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Studies of Behavioural and Neural Mechanism of Aridanim Isolated from Tetrapleura tetraptera in Mice

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Abstract: The aim of the present study was to investigate the central nervous system depressant effect and neural mechanisms of Aridamin in mice. Novelty-induced rearing, head dips, locomotor activity and effect on learning and memory were studied. Aridanin at 5, 10, 20 and 30 mg kg⁻¹ b.wt. reduced novelty-induced rearing and locomotor activity in mice. Head dip reduction was noticed only with the highest dose (30 mg kg⁻¹, i.p.) while in Y-maze, a reduction in number of entrance (locomotion) with no change in percentage alternation on short term working memory was obtained. Aridanin reversed the central excitatory effect of flumazenil in the methods. These results confirm the central depressant properties of Aridanin which may be mediated through GABA_A receptor.

Key words: Aridanin, Tetrapleura tetraptera, Y-maze, hole board, open field, rearing

INTRODUCTION

Tetrapleura tetraptera Taub (Mimosaceae) locally known as Aridan is a large tree growing throughout the rain forest belt of West Africa. It is generally found in the lowland forest of tropical Africa. The fruit consist of a fleshy pulp with small, brownish-black seeds. The plant has many traditional uses mainly in the management of convulsion, leprosy, inflammation and rheumatic pains, schistosomiasis, asthma and hypertension (Ojewole and Adesina, 1983). The dry fruit has a pleasant aroma (Aladesanmi, 2007). It is used as a popular seasoning spice, a medicine and a dietary supplement rich in vitamins in Southern and Eastern Nigeria (Okwu, 2003; Essien et al., 1994). The fruit is used to prepare soup for mothers from the first day of birth to prevent post partum contraction (Nwawu and Akali, 1986). The root extract has been proven to be useful for the treatment of gastrointestinal related clinical problem (Noamesi et al., 1994). The ethanol extract and saponins from the stem bark of Tetrapleura tetraptera exerted an inhibitory effect on luteinizing hormone released by pituitary cells, suggesting its use as contraceptive agent (El-Izzi et al.,

1990). Tetrapleura tetraptera is a natural molluscicides as aqueous extract of it is effective against Bulinus globosus and Lymnaea natalensis (Adewunmi, 1991). The alleopathic potential of Tetrapleura tetraptera has led to its integration into an agro forestry system (Amoo et al., 2008). Tetrapleura tetraptera has been shown to improve the foaming ability of soaps (Adebayo et al., 2000). Tetrapleura tetraptera has no influence on cell proliferation and neither induced chromosomal aberration nor sister chromatid exchanges in Chinese hamster ovary cells (no genotoxic effect) (Adewunmi et al., 1991). Tetrapleura tetraptera has been shown to cause elevation in serum AST and alteration of various metabolites parameters and did not induce any marked pathological lesion in the liver (Lawal et al., 2009). The and analgesic effect of sedative, anticonvulsant Aridanin in mice have been reported by Aderibigbe et al. (2007 a, b) and Ojewole (2005). The aqueous extract of Tetrapleura tetraptera fruit have been shown to possessed anti-inflammatory hypoglycaemic and properties (Ojewole and Adewunmi, 2004). The ethanolic extract of Tetrapleura tetraptera fruit possessed antiplasmodial activity in mice (Okonkon et al., 2007). One

of the active constituents isolated from *Tetrapleura* tetraptera fruit is a mono-N-acetylglycoside of oleanoic acid $(3\beta$ -hydroxyolean-12-en-28-oic) called Aridanin (Adesina and Reish, 1985). The present study was carried out in order to investigate further the neurobiology properties of Aridanin.

MATERIALS AND METHODS

Plant material: Structural elucidation and characterization of Aridanin (Fig. 1) from *Tetrapleura tetraptera* was carried out by Prof. S. K. Adesina (Adesina and Reish, 1985) of Drug Research and Product Unit, Faculty of Pharmacy, Obafemi Awolowo University, Ile-Ife. Sample used for this experiment was collected from him.

Animals: Swiss albino male mice weighing between (20-25 g) were obtained in March 2009 from the animal house of the Faculty of Pharmacy, Obafemi Awolowo University, Ile-Ife. The animals were divided into five mice in each cage and were fed with a standard laboratory diet and tap water *ad libitum*. The animals were maintained at 25+1°C under natural 12 h daylight night conditions. All experiment was carried out in compliance with Obafemi Awolowo University Ethics Committee on research in animals and in accordance with NIH guide for the care and use of laboratory animals.

