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Review Article

Multifunctional Nanomaterials for Multifaceted Applications in Biomedical Arena

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Abstract

Nanotechnology is the technology having massive capacity in the areas of biology, biotechnology and medicine technology etc. It includes understanding and controlling materials ordinarily in the size range of 1-100 nm. Owing to their nanoscale effects and enhanced surface area. Nanomaterials have been explored as promising tools for the progression of medication and gene delivery, diagnostic biosensors and biomedical imaging. In contrast with their larger counterparts, nanomaterials have inimitable physicochemical and biological properties. These nanomaterials are at the leading edge in the field of nanotechnology. Numerous properties of the nanomaterials, for instance, size, shape, surface structure, chemical composition and charge significantly influence their interactions with biomolecules and cells. Nanoparticles with size-tunable light emanation have been utilized to create uncommon pictures of tumor destinations. Single-walled carbon nanotubes, having distanced across practically identical to the width of DNA atoms. They have exhibited a great potential as high-efficiency delivery conveyance transporters for biomolecules into cells. Thus, in this review, a brief account of the diverse types of nanosystems is discussed. The applications of various nanomaterials in the biomedical area have been explained in detail. The diverse applications of nanomaterials in drug delivery, gene delivery, etc. have been clearly discussed. Therefore, this review would help the readers to better understand different types of nanomaterials along with the diverse applications of these nanometers in the biomedical field.

Key words: Nanotechnology, nanomaterials, nanomedicine, biomedical applications

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INTRODUCTION

The idiom "Nanotechnology" was first coined in 1974 in the Tokyo Science University by Norio Taniguchi. Nanotechnology was abbreviated to "Nanotech". It is the learning of manipulating materials on an atomic and molecular scale^{1,2}. Nanotechnology deals with a dimension of 0.1 nm to 100 nm materials³. The concept of nanotechnology relates to Richard Feynman's lecture in 1959 at California Institute of Technology which was given on the topic, there's plenty of room at the bottom⁴. However, at that time the practical methods of implementing Feynman's ideas had not been discovered⁵. But, in 1982 with discovery of the Scanning Tunneling Microscope (STM)⁶ and the Atomic Force Microscope (AFM) in 1986⁷ paved the way milestone for the enlargement of nanotechnology. With the help of these two techniques, it becomes probable to observe structures on the atomic scale. The further increment in the development of nanotechnology was achieved by the Eric Drexler's book Engines of Creation, (1986)⁸. The main developments were,

fullerene (1985)⁹ and carbon nanotubes (1991)¹⁰ for the nanostructures synthesis. The first application of nanomaterials in the treatment of cancer has been seen in 2003¹¹. The various stages of development in the field of nanotechnology are tabulated in Table 1. Nanotechnology is an interdisciplinary branch of science, with combining elements of physics, chemistry and technical sciences. Nanotechnology is changing the direction of science by giving us a wide range of applications¹²⁻¹⁴. The application of nanomaterials can be historically traced back to even before the generation of modern science and technology. Michael Faraday explained how metal nanoparticles affect the color of church windows, in his paper in 1857¹⁵.

Nanotechnology includes practical applications in medical and pharmaceutical sciences¹⁶⁻²⁰. Nanotechnology is useful in medicine and related sciences. For example, in drug delivery on the cellular level, cell Bioimaging in the therapy of cancer, targeted therapy and in the regeneration of organs and tissues²¹. Figure 1 outlines the practical application of nanotechnology in pharmacy, medication and medical

Table 1: Periodical development of nanotechnology

Year	Improvement in nanotechnology
1959	R. Feynman began the process
1974	The first-time nanotechnology term was utilized by Taniguchi
1981	IBM Scanning Tunneling Microscope
1985	"Bucky Ball"
1986	The first book "Engines of Creation" on nanotechnology published by K. Eric Drexler, Atomic Force Microscope
1989	IBM logo was made with individual atoms.
1991	First time Carbon nanotubes discovered by S. Iijima
1999	R. Freitas publishes first "Nanomedicine" book
2000	The National Nanotechnology Initiative was established
2001	Preparation and characterization of CNTs Feynman awarded for developing theory for nanometer-scale nanotechnology
2002-04	Prize was awarded to Feynman in the field of Nanotechnology
2005-10	3D Nanosystems like robotics 3D, networking
2011	Era of molecular nanotechnology began

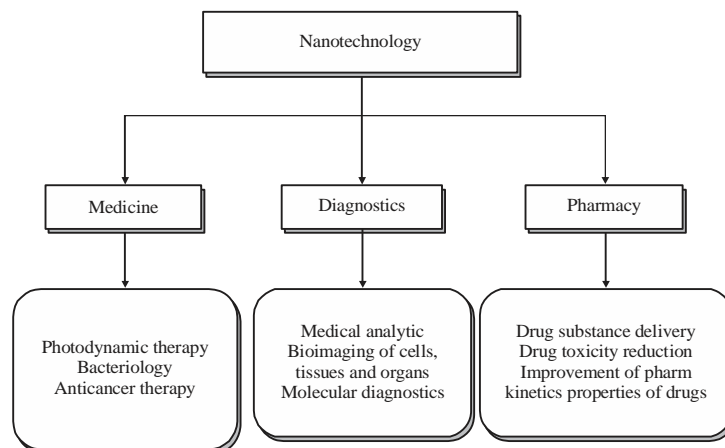


Fig. 1: Application of nanotechnology in medical science

diagnostics. Nanotechnology also useful in many diseases like Alzheimer's disease, cancer, cardiovascular diseases, diabetes, as well as different types of severe infectious diseases like HIV. Damaged tissues can be reproduced or doctored with the aid of nanotechnology. Superior biosensors can be synthesized by means of nanotubes with novel properties. These advanced nanomaterials are used for Astrobiology and help to study the origins of life. This technology also leads in stem cell research like Quantum dots have been employed for molecular imaging and tracking of stem cells, not-fluorescent carbon nanotubes, as well as fluorescent CNTs, have been used in the stem cell research^{22,23}. In stem cell, research nanodevices have been used in imaging and tracking them. Nanodevices also have an application of basic science and translational medicine. Nanodevices also have been used for intelligent delivery, intracellular access and for the biomolecules sensing. Nanotechnology holds a big influence in tissue technology and stem cell microenvironment and has a promising potential for biomedical applications²⁴. Nanotechnology contributes to the growth of every field of science.

PROPERTIES

Enhanced relative surface area and quantum effects are the two important principal properties of nanomaterials, which distinguish theirs from other materials. Due to these extraordinary attributes of minute size and high surface area to volume ratio and the capacity of surface changes. Nanomaterials exhibit unique biological, magnetic and optical properties. The biological properties assist us to adjust processes taking place on a cellular stage. The electrical properties of the nanomaterials rely upon the diameter of the materials. These properties diverge between metallic to semiconducting materials. Nanomaterials have extremely high electrical conductivity, because of the fewer defects in the crystal. Nanomaterials have enhanced thermal conductivity, owing to the heavy vibration of covalent bonds. These have 10X higher thermal conductivity than the metal and this is because of fewer defects in the quartz. They are exceedingly solid and hold out extreme strain. Again, due to fewer defects in the crystal structures, materials do not fracture on bending²⁵.

CLASSIFICATION

The classification of nanomaterials can be managed according to their chemical composition, dimensionality or application and part of the social system. Based on the structure type, nanomaterials can be divided into

Table 2: Nanomaterials based on different phases

Phase	Examples
Single phase solids	Crystalline, amorphous particles and layers
Multi-phase solids	Coated particles, matrix composites
Multi-phase	Aerogels, colloids, ferrofluids

nanoparticles, nanotubes, dendrimers, quantum dots and micelle formations. Based on the chemical classification, nanomaterials can be either inorganic or organic. Organic nanomaterials include carbon structures (CNTs, fullerenes, graphene), dendrimers or polymer nanoparticles and an inorganic nanomaterial include metal oxide nanoparticles, semimetal oxides, metal nanoparticles and semiconductor quantum dots. Figure 2 outlines the classification of nanomaterials in terms of structure type and chemical composition.

Nanomaterials can also be classified by dimensions (0D, 1D, 2D and 3D) like nanorods, nanowires, tubes, fibers, platelets and particles and quantum dots. Nanomaterials can also classify based on phases, like single phase solids and multiphase solids (Table 2).

BIOMEDICAL APPLICATION OF NANOMATERIALS

Different types of nanomaterials are used for biomedical applications. They may be dendrimers, liposomes, carbon nanotube, nanocrystals, nanoparticles, inorganic nanoparticles, metal-based nanoparticles and polymeric nanoparticles etc²⁶⁻³³.

