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An Exhaustive Review on Recent Advancement in Pharmaceutical Bioadhesive Used for Systemic Drug Delivery Through Oral Mucosa for Achieving Maximum Pharmacological Response and Effect

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Abstract: As compared to both injectable and oral delivery, the buccal delivery is preferred to and advantageous for enhancement of bioavailability of drug. The mucosal surfaces are usually rich in blood supply and provide the means for rapid drug transport to the systemic circulation avoiding drug degradation in harsh gastric environment and first-pass hepatic metabolism. Besides, it can prolong absorption and residence time due to prolonged contact with absorption surface and with the site of application allowing once or twice daily dosing. A rapid onset of action with comfort and convenience in delivery of certain drugs has been observed. Many drugs have been tried with buccal route, a few are also available commercially. Clinical need should be significant and must be high enough to counterbalance the high cost associated with development of a buccal product. Buccal drug delivery assures four times the absorption rate from the skin. The review aims the development in the buccal adhesive drug delivery systems to provide basic principles to the young researchers which will be helpful in overcoming the difficulties associated with the formulation design.

Key words: Buccal drug delivery, mucoadhesive polymer, mucoadhesion, bioadhesion, buccal mucosa, buccal absorption

INTRODUCTION

There are various route of drug administration route has its own limitations. (Hoogstraate and Wertz, 1998). Oral route for delivery of drug, although more convenient, causes major problems like hepatic first pass metabolism (Patel et al., 2011a; Abdollahi et al., 2003) degradation of drug in harsh gastrointestinal environment (Kulkarmi and Desai, 2010) and poor bioavailability (Dharam and Shayeda, 2010; Jafar and Ali, 2011). This route shows inadequate and erratic absorption (Sudhakar et al., 2006). Parenteral route for drug administration avoids this problem but it has also some drawback like pain at the site of administration, anaphylaxis and extravasation infection (Scholz et al., 2008; Qureshi et al., 2006), so oral cavity is selected for administration of macromolecules such as oligopeptides (Veuillez et al., 2001), unstable proteins polysaccharides (Gandhi et al., 2011). Administration through oral cavity can be used as an attractive and alternative site for drug delivery which can facilitate particularly in overcoming deficiencies associated with frequent dosing (Wong et al., 1999).

The oral mucosa is comparatively permeable and enriched with blood supply (Shojaei, 1998), it is

highly vascularized (Rossi *et al.*, 2005), vigorous and demonstrate short recovery time after stress or damage (Yamamoto *et al.*, 2001). For drug absorption the total surface area of membrane of oral cavity available is 100-170 cm² (Tayal and Jain, 2011; Miller *et al.*, 2005) of which 50 cm² represent buccal cavity consisting of liner mucosa (Lee *et al.*, 2000). The thickness of buccal epithelium is approximately 500-800 µm (Harris and Robinson, 1992).

Local therapy is used to treat condition such as gingivitis, oral candidiasis (infection by fungus *Candida albicans*), oral lesions (breaking of mucous membrane due to disease), dental carries (decaying of teeth), xerostomia (dryness of mouth due to lack of saliva), oral cancer, mucositis and neuropathic pain (Galati *et al.*, 2000; Smart, 2004; Pourhashemi *et al.*, 2007; Taliyan and Sharma, 2010).

For transmucosal drug delivery buccal and sublingual or floor of mouth (Hoogstraate *et al.*, 1996; Kowsalya *et al.*, 2011), areas are most commonly used route. Sublingual mucosa is more permeable, thinner with high blood flow than buccal mucosa. Due to its smooth and relatively immobile surface, the buccal mucosa offer sustained and controlled delivery of drug (Madhav *et al.*, 2009) and less permeability (Shakya *et al.*, 2011).

Flexible, elastic and soft patches are used for buccal delivery (Patel *et al.*, 2001; Patel and Poddar, 2009) for modified release dosage form (Parmar *et al.*, 2010). Compared to tablet buccal patches are of small size and with adequate thickness to provide better patient compliance (Morales and McConville, 2011).

ADVANTAGES OF BUCCAL DRUG DELIVERY

- As it by passes the GIT and hepatic portal system, it protects the drug from degradation due to pH and various enzyme present in GIT. The bioavailability of orally administrated drug is increased (Vinod et al., 2010)
- Patient compliance is improved due to the elimination of associated pain with injections. Drugs can be administered to mentally ill, disabled and uncooperative, or unconscious or incapacitated patients conveniently (Miller et al., 2005)
- Allows localization of the drug for a prolonged period of time as required for sustained and controlled drug delivery (Grabowski et al., 1992; Trivedi et al., 2011)
- Dose dependent side effect are reduced due to dose reduction
- Unlike other surfaces for transdermal drug delivery, mucosal surfaces exhibit a faster initiation and decline of delivery than other transdermal patches as it does not have stratum corneum, a major barrier layer for drug transport (Gupta et al., 2011; Varshosaz et al., 2006)
- Flexibility in physical state, shape, size and surface
- Low metabolic activity (Gangwar, 2011)
- Very rare incidents of nausea and vomiting have been reported (Mohammadi and Seyedi, 2008)
- Drugs with poor bioavailability and the drugs which are unstable at different pH can be administrated conveniently by this route (Amarji et al., 2007; Giri et al., 2010a)
- Though, less permeable than the sublingual area, drugs can be rapidly absorbed into the venous system below the oral mucosa because it is well vascularized,. Unlike rectal and transdermal route the drug dissolution is relatively large due to the presence of saliva (Patel et al., 2011b)

LIMITATION OF BUCCAL DELIVERY

- Drugs which are unstable at buccal pH (5.5-7) cannot be administered (Vyas and Khar, 2008)
- Restriction in eating and drinking may be required (Lalla and Gurnancy, 2002)

- Due to over hydration, the structural integrity of the formulation may get disrupted leading to slippery surface and hydration of bioadhesive polymer (Jain, 2002; Shanker et al., 2009)
- The area of absorptive membrane is relatively lesser. If the effective area for absorption is dictated by the dimensions of a delivery system, this area then becomes even smaller (Alur *et al.*, 2002)
- Drugs with large dose are difficult to be administered (Khairnar and Sayyad, 2010)
- Only those drugs which are absorbed by passive diffusion can be administered by this route
- Frequent dosing may be required for drugs intended for local action but may face rapid elimination due to the flushing action of saliva or the ingestion of foods stuffs (Yousefzadeh et al., 2006; Wani et al., 2007)
- Low permeability of the buccal membrane, specifically when compared to the sublingual membrane (Peppas and Buri, 1985)

OVERVIEW OF ORAL MUCOSA

Anatomy of oral mucosa: The oral cavity comprises the lip, cheek, tongue, hard palate, soft palate and floor of mouth as shown in Fig. 1. The oral mucosa divided in three distinctive layer which are outer epithelium, middle basement and inner connective tissues as shown in Fig. 2.

