calf serum (Gibco); 0.1 mL Antibiotic-antimycotic mixture (Gibco); Phytohaemagglutinin-M (Gibco);

Colchicine (Microlab); Hoechst 33258 stain (Fluka); 3% Giemsa solution in phosphate buffer, pH 6.8 (Merck); 5-Bromo-2-deoxyuridine, BrdU (Sigma); Dimethylsulphoxide, DMSO (Merck); Mitomycin C (Merck).

Lymphocyte culture and chromosomal aberration analysis: Peripheral blood cultures were done in duplicate (Carballo *et al.*, 1993). Heparinized blood (0.5 mL) samples were taken from healthy female donors and were placed in sterile tubes containing 7 mL of RPMI-1640, supplemented with 1.5 mL of fetal calf serum and 0.1 mL of phytohaemagglutinin and incubated at 37°C for 24 h. Norethandrolone and Oxandrolone, both dissolved in dimethyl sulphoxide (DMSO), were added separately to different cultures at 5, 10, 20, 30 and 40 μM . DMSO (5 μl mL $^{-1}$) served as negative control.

Two hours before harvesting, 0.2 mL of colchicine (0.2 µg mL⁻¹) was added to the culture tubes. Cells were centrifuged at 800-1000 rpm for 10 min. The supernatant was removed and 5 mL of pre-warmed (37°C) 0.075 M KCl hypotonic solution was added. Cells were resuspended and incubated at 37°C for 15 min. The supernatant was removed by centrifugation and 5 mL of fixative (methanol: glacial acetic acid, 3:1) was added. The fixative was removed by centrifugation and the procedure was repeated twice. The slides were stained in Giemsa solution in phosphate buffer for 15 min. At least 300 metaphases were analyzed for different types of chromosome breakage frequencies. Classification of aberrations was done according to the guidelines of the International Programme on Chemical Safety (1985) WHO, Geneva, for the study of genetic defects in human population.

Sister chromatid exchange analysis: For SCE analysis, BrdU (10 μg mL⁻¹) was added at the beginning of the culture. After 24 h, same treatments were given as for CA analysis. Two hours before harvesting, 0.2 mL of colchicine (0.2 μg mL⁻¹) was added, followed by hypotonic treatment and fixation and processing of slides (Perry and Wolff, 1974) was done as for CA analysis. Slides were stained for 20 min in 0.05% (w/v) Hoechst 33258 solution, rinsed with tap water and placed under a UV lamp for 90 min covered with Sorensen's buffer (pH 6.8) and stained with Giemsa solution in phosphate buffer for 15 min. The sister chromatid exchange average was taken from an analysis of 50 metaphases.

Statistical analysis: Student's t-test was used for calculating the statistical significance in CAs and SCEs. The level of significance was tested using standard statistical tables (Fisher and Yates, 1963).

RESULTS

Both Genistein and Gingerol proved to be equally effective in reducing the number of abnormal cells with CAs induced by Norethandrolone and Oxandrolone at all their tested doses. A significant increase in CAs was

observed at 30 and 40 μM of both norethandrolone and oxandrolone (Table 1, 2). The dose-dependent increase in CAs was not much from 30 to 40 μM .

A dose-dependent decrease in number of abnormal metaphases was observed when the genotoxic doses of norethandrolone were treated with 30 and 40 μ M of

Table 1: Ameliorative action of genistein and gingerol on CAs induced by norethandrolone in cultured human lymphocytes

	No. of abnormal cells (%±SE)	Total CAs				
Treatments		Gaps	CTB	CSB	CTE	DIC
Norethandrolone (µM)						
5	7 (2.33±0.87)	2	4	3	-	-
10	8 (2.67±0.93)	3	5	3	-	-
20	10 (3.33±1.04)	4	6	3	-	-
30	21 (7.00±1.47) ^b	8	13	7	-	-
40	27 (9.00±1.65)b	11	18	6	-	-
Genistein (μM)						
30	4 (1.33±0.66)	1	2	2	-	-
40	5 (1.67±0.74)	2	3	2	-	-
Genistein + Norethandrolone						
30 + 30	15 (5.00±1.26)°	5	9	6	-	-
30 + 40	21 (7.00±1.47)°	8	13	8	-	-
40 + 30	11 (3.67±1.09)°	4	7	4	-	-
40 + 40	17 (5.67±1.34)°	6	11	6	-	-
Gingerol (μM)						
20	3 (1.00±0.57)	1	2	1	-	-
30	4 (1.33±0.66)	2	3	1	-	-
Gingerol + Norethandrolone						
20 + 30	13 (4.33±1.18) ^c	5	8	5	-	-
20 + 40	18 (6.00±1.37)°	6	10	8	_	-
30 + 30	9 (3.00±0.98)°	3	5	4	_	-
30 + 40	11 (3.67±1.09)°)	4	7	4	-	-
Untreated	3 (1.00±0.57)	2	2	1	-	-
Negative control (DMSO, 5 μl mL ⁻¹)	3 (1.00±0.57)	1	2	1	-	-
Positive control (Mitomycin C, 0.3 µg mL ⁻¹)	60 (20.00±2.31°)	28	45	20	4	2