Carbon Nanotubes (CNTs): Ever since the discovery of carbon nanotubes (1991), they have been broadly utilized for the probable applications in the biomedical area. Owing to their capacity to conduct electricity with less resistance they can be utilized in devices and sensors. The tremendously high length-to-diameter ratio makes them suitable as nanocarriers^{34,35}. While their outstanding spectroscopic properties make them attractive for photothermal therapy and/or for medical imaging³⁶. In comparison to the other nanomaterials i.e., spherical ones, CNTs have extremely high absorption properties. The indomitable strength of CNTs because of the sp² character of the C-C bonds and owing to their very less density they are very light materials. Thus, CNTs can be precious materials for diverse of biomedical applications³⁷. Various studies have been demonstrated the superb biocompatibility³⁸⁻⁴⁰ and high capability of adequately functionalized CNTs to selectively target cells after intravenous injection⁴¹. The blend of these properties and their performance in biological systems makes carbon nanotubes as a fascinating nanomaterial for biomedical applications. The

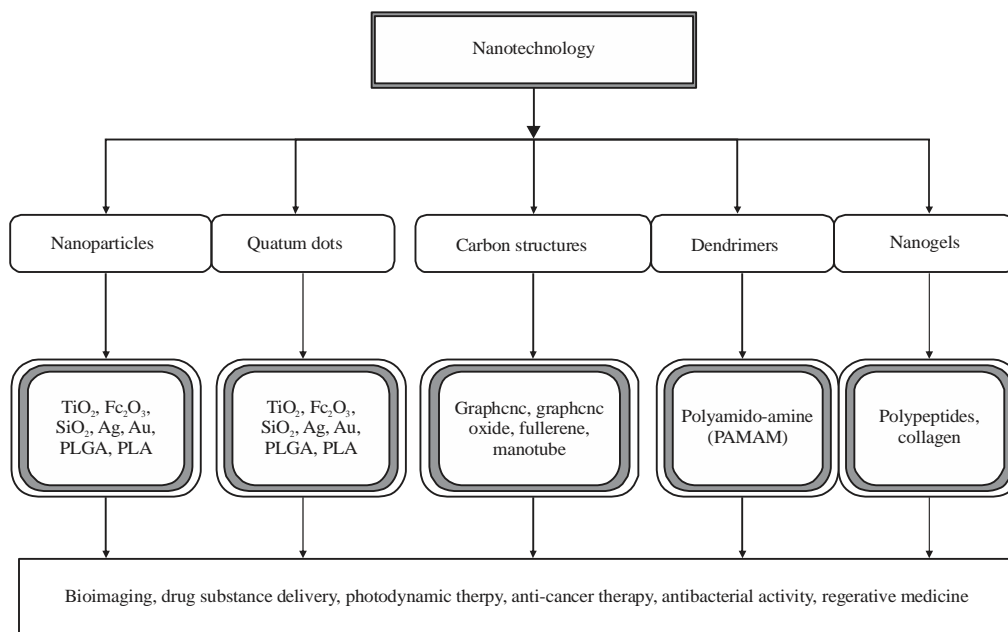


Fig. 2: Outline classification of nanomaterials in terms of structure type and chemical composition. [PLGA- poly (lactic-co-glycolic acid), PLA- polylactic acid, PAMAM- polyamide amine]

classification of CNTs can be done based on the number of their layers. Single-Walled Carbon Nanotubes (SWCNTs) are characterized as one-atom-thick sheets of carbon atoms. These sheets are associated by means of sp^2 bonds arranged in hexagonal structural and rolled up into a cylinder characterized by a high aspect ratio. Multi-walled CNTs (MWCNTs) can have rolled-up structures of multiple concentric sheets. The diameter SWCNTs has 0.4-2 nm and length up to 1 μ m while the MWCNTs has the diameter from 2-100 nm and lengths up to several micrometres⁴²⁻⁴⁴. The syntheses of SWCNTs involve catalyst and generally have a higher amount of impurities and are more for defects during functionalization. The producer of MWCNTs can be achieved in the absence of a catalyst and is having a higher purity as well as fewer defects⁴⁵.

CNT in cancer therapy: The CNTs have been utilized as a therapeutic tool and as a diagnostic in the cancer treatment. Kesharwani *et al.*⁴⁶ thoroughly examined the utilization of diverse cell lines for the testing of CNTs. The most well-known model of anticancer drug that is utilized to show the capacity of new drug delivery systems is the Doxorubicin (DOX)⁴⁷.

Oxidized CNTs are usually favored, over pristine and other modified tubes, as drug vehicles. They contain surface carboxylic gatherings that allow the simple functionalization, specifically for the addition of passive and active targeting moieties. In addition, oxidized carbon tubes possess fewer

impurities of metal and cross cell hindrances more productively because of their abbreviated size. The CNTs functionalized with Folic Acid (FA) and wrapped in hydrophilic polymers can enter cancer cells by means of energy-dependent mechanism mediated by the means of folic acid receptor⁴⁸⁻⁵⁰. This addition of FA with CNTs is effective on numerous human tumor cells. This is a well-organized approach to enhance the cellular uptake of drug loaded carbon tubes.

The functionalized carbon nanotubes along with the paramagnetic particles were reported by Chen *et al.*⁵¹ for building up a drug delivery system, the proper conditions must be chosen. Cisplatin is extensively utilized as an anticancer drug that can be utilized against various distinctive strong tumors. With respect to DOX, cisplatin is a very high degree cytotoxic and requires specific delivery to diminish harmful impacts. With the assistance of DOX, cisplatin tranquilizes loads inside the CNT. Guven *et al.* encouraged the synthesized ultrashort carbon tubes for the delivery of cisplatin, which could avoid the reticuloendothelial system. Li *et al.*⁵² worked on MWNTs which are capped with C18-coated GNPs, in this way the cisplatin molecules retain within MWNTs. Tan *et al.*⁵³ further worked on the coating of carbon tubes with a biocompatible polymer. These Functionalized tubes not only just benefits the general biocompatibility of the systems, but also synergistically improve the thermal properties of the nanocomposite⁵³.

CNTs in tissue engineering and in bone regeneration: Tissue engineering and regenerative medicine are innovative approaches in medical science. In future, for developing engineered artificial tissues for applications in replacement grafts and tissue models for *in vitro* disease studies and drug discovery⁵⁴⁻⁵⁸.

Adhesion, differentiation, migration and proliferation, migration of cells within the ECM form the basis of tissue regeneration. Tonelli et al. reported the tissue engineering applications of CNTs. They focused on how these materials interact with osteoblasts with regeneration of bones, myocytes with regeneration of muscles and neurons for regeneration of neural tissue⁵⁹. Many polymers and polymeric hydrogels have been used for matrix forming materials. On recent advances CNTs enhance the strength of matrices and increase the porosity of matrices⁶⁰, because of its high tensile properties. It was demonstrated by Li *et al.*⁶¹; that CNTs favor subsequent cell adhesion, migration and proliferation. Due to their nanostructure, enhanced surface area and high capacity to absorb proteins⁶¹. The CNTs with biodegradable polymers are effective for bone regeneration matrices. The CNTs with polymer matrices are also the system with biocompatibility together with controllable structure. Ciapetti *et al.*⁶² demonstrated the disputable impact of the introduction of carbon tubes in PLLA/HA develop. PLLA scaffolds stacked with SWNTs indicated poor osteoconductive properties when tried *in vitro* with human bone marrow-derived mesenchymal stromal cells⁶². Hirata *et al.*⁶³ reported the PLLA polymer surface coated with MWNTs which enhanced cell attachment. This might be due to carboxylated groups, so more hydrophilic⁶³ and in this case, differentiation and migration were not affected. Strengthening properties of CNTs are more beneficial for bone regeneration. CNTs with surface characteristics were capable of favouring cell attachment, migration, differentiation and proliferation.

Shimizu *et al.*⁶⁴ reported first the significance of the nanotubes surface modification. These modified carbon nano tubes utilized in the induction of bone calcification and studies the underlying effects of CNTs on bone regeneration⁶⁴. Oxidized MWNTs have surface carboxyl group attract Ca^{2+} ions from physiological fluids. This enhanced the extracellular Ca^{2+} concentration which serves to favors osteoblasts differentiation. This differentiated cell discharge alkaline phosphatase which advances calcification. Pan *et al.*⁶⁵ also developed an ideal weight ratio of CNT/ polymer that permits the synthesis of scaffolds with promising properties for biomedical applications⁶⁵.

CNTs as biosensors: Research in the field of biosensors is to synthesize cost effective sensors for the detection of glucose

in vivo. Owing to the enhanced surface area, electrical conductivity and electrochemical stability, CNTs displayed a noteworthy application in electrochemical glucose⁶⁶. The method utilized for the determination of glucose can be enzyme based or nonenzymic. Wang *et al.*⁶⁰ detailed the preparation of enzyme based detectors that utilized glucose oxidase (GOX) and glucose dehydrogenase (GDH). Electrostatic self-assembly method was employed for the synthesis of the sensor by Fu *et al.*⁶⁷. The synthesized biosensor not only displayed enhanced sensitivity, fast response and less detection limit but also the improved stability. Because of the formation of strong covalent bonds in comparison with the non-photo-cross-linked biosensors.

Hoshino *et al.*⁶⁸ developed multilayer amperometric biosensors by the dry method. The developed sensor comprised of SWNTs, nano-thin plasma, the electron transfer mediator phenothiazine (PT), GDH and polymerized film (PPF). The synergy between CNTs and the electron transfer mediator increases the sensitivity to glucose and decreases the working potential.

Researchers have also synthesized biosensors which can detect other biomolecules and species. Hu *et al.*⁶⁹ prepared detector to identify concanavalin A (ConA), by immobilized D-glucose to MWNTs-polyanilin. The prepared sensors demonstrate a detection limit of 1 pM. This strategy can be employed in the development of fast, easy, cost-effective and miniaturized electrochemical biosensors for biological binding assays.