There are two parts of oral cavity; the space between teeth and cheeks or lips is outer oral vestibule while the space between teeth and pharynx is interior oral vestibule (Tortora, 2002). The outer layer of buccal mucosa is composed of approximately 40-50 layers of stratified squamous epithelial cell while the sublingual epithelium contains fewer layers (Gandhi and Robinson, 1988). This layer serves as protective covering for the tissue and acts as a barrier to the entry of foreign material (Ghosh and Pfister, 2005), like antigens, carcinogens, microbial toxin and enzyme from food and beverages. The middle layer, a basement membrane, is a continuous layer of extracellular material and forms a boundary between the basal layer of epithelium and connective tissue (Senthil et al., 2007; Shojaei, 1998; Basu et al., 2010). The epithelium in masticatory mucosa are keratinized and lining mucosa are non-keratinized. The lining mucosa contributes approximately 60%, the masticatory mucosa contributes 25% and the specialized mucosa occupies approximately 15% of the total surface area of oral mucosa lining in an adult human. As compared to masticatory mucosa, the lamina propria of

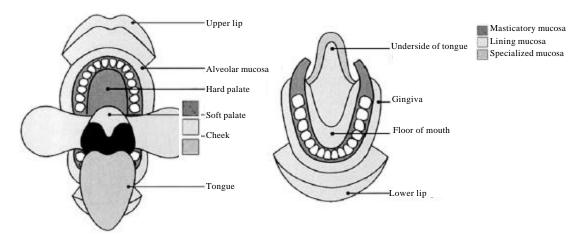


Fig. 1: Schematic representation of the different linings of mucosa in mouth (Patel et al., 2011a)

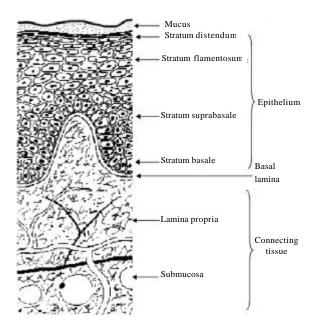


Fig. 2: Cross-section of buccal mucosa (Sudhakar *et al.*, 2006)

lining mucosa is lesser in thickness and elasticity. The specialized mucosa possesses well papilated surface both keratinized and non-keratinized (Collins and Dawes, 1987; Ahmed *et al.*, 2011).

Biochemistry of oral mucosa: The keratinized cell of epidemis of skin and masticatory mucosa of oral cavity is a barrier for drug permeability. In general, keratinized and non-keratinized epithelium of oral cavity occupies about 50% and 30%, respectively (Collins and Dawes, 1987). The composition and state of keratinization of oral mucosa is shown in Table 1.

Secretion of oral cavity

Saliva: The physiological environment of the oral cavity in terms of pH, fluid volume and composition, depends on the secretion of saliva (Herrera et al., 1988; Pajournand et al., 2003). There is a salivary coating over mucosal surface having thickness of about 70 µm. This acts like a stagnant layer. Three major salivary glands-parotid, submandibular and sublingual and minor salivary or buccal glands situated in or immediately beneath the mucosa secrete the saliva (Slomiany et al., 1996; Abdollahi et al., 2003). The paired parotid glands, the major salivary glands, are located opposite to the maxillary first molars and the submandibular and sublingual glands as shown in Fig. 3 are located in the floor of the mouth. Minor salivary or buccal glands are found in the lower lip, tongue, palate, cheeks and pharynx (Humphrey and Russell, 2001). Watery secretion is produced by the parotid and submaxillary glands while the main viscous saliva with limited enzymatic activity is secreted by sublingual glands(Slomiany et al., 1996; Herrera et al., 1988). Except for the gums and the anterior part of the hard palate these glands are placed at every region of the mouth (Llena-Puy, 2006; Giradkar et al., 2010).

There are several compounds in the saliva that control the mouth hemiparasites. The major constituent, about 99% is water with 1% organic and inorganic materials. The composition of the saliva depends mostly on the flow rate along with three other factors: the time of day, the type of stimulus and the degree of stimulation. Its pH varies from 5.5-7 depending on the flow rate. The concentrations of sodium and bicarbonate increase when flow of saliva increases resulting to an increase in the pH. Saliva imparts major effect on many activities like mastication, speech and tissue lubrication

Table 1: The composition and state of keratinisation of oral mucosa

State of keratinization	on Tissue	Composition	Characteristic features	References
Non-keratinized	Buccal mucosa	It do not contain acylceramide with small	These epithelia are more	Squier et al. (1991)
	Sublingual mucosa	amount of ceramide including Small amount	permeable to water	Squier and Wertz (1996)
		of neutral but polar lipid. cholesterol sulphate		
		and glucosyl ceramide are also present		
Keratinized	Palatal mucosa	Neutral lipid like acylceramide	These epithelia are relatively	Wertz and Squier (1991)
	Gingival mucosa	Ceramide	impermeable to water	

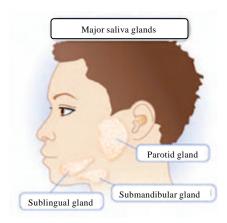


Fig. 3: Major salivary gland (Patricia et al., 2008)

(Tabak et al., 1982; Malekipour et al., 2008; Rohaya et al., 2010). Saliva contains a high molecular weight mucin called MG1 that attaches to the surface of the oral mucosa and maintains hydration, provides lubrication and concentrates protective molecules such as secretory immunoglobulins and also limits the attachment of microorganisms.

Salivary glands consists of acinar and ductal cells. Acinar cells, present in parotid gland, are responsible for production of serous secretion. Most of the α-amylase is synthesized in this gland. The submandibular and sublingual glands produce mainly mucin while proline and histin are produced by parotid and submandibular glands. Mucous are essentially produced by the minor salivary glands (Nauntofte and Lagerlof, 2003; Saad *et al.*, 2005; Chatterjee, 1985).

Clinical diseases and progonostic monitoring on human saliva has a great potential (Jessie *et al.*, 2008). Saliva is complex mixture of fluids containing oral bacteria and food debris. The fluids along with gingival crevicular fluid come from the major and minor salivary glands. The average daily flow of whole saliva varies from 1-1.5 L depending on the health. The contributions of the salivary glands for usual flow are: 20% from parotid, 65% from submandibular, 7-8 % via sublingual and less than 10% via humerous minor gland, shown in (Fig. 4) and when stimulated flow rate can increase by more than 50% of total salivary secretion (Edgar, 1990).

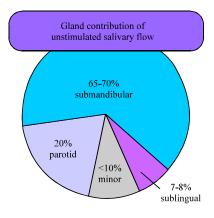


Fig. 4: Percentage contributions of the different salivary glands during unstimulated (Patricia *et al.*, 2008)

Mucus: The intercellular ground substance called mucus surrounds the epithelial cells of buccal mucosa varies in thickness ranging from 40-300 μm (Allen *et al.*, 1984). Mucus, a translucent and viscid secretion, forms a thin, continuous gel cover over the mucosal epithelial surface like a blanket. It is secreted by the goblet cells lining the epithelia or by special exocrine glands with mucus cells acini. Mucus consists of about 95% water and the major organic component in mucus is glycoproteins (Ehrhardt and Kim, 1995; Zakaria *et al.*, 2004).