*Significant difference with respect to untreated (p<0.05), *Significant difference with respect to untreated (p<0.01), Significant with respect to Norethandrolone (p<0.05), SE: Standard Error, CTB: Chromatid Break, CSB: Chromosome Break, CTE: Chromatid Exchange, DIC: Dicentric Chromosome

Table 2: Ameliorative action of genistein and gingerol on CAs induced by Oxandrolone in cultured human lymphocytes

Treatments	No. of abnormal cells (%±SE)	Total CAs					
		 Gaps	СТВ	CSB	CTE	DIC	
Oxandrolone (µM)	cens (/a=SE/)	Оцра	CIB	CDD	CIL	DIC	
5	5 (1.67±0.74)	3	4	1	_	_	
10	7 (2.33±0.87)	3	4	3	_	_	
20	10 (3.33±1.04)	4	6	4	_	_	
30	15 (5.00±1.26) ^b	7	10	5	_	_	
40	19 (6.33±1.41) ^b	9	15	4	_	_	
Genistein (μM)	19 (0.33±1.41)	,	13	7	_	_	
30	3 (1.00±0.57)	1	2	1			
40	4 (1.33±0.66)	2	3	1	-	-	
Genistein + Oxandrolone	4 (1.33±0.00)	2	3	1	-	-	
30 + 30	10 (3.33±1.04)°	2	6	4			
30 ± 30 30 ± 40	14 (4.67±1.22)°	3	9	5	-	-	
40 + 30		4	9	_	-	-	
	7 (2.33±0.87)°	2	3	2	-	-	
40 + 40	11 (3.67±1.09)°	3	8	3	-	-	
Gingerol (μM)			_				
20	2 (0.67±0.47)	1	1	1	-	-	
30	3 (1.00±0.57)	1	2	1	-	-	
Gingerol + Oxandrolone							
20 + 30	8 (2.87±0.93)°	3	6	2	-	-	
20 + 40	12 (4.00±1.13)°	5	10	2	-	-	
30 + 30	6 (2.00±0.81)°	2	4	2	-	-	
30 + 40	9 (3.00±0.98)°	2	6	3	-	-	
Untreated	3 (1.00±0.57)	1	2	1	-	-	
Negative control (DMSO, 5 μl mL ⁻¹)	3 (1.00±0.57)	1	2	1	-	-	
Positive control (Mitomycin C, 0.3 µg mL ⁻¹)	48 (16.00±2.12) ^a	22	38	10	5	1	
201 101 1100 24 4 4 1 / 4	0.04) hat tall 1100 til		/ -0.05\ 00'	: a : . : . : . :			

*Significant difference with respect to untreated (p<0.01), 'Significant difference with respect to untreated (p<0.05), 'Significant with respect to Oxandrolone (p<0.05), SE: Standard Error, CTB: Chromatid Break, CSB: Chromosome Break, CTE: Chromatid Exchange, DIC: Dicentric Chromosome

Table 3: Ameliorative action of genistein and gingerol on SCEs induced by Norethandrolone in cultured human lymphocytes

	SCEs/Cell		
Treatments	(Mean±SE)		
Norethandrolone (µM)			
5	0.79 ± 0.35		
10	1.07 ± 0.58		
20	1.33 ± 0.64		
30	4.88 ± 1.30^{b}		
40	5.96±1.53 ^b		
Genistein (µM)			
30	1.35 ± 0.67		
40	1.39 ± 0.68		
Genistein + Norethandrolone			
30 + 30	3.22±1.12°		
30 + 40	4.12±1.27°		
40 + 30	$3.01\pm1.03^{\circ}$		
40 + 40	3.52±1.23°		
Gingerol (μM)			
20	1.10 ± 0.57		
30	1.28 ± 0.62		
Gingerol + Norethandrolone			
20 + 30	$3.02\pm1.14^{\circ}$		
20 + 40	$3.78\pm1.26^{\circ}$		
30 + 30	2.92±1.08°		
30 + 40	3.22±1.21°		
Untreated	1.00±0.56		
Negative control (DMSO, 5 μl mL ⁻¹)	1.33±0.66		
Positive control (Mitomycin C, 0.3 µg mL ⁻¹)	13.22 ± 2.02^a		