Chen *et al.*⁷⁰ synthesized an amperometric sensor, for the detection of hydrogen peroxide. The sensor comprised of MWNTs and bimetallic nanoparticles in a Nafion film. The proposed sensor could operate at reduced working potential achieving high sensitivity and quick amperometric response⁷¹. Cao *et al.*⁷² prepared a luminal electrochemiluminescence biosensor, for the determination of A-1-fetoprotein. On integrating the enhanced specific surface area of MWNTs and conductivity of ionic liquids, Bai *et al.*⁷³ synthesized an electrochemical sensor. The integration of carbon tubes into biosensors has an influence on their stability, sensitivity and reproducibility by increasing their interfacial properties.

Dendrimers: Dendrimers are extremely branched synthetic polymers in nano range 1-10 nm. The major advantages of the dendrimers are their control over the size, predictability and number of functional groups present for its amendments. This provides a greater chance of incorporation of drugs. Hence, it enables reproducible pharmacokinetics, which makes dendrimers an interesting drug delivery system for Photo. The action of dendrimer in drug delivery is shown in Table 3.

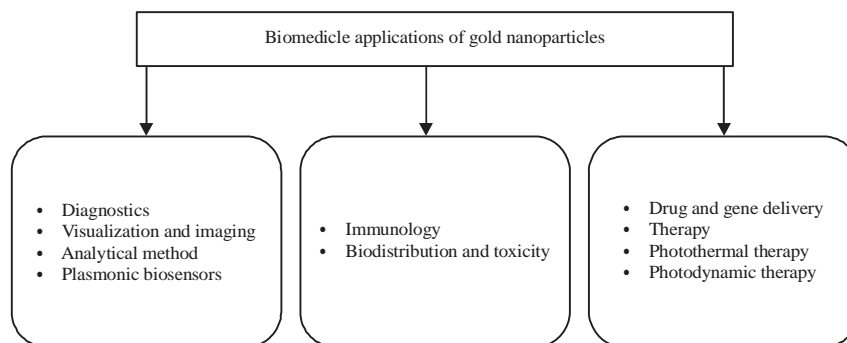


Fig. 3: Biomedical applications of GNPs

Table 3: Different therapeutic moieties studied using dendrimers platform

Drugs	Pharmacology	Dendrimers type	Application	References
10 hydroxycamptothecin	Anticancer	Carboxylated poly(glycerolsuccinic acid)	High toxicity	Morgan <i>et al.</i> ⁷⁴
7 butyl 10 aminocamptothecin	Anticancer	G4.5 poly (glycerol succinic acid) COONa	Increased aqueous solubility and 16-fold increased cellular uptake	Morgan <i>et al.</i> ⁷⁵
Paclitaxel	Anticancer	Polyglycerol (G4 and G5) Increased aqueous solubility ³⁰	Increased aqueous solubility, enhanced	Ooya <i>et al.</i> ⁷⁶
Methotrexate		G4 PAMAM	Cytotoxicity	
		G2.5 and G3 PAMAM 24 fold increment in cytotoxicity ³⁴	24 fold increment in cytotoxicity ³⁴	Gurdag <i>et al.</i> ⁷⁷
		G5 PAMAM	G5 PAMAM Targeted delivery	
Doxorubicin		G4 PAMAM	Improved cytotoxicity	Papagiannaros <i>et al.</i> ⁷⁸
Famotidine		G5 PPI	Improved solubility	Kumar <i>et al.</i> ⁷⁹
Indomethacin	NSAIDs	G5 PPI	Improved solubility	Mutlu <i>et al.</i> ³⁹
Rifampicin	Antitubercular	Mannosylated PPI	Sustained release and targeted delivery	Dutta <i>et al.</i> ⁸⁰
Lamivudine	Anti HIV	Mannosylated PPI	Prolonged drug release up to 144 h	Dutta <i>et al.</i> ⁸¹
Efavirenz	Anti HIV	PPI	Targeted delivery	Dutta <i>et al.</i> ⁸¹
Furosemide	Diuretic	G4 PAMAM	Increased solubility and sustained release	Devarakonda <i>et al.</i> ⁸²
Etoposide	Anticancer	PAMAM	High loading capacity	Wang <i>et al.</i> ⁸³
Erythromycin	Bactericidal			
Antibiotic	G4 PAMAM	Sustained release and improved activity	Bosnjakovic <i>et al.</i> ⁸⁴	
Zidovudine	Anti HIV	G4 PPI	Targeted delivery	Gajbhiye <i>et al.</i> ⁸⁵
Ketoprofen	NSAIDs	PAMAM	Improvement of drug permeation through Skin	Cheng <i>et al.</i> ⁸⁶
Diflunisal	NSAIDs	PAMAM	Improvement of drug permeation through Skin	Zharov <i>et al.</i> ⁸⁷

Metal-based nanoparticles

Gold Nanoparticles (GNPs): The GNPs have found diverse applications in biomedical applications with high sensitivity diagnostic assay, drug and gene delivery, radiotherapy and photothermal therapy enhancement. Outline applications of GNPs are shown in Fig. 3. To make effective GNPs for the biomedical applications GNPs have also been functionalized. In 2003, GNPs were first used in photothermal therapy. Photothermal therapy with GNPs is also called as the plasmonic photothermal therapy⁸⁸. Pitsillides *et al.*⁸⁹ reported the use of 20 and 30 nm gold nanosphere for selective damage to target cells. Khlebtsov *et al.*⁹⁰ reported that the effectiveness of GNPs for the photothermal therapy is based on the shape, size; aggregation extent and the structure of

GNPs. Huang *et al.*⁹¹ showed that 30 nm small aggregates of GNPs were capable of destroying cancerous cells.

GNPs as a therapeutic agent: The GNPs have been increasingly used for the direct therapeutic purposes. Abraham and Himmel⁹² demonstrated the utility of gold nanospheres (GNSs) for the cure of rheumatoid arthritis. Tsai *et al.*⁹³ reported the utilization of gold colloid in treatment collagen-induced arthritis in rats. Further, Brown *et al.*^{94,95} subcutaneously injected GNPs into rats with collagen and pristane-induced arthritis. The antiangiogenic property of GNPs is explained by Bhattacharya *et al.*⁹⁶ and Mukherjee *et al.*⁹⁷. These GNPs mediate angiogenesis, together with that in tumor tissues and reduce tumor activity⁹⁸.

Table 4: Antitumor substances conjugated with GNPs

Drugs	Particle size (nm)	Method of functionalization	Auxiliary substance	References
Paclitaxel	GNSs, 26 nm	Paclitaxel-SH	PEG-SH, TNF	Llevot <i>et al.</i> ¹⁰³
Methotrexate	GNSs, 13 nm	Physical adsorption	-	Paciotti <i>et al.</i> ¹⁰⁴
Daunorubicin	GNSs, 5nm,16 nm	3-mercaptopropionic acid as a linker	-	Chen <i>et al.</i> ¹⁰⁵
Gemcitabine	GNSs, 5 nm	Physical adsorption	Cetuximab	Li <i>et al.</i> ¹⁰⁶
6-mercaptopurine	GNSs, 5 nm	Physical adsorption	-	Patra <i>et al.</i> ¹⁰⁷
Dodecylcysteine	GNSs, 3-6 nm	Physical adsorption	-	Podsiadlo <i>et al.</i> ¹⁰⁸
5-fluorouracil	GNSs, 2 nm	Thiol ligand	-	Azzam <i>et al.</i> ¹⁰⁹
Doxorubicin	GNSs, 12 nm	Physical adsorption	Folate-modified PEG	Agasti <i>et al.</i> ¹¹⁰
Cisplatin	GNSs, 5 nm	PEG-SH linker	PEG-SH, Folic acid	Dhar <i>et al.</i> ¹¹¹
Oxaliplatin	GNSs, 30 nm	PEG-SH linker	PEG-SH	Patra <i>et al.</i> ¹¹²
Kahalalide F	GNSs, 20, 40 nm	Physical adsorption	-	Brown <i>et al.</i> ¹¹³
Tamoxifen	GNSs, 25 nm	PEG_SH as a linker	PEG-SH	Hosta <i>et al.</i> ¹¹⁴
Herceptin	GNSs,	11-mercaptoundecanoic acid as a linker	-	Dreaden <i>et al.</i> ¹¹⁵
B-Lapachon	GNSs, 25 nm	Physical adsorption	Cyclodextrin as a drug pocket, anti-EGFR, PEG-SH	Eghtedari <i>et al.</i> ¹¹⁶
Prospidin	GNSs, 50 nm	Physical adsorption	-	Park <i>et al.</i> ¹¹⁷

Wang *et al.*⁹⁹ reported the PEG-coated gold nanorods (GNRs), they have the unique ability to induce tumor cell death and subsequently damage them.

GNPs as drug carriers: The GNPs have shown potential applications in medicines as well as targeted drug delivery^{100,101}. The most popular objects for targeted delivery are antitumor preparations¹⁰² and antibiotics. The GNPs have been complexes with a number of antitumor substances¹⁰³⁻¹¹⁷ which are shown in Table 4.