The thickness of the mucus depends on its location (Marriot and Hughes, 1990). The thickness of the mucus blanket is controlled by the balance between the rate of secretion and the rate of degradation and shedding. Mucus secretion is greatly stimulated by and irritating substances, the thickness of the mucus blanket is increased while the irritants go away from the epithelium efficiently and rapidly moving (Thomas and Moridani, 2009; Puchelle, 1987). The major and minor salivary glands secret mucus as part of saliva (Sangeetha et al., 2010). Up to 70% of the total mucin found in saliva is secreted by the minor salivary glands (Rathbone et al., 1994). The mucus network carries a negative charge at physiological pH due to the presence of sialic acid and sulfate residues which also play a role in mucoadhesion. At this pH mucus a strongly cohesive gel structure can be formed that will combine with the

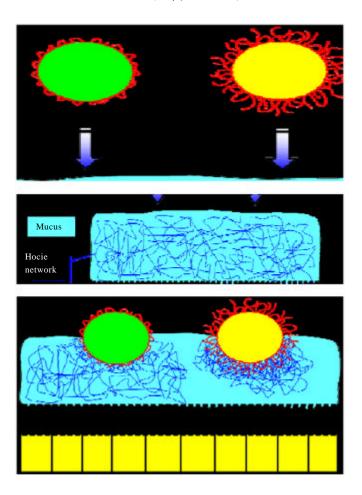


Fig. 5: Mucus interaction with drug delivery system (Madhav et al., 2009)

epithelial cell surface as a gelatinous layer. The interaction of mucus with drug delivery system is shown as Fig. 5 (Gandhi and Robinson, 1988).

ORAL MUCOSA A BARRIER TO PERMEABILITY

The rate and extent of drug absorption through the buccal mucosa can be retarded by the saliva, mucus, membrane coating granules, basement membrane, etc., which also act as barriers (Gandhi and Robinson, 1988). The oral mucosa in general is a somewhat spongy epithelia intermediate between that of the epidermis and intestinal mucosa. It has been observed that the permeability of the buccal mucosa is 4-4000 times greater than that of the skin (Galey et al., 1976). However, this value is not absolute, there are considerable differences in permeability between different regions of the oral cavity due to difference in structures and functions of the different oral mucosa (Harris and Robinson, 1992;

Akhionbare and Ojehanon, 2007). In general, the permeability of the oral mucosa decreases in the order: sublingual

sublingual

palatal (Harris and Robinson, 1992). The permeability coefficient has a direct relation with the membrane thickness (i.e., inverse to its thickness) degree of keratinisation of these tissues and with the physicochemical properties of the drug like molecular weight, size and lipophilicity. The characteristic features of oral mucosa are shown in Table 2 (Squlcr and Johnson, 1975).

Sublingual mucosa is relatively thin and non-keratinized, the buccal mucosa is thicker and non keratinized and the palatal mucosa is intermediate in thickness and is keratinized (Wertz and Squier, 1991). The permeability barrier property of the oral mucosa is mainly due to intercellular materials derived from the so called membrane coating granules (Gandhi and Robinson, 1994). Membrane coating granules are spherical or oval organelles with size range from 100-300 nm and are found in both keratinized and non-keratinized epithelia

Table 2: Characteristics features of oral mucosa

	Tissue				
Characteristics	Buccal	Sublingual	Gingival	Palatal	References
Nature of mucosa	NK	NK	K	K	Harris and Robinson (1992)
Thickness (micrometer)	500-800	100-200	100-200	100-200	Gandhi and Robinson (1994)
Turnover time (in days)	5-6	20	10	24	Harris and Robinson (1992)
Surface area (cm ² ±SD)	50.2±2.9	26.5±4.2		20.1 ± 1.9	Collins and Dawes (1987)
Permeability	Intermediate	Very good	Poor	Poor	De Vries et al. (1991)

NK: Non-keratinized, K: Keratinized

(Hayward, 1979). The membrane coating lipids granules of keratinized epithelia include sphingomyelin, glucosyl ceramide, ceramide and other non-polar lipid. However, for non-keratinized epithelia the major membrane coating granules are lipid components e.g., cholesterol ester, cholesterol and glycosphingolipids (Ganem-Quintanar et al., 1997a). The membrane coating granules present on the basement membrane exhibit some resistance to permeation, however the outer epithelium is considered as rate limiting to mucosal penetration. The relatively large molecules cannot get excluded because of less dense structure of the basement membrane. (Slomiany et al., 1996).

DRUG DELIVERY VIA THE ORAL MUCOSA

The function and detailed description of the oral mucosa is available elsewhere but only those details which are relevant to the oral mucosal delivery of drug have been included here. Lips, cheek (buccal), tongue, hard palate, soft palate and floor of mouth are the parts of oral cavity (Dawes, 2007; Al-Bassiouny, 2009). Buccal and sublingual areas are the most appropriate site for drug delivery and these may be used for the treatment of local (Chiappin et al., 2007) disease like toothaches (Ishida et al., 1982), periodontal disease (Collins et al., 1989; Elkayam et al., 1988), bacterial and fungal infections (Samaranayake and Ferguson, 1994), aphthous (Nagai and Machida, 1985) and dental stomatitis (Nagai, 1985) or systemic diseases. The surface area of oral mucosa, skin and gastrointestinal tract are about 200, 20000 and 350000 cm², respectively. However, the oral mucosa is highly vascularized and therefore any drug diffusing into the oral mucosal membranes has direct access to the systemic circulation via capillaries and venous drainage (Squier and Wertz, 1993; Hakan et al., 1990; El-Kamel et al., 2007).

Drug delivery via sublingual route: The sublingual mucosa is generally well accepted because it is relatively permeable, gives rapid absorption, provide acceptable bioavailability of many drugs and is convenient. The sublingual route is capable of producing a rapid onset of action due to high permeability and rich blood supply which makes it appropriate for drugs with short delivery period (Squier and Wertz, 1996; Giri *et al.*, 2010b).

Among all routes, the sublingual route is studied widely. There are two different design of sublingual dosage form, one composed of rapidly disintegrating tablets and the other consisting of soft gelatin capsules filled with liquid drug, such system before absorption to mucosa create a very high drug conc. in sublingual region (Ishida *et al.*, 1982). Generally, sublingual route is used for drug delivery in treatment of acute disorders, but due to washing of its surface by saliva and tongue activity it makes it difficult to keep the dosage form in contact for an extended period of time with mucosa (Chiappin *et al.*, 2007; Ryan *et al.*, 1997).

Drug delivery via buccal route: The buccal mucosa is less permeable than sublingual area and they generally do not provide rapid absorption as sublingual administration. Buccal mucosa is more fitted for sustained and controlled delivery application, delivery of less permeable molecules and peptide drugs because of immobile mucosa. (Aungst and Rogers, 1989; Singh et al., 2011a). Both application and removal of a drug from buccal mucosal site are very convenient (Verma et al., 2011). The buccal mucosa is a useful route for the treatment of either local or systemic therapies overcoming the drawbacks of conventional administration routes. The buccal mucosa for its permeability and robustness in comparison to other mucosal tissues, is more tolerant to potential allergens and has a reduced tendency to irreversible irritation or damage. So, it has been largely investigated as a potential site for controlled drug delivery in various chronic systemic therapies (Chiappin et al., 2007).