Drugs: Diazepanı (Roche, Basel Switzerland), Atropine, Flumazenil, Yohimbine, Naloxone, Cyproheptadine (Sigma Chemicals Co. St. Louis, Missouri, USA).

Drug dissolution: Aridanin was dissolved in 5% Tween 80. Diazepani, flumazenil, atropine, yohimbine, naloxone and cyproheptadine were dissolved in normal saline. Tween 80 at 5% concentration did not affect behavioural studies in rodents (Castro *et al.*, 1995). The resulting solution, control vehicle and test materials were usually administered intraperitoneally (i.p.).

Fig. 1: The chemical structure of Aridanin

Toxicity: Acute toxicity studies of Aridanin in mice were carried out as described by Miller and Tainter (1944) and the lethal dose was calculated by the method of Lichtfield and Wilcoxon (1949). It was carried out by injecting Aridanin intraperitioneally (i.p.) into 5 groups of male mice containing 5 animals with the following dose levels 25, 37.5, 50, 75 and 100 mg kg⁻¹. The animals were observed for over 24 h and the LD₅₀ was calculated.

Novelty-induced rearing: The behavioural profile of albino mice under the influence of Aridanin was assessed singly in a Plexiglass cage measuring (45×25×25 cm) containing wood shavings. Behavioural measurements were carried out after i.p. administration of 5% Tween 80 (0.2 mL/20 g) group 1 and Aridanin (5, 10, 20 and 30 mg kg⁻¹, i.p.) groups 2-5. Aridanin was administered for 30 min into animals before been placed singly into an opaque plexiglass observation cage with only one side transparent for observation. Each animal was used only once, with the saw dust bedding changed after each assessment to remove olfactory cue from one animal to the other. The time of the experiment was kept constant (9.00 a.m.-1.00 p.m.) daily to avoid changes in biological rhythm. The behavioural component employed in this observational analysis is rearing. Rearing is defined as the number of times the animal stood on its hind legs or with its forearm against the wall of the observation cage or in the free air (Ajayi and Ukponmwan, 1994). The frequency of rearing episodes was counted manually for 5 min.

Locomotor activity: Motor activity was measured in an open field apparatus consisting of a white plexiglas box $(28 \times 28 \times 25 \text{ cm})$ with a painted back grid dividing the floor into $16 (7 \times 7 \text{ cm})$ equal squares. The animals were divided into six groups. Group 1 was given 5% Tween 80 (0.2 mL/20 g i.p.), while groups 2-5 was given Aridanin $(5, 10, 20 \text{ and } 30 \text{ mg kg}^{-1} \text{ i.p.})$. Aridanin was administered for 30 min into animals and were placed singly in one of the corners of the box; the number of squares crossed with all four paws was counted for 5 min. The cages were cleared with 70% ethanol at intervals when the animal is removed (Mandal *et al.*, 2001). Diazepam $(2.0 \text{ mg kg}^{-1}, \text{i.p.})$ group 6 served as reference drug.

Head dips: The effect of Aridanin on the rate of head dipping was determined in the holeboard with a number of holes (usually 16) in the floor through which the animal can poke its head. The animals were divided into six groups. Group 1 was given 5% Tween 80

(0.2 mL/20 g i.p.), while groups 2-5 was given Aridamin (5, 10, 20 and 30 mg/kg i.p.). Aridamin was administered for 30 min into animals and placed on top of a wooden box with 16 evenly spaced hole. The number of times that each animal dipped their head into the holes in 5 min was counted (Dorr *et al.*, 1971). 70% ethanol was used to clean the cages at intervals. Diazepam (2.0 mg kg⁻¹, i.p.) group 6 served as reference drug.