Gu *et al.*¹¹⁸ reported a gold colloid with vancomycin. A gold vancomycin-colloid is effective toward various enteropathogenic strains of *enterococcus faecal*, *Enterococcus faecium* and *E. coli*. Rosemary *et al.*¹¹⁹ reported a similar complex, which was synthesized by gold nanoshells and ciprofloxacin. This complex with gold nanoshells shows elevated antibacterial activity against *E. coli*/bacteria. Selvaraj and Alagar further reported a conjugated colloidal gold complex with antileukemic drug 5-fluorouracil¹²⁰. The gold colloid complex exhibited unexpectedly high antifungal and antibacterial activities against *A. niger*, *Aspergillus fumigates*, *E. coli* and *Pseudomonas aeruginosin*. With the help of GNPs, the antibacterial activity of the antibiotics is improved. Burgin *et al.* reported the synthesis of a stable composite of GNPs coated with antibiotic molecules which enhance antibacterial activity¹²¹. The GNP coated antibiotic composite had extra high antibacterial activity against *E. coli* and *S. aureus*. Nie *et al.*¹²² reported GNPs complexed with tocopherol which has high antioxidant activity. Bowman *et al.*¹²³ reported the preparation of the conjugate of GNPs with TAK-779 which exhibits more prominent activity against HIV than the native preparation at the cost of the high local concentration. Finally, Chamberland *et al.*¹²⁴ reported a therapeutic effect of GNRs conjugated with antirheumatic drug etanercept.

Iron oxide nanoparticles (IONPs): Magnetic iron oxide nanoparticles have been in the limelight due to the exclusive properties stemmed from their exceptionally minute size and large specific surface area¹²⁵. They have been broadly investigated in current years for the promising biomedical applications, such as drug delivery^{126,127}, immunoassay analyser^{128,129}, magnetic resonance imaging^{130,131} and cancer hyperthermia¹³². For most of these applications, surface modification of iron oxide nanoparticles plays an important role in enhancing their hydrophilicity, biocompatibility and conjugation of bioactive functional groups. To this end, many materials have been employed to design stable surface coatings for iron oxide nanoparticles, such as polymers, surfactants, or inorganic shells^{133,134}.

Magnetic Resonance Imaging (MRI): The MRI drugs help to enlarge the contrast of the image between disease and the normal tissue to distinguish pathological and healthy tissues. A number of different contrast agents have been used for this function. Paramagnetic gadolinium chelates work as a contrast agent in MRI studies up to¹³⁵. These MRI agents work by reduction of the longitudinal relaxation time (T_1 relaxation time) of water in tissues. While superparamagnetic iron oxide nanoparticles (SPIONs) enhance the specificity and diagnostic sensitivity due to their advanced properties. They have the capability to decrease both longitudinal relaxation time (T_1) and transversal relaxation time (T_2) and have the higher molar relaxivities¹³⁶. The effectiveness of these particles can be further improved by the amendment of their surface. These modifications can be done with the help of biologically active antibodies, polysaccharides, proteins etc. The efficiency of these particles depends on their charge, size and coating properties of particles¹³⁷. Yu *et al.*¹³⁸ developed a new contrast agent for MRI in cancer imaging. These particles increase the

permeability and retention effect of the coating, which helps in tumor determination. So, this MRI agent has high efficiency to deliver anti-cancer drugs to tumors also. Thus, this contrast agent is beneficial in both cancer imaging and therapy¹³⁸. Yang *et al.*¹³⁹ further synthesis the multifunctional SPIONs. These are used for MRI or positron emission tomography (PET), targeted drug delivery and utilized for the cancer therapy¹³⁹.

Gene and drug delivery: The IONPs have also been utilized in gene and drug delivery *in vivo* and *in vitro*. Magnetic nanoparticles attach to therapeutic gene and these particles targeted towards the specific target via high gradient external magnets. This approach is known as magnetoreception. After reaching the specific target the gene release from the IONPs. The release of the gene with the IONPs can possibly through the hydrolysis of the polymer which is coating around NPs and through the enzymatic degradation¹⁴⁰. Mah *et al.*¹⁴¹ used Adeno-Associated Virus (AAV), cleavable heparin and green fluorescent protein attach to magnetic particles. These reported for the targeted delivery of DNA¹⁴¹. Lubbe *et al.* and Wilson *et al.* utilized IONPs for the delivery of epidoxorubicin¹⁴².

Basuki *et al.*¹⁴³ reported the synthesis of IONPs with exceptional colloidal stability that is stabilized with functional polymers with a capability to attach DOX through a pH-onsite imine bond¹⁴³. Laurent *et al.*¹⁴⁴ reported the IONPs covered the starch derivative and this can be further functionalized with phosphate groups which help in chemotherapy by targeting mitoxantrone to the tumor location¹⁴⁴. In addition, SPIONs and quantum dots have also been used to accomplish targeted delivery of an anti-cancer agent¹⁴⁵. Recently, Kebede *et al.*¹⁴⁶ reported the synthesis the composite of iron oxide with chitosan and loaded with insulin and investigated the use in type II diabetes through oral delivery¹⁴⁶. The iron oxide-chitosan nanocomposite loaded with insulin can lower down the 51% blood glucose levels in mild diabetic, sub diabetic and severely diabetic rats. Shen *et al.* synthesized the similar composite of hybrid nanogels composed from SPIONs with chitosan and CdTe quantum dots¹⁴⁷. Spherical hybrid nanogels, smaller than 160 nm, were used for insulin loading. These hybrid nanoparticles could be capable of both insulin delivery purposes and cell imaging. IONPs have been also used as biosensors to detect proteins¹⁴⁸, cells¹⁴⁹, nucleotides¹⁵⁰ and pathogens¹⁵¹ in a biological sample. Perez *et al.*¹⁵² reported avidin conjugated SPIONs for the successful detection technique for Green Fluorescent Protein (GFP)¹⁵². For detection of GFP, nanoparticles were first conjugated with avidin and then the biotinylated anti-GFP polyclonal antibody was attached to the nanoparticles surfaces. With this

biosensor molecule, GFP can determine in less than 30 min. Tumor cells, progenitor cells, or stem cells can also be targeted by modification of IONPs with the functionalization^{153,154}.

Graphene: Graphene has sp² hybridized carbon atoms arranged in a 2D array. Graphene has many properties suitable for electronic, mechanical, optical and thermal applications^{155,156}.

Graphene has extensive biomedical applications, like drug and gene delivery, imaging and biosensing. The utilization of graphene and its derivatives such as graphene oxide are due its attractive properties like;

- High specific surface area, i.e., 2630 m²/g
- High thermal conductivity, i.e., ~5000 W/m/K
- High electronic conductivity, i.e., 200,000 cm² V⁻¹ s⁻¹
- Mechanical strength, ~1100 Gpa of graphene

In addition to the above exceptional properties, graphene and graphene oxide has intrinsic biocompatibility. It is cost effective and high production and facile biological/chemical functionalization of GO^{157,158}.

Graphene and graphene oxide in drug delivery: Graphite on vigorous oxidation produces graphene oxide by Hummers method¹⁵⁹. The GO is a proficient nanocarrier for drug and gene delivery. Graphene oxide having 1-2 nm broad layer and size in the range of 1-100 nm is used for drug delivery¹⁶⁰⁻¹⁶². The GO has exclusive structural features, like large and planar sp² hybridized carbon domain, enriched oxygen-containing groups, high specific surface area (2630 m²/g) and. The GO has an outstanding physiological solubility and stability. The potential of loading drugs or genes via chemical or physisorption and biocompatibility. In addition to the reactive carboxyl and hydroxyl groups GO makes easy conjugation with a variety of systems. Systems like as polymers¹⁶³, biomolecules, DNA¹⁶⁴, protein¹⁶⁵⁻¹⁶⁸, QD¹⁶⁹, iron oxide nanoparticles¹⁷⁰ and others¹⁷¹. Imparting GO multi-modalities and multi-functionalities for miscellaneous medical and biomedical applications. First-time Nanoscale Graphene Oxide (NGO) synthesized by Dai *et al.*¹⁷¹ which was inspired by the ideas of CNT-based drug delivery. For the efficient delivery of water insoluble aromatic anticancer drug into cells, NGOs are the novel nanocarriers¹⁷². Depan *et al.*¹⁷³ also studied the pH-sensitive drug release behavior from different GO-based drug delivery systems¹⁷³. The advance investigation of drug delivery by GO advantages from anticancer drugs to other drugs also for non-cancer diseases treatment¹⁷⁴. Rana *et al.*¹⁷⁵ demonstrated the chitosan-grafted GO for the

anti-inflammatory drug, delivery i.e., Ibuprofen. To further improve the anticancer drug effect, Yang *et al.*¹⁷⁶ synthesized GO-Fe₃O₄ NPs hybrid which is a bio and magnetic-double targeting drug delivery mechanism¹⁷⁶.

Graphene and graphene oxide in gene delivery: Gene therapy is a new and shows the potential way to cure a variety of severe diseases. Diseases which are especially due to the genetic disorders like cancer, cystic fibrosis and Parkinson's disease¹⁷⁷. For performing an excellent gene therapy, a gene vector plays an important role to defend DNA from nuclease degradation and helps to facilitate cellular uptake of DNA with very high transfection efficiency¹⁷⁸. The most important problem in front of the growth of gene therapy is the need of safe and efficient vectors for the gene¹⁶⁹. Liu *et al.*¹⁷⁶ studied the gene delivery using graphene oxide derivative with the polyethyleneimine (PEI)-modified i.e., PEI-GO. The GO hybrid with positively charged PEI and with the electrostatic interaction between cationic polymer and DNA. It allows for the condensation of plasmid DNA onto the surface of GO sheet. The transfection efficiency of GO-PEI-10K and GO-PEI-1.2K was also studied by Liu and co-workers. They compared the transfection efficiency with the free polymers of PEI-10K and PEI-1.2K, respectively^{179,180}. The chitosan-functionalized graphene oxide (GO-CS) sheets were reported for the application for drug and gene delivery by the research group of Singapore. This work of this group shows that GO-CS sheets have a high drug payload and improved cancer cell killing ability of the CPT-loaded GO-CS as compared to the pure CPT¹⁸¹.