BUCCAL ABSORPTION

Mechanism: In the oral mucosa both the hydrophilic and lipophilic regions are coexisting, for which there are two routes for drug transport, i.e., the paracellular and the transcellular routes. For hydrophilic compounds the paracellular route is the primary route, it is difficult for a hydrophilic compound to penetrate into the lipophilic cell membrane and thus, the intercellular space is the preferred route for drug transport. (Harris and Robinson, 1992; Shojaei and Li, 1997; Alur *et al.*, 2002; Lorenza *et al.*, 2008).

Lipophilic compounds would have low solubilities in the hydrophilic intercellular spaces and cytoplasm

(Peppas and Buri, 1985). The cell membrane, although, is relatively lipophilic in nature hydrophilic solutes will have difficulty permeating through the cell membrane due to a low partition coefficient (Harris and Robinson, 1992). Some of hydrophilic macromolecular therapeutic agents like polysaccharides, oligonucleotides, peptides, can be delivered as controlled delivery via buccal mucosa. To overcome low permeability the absorption enhancer may be required which can improve bioavailability of the high molecular weight drugs (Senel and Hincal, 2001; Lu and Low, 2002).

Dynamic: The first order rate process describes oral mucosal absorption of drugs adequately. Various potential barriers have been identified as obstacles to oral mucosa drug absorption. They mainly include mucus layer, keratinized layer, intercellular lipid of epithelium, basement membrane and lamina propria. The absorptive membrane thickness, blood supply/lymph drainage cell renewal and enzyme content helps in reduction of rate and amount of drug entering the systemic circulation.

The systemic circulation pointed out that salivary secretion alters the buccal absorption kinetics from drug solution by changing the concentration of drug in the mouth. They proposed a linear relationship between salivary secretion and time thus:

dM-KC dt ViVt

where, 'm' and 'C' are the mass and concentration of drug in mouth at time 't', Vi, the volume of solution put into mouth cavity and 'V' is salivary secretion rate (Kumar *et al.*, 2004).

FACTORS AFFECTING BUCCAL ABSORPTION

The oral cavity is a complex environment for drug delivery, as there are many interdependent and independent factors which reduce the absorbable concentration at the site of absorption.

Physicochemical factors affecting buccal absorption:

- Size: Smaller molecules (75-100 Da) generally exhibit rapid transport across the mucosa, with decreasing permeability as molecular size increases (Vaughan, 2003)
- Partition coefficient: When partition coefficient increase, permeability of drug through lipoidal membrane also increase

- pH: pH at the site of drug absorption can influence the partition coefficient. With increasing pH, the partition coefficient of acidic drugs decreases while that of basic drugs increases (Rao et al., 1998)
- Ionization of drug: Both pKa and pH at the mucosal surface influence the ionization of a drug. Lipid solubility is exhibited appreciably only by the nonionized form of many weak acids and weak bases and thus the ability to cross lipoidal membranes (Ajazuddin, 2010)

Physiological factors affecting buccal absorption:

- Inherent permeability of the epithelium
- Thickness of epithelium (Swarbrick, 1999)
- Blood supply
- · Metabolic activity
- Saliva and mucus
- Ability to retain delivery system
- Species differences
- Transport routes and mechanisms (Chatterjee, 1985)

MUCOADHESION/BIOADHESION

Definition

Adhesion: Adhesion can be defined as the bond produced by contact between a pressure sensitive adhesive and a surface (Jimenez *et al.*, 1993; Suryawanshi *et al.*, 2010).

Mucoadhesion: Mucoadhesion can be defined as the phenomenon of the attachment of natural or synthetic macromolecule to mucin layer of mucosal surface or epithelial surface. (Longer and Robinson, 1986; Laura *et al.*, 2009).

Bioadhesion: Bioadhesion is an Interfacial phenomenon in which two materials, at least one of which is biological nature are held together for extended periods of time by means of interfacial forces (Mathias and Hussain, 2010).

Stages of mucoadhesion: Dry or partially hydrated dosage form involve two basic steps in mucoadhesion shown in (Fig. 6). First step involve contact stage, where intimate contact is formed between the mucoadhesive and mucous membrane. The buccal patch is placed within the buccal cavity in contact with required mucosa and to the place to allow adhesion to occur. Second step involve consolidation stage where various physiochemical interaction occur to consolidate and strengthen the adhesive joint which prolongs adhesion (Smart, 2005; Omari *et al.*, 2012; Patel *et al.*, 2007a,b; Huang *et al.*, 2000).

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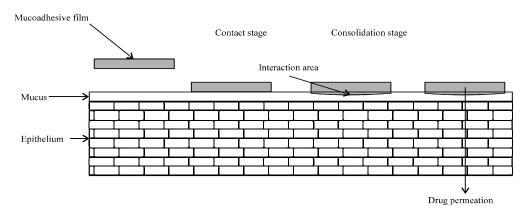


Fig. 6: Contact and consolidation stages of mucoadhesion (Morales and McConville, 2011)

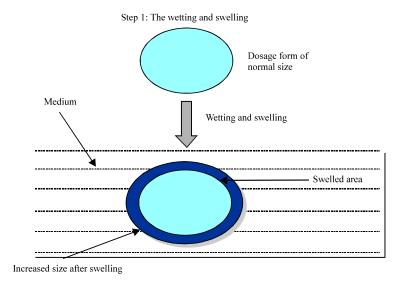


Fig. 7: Wetting and swelling of polymer (Aidoo and Sheila, 2009)

Mechanism of mucoadhesion: The mechanisms responsible in the formation of bioadhesive bonds are not fully known, however most research has described bioadhesive bond formation as a three step process:

- Step 1: Wetting and swelling of polymer
- **Step 2:** Interpenetration between the polymer chains and the mucosal membrane (Varum *et al.*, 2008)
- **Step 3:** Formation of Chemical bonds between the entangled chains (Smart, 2005)
- Step 1: The wetting and swelling step occurs when
 the polymer spreads over the surface of the biological
 substrate or mucosal membrane in order to develop
 an intimate contact with the substrate as shown in
 (Fig. 7) (Bhatt, 2009; Hagerstrom, 2003). Swelling of

- polymers occur because the components within the polymers have an affinity for water (Aidoo and Sheila, 2009)
- Step 2: Glycoproteins the high molecular weight polymers composes the surface of mucosal membrane. A great area of contact shown between chains of mucoadhesive polymer and mucous gel network as shown in (Fig. 8) (Hagerstrom, 2003; Alexander et al., 2011a; Sharma et al., 2009)
- Step 3: This step include entanglement and formation
 of weak chemical bonds along with secondary bonds
 between the polymer chains. Primary bonds such as
 covalent bond and weaker secondary interactions
 such as Van der Waals interactions and hydrogen
 bonds are formed in mucin molecules which is also
 showed in (Fig. 9). Strong adhesion between

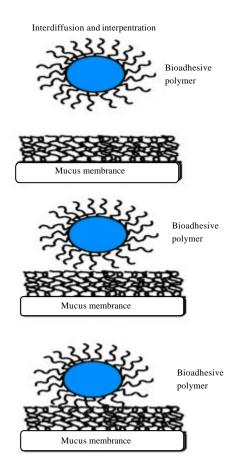


Fig. 8: Interdiffusion and interpenetration of polymer and mucus (Smart, 2005)