 $^{\circ}$ Significant difference with respect to untreated (p<0.01), $^{\circ}$ Significant difference with respect to untreated (p<0.05), $^{\circ}$ Significant with respect to Norethandrolone (p<0.05), SE: Standard Error

Table 4: Ameliorative action of genistein and gingerol on SCEs induced by Oxandrolone in cultured human lymphocytes

	SCEs/Cell
Treatments	(Mean±SE)
Oxandrolone (µM)	
5	0.75 ± 0.32
10	1.08 ± 0.56
20	1.26 ± 0.60
30	4.36 ± 1.18^{b}
40	5.22±1.24 ^b
Genistein (µM)	
30	1.37 ± 0.68
40	1.42 ± 0.69
Genistein + Oxandrolone	
30 + 30	3.32±1.10°
30 + 40	4.02±1.16°
40 + 30	$3.04\pm1.06^{\circ}$
40 + 40	3.64±1.12°
Gingerol (µM)	
20	1.12±0.56
30	1.32 ± 0.58
Gingerol + Oxandrolone	
20 + 30	3.02±1.06°
20 + 40	3.72±1.25°
30 + 30	2.78±0.98°
30 + 40	3.34±1.02°
Untreated	1.00 ± 0.56
Negative control (DMSO, 5 μl mL ⁻¹)	1.33 ± 0.66
Positive control (Mitomycin C, 0.3 μg mL ⁻¹)	14.26±1.96°

*Significant difference with respect to untreated (p<0.01), 'Significant difference with respect to untreated (p<0.05), 'Significant with respect to Oxandrolone (p<0.05), SE: Standard Error

genistein and 20 and 30 μM of gingerol (Table 1). The selected dosage of genistein and gingerol was not

genotoxic itself (Table 1, 2). Similar dose-dependent decrease in number of abnormal metaphases was observed when the genotoxic doses i.e., 30 and 40 μ M of oxandrolone was treated with 30 and 40 μ M of genistein and 20 and 30 μ M of gingerol, respectively (Table 2).

For sister chromatid exchange analysis, a significant increase in SCEs/cell was observed at 30 and 40 μ M of norethandrolone and oxandrolone, respectively (Table 3, 4). A significant decrease in SCEs/cell was observed when 30 and 40 μ M of norethandrolone was treated with 30 and 40 μ M of genistein and 20 and 30 μ M of gingerol (Table 3). Similar trend was observed when 30 and 40 μ M of oxandrolone was treated with 30 and 40 μ M of genistein and 20 and 30 μ M of gingerol, respectively (Table 4). The selected dosage of gingerol and genistein did not induce SCEs/cell significantly as compared to the untreated (Table 3, 4).

DISCUSSION

Genistein is particularly effective in quenching free radicals produced by toxic agents and protects cells against oxidative damage especially with respect to DNA (Foti et al., 2005; Lee et al., 2000). It is a common precursor in the biosynthesis of antimicrobial phytoalexins and phytoanticipins in legumes and an important nutraceutical molecule. It is also capable of inhibiting lipoprotein oxidation in vitro and suppressing formation of plasma lipid oxidation products in vivo. Extracts of Ginger (including Gingerol) have antioxidant activity through scavenging of superoxide and hydroxyl radicals and by inhibiting lipid peroxidation (Kikuzaki and Nakatam, 1993). It is also antibacterial, antifungal and used for common cold (Bode et al., 2001). The pungency of Ginger is due to Gingerol which is an alcohol of oleoresin and the aroma is due to its oil (Hasenöhrl et al., 1998). Gingerol is the major pharmacologically active component inducing apoptosis (Lee and Surh, 1998; Lee et al., 1998) in cancer cells. The results of the present study reveal that the selected dosages of genistein and gingerol are not genotoxic per se but reduced the genotoxic damage caused by oxandrolone norethandrolone in human lymphocytes in vitro. The International Agency on Cancer (IAC), mainly on the basis of epidemiological studies classifies steroidal estrogens and estrogen progestin combinations among agents carcinogenic to humans (Group 1), progestins as possibly carcinogenic (Group 2) and androgenic anabolic steroids, as probably carcinogenic (Group (Martelli et al., 2003).