Graphene oxide in cancer: Lu and colleagues studied the use of PEGylated GO first time in the photothermal therapy and *in vivo* tumor uptake. This study done with the help of xenograft tumor mouse models. The PEG-modified GO shows very high tumor uptake ability due to extremely efficient tumor passive targeting of graphene oxide caused by EPR effect^{182,183}. Markovic *et al.*¹⁸⁴ reported the comparison of the photothermal anticancer activity of CNTs and NIR-excited graphene.

Zhang *et al.* synthesized the NGO-PEG-DOX for anti-tumor effect by the combination of chemo and photothermal therapies in both *in vitro* and *in vivo*. The combined experiment of both chemo-photothermal therapies exhibited synergistic effect which led to enhanced cancer killing effect as compared to photothermal or chemotherapy¹⁸⁵.

Huang and coworkers demonstrated the function of sulfonic acid and folic acid-conjugated graphene oxide laden with porphyrin photosensitizers for targeting PDT¹⁸⁶. Tian and

colleagues reported the GO loaded with a photosensitizer in the application of photo-thermally assisted photodynamic therapy. This combined treatment yields extraordinarily enhanced cancer-killing effect¹⁸⁷.

GO-based antibacterial material: Peng *et al.*¹⁸⁸ developed macroscopic freestanding GO and reduced graphene oxide (rGO) paper from their suspension by vacuum filtration technique. These GO papers show a very high antibacterial effect. Akhavan and Ghaderi¹⁸⁹ reported the antibacterial effect of nanosheets of graphene. These sheets show positive effect for both Gram-positive and Gram-negative models of bacteria in the form of nanowalls. These nanowalls are deposited on stainless steel substrates¹⁸⁹. Liu *et al.*¹⁹⁰ reported the antibacterial mechanism of graphene and its derivative, graphite (Gt), graphite oxide (GtO), GO and rGO. GO, rGO, Gt and GtO are the order decreasing order of antibacterial activity¹⁹⁰.

Nanocomposites: Nanocomposites are the hybrid which formed by the composition of a polymer or copolymer in which the NPs spread over the matrix of the polymer. These are used for the applications of anti-HIV drug delivery. PLA/chitosan are considerably useful for the anti-HIV drug delivery applications¹⁹¹. Controlled releasing of the drug is the key factor for the drug delivery. Metal oxide nanocomposites are the appropriate e.g., for the capable transport system in the delivery applications¹⁹².

Polymeric nanoparticles: Polymeric NPs have potential applications in drug delivery. Biodegradable polymers, such as poly(alkyl cyanoacrylates), polyesters, its copolymers and also the polysaccharides are useful for the drug delivery. Polysaccharides like polyglycolic acid, polylactic acid, poly(methylidene malonate), poly(ϵ -caprolactone) are utilized for drug delivery. Polymeric NPs are used in cancer for a delivery remedy to tumor cells with superior capability and decreased cytotoxicity on marginal tissues^{193, 194}.

Inorganic materials: Metals, metal oxides and metal sulfides are the inorganic nanomaterials utilized to synthesize innumerable nanomaterials with a different shape, size and porosity. The Si NPs have been widely used in a drug delivery system. Because of its acquired outstanding properties like high pore volume, tunable pore structures, physicochemical stability and high specific area. The Si NPs were also used for prohibited delivery of a variety of hydrophobic or hydrophilic active agents. The Si NPs have surface properties like surface

functionalization and PEGylation and can function as a drug delivery vehicle for cancer treatment. Douroumis¹⁹⁵ and Wu López *et al.*¹⁹⁶. demonstrated delivery of methotrexate anticancer drug.

CONCLUSION

Nanomaterials have been utilized as a promising tool in biomedical imaging, diagnostic biosensors and cancer therapy etc. Owing to the enhanced surface area and nanoscale effects, nanomaterials have distinct biological properties in comparison to their larger counterparts. The properties of nanomaterials significantly affect their interactions with biomolecules and cells. This is because of their small size, shape, surface structure, charge and solubility etc. There is a splendid upcoming to nanotechnology, by its converging with different technologies and the subsequent emergence of complex and innovative hybrid technologies. Science dependent advances are entwined with nanotechnology are as of now utilized to control hereditary material. Advance investigate in nanotechnology, can help explore its application in other areas. Medicine, regenerative medicine, stem cell research and nutraceuticals are areas which required further amendment by the nanotechnology innovations.

SIGNIFICANCE STATEMENTS

This review focuses on the most relevant and popular nanomaterials. It explores the biomedical applications of nanomaterials such as CNTs, graphene etc. The review will facilitate the readers to know about the various nanomaterials and their utilization in severe diseases, for instance, cancer etc. in detail.

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REFERENCES

1. Deulkar, S.H., H.J. Yo and J.L. Huang, 2010. Tem-based investigations on CVD-assisted growth of ZnO nanowires inside nanochannels of anodized aluminum oxide template. *Int. J. Nanosci.*, 9: 225-235.

2. Arivalagan, K., S. Ravichandran, K. Rangasamy and E. Karthikeyan, 2011. Nanomaterials and its potential applications. *Int. J. Chem. Tech. Res.*, 3: 534-538.
3. Sahoo, S.K., S. Parveen and J.J. Panda, 2007. The present and future of nanotechnology in human health care. *Nanomed.: Nanotechnol. Biol. Med.*, 3: 20-31.
4. Shea, C.M., 2005. Future management research directions in nanotechnology: A case study. *J. Eng. Technol. Manage.*, 22: 185-200.
5. Miyazaki, K. and N. Islam, 2007. Nanotechnology systems of innovation: An analysis of industry and academia research activities. *Technovation*, 27: 661-675.
6. Binnig, G., H. Rohrer, C. Gerber and E. Weibel, 1982. Surface studies by scanning tunneling microscopy. *Phys. Rev. Lett.*, 49: 57-61.
7. Binnig, G., C.F. Quate and C. Gerber, 1986. Atomic force microscope. *Phys. Rev. Lett.*, 56: 930-933.
8. Toumey, C.P., 2007. Expeditions to Na-no-tech. *Anthropol. Today*, 23: 23-25.
9. Kroto, H.W., J.R. Heath, S.C.O. Brien, R.F. Curl and R.E. Smalley, 1985. C₆₀: Buckminsterfullerene. *Nature*, 318: 162-163.
10. Iijima, S., 1991. Helical microtubules of graphitic carbon. *Nature*, 354: 56-58.
11. Rajkumar, N., D. Umamaheswari and K. Ramachandran, 2010. Photoacoustics and magnetic studies of Fe₃O₄ nanoparticles. *Int. J. Nanosci.*, 9: 243-250.
12. Fang, M., M. Chen, L. Liu and Y. Li, 2017. Applications of quantum dots in cancer detection and diagnosis: A review. *J. Biomed. Nanotechnol.*, 13: 1-16.
13. Woiski, T.D., L. de Castro Poncio, J. de Moura, A. Orsato, A.G. Bezerra, Jr., J.C. Minozzo and B.C. de Figueiredo, 2017. Anti-hMC2RL1 functionalized gold nanoparticles for adrenocortical tumor cells targeting and imaging. *J. Biomed. Nanotechnol.*, 13: 68-76.
14. Menaa, B., 2011. The importance of nanotechnology in biomedical sciences. *J. Biotechnol. Biomater.*
15. Artman, G.D., A.W. Grubbs and R.M. Williams, 2007. Concise, asymmetric, stereocontrolled total synthesis of stephacidins A, B and notoamide B. *J. Am. Chem. Soc.*, 129: 6336-6342.
16. Nikalje, A.P., 2015. Nanotechnology and its applications in medicine. *Med. Chem.*, 5: 081-089.
17. Tans, S.J., A.R. Verschuere and C. Dekker, 1998. Room-temperature transistor based on a single carbon nanotube. *Nature*, 393: 49-52.
18. Bachtold, A., P. Hadley, T. Nakanishi and C. Dekker, 2001. Logic circuits with carbon nanotube transistors. *Science*, 294: 1317-1320.
19. Zolnik, B.S. and N. Sadrieh, 2009. Regulatory perspective on the importance of ADME assessment of nanoscale material containing drugs. *Adv. Drug Delivery Rev.*, 61: 422-427.
20. Valavanidis, A. and T. Vlachogianni, 2016. Engineered nanomaterials for pharmaceutical and biomedical products new trends, benefits and opportunities. *J. Pharma Rep.*, Vol. 1.