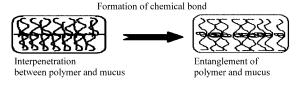


Fig. 9: Entanglement of Polymer and Mucus by Chemical bonds (Hagerstrom, 2003)

polymers is formed during the manufacturing of bioadhesive formulations as primary and secondary bonds are exploited (Hagerstrom, 2003; Alexander *et al.*, 2011b; Fasina *et al.*, 2007)

Theories of mucoadhesion: Mucoadhesion is described as a complex process and various other theories are explained to describe mechanisms are known but other numerous theories should be considered as supplement of mucus/substrate interaction. The diffusion theory of adhesion is shown in (Fig. 10) (Andrews *et al.*, 2009; Alexander *et al.*, 2011a; Sharma *et al.*, 2011):

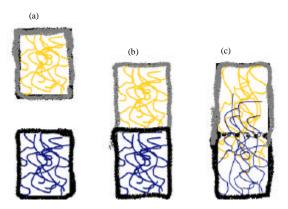


Fig. 10(a-c): The diffusion theory of adhesion, (a) Top (polymer) layer and bottom (mucus) layer before con-tact, (b) Top layer and bottom layer immediately after contact and (c) Top layer and bottom layer after contact (Punitha and Girish, 2010)

- Wettability theory
- The electronic theory
- The fracture theory
- The adsorption theory
- The diffusion-interlocking theory (Lee and Kellaway, 2000)

FACTOR WHICH AFFECT MUCOADHESION IN ORAL CAVITY

The bioadhesive power of a polymer or of a progression of polymers is affected by the nature of the polymer and also by the nature of the surrounding media.

Polymer related factor:

- Molecular weight (Chen and Cyr, 1970)
- Flexibility
- Hydrogen bonding capacity (Peppas and Buri, 1985)
- Cross linking density
- Charge (Lehr *et al.*, 1992)
- Concentration of active polymer (Khairnar et al., 2009)
- Hydration (Swelling) (Gu et al., 1998)

Environmental factor (Khar et al., 1997):

- Applied strength
- pF
- · Initial contact time

Physiological variables

Mucin turnover: These molecules interact with the mucoadhesive before they have a chance to act together with the mucus layer. Mucin turnover may depend on presence of food.

Disease states: The physiochemical properties of mucus are known to adjust during disease conditions such as common cold, gastric ulcers and ulcerative colitis, bacterial and fungal infections of the female reproductive tract (Khar *et al.*, 1997; Rejaie, 2009).

MUCOADHESIVE POLYMER USED IN ORAL CAVITY

Ideal characteristics of mucoadhesive polymer

- Polymer and its degradation products should be nontoxic, non-irritant and free from leachable impurities (Mishra et al., 1996)
- Should have good spreadability, wetting, swelling and solubility and biodegradability properties
- The polymer and its degradation products should be non-hazardous and should be non absorbable from the mucus layer (Longer and Peppas, 1981)
- It should allow daily incorporation to the drug and offer no hindrance to its release (Jimenez et al., 1993)
- Should not aid in development of secondary infections such as dental caries (Wise, 2000)

Classification of mucoadhesive polymer: (Savage, 1977).

In general, adhesive polymers can be classified as synthetic vs. natural, water-soluble vs. water insoluble and charged vs. uncharged polymers (Rao *et al.*, 2011). Examples of the recent polymers classified in these categories are listed in Table 3.

New generation of mucoadhesive polymer: In a recent mini-review current bioadhesive polymers are classified as first generation and second generation (Lee *et al.*, 2000). Newer polymers are capable of forming covalent bonds with the mucus and the underlying cell layers and hence, exhibit improved chemical interactions (Langoth *et al.*, 2003).

The new generation polymer are:

- Thiolated mucoadhesive polymers
- Target-specific, lectin-mediated bioadhesive polymers
- Bacterial adhesion
- Mucoadhesive polymers as enzyme inhibitors and permeation enhancers

PERMEATION ENHANCER

Substances that facilitate the permeation through buccal mucosa are referred as permeation enhancers (Chattarajee and Walker, 1995). As most of the penetration enhancers were originally designed for purposes other than absorption enhancement, The goal of designing penetration enhancers, with improved efficacy and reduced toxicity profile is possible by understanding the relationship between enhancer structure and the effect induced in the membrane and of course, the mechanism of action (Shojaei,1998). The degree of enhancement depended on a number of factors. including the characteristics of the permeant, the composition of the delivery vehicle and whether the tissue was pretreated with enhancer (Ganem-Quintanar et al., 1997b). In some cases usage of enhancers in combination has shown synergistic effect than the individual enhancers.

The efficacy of enhancer in one site is not same in the other site because of differences in cellular morphology, membrane thickness, enzymatic activity, lipid composition and potential protein interactions are structural and functional properties. Penetration enhancement to the buccal membrane is drug specific (Shojaei, 1998).

Mechanism of permeation enhancer: Mechanisms by which penetration enhancers are thought to improve mucosal absorption are as follows:

- By changing mucus rheology in reducing the viscosity and/or elasticity of mucus layer
- By increasing fluidity of lipid bilayer
- By facilitating paracellular transport
- By passing the enzymetic barrier for peptides (Bernkop-Schnurch, 2005)
- By accelerating the thermodynamic activity of peptide drugs
- Alteration of partition coefficient is achieved by increasing the solubility of drug via some enhancer which leads to better absorption with increased thermodynamic activity (Rathbone and Tucker, 1993)
- By acting on the components at tight junctions (Siegel and Gordon, 1985)

Categories of permeation enhancer The categories and examples of various permeation enhancers:

 Bile salts: sodium glycocholate, sodium deoxycholate, sodium taurocholate, sodium glycodeoxycholate, sodium taurodeoxycholate. sodium taurocholate

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Table 3: Comparative study of buccal patch and buccal tablet Bioavai	.	oatch and	buccal table Bioavai	Protein		Amt. of drug	Amt. of drug	Polymer	ease	Polymer	% drug	
Half -lability binding	-lability binding	-lability binding		Oral dos		loadedin	loaded in	usedin	for buccal	nsedin	release for buccal	·
life (h) (%) (%)	(b) (%) (%)	(%) (%)		buccal tabl		buccal patch	Duccal		tablet (h)	buccal patch	patch (h)	References
T, 1.6 50% 10%	50% 10%	50 % 10 %		For adult and 12 yr 2.4 mg day if necess increase 8 mg	r day.		1.6	A M	۵. د:	PVP, Chito-san- PVA	٢	Patel et al. (2009) Arya et al. (2011)
Soluble in dimethyl 7-10 25-35 98 % 25 mg twice a day sulfoxide, methanol, Ethanol methylene chloride, isopropranolol.	25-35 98 %	25-35 98 %		25 mg twice		6.25 mg	15 mg	Methy-β Cyclo-dextrin	ရ	Chito-san and PVP	∞	Hirlekar and Kadam (2009)
Free soluble in 2 90 % actione, chloro-form, ethanol 95%.	:	:		% 06		10 mg	1.5 mg	HPMC-K4M	1.15	Sodium	œ	(1996)
Soluble in water, 2-3 34-40 30 2 mg TDS, max methanol, practically insoluble in acetone, chloroform and ethyl acetate.	34-40 30	30		2 mg TDS, 24 mg per c		45 mg	4.43/ 0.9988 cm2	HPMC-K4M	1.15	Sod CMC	9	Shanker <i>et al.</i> (2009)
Very soluble in water, 3-4 38 - 50 12 25/50/100 mg free soluble in ethanol (95 %), chloroform and in dichloromethane, slightly soluble in acetome, practically insoleble in ether.	3-4 38 - 50 12	77		25/50/100 mg in tablet		50 mg	25 mg	Carbopol and HPMC-K4M	∞	PVP-K30	অ	Rraju et al. (2011)
Free soluble in 2-8 20 90 40/80 mg in tablet chloroform, soluble in, Spaningly soluble in ethanol(95%), practically insoluble in ether	20 90	06		40/80 mg in 1		50 mg	50 mg	Hydroxy ethyl cellu-lose	_∞	PVP-K30	v	(2009)
Soluble in water and 3-4 15 - 23 More than Oral antiarrhyhmic dose in ethanol (9596), 90% is 40-80 mg 2-4 times slightly soluble in chloroform, practically insoluble in ether	3-4 15 - 23		More than Oral antiarthy 90% is 40-80 mg 2 a day	Oral antiarthy is 40-80 mg 2 a day		160 mg	20 mg	HPMC-K4M, SCMC	9	SCMC, CP-934	60	Patel et al. (2007b)