Effect on learning and memory: The Y-maze test can be used as a measure for short term working memory and locomotor activity. Spontaneous alternation is a measure of spatial working memory. To alternate among spatial location, a mouse must remember its previous location. Spontaneous alternation performance was assessed using a Y-maze composed of three equally spaced arms (120°, 41 cm long × 15 cm high). The floors of each arm consist of wood (5 cm wide). This test was carried out using this apparatus to obtain results for spontaneous alternation performance (memory) and locomotor activity (total arm entries). The animals were divided into six group with (n = 5). Group 1 was given control solution 5% Tween 80 (0.2 mL/20 g, i.p.), while groups 2-5 was given Aridanin at the doses of (5, 10, 20 and 30 mg kg⁻¹, i.p.) for 30 min. Each mouse was placed in one of the arm compartments usually arm A for consistency and was allowed to move freely for 5 min without rein forcers. An arm entry is defined as the body of a mouse except for its tail completely entering into an arm compartment. The sequence of arm entries is manually recorded. An alternation is defined as an entry into all three arms on consecutive devices. The percentage alternation was expressed as the ratio of actual alternations to possible alternations (defined as the total number of arm entries minus two) multiplied by 100. 70% of ethanol was used to clean the Y-maze at interval (Akanmu et al., 2007). Diazepam (2.0 mg kg⁻¹, i.p.) group 6 was used as reference drug.

Mechanism of action: In another set of experiment, mice were pre-treated i.p. for 15 min with neurotransmitter blockers to evaluate the mode of actions of Aridanin on novelty-induced rearing (NIR) behaviour, locomotor activity, head dip and Y-Maze in mice. The following receptor blockers were used; atropine (muscarinic antagonist 0.5 mg kg⁻¹), yohimbine (α-2-adrenergic antagonist, 1.0 mg kg⁻¹), naloxone (μ-opioid antagonist, 2.0 mg kg⁻¹) and cyproheptadine (5-HT antagonist, 0.5 mg kg⁻¹). The doses administered are the doses that have been found not to induce behavioural studies of their own in

experimental animals and as such they only block the receptors involved (Ayoka et al., 2006).

Statistic analysis: All behavioural data was analysed by one way ANOVA and Post hoc tests (Student-Newman-Keuls) were carried out to determine the source of a significant mean effect or interaction. Results are expressed as Mean±SEM., p<0.05 is taken as accepted level of significant difference from control. In all these statistical determination, a computer programme the primer of biostatistics (version 3.01) was used (Glantz, 1992).

RESULTS

Results of toxicity testing: The intraperitoneal LD_{50} of Aridanin in mice was calculated to be 60.0 mg kg⁻¹.

Effect of Aridanin on novelty-induced rearing, locomotor activity and head dip in mice: Aridanin induced a decrease in NIR, locomotor activity and head dips in mice [F(5, 24) = 225.2, p<0.001]; [F(5, 24) = 37.3, p<0.001]; [F(5, 24) = 54.1, p<0.001] (Table 1).

Effect of Aridanin on learning and memory in mice: Aridanin (5-30 mg kg $^{-1}$, i.p.) induced a reduction in total arm entries (locomotion) [F (5, 24) = 68.7, p<0.001] (Table 2). The doses administered gave a percentage alternation that is approximately equal to that of 5% Tween 80 control (Table 2).

Table 1: Effect of Aridanin on novelty-induced rearing, locomotor activity and head dip in mice

Treatments	Dose	NIR/5min	LA/5min	HD/5min
5% TW80	0.2 mL / 20 g	48.4±1.6	130.8 ± 2.0	39.8±1.7
Aridanin	5 mg kg^{-1}	22.4±1.5*	83.4±4.1*	39.4±0.9
Aridanin	$10{ m mgkg^{-1}}$	17.0±0.7*	76.8±9.2*	36.4±1.9
Aridanin	$20 { m mg kg^{-1}}$	10.1±0.7*	58.2±4.0*	35.4 ± 1.0
Aridanin	$30 \mathrm{mg kg^{-1}}$	$6.0\pm0.7*$	42.8±6.1*	26.2±1.5*
Diazepam	$2~\mathrm{mg~kg^{-1}}$	7.0±0.7*	48.0±3.1*	13.0±1.2*

Results are expressed as Mean \pm SEM, (n = 5-7). One way ANOVA revealed that there is significant difference between various treatment groups. NIR: Novelty-Induced Rearing; LA: Locomotor activity; HD: Head dip; 5%TW 80: 5% Tween 80. *indicate significant difference from 5% Tween 80 control. p<0.05