An increase in the frequency of chromosomal aberrations in peripheral blood lymphocytes is associated with an increased overall risk of cancer (Hagmar et al., 1994, 1998). The readily quantifiable nature of sister chromatid exchanges with high sensitivity for revealing toxicant-DNA interaction and the demonstrated ability of genotoxic chemicals to induce significant increase in sister chromatid exchanges in cultured cells has resulted in this endpoint being used as indicator of DNA damage in blood lymphocytes of individuals exposed to genotoxic carcinogens (Albertini et al., 2000). The above genotoxic endpoints are well known markers of genotoxicity and any reduction in the frequency of these genotoxic endpoints gives us indication of the antigen toxicity of a particular compound (Albertini et al., 2000). Many plant products protect against xenobiotics either by inducing detoxifying enzymes or by inhibiting oxidative enzymes (Morse and Stoner, 1993).

The protective effect observed in the present study i.e., significant reduction in the frequency of cells with chromosomal damage and sister chromatid exchanges may be due to the direct action of the compounds (i.e., Genistein and Gingerol).

The outcome of this experiment shows that Norethandrolone and Oxandrolone have the potential to be genotoxic and cytotoxic, especially at 30 and 40 µM, in cultured human lymphocytes and their genotoxicity is reduced significantly on applying Genistein and Gingerol separately, at appropriate dosage. The evaluation of these genotoxicity tests is a useful tool for determining the toxic effects of potentially genotoxic chemicals, leading to identification of such carcinogenic agents. It is advisable to use the steroids studied here at their lowest effective dosage so that the risk to public health could be minimized. The risk of damage to human genetic material is very likely at higher doses of these drugs. The effectiveness of Genistein and Gingerol as antimutagenic agents is an attribute that can be effectively used in making anticancer drugs.

ACKNOWLEDGMENT

We are thankful to the Chairman, Department of Zoology, AMU, for the indispensable laboratory facilities endowed for this experiment.

REFERENCES

Ahmad, M.S., Sheeba and M. Afzal, 2004. Amelioration of genotoxic damage by certain phytoproducts in human lymphocyte cultures. Chem. Biol. Interact, 149: 107-115.

- Albertini, R.J., D. Anderson, G.R. Douglas, L. Hagmar, K. Heminiki, F. Merelo, A.T. Natrajan, H. Norppa, D.E.G. Shuker, R. Tice, M.D. Walters and A. Aitio, 2000. IPCS guidelines for the monitoring of genotoxic effects of carcinogens in humans. Mutat. Res., 463: 111-172.
- Beg, T., Y.H. Siddique, G. Ara and M. Afzal, 2007. Antimutagenic evaluation of Genistein, a polyphenol, in cultured human peripheral blood lymphocytes. Biomed. Res., 18: 141-145.
- Bode, A.M., W.Y. Ma, Y.J. Surh and Z. Dong, 2001. Inhibition of epidermal growth factor-induced cell transformation and activator protein 1 activation by [6]-gingerol. Cancer Res., 61: 850-853.
- Carballo, M.A., S. Aluarez and A. Boveris, 1993. Cellular stress by light and rose bengal in human lymphocytes. Mutat. Res., 288: 215-222.
- Dewhurst, J. and R.R. Gordon, 1984. Fertility following change of Sex: A follow-up. Lancet, 22: 1461-1462.
- Dhillon, V.S., J.R. Singh, H. Singh and R.S. Kler, 1994.
 In vitro and in vivo genotoxicity evaluation of hormonal drugs V, Mestranol. Mutat. Res., 322: 173-183.
- Dixon, R.A. and D. Ferreira, 2002. Genistein. Phytochemistry, 60: 205-211.
- Ferretti, G., T. Bacchetti, F. Menanno and G. Curatola, 2004. Effect of genistein against copper-induced lipid peroxidation of human high density lipoproteins (HDL). Atherosclerosis, 172: 55-61.
- Fisher, R.A. and Y. Yates, 1963. Statistical Table for Biological, Agricultural and Medical Research. 6th Edn., Oliver and Boyd, Edinburgh.
- Foti, P., D. Erba, P. Riso, A. Spadafrance, Criscuoli and G. Testolin, 2005. Comparison between daidzein and genistein antioxidant activity in primary and cancer lymphocytes. Arch. Biochem. Biophys., 433: 421-427.
- Gannon, T., 1994. Dermatologic emergencies. When early recognition can be lifesaving. Postgrad. Med., 96: 28-30.
- Hagmar, L., A. Brogger, I.L. Hansteen, S. Heims,
 B. Hoggstedt, L. Knudsen, B. Lambert, K. Linnaimaa,
 F. Mitelman, I. Nordenson, C. Reuterwall, S. Salomaa,
 S. Skerfving and M. Sorsa, 1994. Cancer risk in humans predicted by increased level of chromosomal aberrations in human lymphocytes: Nordic study group on the health risk of chromosome damage.
 Cancer Res., 54: 2919-2922.
- Hagmar, L., S. Bonassi, U. Stromberg, A. Brogger, J.E. Knudson, H. Norppa and C. Reuterwall, 1998. Chromosomal aberration in human lymphocytes predict human cancer: A report from the European Study Group on Cytogenetic Biomarkers and Health (ESCH). Cancer Res., 58: 4117-4121.