- Chelators: Disodium EDTA, citric acid, sodium salicylate, methoxy salicylates (Kurosaki et al., 1989; Pramod and Lingappa, 2008)
- Surfactants: Sodium lauryl sulfate, polyoxyethylene, Polyoxyethylene-9-laurylether, Polyoxythylene-20cetyl ether, Benzalkonium chloride, 23-lauryl ether, cetylpyridinium chloride, cetyl trimethyl ammonium bromide (Aungst, 1994)
- Non-surfactants: Unsaturated cyclic urea
- Steroidal surfactants: Sodium cholate
- Fatty acids: Oleic acid, capric acid, lauric acid, propylene glycol, cod liver oil methyl oleate, lysophosphatidylcholine, phosphatidylcholine (Hinton et al., 2009)
- Inclusion complexes: Cyclodextrins
- Thiolated polymers: Chitosan-4-thiobutylamide, chitosan- 4-thiobutylamide/GSH, chitosan-cysteine, Poly (acrylic acid)-homocysteine, polycarbophilcysteine, polycarbophil- cysteine/GSH, chitosan-4thioethylamide/GSH, chitosan-4-thioglycolic acid
- Others: Aprotinin, azone, cyclodextrin, dextran sulfate, menthol, polysorbate 80, sulfoxides and various alkyl glycosides (Zhang *et al.*, 1994; Chen *et al.*, 2011)

Buccal formulation: In the past decades, different drug delivery systems intended for buccal administration have been developed, The size of the delivery system varies with the type of formulation, i.e., a buccal tablet may be approximately 5-8 mm in diameter, whereas a flexible buccal patch may be as large as 10-15 cm² in area. Mucoadhesive buccal patches with a surface area of 1-3 cm² are most acceptable. Comparative study of buccal patch and buccal tablet are shown in (Table 4). It has been estimated that the total amount of drug that can be delivered across the buccal mucosa from a 2 cm² system in 1 day is approximately 10-20 mg (Rathbone *et al.*, 1996; Chaudhary *et al.*, 2010).

Matrix tablets/Bioadhesive buccal tablets: Monolithic tablets in their simplest version consist of a mixture of drug with a swelling bioadhesive/sustained release polymer with a bidirectional release. They can be coated on the outer or on all sides but one face with water impermeable hydrophobic substances to allow an unidirectional drug release for systemic delivery (Rathbone et al., 1996; Bose et al., 2011). In case of bilayered tablets, drug can be incorporated in the adhesive layer which comes in contact with the mucosal surface. This drug containing mucoadhesive layer is then protected from the oral cavity environment by a second upper inert layer which faces into the oral cavity. Alternatively, the drug can be incorporated into the upper non adhesive layer to release the drug into the oral cavity. Various types of matrix tablets shown in (Fig. 11) (Gupta et al., 2010; Chandira et al., 2009).

Semisolid preparations (ointments and gels): Solid dosage adhesive forms are used widely than bioadhesive gels or ointments, because they are most used for localized drug therapy within oral cavity. "ORABASE" the original oral mucosal adhesive delivery system consist of finely ground pectin, gelatin and sodium carboxymethyl cellulose dispersed in a poly (ethylene) and a mineral oil gel base. It can be maintained at the site of application for 15-150 min.

Powders: Yamamoto *et al.* (2001) described a hydroxypropyl cellulose and beclomethasone-diproprionate containing powder that was sprayed onto the oral mucosa of rats. A significant increase in the residence time relative to an oral solution was seen and 2.5% of beclomethasone was retained on buccal mucosa for over 4 h (Senel and Hincal, 2001; Ch'Ng *et al.*, 1985; Kaur and Kaur, 2011).

Table 4: Classification of polymers used in formulation of buccal patch (Savage, 1977)

Criteria	Categories	Examples
Source	Seminatural/natural	Agarose, chitosan, gelatin, Hyaluronic acid, Various gums (guar, xanthan, gellan, carragenan, pectin and sodium alginate)
	Synthetic	Cellulose derivatives
		[CMC, thiolated CMC, sodium CMC, HEC, HPC, HPMC, MC, Methyl hydroxyl ethyl cellulose] Poly(acrylic acid)-
		based polymers [CP, PC, PAA, polyacrylates, poly(methyl vinyl ether-co-methacrylic acid), poly(2- hydroxyethyl
		methacrylate), poly(acrylic acid-co-ethylhexyl acrylate), poly(methacrylate), poly(alkyl cyanoacrylate), poly (isohexyl
		cyanoacrylate), poly(isobutyl cyanoa crylate), copolymer of acrylic acid and PEG] Others: polyoxyethylene, PVA, PVP,
		thiolated polymers
Aqueous	Water-soluble	CP, HEC, HPC (waterb38 8C), HPMC (cold water), PAA, sodium CMC, sodium alginate
Solubility	Water-insoluble	Chitosan (soluble in dilute aqueous acids), EC, PC
Charge	Cationic	Aminodextran, chitosan, (DEAE)-dextran, TMC
	Anionic	Chitosan-EDTA, CP, CMC, pectin, PAA, PC, sodium alginate, sodium CMC, xanthan gum
	Non-ionic	Hydroxyethyl starch, HPC, poly(ethylene oxide), PVA, PVP, scleroglucan Potential Bioadhesive forces
	Covalent	Cyanoacrylate
	Hydrogen bond	Acrylates [hydroxylated methacrylate, poly(methacrylic acid)], CP, PC, PVA
	Electrostatic interaction	Chitosan

Buccal patches: The drawbacks of other dosage form are overcomed by development of flexible adhesive patches. Transmucosal delivery patches include unique characteristic like rapid onset of drug delivery, sustained drug release and rapid decline in the serum drug when the patch is removed. The less inter and intraindividual variability absorption profile is showed by a buccal patch

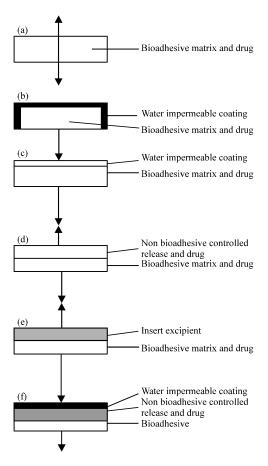


Fig. 11(a-f): Schematic representation of different matrix tablets for buccal delivery. Arrows indicate the direction of drug release (Rossi *et al.*, 2005)

which it confined to the buccal area over which it is attached. List of commercial buccal patches for oral mucosal drug delivery are shown in (Table 5). In general patches with a dissolvable matrix, patches with a non-dissolvable backing and patches with a dissolved backing are the 3 categories for classification of oral mucosal patches (Dixit and Puthli, 2009; Alagusundaram *et al.*, 2009).