Table 2: Effect of Aridanin on learning and memory in mice

Treatments	Dose	No. of entrance/5 min	% Alternation
5% TW	800.mL/20 g	23.0±0.7	60.0±2.7
Aridanin	5 mg kg ⁻¹	11.0±0.7*	60.0±2.7
Aridanin	$10 \ { m mg \ kg^{-1}}$	10.4±0.5*	65.3±4.7
Aridanin	$20 { m mg \ kg^{-1}}$	10.6±0.5*	60.0 ± 2.7
Aridanin	$30 { m mg kg^{-1}}$	8.0±0.7*	66.2±5.1
Diazepam	$2~\mathrm{mg~kg^{-1}}$	12.4±0.7*	41.0±1.5*

Results are expressed as Mean \pm SEM, (n = 5-7). One way ANOVA revealed that there is significant difference between various treatment groups. 5%TW 80:5% Tween 80 *indicate significant difference from 5% Tween 80 control. p<0.05

Table 3: Effect of Aridanin on novelty-induced rearing, locomotor activity and head dip in the presence of antagonists

Treatments	Dose	NIR/5 min	LA/5 min	HD/5 min
5% TW80	0.2 mL/20 g	48.4±1.6	130.8±2.0	39.8±1.7
Aridanin	$10.0 \ { m mg \ kg^{-1}}$	$17.0\pm0.7*$	76.8±9.2*	36.4 ± 1.9
ATR	$0.5 \; { m mg \ kg^{-1}}$	47.0 ± 3.2	127.6 ± 2.3	36.1 ± 1.7
ATR+ARI	$10.0~{ m mg~kg^{-1}}$	14.8±2.0*	55.2±5.1*	32.0 ± 0.9
YOH	$1.0~{ m mg~kg^{-1}}$	40.1 ± 1.3	120.0 ± 7.1	32.5 ± 2.2
YOH+ARI	$10.0~{ m mg~kg^{-1}}$	6.8±0.6*	50.2±7.0*	32.2 ± 0.9
FLU	$2.0 \ { m mg \ kg^{-1}}$	50.6 ± 2.7	160.0 ± 1.3	52.2±3.5*
FLU+ARI	$10.0~{ m mg~kg^{-1}}$	5.0±0.3*	33.4±1.6*	20.6±1.1*
NAL	$2.0 \ { m mg \ kg^{-1}}$	41.7 ± 1.4	124.0 ± 2.7	31.4 ± 2.4
NAL+ARI	$10.0~{ m mg~kg^{-1}}$	$2.6\pm0.4*$	13.8±7.3*	15.6±1.7*
CYP	$0.5 \; { m mg \ kg^{-1}}$	40.6 ± 1.4	127.1 ± 4.2	33.1 ± 2.1
CYP+ARI	$10.0~{ m mg~kg^{-1}}$	7.0±0.7*	32.4±1.4*	28.6±1.3*

Result are expressed as mean±SEM, (n = 5-7). One way ANOVA revealed that there is significant difference between control and treatment groups. NIR: Novelty-Induced Rearing; LA: Locomotor activity; HD: Head dip; 5%TW 80: 5% Tween 80; ATR: Atropine; YOH: Yohimbine; FLU: Flumazenil; NAL: Naloxone; CYP: Cyproheptadine: ARI: Aridanin.*indicate significant difference from 5% Tween 80 control. n<0.05

Table 4: Effect of Aridanin on learning and memory in the presence of

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Treatments	Dose	No. of entrance/5 min	% Alternation
5% TW80	0.2 mL/20 g	23.0 ± 0.7	60.0 ± 2.7
Aridanin	$10.0~{ m mg~kg^{-1}}$	10.4±0.5*	65.3±4.7
ATR	$0.5 \; { m mg \ kg^{-1}}$	24.2 ± 0.8	63.1±3.1
ATR+ARI	$10.0~{ m mg~kg^{-1}}$	8.6±1.1*	63.7±3.3
YOH	$1.0~{ m mg~kg^{-1}}$	24.2 ± 1.3	62.0±2.9
YOH+ARI	$10.0~{ m mg~kg^{-1}}$	6.2±0.6*	63.4±3.2
FLU	$2.0 { m mg \ kg^{-1}}$	45.2±2.1*	63.4±3.2
FLU+ARI	$10.0~{ m mg~kg^{-1}}$	5.2±0.4*	64.4±4.5
NAL	$2.0 { m mg \ kg^{-1}}$	22.7 ± 0.9	61.0±2.7
NAL+ARI	$10.0~{ m mg~kg^{-1}}$	6.2±0.6*	65.8±4.7
CYP	$0.5 \; { m mg \ kg^{-1}}$	21.9 ± 0.7	61.4±2.7
CYP+ARI	$10.0~{ m mg~kg^{-1}}$	5.8±0.4*	64.6±4.7