21. Gao, J. and B. Xu, 2009. Applications of nanomaterials inside cells. *Nano Today*, 4: 37-51.
22. Wang, Z., J. Ruan and D. Cui, 2009. Advances and prospect of nanotechnology in stem cells. *Nanoscale Res. Lett.*, 4: 593-605.
23. Yue, Y., D. Yuchi, P. Guan, J. Xu, L. Guo and J. Liu, 2016. Atomic scale observation of oxygen delivery during silver-oxygen nanoparticle catalysed oxidation of carbon nanotubes. *Nature Commun.*, Vol. 7.
24. Deb, K.D., M. Griffith, E. De Muinck and M. Rafat, 2012. Nanotechnology in stem cells research: Advances and applications. *Front Biosci.*, 17: 1747-1760.
25. Fadeel, B. and A.E. Garcia-Bennett, 2010. Better safe than sorry: Understanding the toxicological properties of inorganic nanoparticles manufactured for biomedical applications. *Adv. Drug Deliv. Rev.*, 62: 362-374.
26. Strasak, T., J. Maly, D. Wrobel, M. Maly and R. Herma *et al.*, 2017. Phosphonium carbosilane dendrimers for biomedical applications-synthesis, characterization and cytotoxicity evaluation. *RSC Adv.*, 7: 18724-18744.
27. Taylor, A., K.M. Wilson, P. Murray, D.G. Fernig and R. Levy, 2012. Long-term tracking of cells using inorganic nanoparticles as contrast agents: Are we there yet? *Chem. Soc. Rev.*, 41: 2707-2717.
28. Panahi, Y., M. Farshbaf, M. Mohammadhosseini, M. Mirahadi, R. Khalilov, S. Saghfi and A. Akbarzadeh, 2017. Recent advances on liposomal nanoparticles: Synthesis, characterization and biomedical applications. *Artif. Cells Nanomed. Biotechnol.*, 45: 788-799.
29. Mattea, F., J. Vedelago, F. Malano, C. Gomez, M.C. Strumia and M. Valente, 2017. Silver nanoparticles in X-ray biomedical applications. *Radiat. Phys. Chem.*, 130: 442-450.
30. Cabuzu, D., A. Cirja, R. Puiu and A.M. Grumezescu, 2015. Biomedical applications of gold nanoparticles. *Curr. Top. Med. Chem.*, 15: 1605-1613.
31. Bajwa, N., N.K. Mehra, K. Jain and N.K. Jain, 2016. Pharmaceutical and biomedical applications of quantum dots. *Artif. Cells Nanomed. Biotechnol.*, 44: 758-768.
32. Jorfi, M. and E.J. Foster, 2015. Recent advances in nanocellulose for biomedical applications. *J. Applied Polym. Sci.*, Vol. 132.
33. McNamara, K. and S.A.M. Tofail, 2017. Nanoparticles in biomedical applications. *Adv. Phys.*, 2: 54-88.
34. Boczkowski, J. and S. Lanone, 2007. Potential uses of carbon nanotubes in the medical field: How worried should patients be? *Nanomedicine*, 4: 407-410.
35. Beg, S., M. Rizwan, A.M. Sheikh, M.S. Hasnain, K. Anwer and K. Kohli, 2011. Advancement in carbon nanotubes: Basics, biomedical applications and toxicity. *J. Pharm. Pharmacol.*, 63: 141-163.
36. De Volder, M.F., S.H. Tawfik, R.H. Baughman and A.J. Hart, 2013. Carbon nanotubes: Present and future commercial applications. *Science*, 339: 535-539.
37. Heister, E., E.W. Brunner, G.R. Dieckmann, I. Jurewicz and A.B. Dalton, 2013. Are carbon nanotubes a natural solution? Applications in biology and medicine. *ACS Applied Mater. Interfaces*, 5: 1870-1891.
38. Yan, L., F. Zhao, S. Li, Z. Hu and Y. Zhao, 2011. Low-toxic and safe nanomaterials by surface-chemical design, carbon nanotubes, fullerenes, metallofullerenes and graphenes. *Nanoscale*, 3: 362-382.
39. Mutlu, G.M., G.S. Budinger, A.A. Green, D. Urich and S. Soberanes *et al.*, 2010. Biocompatible nanoscale dispersion of single-walled carbon nanotubes minimizes *in vivo* pulmonary toxicity. *Nano Lett.*, 10: 1664-1670.
40. Jain, S., V.S. Thakare, M. Das, C. Godugu and A.K. Jain *et al.*, 2011. Toxicity of multiwalled carbon nanotubes with end defects critically depends on their functionalization density. *Chem. Res. Toxicol.*, 24: 2028-2039.
41. Kotagiri, N. and J.W. Kim, 2014. Stealth nanotubes: Strategies of shielding carbon nanotubes to evade opsonization and improve biodistribution. *Int. J. Nanomed.*, 9: 85-105.
42. Elhissi, A., W. Ahmed, I.U. Hassan, V.R. Dhanak and A. D'Emanuele, 2012. Carbon nanotubes in cancer therapy and drug delivery. *J. Drug Delivery*, Vol. 2012.
43. Ma-Hock, L., S. Treumann, V. Strauss, S. Brill and F. Luizi *et al.*, 2009. Inhalation toxicity of multi-wall carbon nanotubes in rats exposed for 3 months. *Toxicol. Sci.*, 112: 468-481.
44. Bottini, M., N. Rosato and N. Bottini, 2011. PEG-modified carbon nanotubes in biomedicine: Current status and challenges ahead. *Biomacromolecules*, 12: 3381-3393.
45. He, H., L.A. Pham-Huy, P. Dramou, D. Xiao, P. Zuo and C. Pham-Huy, 2013. Carbon nanotubes: Applications in pharmacy and medicine. *Bio. Med. Res. Int.*, Vol. 2013.
46. Kesharwani, P., R. Ghanghoria and N.K. Jain, 2012. Carbon nanotube exploration in cancer cell lines. *Drug Discov. Today*, 17: 1023-1030.
47. Fabbro, C., H. Ali-Boucetta, T. Da Ros, K. Kostarelos, A. Bianco and M. Prato, 2012. Targeting carbon nanotubes against cancer. *Chem. Commun.*, 48: 3911-3926.
48. Meng, L., X. Zhang, Q. Lu, Z. Fei and P.J. Dyson, 2012. Single walled carbon nanotubes as drug delivery vehicles: Targeting doxorubicin to tumors. *Biomaterials*, 33: 1689-1698.
49. Heister, E., V. Neves, C. Lamprecht, S.R.P. Silva, H.M. Coley and J. McFadden, 2012. Drug loading, dispersion stability and therapeutic efficacy in targeted drug delivery with carbon nanotubes. *Carbon*, 50: 622-632.
50. Ji, Z., G. Lin, Q. Lu, L. Meng and X. Shen *et al.*, 2012. Targeted therapy of SMMC-7721 liver cancer *in vitro* and *in vivo* with carbon nanotubes based drug delivery system. *J. Colloid Interface Sci.*, 365: 143-149.
51. Lu, Y.J., K.C. Wei, C.C.M. Ma, S.Y. Yang and J.P. Chen, 2012. Dual targeted delivery of doxorubicin to cancer cells using folate-conjugated magnetic multi-walled carbon nanotubes. *Colloids Surf. B: Biointerfaces*, 89: 1-9.