- Hasenöhrl, R.U., B. Topic, C. Frisch, R. Häcker, C.M. Mattern and J.P. Huston, 1998. Dissociation between anxiolytic and hypomnestic effects for combined extracts of *Zingiber officinale* and Ginkgo biloba, as opposed to diazepam. Pharmacol. Biochem. Behav., 59: 527-535.
- Haynes, R.C. Jr. and F. Murad, 1985. Adrenocorticotropic Hormone, Adrenocortical Steroids and their Synthetic Analogs, Inhibitors of Adrenocortical Steroid Biosynthesis. In: The Pharmacological Basis of Therapeutics, Gilman, A.G., L.S. Goodman, T.W. Rall and F. Murad (Eds.) Macmillan Publishing Co., New York, pp. 1459-1489.
- Huang, Q., H. Matsuda, K. Sakai, J. Yamahara and Y. Tamai, 1990. The effect of ginger on serotonin induced hypothermia and diarrhea. Yakugaku Zasshi, 110: 936-942.
- Hundal, B.S., V.S. Dhillon and I.S. Sidhu, 1997. Genotoxic potential of estrogens. Mutat. Res., 389: 173-181.
- International Programme on Chemical Safety, 1985.
 Environmental Health Criteria 46. Guidelines for the study of genetic effects in human population. WHO, Geneva, 25-54.
- International Programme on Chemical Safety, 1993.
 Poisons Information Monograph 918,
 Pharmaceutical. WHO, Geneva. PIM G007.
- Joosten, H.F.P., F.A.A. van Acker, D.J. van den Dobbelsteen, G.J.M.J. Horbach and E.I. Krante, 2004. Genotoxicity of hormonal steroids. Toxicol. Lett., 151: 113-134.
- Kikuzaki, H. and N. Nakatami, 1993. Antioxidant effect of some ginger constituents. J. Food Sci., 58: 1407-1410.
- Lee, E. and Y.J. Surh, 1998. Induction of apoptosis in HL-60 cells by pungent vanilloids, [6]-gingerol and [6]-paradol. Cancer Lett., 134: 163-168.
- Lee, E., K.K. Park, J.M. Lee, K.S. Chun, J.Y. Kang, S.S. Lee and Y.J. Surh, 1998. Suppression of mouse skin tumor promotion and induction of apoptosis in HL-60 cells by *Alpinia oxyphylla* Miquel (Zingiberaceae). Carcinogenesis (Lond.), 19: 1377-1381.
- Lee, R., Y.J. Kim, Y.J. Lee and H.W. Chung, 2000. The selective effect of genistein on the toxicity of bleomycin in normal lymphocytes and HL-60 cells. Toxicology, 195: 87-95.
- Martelli, A., F. Mattioli, M. Angiola, R. Reimann and G. Brambilla, 2003. Species, sex and interindividual differences in DNA repair induced by nine sex steroids in primary cultures of rats and human hepatocytes. Mutat. Res., 536: 69-78.
- Mooradian, A.D., J.E. Morley and S.G. Korenman, 1987. Biological actions of androgens. Endo. Rev., 8: 1-28.