Result are expressed as mean \pm SEM, (n = 57). One way ANOVA revealed that there is significant difference between control and treatment groups. 5%TW 80: 5% Tween 80; ATR: Atropine; YOH: Yohimbine; FLU: Flumazenil; NAL: Naloxone; CYP: Cyproheptadine: ARI: Aridanin *indicate significant difference from 5% Tween 80 control. p<0.05

Effect of Aridanin on novelty-induced rearing, locomotor activity and head dip in the presence of antagonists:

Pretreatment of mice with atropine yohimbine, naloxone, flumazenil and cyproheptadine did not reverse the decrease in NIR, locomotor activity and head dips of Aridanin in mice. The antagonists potentiated the decrease in NIR and locomotor activity in mice [F (11, 48) = 143.8, p<0.001]; [F (11, 48) = 95.7, p<0.001] (Table 3). However, flumazenil and naloxone potentiated the reduction in head dip induced by 10 mg kg $^{-1}$ of Aridanin in mice [F (11, 48) = 22.6, p<0.001] (Table 3).

Effect of Aridanin on learning and memory in the presence of antagonists: Pretreatment of mice with atropine, yohimbine, naloxone, flumazenil and cyproheptadine potentiated the reduction in total arm entries (locomotion) induced by Aridanin [F (11, 48) = 164.7; p<0.001] (Table 4).

DISCUSSION

Aridanin decreased the NIR behaviour in mice at (30 mg kg⁻¹) but with the other doses the decrease is not dose dependent. On exposure to a new environment mice novelty-induced behaviour consisting of rearing, grooming, scratching and wet dog shakes. In this study, NIR which is a measure of central nervous system excitation (Ajayi and Ukponmwan, 1994; Labella et al., 1979) was used to test the sedative properties of Aridanin. This inhibition of NIR behaviour suggests that Aridanin possesses sedative action. The work is in agreement with some plants that have been shown to possess strong sedative effect such as Cissus quadrangulensis, Spondia mombin, Ficus thoningii, Stachys lavandufolia, Nigella sativa L. and Cryptolepsis sanguinolenta (Viswanatha Swaniy et al., 2006; Musa et al., 2006; Rabbani et al., 2003; Al-Naggar et al., 2003; Ayoka et al., 2006; Ansah et al., 2008).

Aridanin produced a reduction in locomotor activity at (30 mg kg⁻¹) but with the other doses the reduction is not dose dependent. Locomotion is mediated mainly through dopantinergic pathway (Rang *et al.*, 1999) but other neurochemical pathways have been reported to modulate locomotive activities in animals. This reduction in locomotor activity also confirms the central depressant properties of Aridanin. The reduction in locomotor activity is in agreement with plants that have been reported to inhibit locomotor activity such as *Cissus quadrangulensis*, *Spondia mombin* and *Ficus thoningii* (Viswanatha Swanıy *et al.*, 2006; Musa *et al.*, 2006; Ayoka *et al.*, 2006).

Aridanin at a dose of 5, 10 and 20 mg kg⁻¹, i.p. produced an increase in number of head dips comparable to 5% Tween 80 control; this shows that Aridanin may possess anxiolytic properties. However, 30 mg kg⁻¹ i.p. of Aridanin produced a significant reduction in head dip when compared to 5% Tween 80 control, showing that Aridanin may be anxiogenic. Hence, at higher dose of Aridanin a sedative effect was produced. The effect of Aridanin is similar to benzodiazepine which at low doses has anxiolytic action and at high doses has anxiogenic effect and plants such as Cissus cornifolia, Careya anboree which has both anxiolytic and anxiogenic properties (Onaivi et al., 1992; Musa et al., 2008; Kumar et al., 2008). The study is also in conformity with a number of scientific reports which indicated that troterpenoids produced CNS depressant action, since Aridanin is a triterpenoid saponins (Chattopadhyay et al., 2003).