Patches with a dissolvable matrix shown in Fig. 12 are designed to release drug into the oral cavity. The mucoadhesive layer (either in drug matrix or attached to drug matrix) would prolong the duration of drug matrix in the oral cavity. Hence, in comparison to other dosage these systems are longer acting and can forms, potentially deliver more drug quantities. Patches with non-dissolvable backing shown in Fig. 13 are usually designed for systemic delivery. Since, they are closed systems and the formulations are protected from saliva, the drug concentrations are controlled and drug is continuously delivered for 10-15 h. The system include disadvantages as removal of backing by the patient after drug administration and use of only a small area. The entire patch get dissolved in oral cavity which have dissolvable backing. But these patches show shorter acting as compared to non-dissolvable backing membrane. Basically buccal patch consist of drug substance, bioadhesive polymer, plasticizers, backing membrane and permeation enhancer. The commonly used



Fig. 12: Buccal patch with a dissolvable matrix (Veillard et al., 1987)

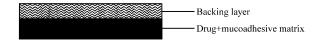


Fig. 13: Buccal patch with a non dissolvable backing (Kurosaki *et al.*, 1989)

	Table 5: List of	commercial	buccal	patches t	for ora	l mucosal	drug (delivery
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Product name/brand name	API (Active pharmaceutical ingredient)	Name of manufacturer	Strength (mg)	References
Triaminic	Dextromethorphan HBr	Novartis	7.5	Bhyan et al. (2011)
Triaminic	Diphenhydramine HCl	Novartis	12.5	
Theraflu	Dextromethorphan HBr	Novartis	15.0	
Gas-X	Simethicone	Novartis	62.5	
Sudafed	Pheny lephrine HCl	Pfizer	10.0	
Benadryl	Diphenhydramine HCl	Pfizer	12.5	
Chloraseptic	Benzocaine Menthol	Prestige	3/3.0	
Suppress	Menthol	InnoZen	2.5	
Orajel	Menthol/Pectin	Del	2/30.0	
Listerine	Cool mint	Pfizer		
DentiPatch	Lidocaine	Noven Pharmaceuticals	46.1	
Onsolis	Fentanyl citrate	Media pharmaceutical		Pather et al. (2008)

Table 6: List of active pharmaceutical ingredients (Drugs) and polymer used in buccal patches

Table 6: List of active p	harmaceutical ingredients (Drugs) and polyn	ner used in buccal patches	
Drug	Category	Polymer	References
Felodipine	Antihypertensive (dihydropyridine	PVP, PVA	Martindale (2005)
Ondansetron Hydrochloride	derivative calcium channel blocker) Antiemetic (5HT-3 inhibitor)	HPMC	Vamshi <i>et al.</i> (2007) and Steven (2002)
Metoprolol tartrate	Selective β -1 adrenergic antagonist	Poly (ethylacrylate methyl methacrylate) copolymer (Eudragit NE40D)	Robertson (1983) and Lanzara (2005)
Metoprolol succinate	Selective β-1 adrenergic antagonist	PVP, PVA	Lehr et al. (1992)
Verapamil HCL	Calcium channel blocker and a class IV antiarrhythmic agent	Chitosan, PVP K-30	Patel et al. (2007b)
Atenolol	β1-receptor selective antagonist	Carbopol 934P, Sodium carboxymethyl cellulose and HPMC	Jug et al. (2009)
Clotrimazole	First line broad-spectrum antifungal agent	Sodium carboxymethylcellulose and carbopol 974P (CP 974P)	Ahuja <i>et al</i> . (1997) and Martin (1990)
Enalapril maleate	Antihypertensive and antianginal	Sodium CMC, HPMC, HEC and polyvinyl pyrrolidone K-90	Houghton (2002) and Singh <i>et al</i> . (2011a,b)
Carvedilol	Non selective β -adrenergic blocking agent with α -1 blocking activity	Chitosan, PVP	Riikka et al. (2009) and Rejaie (2009)
Propranolol hydrochloride	Non-selective beta-adrenergic blocking agent.	Eudragit L-100, Polyvinylpyrrolidone K-30	Gampimol et al. (2002)
Metoprolol tartrate	selective β-1 adrenergic antagonist	Chitosan, PVP K-30 and Ethyl Cellulose	Senel et al. (2000)
Sumatriptan succinate	HT1 receptor agonist used in the treatment of migraine	Ethyl cellulose, PVP	Ryan (1997)
Zolmitriptan	serotonin (5-HT1) agonist	NaCMC, HPMC, HEC, chitosan	Rao et al. (2011)
Lidocaine	Local anaesthetic	HPC, Methocel K-15 HPMC	Repka et al. (2005)
Cetylpyridinium chloride	Quaternary antiseptic	(PVA), hydroxyethyl cellulose (HEC) and chitosan	Noha et al. (2003)
Oxybutynin HCl	Antispasmodic, anticholinergic agent	HPMCP, HPMCK4M, chitosan	Bose et al. (2011)
Fexofenadine HCl	H1 antagonist, is a potent antihistamine	HPMC, Carbopol 934, eudragit RS 100 and Ethylcellulose	Breier et al. (2005)
Methotrexate (MXT)	Anticancer	(EC), sodium alginate, carbopol 934, Na CMC, PVP, HPMC and PVP	Saty abrata et al. (2010)
Miconazole nitrate	Antifungal	Carbopol 934 and HPMC	Vinod et al. (2010)
Famotidine	H2-receptor antagonist (also called H2-blocker) which decreases the amount	HPMC, sodium CMC, PVA	Kumar et al. (2010) and
Lidocaine	of acid produced by the stomach Local anaesthetic	HPMC E-15, NaCMC	Ramchandran <i>et al.</i> (2011) Varshosaz <i>et al.</i> (2006)
Hydrochloride Tizanidine	Skeletal muscle relaxant	NaCMC, Carbopol 934 (CP 934)	Giradkar et al. (2010)
Hydrochloride (TZH) Aceclofenac	NSAID posses good antiinflammatory,	Poly-Sodium CMC and Polyvinyl Alcohol.	Kharinar et al. (2009) and
Meloxicam	analgesics and anti-pyretic. Non-steroidal anti-inflammatory drug is widely used in the treatment of	Chitosan, Carbopol 934p	Gupta et al. (2010) Jafar et al. (2011)
Ibuprofen	rheumatoid arthritis NSAID	NaCMC, PVP	Luana et al. (2004) and
Ranitidine	H2- antagonist	HPMC, PVP	Giri et al. (2010a,b) Alagusundaram et al. (2009)
Risperidone	A benzisoxazole derivative, is a novel antipsychotic	Chitosan, HPMC, PVP, PVA	and Ramchandran et al. (2011) Bingi et al. (2010)
Pimozide	A diphenylbutylpiperidine derivative, an antipsychotic agent.	Carbopol 934 and hydroxypropylmethylcellulose (15 cps) (HPMC), Poly vinyl alcohol (PVA) and poly vinyl pyrrolidone, HPMC 47 cps	Basu et al. (2010)
Methotrexate	Anticancer	Ethyl cellulose	Choudhary et al. (2010)
Carvedilol	Non selective β -adrenergic blocking agent with α -1 blocking activity	Chitosan	Kaur and Kaur (2011)
Triamcinolone Acetonide	A synthetic corticosteroid.	Carbopol 934 and (HPMC)	Singh <i>et al.</i> (2011a,b)
Tetracy cline HCL	Broad-spectmm antibiotics	Carbopol 934, Carvacrol and ethyl cellulose.	Rana et al. (2011)