52. Li, J., S.Q. Yap, S.L. Yoong, T.R. Nayak and G.W. Chandra *et al.*, 2012. Carbon nanotube bottles for incorporation, release and enhanced cytotoxic effect of cisplatin. *Carbon*, 50: 1625-1635.
53. Tan, A., S.Y. Madani, J. Rajadas, G. Pastorin and A.M. Seifalian, 2012. Synergistic photothermal ablative effects of functionalizing carbon nanotubes with a POSS-PCU nanocomposite polymer. *J. Nanobiotech.*
54. Barthes, J., H. Ozcelik, M. Hindie, A. Ndreu-Halili, A. Hasan and N.E. Vrana, 2014. Cell microenvironment engineering and monitoring for tissue engineering and regenerative medicine: The recent advances. *BioMed Res. Int.*
55. Hasan, A., A. Khattab, M.A. Islam, K.A. Hweij and J. Zeitouny *et al.*, 2015. Injectable hydrogels for cardiac tissue repair after myocardial infarction. *Adv. Sci.*
56. Hasan, A., A. Memic, N. Annabi, M. Hossain and A. Paul *et al.*, 2014. Electrospun scaffolds for tissue engineering of vascular grafts. *Acta Biomaterialia*, 10: 11-25.
57. Hasan, A., A. Paul, N.E. Vrana, X. Zhao and A. Memic *et al.*, 2014. Microfluidic techniques for development of 3D vascularized tissue. *Biomaterials*, 35: 7308-7325.
58. Hasan, A., K. Ragaert, W. Swieszkowski, S. Selimovic and A. Paul *et al.*, 2014. Biomechanical properties of native and tissue engineered heart valve constructs. *J. Biomech.*, 47: 1949-1963.
59. Tonelli, F.M.P., A.K. Santos, K.N. Gomes, E. Lorencon, S. Guatimosim, L.O. Ladeira and R.R. Resende, 2012. Carbon nanotube interaction with extracellular matrix proteins producing scaffolds for tissue engineering. *Int. J. Nanomed.*, 7: 4511-4529.
60. Wang, W., Y. Zhu, F. Watari, S. Liao and A. Yokoyama *et al.*, 2012. Carbon nanotubes/hydroxyapatite nanocomposites fabricated by spark plasma sintering for bonegraft applications. *Applied Surf. Sci.*, 262: 194-199.
61. Li, X., H. Liu, X. Niu, B. Yu and Y. Fan *et al.*, 2012. The use of carbon nanotubes to induce osteogenic differentiation of human adipose-derived MSCs *in vitro* and ectopic bone formation *in vivo*. *Biomaterials*, 33: 4818-4827.
62. Ciapetti, G., D. Granchi, K.N.V. Devescovi, S.R.E. Baglio and E. Leonardi *et al.*, 2012. Enhancing osteoconduction of PLLA-based nanocomposite scaffolds for bone regeneration using different biomimetic signals to MSCs. *Int. J. Mol. Sci.*, 13: 2439-2458.
63. Hirata, E., T. Akasaka, M. Uo, H. Takita, F. Watari and A. Yokoyama, 2012. Carbon nanotube-coating accelerated cell adhesion and proliferation on poly (L-lactide). *Applied Surf. Sci.*, 262: 24-27.
64. Shimizu, M., Y. Kobayashi, T. Mizoguchi, H. Nakamura and I. Kawahara *et al.*, 2012. Carbon nanotubes induce bone calcification by bidirectional interaction with osteoblasts. *Adv. Mater.*, 24: 2176-2185.
65. Pan, L., X. Pei, R. He, Q. Wan and J. Wang, 2012. Multiwall carbon nanotubes/polycaprolactone composites for bone tissue engineering application. *Colloids Surf. B: Biointerfaces*, 93: 226-234.
66. Liu, F., Y. Piao, K.S. Choi and T.S. Seo, 2012. Fabrication of free-standing graphene composite films as electrochemical biosensors. *Carbon*, 50: 123-133.
67. Fu, G. and Z. Dai, 2012. Efficient immobilization of glucose oxidase by in situ photo-cross-linking for glucose biosensing. *Talanta*, 97: 438-444.
68. Hoshino, T., S.I. Sekiguchi and H. Muguruma, 2012. Amperometric biosensor based on multilayer containing carbon nanotube, plasma-polymerized film, electron transfer mediator phenothiazine and glucose dehydrogenase. *Bioelectrochemistry*, 84: 1-5.
69. Hu, F., S. Chen, C. Wang, R. Yuan, Y. Xiang and C. Wang, 2012. Multi-wall carbon nanotube-polyaniline biosensor based on lectin-carbohydrate affinity for ultrasensitive detection of Con A. *Biosens. Bioelectron.*, 34: 202-297.
70. Chen, K.J., K.C. Pillai, J. Rick, C.J. Pan, S.H. Wang, C.C. Liu and B.J. Hwang, 2012. Bimetallic PtM (M = Pd, Ir) nanoparticle decorated multi-walled carbon nanotube enzyme-free, mediator-less amperometric sensor for H₂O₂. *Biosens. Bioelectron.*, 33: 120-127.
71. Li, L., B. Liang, J. Shi, F. Li, M. Mascini and A. Liu, 2012. A selective and sensitive D-xylose electrochemical biosensor based on xylose dehydrogenase displayed on the surface of bacteria and multi-walled carbon nanotubes modified electrode. *Biosens. Bioelectron.*, 33: 100-105.
72. Cao, Y., R. Yuan, Y. Chai, L. Mao, H. Niu, H. Liu and Y. Zhuo, 2012. Ultrasensitive luminol electrochemiluminescence for protein detection based on in situ generated hydrogen peroxide as coreactant with glucose oxidase anchored AuNPs@MWCNTs labeling. *Biosens. Bioelectron.*, 31: 305-309.
73. Bai, L., D. Wen, J. Yin, L. Deng, C. Zhu and S. Dong, 2012. Carbon nanotubes-ionic liquid nanocomposites sensing platform for NADH oxidation and oxygen, glucose detection in blood. *Talanta*, 91: 110-115.
74. Morgan, M.T., M.A. Carnahan, C.E. Immoos, A.A. Ribeiro and S. Finkelstein *et al.*, 2003. Dendritic molecular capsules for hydrophobic compounds. *J. Am. Chem. Soc.*, 125: 15485-15489.
75. Morgan, M.T., Y. Nakanishi, D.J. Kroll, A.P. Griset and M.A. Carnahan *et al.*, 2006. Dendrimer-encapsulated camptothecins: Increased solubility, cellular uptake and cellular retention affords enhanced anticancer activity *In vitro*. *Cancer Res.*, 66: 11913-11921.
76. Ooya, T., J. Lee and K. Park, 2004. Hydrotropic dendrimers of generations 4 and 5: Synthesis, characterization, and hydrotropic solubilization of paclitaxel. *Bioconj. Chem.*, 15: 1221-1229.

77. Gurdag, S., J. Khandare, S. Stapels, L.H. Matherly and R.M. Kannan, 2006. Activity of dendrimer-methotrexate conjugates on methotrexate-sensitive and -resistant cell lines. *Bioconj. Chem.*, 17: 275-283.
78. Papagiannaros, A., K. Dimas, G.T. Papaioannou and C. Demetzos, 2005. Doxorubicin-PAMAM dendrimer complex attached to liposomes: cytotoxic studies against human cancer cell lines. *Int. J. Pharm.*, 302: 29-38.
79. Kumar, P.V., A. Asthana, T. Dutta and N.K. Jain, 2006. Intracellular macrophage uptake of rifampicin loaded mannosylated dendrimers. *J. Drug Target*, 14: 546-556.
80. Dutta, T. and N.K. Jain, 2007. Targeting potential and anti-HIV activity of lamivudine loaded mannosylated poly(propyleneimine) dendrimer. *Biochimica Biophysica Acta (BBA)-Gen. Subj.*, 1770: 681-686.
81. Dutta, T., M. Garg and N.K. Jain, 2008. Targeting of efavirenz loaded tuftsin conjugated poly(propyleneimine) dendrimers to HIV infected macrophages *in vitro*. *Eur. J. Pharm. Sci.*, 34: 181-189.
82. Devarakonda, B., D.P. Otto, A. Judefeind, R.A. Hill and M.M. de Villiers, 2007. Effect of pH on the solubility and release of furosemide from polyamidoamine (PAMAM) dendrimer complexes. *Int. J. Pharmaceut.*, 345: 142-153.
83. Wang, F., T.K. Bronich, A.V. Kabanov, R.D. Rauh and J. Roovers, 2005. Synthesis and evaluation of a star amphiphilic block copolymer from poly(ϵ -caprolactone) and poly(ethylene glycol) as a potential drug delivery carrier. *Bioconj. Chem.*, 16: 397-405.
84. Bosnjakovic, A., M.K. Mishra, W. Ren, Y.E. Kurtoglu, T. Shi, D. Fan and R.M. Kannan, 2011. Poly(amidoamine) dendrimer-erythromycin conjugates for drug delivery to macrophages involved in periprosthetic inflammation. *Nanomedicine*, 7: 284-294.
85. Gajbhiye, V., N. Ganesh, J. Barve and N.K. Jain, 2013. Synthesis, characterization and targeting potential of zidovudine loaded sialic acid conjugated-mannosylated poly(propyleneimine) dendrimers. *Eur. J. Pharm. Sci.*, 48: 668-679.
86. Cheng, Y., N. Man, T. Xu, R. Fu, X. Wang, X. Wang and L. Wen, 2007. Transdermal delivery of nonsteroidal anti-inflammatory drugs mediated by polyamidoamine (PAMAM) dendrimers. *J. Pharm. Sci.*, 96: 595-602.
87. Zharov, V.P., V. Galitovsky and M. Viegas, 2003. Photothermal detection of local thermal effects during selective nanophotothermolysis. *Applied Phys. Lett.*, 83: 4897-4899.
88. Huang, X., P.K. Jain, I.H. El-Sayed and M.A. El-Sayed, 2008. Plasmonic Photothermal Therapy (PPTT) using gold nanoparticles. *Lasers Med. Sci.*, 23: 217-228.
89. Pitsillides, C.M., E.K. Joe, X. Wei, R.R. Anderson and C.P. Lin, 2003. Selective cell targeting with light-absorbing microparticles and nanoparticles. *Biophys. J.*, 84: 4023-4032.
90. Khlebtsov, B.N., A.G. Melnikov, V. Zharov and N. Khlebtsov, 2006. Absorption and scattering of light by a dimer of metal nanospheres: comparison of dipole and multipole approaches. *Nanotechnology*, 17: 1437-1445.
91. Huang, X., W. Qian, I.H. El-Sayed and M.A. El-Sayed, 2007. The potential use of the enhanced nonlinear properties of gold nanospheres in photothermal cancer therapy. *Lasers Surg. Med.*, 39: 747-753.
92. Abraham, G.E. and P.B. Himmel, 1997. Management of rheumatoid arthritis: Rationale for the use of colloidal metallic gold. *J. Nutr. Environ. Med.*, 7: 295-305.
93. Tsai, C.Y., A.L. Shiau, S.Y. Chen, Y.H. Chen and P.C. Cheng *et al.*, 2007. Amelioration of collagen-induced arthritis in rats by nanogold. *Arthritis Rheum.*, 56: 544-554.
94. Brown, C.L., G. Bushell, M.W. Whitehouse, D.S. Agrawal, S.G. Tupe, K.M. Paknikar and E.R.T. Tiekink, 2007. Nanogold-pharmaceutics. *Gold Bull.*, 40: 245-250.
95. Brown, C.L., M.W. Whitehouse, E.R.T. Tiekink and G.R. Bushell, 2008. Colloidal metallic gold is not bio-inert. *Inflammopharmacology*, 16: 133-137.
96. Bhattacharya, R., P. Mukherjee, Z. Xiong, A. Atala, S. Soker and D. Mukhopadhyay, 2004. Gold nanoparticles inhibit VEGF165-induced proliferation of HUVEC cells. *Nano Lett.*, 4: 2479-2481.
97. Mukherjee, P., R. Bhattacharya, P. Wang, L. Wang and S. Basu *et al.*, 2005. Antiangiogenic properties of gold nanoparticles. *Clin. Cancer Res.*, 11: 3530-3534.
98. Bhattacharya, R. and P. Mukherjee, 2008. Biological properties of Adv. *Drug Deliv. Rev.*, 60: 1289-1306.
99. Wang, L., Y. Liu, W. Li, X. Jiang and Y. Ji *et al.*, 2011. Selective targeting of gold nanorods at the mitochondria of cancer cells: Implications for cancer therapy. *Nano Lett.*, 11: 772-780.
100. Alanazi, F.K., A.A. Radwan and I.A. Alsarra, 2010. Biopharmaceutical applications of nanogold. *Saudi Pharm. J.*, 18: 179-193.
101. Duncan, B., C. Kim and V.M. Rotello, 2010. Gold nanoparticle platforms as drug and biomacromolecule delivery systems. *J. Control. Release*, 148: 122-127.
102. Pissuwan, D., T. Niidome and M.B. Cortie, 2011. The forthcoming applications of gold nanoparticles in drug and gene delivery systems. *J. Control. Release*, 149: 65-71.
103. Llevot, A. and D. Astruc, 2011. Applications of vectorized gold nanoparticles to the diagnosis and therapy of cancer. *Chem. Soc. Rev.*, 41: 242-257.
104. Paciotti, G.F., D.G.I. Kingston and L. Tamarkin, 2006. Colloidal gold nanoparticles: A novel nanoparticle platform for developing multifunctional tumor-targeted drug delivery vectors. *Drug Dev. Res.*, 67: 47-54.
105. Chen, Y.H., C.Y. Tsai, P.Y. Huang, M.Y. Chang and P.C. Cheng *et al.*, 2007. Methotrexate conjugated to gold nanoparticles inhibits tumor growth in a syngeneic lung tumor model. *Mol. Pharm.*, 4: 713-722.
106. Li, J., X. Wang, C. Wang, B. Chen and Y. Dai *et al.*, 2007. The enhancement effect of gold nanoparticles in drug delivery and as biomarkers of drug-resistant cancer cells. *Chem. Med. Chem.*, 2: 374-378.