In the Y-maze test, Aridanin at (30 mg kg⁻¹) decrease total arm entries (locomotor activity) in mice, but with the other doses the decrease in total arm entries is not dose dependent. Aridanin at the doses administered do not affect percentage alternation in mice. Memory is a highly complex process that involved several brain structures as well as the role of several neurotransmitters and neuropeptides (Akanmu *et al.*, 2007; Steckler *et al.*, 1998). Aridanin at doses of (5-30 mg kg⁻¹ i.p.) has no effect on working memory as the percentage alternation produced is not different from that of control.

The NIR behaviour response is regulated by multiple neurotransmitter systems; such transmitters include Ganıma-Amino-Butyric-Acid (GABA), opioid and dopanine receptors (Walting, 1998). The administration of atropine, vohimbine, naloxone and cyproheptadine did not reverse the inhibitory effect of Aridanin on NIR, locomotor activity, and Y-Maze, this suggest that muscarinic, adrenergic, opioid and serotonin receptors are not involved in the inhibitory effect of Aridanin on NIR, locomotor activity and Y-Maze. However, yohimbine and naloxone potentiated the inhibitory effect of Aridanin on NIR. However the presence of atropine, yohimbine, naloxone and cyproheptadine potentiated the inhibitory effect of Aridanin on locomotor activity. Flumazenil, a GABA, antagonist increased novelty induced rearing, locomotor activity, head dips and Y-Maze which was blocked by Aridanin. This suggests that Aridanin may facilitate GABA, transmission (Garrett et al., 2003). The administration of atropine and vohimbine produced a reversal in the reduction of head dip produced by Aridanin. Naloxone potentiated the reduction in head dip produced by Aridanin. Cyproheptadine did not reverse the reduction in head dip produced by Aridanin. The results also show that both cholinergic and adrenergic receptor is partially involved in the exploratory head dip of Aridanin.

The results are in line with previous findings where it was shown that the plant has sedative, anticonvulsant and analgesic effect. Since Aridanin facilitate GABAergic transmission, this may the mechanism of the sedative and anticonvulsant effect of Aridanin.

In conclusion, Aridanin significantly reduced NIR, locomotor activity and head dip in mice. The reduction in NIR, locomotor activity and head dip may be related to an interaction with GABA_A neurotransmission. This shows that Aridanin may facilitate GABA-ergic neurotransmission. It is known that the inhibitory action of GABA consists of the GABA receptor opening the chloride ion charmel to allow more chloride ion to enter the cell, thus making the membrane less likely to polarize

(Gottesmann, 2002). It is possible that the sedative effect of Aridanin is mediated by such GABAergic mechanism, since it is known that GABAergic transmission produced profound sedation in mice (Gottesmann, 2002).

REFERENCES

- Adebayo, A.S., I.A. Gbadomosi and C.O. Adewunmi, 2000. Formulation of antimicrobial dried powered herbs in soap basis. Proceedings of the Phyto Medicines in Malaria and Sexually Transmitted Disease, Challenges for the New Millenium, (PMMSTDC'00), Obafemi Awolowo University, Ile-Ife, pp: 97-97.
- Aderibigbe, A.O., E.O. Iwalewa, S.K. Adesina, A.O. Adebanjo and O.E. Ukponmwan, 2007a. Neuropharmacological evaluation of Aridanin a Glycoside isolated from *Tetrapleura tetraptera* fruit. Disc. Innol., 19: 177-181.
- Aderibigbe, A.O., E.O. Iwalewa, S.K. Adesina, A.O. Adebanjo and O.E. Ukponmwan, 2007b. Anticonvulsant, analgesic and hypothermic effects of Aridanin isolated from *Tetrapleura tetrapetra* fruit in mice. J. Boil. Sci., 7: 1520-1524.
- Adesina, S.K. and J.A. Reisch, 1985. Triterpenoid glycoside from *Tetrapleura tetraptera* fruit. Phytochemistry, 24: 3003-3006.
- Adewunmi, C.O., 1991. Plant molluscicides: Potentual of Aridanin, *Tetrapleura tetraptera* for schistosomiasis control in Nigeria. Sci. Total Environ., 103: 21-33.
- Adewunmi, C.O., H.C. Anderson and L. Busk, 1991. Potential molluscicides, aridan (*Tetrapleura tetraptera*), neither induces chromosomal alterations in Chinese hamster ovary cells, nor mutation in *Salmonella typhimurium*. Toxicol. Environ. Chem., 30: 69-74.
- Ajayi, A.A. and O.E. Ukponmwan, 1994. Evidence of angiotensin 11 and endogenous opioid modulation of novelty induced rearing in the rat. Afr. J. Med. Med. Sci., 23: 287-290.
- Akanmu, M.A., S.O. Adeosun and O.R. Ilesanmi, 2007. Neuropharmacological effects of *Oleamide* in male and female mice. Behav. Brain Res., 182: 88-89.
- Al-Naggar, T.B., M.P. Gomez-Serranillos, M.E. Carretero and A.M. Villar, 2003. Neuropharmacological activity of *Nigella sativa* L. extracts. J. Ethnopharmacol., 88: 63-68.
- Aladesanmi, J.A., 2007. Tetrapleura tetraptera: Molluscicidal activity and chemical constituents: A review. Afr. J. Trad. Complementary Alternative Med., 4: 23-36.