- Morse, M.A. and G.D. Stoner, 1993. Cancer chemoprevention: Principles and prospects. Carcinogenesis, 14: 1737-1746.
- Muster, W., S. Albertini, A. Chatelat, B. Miller and E. Gocke, 1997. Mutagenicity evaluation of Genistein, an anticarcinogenic isoflavone present in soybeans. Mutat. Res., 37: S170-S170.
- Overly, W.L., J.A. Dankoff, B.K. Wang and U.D. Singh, 1984. Androgens and hepatocellular carcinoma in an athlete. Ann. Internal Med., 100: 158-159.
- Patel, R.P., B.J. Boersma, J.H. Crawford, N. Hogg, M. Kirk, B. Kalyanaraman, D.A. Parks, S. Barnes and V. Darley-Usmar, 2001. Antioxidant mechanism of isoflavones in lipid systems: Paradoxical effects of peroxyl radical scavenging. Free Radic. Biol. Med., 31: 1570-1581.
- Perry, P. and S. Wolff, 1974. New Giemsa Method for Differential Staining of Sister Chromatids. Nature, 251: 156-158.
- Roberts, J.T. and D.M. Essenhigh, 1986. Adenocarcinoma of Prostate in a 40-year-old Body-builder. Lancet, 2: 742-742.
- Siddique, Y.H. and M. Afzal, 2004a. Evaluation of genotoxic potential of synthetic progestin chlormadinone acetate. Toxicol. Lett., 153: 221-225.
- Siddique, Y.H. and M. Afzal, 2004b. Evaluation of genotoxic potential of ethynodiol diacetate in human lymphocytes *in vitro*. Curr. Sci., 86: 1161-1165.
- Siddique, Y.H. and M. Afzal, 2004c. Induction of chromosomal aberrations and sister chromatid exchanges by cyproterone acetate in human lymphocytes. Int. J. Hum. Genet., 4: 187-191.
- Siddique, Y.H. and M. Afzal, 2005a. Genotoxic potential of cyproterone acetate: A possible role of reactive oxygen species. Toxicol. *In vitro*, 19: 63-68.
- Siddique, Y.H. and M. Afzal, 2005b. Protective role of allicin and L-ascorbic acid against the genotoxic damage induced by chlormadinone acetate in cultured human lymphocytes. Indian J. Exp. Biol., 43: 769-772.
- Siddique, Y.H., T. Beg and M. Afzal, 2005a. Genotoxic potential of ethinylestradiol in cultured mammalian cells. Chem. Biol. Interact., 151: 141-144.
- Siddique, Y.H., T. Beg and M. Afzal, 2005b. Antigenotoxic effects of ascorbic acid against megestrol acetate induced genotoxicity in mice. Hum. Exp. Toxicol., 24: 121-127.
- Siddique, Y.H., G. Ara, T. Beg and M. Afzal, 2006a. Genotoxic potential of medroxyprogesterone acetate in cultured human peripheral blood lymphocytes. Life Sci., 80: 212-218.

- Siddique, Y.H., T. Beg and M. Afzal, 2006b. Protective role of nordihydroguaiaretic acid (NDGA) against norgestrel induced genotoxic damage. Toxicol. *In vitro*, 20: 227-233.
- Siddique, Y.H., G. Ara, T. Beg and M. Afzal, 2007a. Protective role of nordihydroguaiaretic acid (NDGA) against the genotoxic damage induced by ethynodiol diacetate in human lymphocytes *in vitro*. J. Environ. Biol., 28: 279-282.
- Siddique, Y.H., G. Ara, T. Beg and M. Afzal, 2007b. Additive action of vitamin C and E against norgestrel induced genotoxicity. Biomed. Res., 18: 155-160.
- Siddique, Y.H., G. Ara, T. Beg and M. Afzal, 2007c. Antigenotoxic effect of *Ocimum sanctum* L. extract against cyproterone acetate induced genotoxic damage in cultured mammalian cells. Acta Biologica Hungarica, 58: 397-409.

- Siddique, Y.H., G. Ara, T. Beg and M. Afzal, 2008a. Antigenotoxic effect of nordihydroguaiaretic acid against chlormadinone acetate-induced genotoxicity in mice bone-marrow cells. J. Nat. Med., 62: 52-56.
- Siddique, Y.H., G. Ara, T. Beg, M. Faisal, M. Ahmad and M. Afzal, 2008b. Antigenotoxic role of *Centella asiatica* L. extract against cyproterone acetate induced genotoxic damage in cultured human lymphocytes. Toxicol. *In vitro*, 22: 10-17.