materials in backing membrane include carbopol, magnesium stearate, HPMC, HPC, CMC, polycarbophil etc. List of active pharmaceutical ingredients (Drugs) and polymer used in buccal patches are shown in Table 6. (Santos *et al.*, 1999; Satyabrata *et al.*, 2010; Ritthidej *et al.*, 2002; Kumar *et al.*, 2010).

Innovative drug delivery systems: Innovative drug delivery systems, such as lipophilic gel, buccal spray and phospholipid vesicles have been recently proposed to deliver peptides via the buccal route (Lee *et al.*, 2000; Senel *et al.*, 2000). A novel liquid aerosol formulation has been recently developed and it is now in clinical phase II

Table 7: Patents on buccal patches

Patent No.	Inventor	Comments
US 2011/0160634	Malcovati (2011)	An oral mucoadhesive patch were formulated in which the protective layer comprises pressure sensitive adhesive
US 7862802	Kim et al. (2011)	The present invention relates to a dry type tooth whitening patch in which peroxide is contained, as a teeth whitening agent, in a matrix type adhesive layer
US 2010/0239646	Nair (2010)	The inventor had designed the sublingual methotrexate patches
US 2010/0189770	Crutchley et al. (2010)	This invention relates to formulation of bioerodible patch which applied on mucosal or dermal surface of mammals
US 7579019	Tapolsky and Osborue (2009)	The formulator designed the device comprises an adhesive layer and a non-adhesive backing layer for providing localized drug delivery and protection to the treatment site
US 2009/0010997	Haley-Jeffrey (2009)	The invention relates to a lenticular medical patch is designed with a thin, tapered edge with multiple layers by passing a single sheet to increase the bioavailability of the drug
US 2008/0274164	Vollmer (2008)	The invention relates to a plaster composition for administering active agents by buccal patch through buccal cavity, lips or genitalia
US 7306812	Zhang (2007a)	The inventor design a patch consisting dissolvable backing layer for transmucosal drug delivery
US 7276246	Zhang (2007b)	The formulator designed a device comprising a water-dissolvable backing layer, an adhesive layer adjacent to at least a portion of the backing layer and an active layer that is circumscribed by the backing layer and adhesive layer
US 6592887	Zerbe et al. (2003)	This invention relates to a composition which contain the apeutic agents and/or breath freshening agents. This formulation is generally used in oral cavity
US 6552024	Chen et al. (2003)	The present invention relates to designing of dissolving oral film which help administration of drug in pediatric and geriatric patient
US 6375963	Repka et al. (2002)	The present invention includes configuration of hot melt extruded film which contain a therapeutic agent and many other additives
US 6319510	Yates (2001)	In this invention formulator design a gum pad which applied on oral cavity and is used for treatment of systemic disorder
US 6210699	Acharya and Baker (2001)	Inventor formulate a mucoadhesive dosage form which can adhere easily on oral mucosa and provide sustained delivery of drug
US 6197331	Lerner et al. (2001)	This invention relates to formulation of sustained and controlled release pharmaceutical oral patch
US 6177096	Zerbe et al. (2001)	Invention relates to formulation of mucoadhesive dosage form by which we can deliver therapeutic and cosmetic agent

trials. This system allows precise insulin dose delivery via a metered dose inhaler in the form of fine aerosolized droplets directed into the mouth. Levels of drug in the mouth are dramatically increased compared with conventional technology. This oral aerosol formulation is rapidly absorbed through the buccal mucosal epithelium and it provides the plasma insulin levels necessary to control postprandial glucose rise in diabetic patients (Modi et al., 2002). A number of advantages are included in this novel, pain free oral insulin formulation which also show rapid absorption, precise dosing control, a simple administration technique and bolus delivery of drug. The recently devised element for delivery of insulin in buccal cavity are phospholipids, deformable vesicles and transferosomes. These vesicles are prepared from surfactants, such as sodium cholate or sodium deoxycholate. They are similar to liposomes morphologically but they pecularly respond to external stresses and their rapid shape transformation require low energy (Yang et al., 2002; Khanna et al., 1996).

Recent invention in field of buccal patches: Lots of work had been done in the field of bioadhesive drug delivery systems and the innovators had successfully patented their work with new modified systems. Some of the patents on this drug delivery have been reported in (Table 7) (Kim *et al.*, 2011).

CONCLUSION

The ease of access and avoidance of the hepatic metabolism enable buccal drug delivery for being a potential alternative conventional oral drug delivery and parental administration which suffer from certain limitations. Almost 80% of the current commercially available formulations are limited to tablet. Oral buccal dosage forms will continue to be an exciting research focus for improving drug absorption especially for the new generation of the so called 'biologics', however, the palatability, irritancy and formulation retention at the site of application need to be considered during design of such formulation.

This article presents a summary of the investigations conducted by various researchers to explore the possibility of utilizing buccal drug delivery and their observations during last two to three decades. The authors expect that this article can be useful and ready reference to those who shall be interested to design, develop a buccal drug delivery system.

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ABBREVIATION

MCG Membrane coating granules

Centimeter cm

Mass m

C Concentration

Time V Volume Dalton Da Litre Millimeter mm =mg Milligram

Ε Young's modules of elasticity =

Fracture energy 2 Critical crack length L

Calorie ca1

EDTA = Ethylenediaminetetraacetic acid

h Kilogram kg

RH = Relative humidity

Wyt Water vapour transmission

S Surface area = Resolution per min rpm Non-keratinized NK K Keratinized

= Standard deviation SD

Hydroxy propyl methylcellulose HPMC =

Hydroxy propyl cellulose HPC CMC Carboxymethylcellulose PVA Polyvinylalcohol = Polyvinylpyrrolidone PVP

CP Carbopol

Polyethylene glycol PEG =Ethyl cellulose EC =

Hydroxyethylcellulose HEC =

US United state Centipoises cps PCPolycarbonate

DEAE = Diethylaminoethyl cellulose

TMC = Trimethylcellulose Polyacrylic acid PAA CP Conductive polymer

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