- Amoo, S.O., A.U. Ojo and J. van Staden, 2008. Allelpathic potential of *Tetrapleura tetraptera* leaf extracts on early seedling of five agricultural crops. South Afr. J. Bot., 74: 149-152.
- Ansah, C., E. Woode, E.A.A. Mfoafo and M. Duwiejua, 2008. Anxiogenic effect of an aqueous extract of *Cryptolepis sanquinolenta*. Int. J. Pharmacol., 4: 20-26.
- Ayoka, A.O., R.O. Akomolafe, E.O. Iwalewa, M.A. Akanmu and O.E. Ukponwan, 2006. Sedative antiepileptic and antipsychotic effects of *Spondia mombin* L. (Anacardiaceae) in mice and rats. J. Ethnopharmacol., 103: 166-175.
- Castro, C.A., J.B. Hogan, K.A. Benson, C.W. Shehata and M.R. Landauer, 1995. Behavioural effects of vehicles: DMSO, ethanol, Tween-20, Tween-80 and emulphor-620. Pharmacol. Biochem. Behav., 50: 521-526.
- Chattopadhyay, D., G. Arunachalam, S.C. Mandal, R. Bhadra and A.B Mandal, 2003. CNS activity of the methanol extract of malloatus (Geist) muell arg. leaf: An ethnomedicine of onge. J. Ethnopharmacol., 85: 99-105.
- Dorr, M., D. Jayce, R.D. Porsolt, H. Steinberg, A. Summerfield and N. Tomkiewicz, 1971. Persistence of dose related behavior in mice. Nature, 231: 121-123.
- El-Izzi, A., T. Bennie, M. Thieulant and L. Dural, 1990. Inhibitory effect of Saponins from *Tetrapleura tetraptera* on the luteinizing hormone release by pituitary cells. Planta Med., 56: 357-359.
- Essien, E.U., B.C. Izunwane, C.Y. Aremu and O.U. Eka, 1994. Significance for human of the nutrients of the dry fruits of *Tetrapleura tetraptera*. Plant Foods Hum. Nutr., 45: 47-51.
- Garrett, K.M., G. Basnadjian, I.A. Khan, B.T. Shaneberg and T.W. Scale, 2003. Extract of Kava (*Piper methysticum*) induce acute anxiolytic like behaviour in mice. Psychopharmacology, 170: 33-41.
- Glantz, A.S., 1992. Primer of Biostatistics. Version 3.01, McGraw-Hill Inc., USA.
- Gottesmann, C., 2002. GABA mechanism and sleep. Neuroscience, 111: 231-239.
- Kumar, R.S., R.S. Sundram, P. Sivakumar, R. Nethaji, V. Senthil, N.V. Murthy and R. Kanagasabi, 2008. CNS activity of the methanol extracts of *Careya arborea* in experimental animal model. Bangladesh J. Pharmacol., 3: 36-43.
- Labella, F.S., C. Punsky and V. Havlicek, 1979. Morphine derivatives with diminished opiate receptor potency show enhanced control excitatory activity. Brain Res., 174: 263-271.

- Lawal, R.A., S.A. Ojokuku and S.O. Odesanmi, 2009. Effects of ethanolic extract of *Tetrapleura tetraptera* on liver function profile and histopathology in male dutch white rabbits. Int. J. Trop. Med., 4: 136-139.
- Lichtfield, J.T. and F.A. Wilcoxon, 1949. A simplified method of evaluation doses-effects experiments J. Pharmacol. Exp. Ther., 96: 99-113.
- Mandal, S.C., A.K. Dhara and B.C. Manti, 2001. Neuropharmacological activity of *Xanthium strumarium* Linn. extract. J. Herbs Spices Med. Plants, 8: 69-77.
- Miller, L.C. and M.L. Tainter, 1944. Estimation of ED50 and its error by means of Logarithmic probit graph paper. Proc. Soc. Exp. Bio. Med., 57: 261-269.
- Musa, A.M., A.H. Yaro and N.M. Danjuma, 2006. Preliminary phytochemical screening and central nervous system depressant activity of the stem bark of *Ficus thoningii* blums. Biol. Environ. Sci. Trop. J., 3: 1-6.
- Musa, A.M., A.H. Yaro, H. Usman, M.G. Magaji and M. Habu, 2008. Phytochemical and some neuropharmacological studies on the methanolic leaf extracts of *Cissus cornifolia* [Vitaceae] in mice. Int. J. Pharmacol., 4: 145-148.
- Noamesi, B.K., J.F. Mensah, M. Bogale, E. Dagney and J. Adotey, 1994. Antiulcerative properties and acute toxicity profile of some African medicinal plants extracts. J. Ethnopharmacol., 42: 13-18.
- Nwawu, J.I. and P.A. Akali, 1986. Anticonvulsant activity of the volatile oil from the fruit of *Tetrapleura tetraptera*. J. Ethnopharmacol., 18: 103-107.
- Ojewole, J.A.O. and S.K. Adesina, 1983. Mchanism of the hypotensive effect of Scopoletin isolated from the fruit of *rapleura tetraptera*. Planta Med., 49: 46-50.
- Ojewole, J.A.O. and C.O. Adewunmi, 2004. Anti inflammatory and hypoglycaemic effects of *Tetrapleura tetraptera* (Taub.) (Fabaceae) fruit aqueous extract in rats. J. Ethnopharmacol., 95: 177-182.
- Ojewole, J.A.O., 2005. Analgesic and anticonvulsant properties of *Tetrapleura tetrapetra* (Taub) [Fabaceae] fruit aqueous extract in mice. Phytother. Res., 19: 1023-1029.
- Okonkon, J.E., A.E. Udokpoh and B.S. Antia, 2007. Antimalaria activity of ethanolic extract of *Tetrapleura tetraptera* fruit. J. Ethnopharmacol., 111: 537-540.
- Okwu, D.E., 2003. The potentials of *Ocimum gratissimum*, *Pergularia extensa* and *Tetrapleura tetraptera* as spice and flavouring agents. Nig. Agric. J., 35: 143-148.

- Onaivi, E.S., P.A. Maguiri, N.F. Tsai, M.F. Davies and G.H. Locu, 1992. Comparison of behavioural and central BDZ binding profile in three rat lines. Pharmacol. Biochem. Behav., 43: 825-831.
- Rabbani, M., S.E. Sajjadi and H.R. Zarett, 2003. Anxiolytic effect of *Stachys lavandulifolia* Vahl on the elevated plus maze model of anxiety in mice. J. Ethnopharmacol., 89: 271-276.
- Rang, H.P., M.M. Dale and J.M. Ritter, 1999. Pharmacology. 3rd Edn., Churchill Livingstone, USA., pp: 491-665.
- Steckler, T., W.H. Drunkenburg, H. Sahgal and J.P. Aggleton, 1998. Recognition memory in rats-1. Concepts and classification. Prog. Neurobiol., 54: 289-311.
- Viswanatha Swamy, A.H.M., A.H.M. Thippeswamy, D.V. Manjula and C.B. Mehendra Kumar, 2006. Some neuropharmacological effects of the methanolic root extract of *Cissus quadrangularis* in mice. Afr. J. Biomed. Res., 9: 69-75.
- Walting, K.J., 1998. Overview of Central Nervous System Receptors. In: The RBI Handbook of Receptor Classification and Signal Transduction. Walting, K.J. (Ed.). 3rd Edn., RBI, Natick, MA, pp. 2-45.