107. Patra, C.R., R. Bhattacharya, E. Wang, A. Katarya and J.S. Lau *et al.*, 2008. Targeted delivery of gemcitabine to pancreatic adenocarcinoma using cetuximab as a targeting agent. *Cancer Res.*, 68: 1970-1978.
108. Podsiadlo, P., V.A. Sinani, J.H. Bahng, N.W.S. Kam, J. Lee and N.A. Kotov, 2008. Gold nanoparticles enhance the anti-leukemia action of a 6-mercaptapurine chemotherapeutic agent. *Langmuir*, 24: 568-574.
109. Azzam, E.M.S. and S.M.I. Morsy, 2008. Enhancement of the antitumour activity for the synthesised dodecylcysteine surfactant using gold nanoparticles. *J. Surfactants Deterg.*, 11: 195-199.
110. Agasti, S.S., A. Chompoosor, C.C. You, P. Ghosh, C.K. Kim and V.M. Rotello, 2009. Photoregulated release of caged anticancer drugs from gold nanoparticles. *J. Am. Chem. Soc.*, 131: 5728-5729.
111. Dhar, S., W.L. Daniel, D.A. Giljohann, C.A. Mirkin and S.J. Lippard, 2009. Polyvalent oligonucleotide gold nanoparticle conjugates as delivery vehicles for platinum(IV) warheads. *J. Am. Chem. Soc.*, 131: 14652-14653.
112. Patra, C.R., R. Bhattacharya and P. Mukherjee, 2010. Fabrication and functional characterization of gold nanoconjugates for potential application in ovarian cancer. *J. Mater. Chem.*, 20: 547-554.
113. Brown, S.D., P. Nativo, J.A. Smith, D. Stirling and P.R. Edwards *et al.*, 2010. Gold nanoparticles for the improved anticancer drug delivery of the active component of oxaliplatin. *J. Am. Chem. Soc.*, 132: 4678-4684.
114. Hosta, L., M. Pla-Roca and J. Arbiol, 2009. Conjugation of kahalalide F with gold nanoparticles to enhance *in vitro* antitumoral activity. *Bioconj. Chem.*, 20: 138-146.
115. Dreaden, E.C., S.C. Mwakwari, Q.H. Sodji, A.K. Oyelere and M.A. El-Sayed, 2009. Tamoxifen-poly(ethylene glycol)-thiol gold nanoparticle conjugates: Enhanced potency and selective delivery for breast cancer treatment. *Bioconj. Chem.*, 20: 2247-2253.
116. Eghtedari, M., A.V. Liopo, J.A. Copland, A.A. Oraevsky and M. Motamedi, 2009. Engineering of hetero-functional gold nanorods for the *in vivo* molecular targeting of breast cancer cells. *Nano Lett.*, 9: 287-291.
117. Park, C., H. Youn, H. Kim, T. Noh and Y.H. Kook *et al.*, 2009. Cyclodextrin-covered gold nanoparticles for targeted delivery of an anti-cancer drug. *J. Mater. Chem.*, 19: 2310-2315.
118. Gu, H., P.L. Ho, E. Tong, L. Wang and B. Xu, 2003. Presenting vancomycin on nanoparticles to enhance antimicrobial activities. *Nano Lett.*, 3: 1261-1263.
119. Rosemary, M.J., I. MacLaren and T. Pradeep, 2006. Investigations of the antibacterial properties of ciprofloxacin@SiO₂. *Langmuir*, 22: 10125-10129.
120. Selvaraj, V. and M. Alagar, 2007. Analytical detection and biological assay of antileukemic drug 5-fluorouracil using gold nanoparticles as probe. *Int. J. Pharm.*, 337: 275-281.
121. Burygin, G.L., B.N. Khlebtsov, A.N. Shantrokha, L.A. Dykman, V.A. Bogatyrev and N.G. Khlebtsov, 2009. On the enhanced antibacterial activity of antibiotics mixed with gold nanoparticles. *Nanoscale Res. Lett.*, 4: 794-801.
122. Nie, Z., K.J. Liu, C.J. Zhong, L.F. Wang, Y. Yang, Q. Tian and Y. Liu, 2007. Enhanced radical scavenging activity by antioxidant-functionalized gold nanoparticles: A novel inspiration for development of new artificial antioxidants. *Free Radical Biol. Med.*, 43: 1243-1254.
123. Bowman, M.C., T.E. Ballard, C.J. Ackerson, D.L. Feldheim, D.M. Margolis and C. Melander, 2008. Inhibition of HIV fusion with multivalent gold nanoparticles. *J. Am. Chem. Soc.*, 130: 6896-6897.
124. Chamberland, D.L., A. Agarwal, N. Kotov, J.B. Fowlkes, P.L. Carson and X. Wang, 2008. Photoacoustic tomography of joints aided by an Etanercept-conjugated gold nanoparticle contrast agent-an *ex vivo* preliminary rat study. *Nanotechnology*. 10.1088/0957-4484/19/9/095101
125. Yuan, Y., D. Rende, C.L. Altan, S. Bucak, R. Ozisik and D.A. Borca-Tasciuc, 2012. Effect of surface modification on magnetization of iron oxide nanoparticle colloids. *Langmuir*, 28: 13051-13059.
126. Talelli, M., C.J. Rijcken, T. Lammers, P.R. Seevinck, G. Storm, C.F. van Nostrum and W.E. Hennink, 2009. Superparamagnetic iron oxide nanoparticles encapsulated in biodegradable thermosensitive polymeric micelles: Toward a targeted nanomedicine suitable for image-guided drug delivery. *Langmuir*, 25: 2060-2067.
127. Mahmoudi, M., A. Simchi, M. Imani and U.O. Hafeli, 2009. Superparamagnetic iron oxide nanoparticles with rigid cross-linked polyethylene glycol fumarate coating for application in imaging and drug delivery. *J. Phys. Chem. C*, 113: 8124-8131.
128. Beveridge, J.S., J.R. Stephens and M.E. Williams, 2011. The use of magnetic nanoparticles in analytical chemistry. *Annu. Rev. Anal. Chem.*, 4: 251-273.
129. Smith, J.E., K.E. Sapsford, W. Tan and F.S. Ligler, 2011. Optimization of antibody-conjugated magnetic nanoparticles for target preconcentration and immunoassays. *Anal. Biochem.*, 410: 124-132.
130. Daldrup-Link, H.E., D. Golovko, B. Ruffell, D.G. DeNardo and R. Castaneda *et al.*, 2011. MRI of tumor-associated macrophages with clinically applicable iron oxide nanoparticles. *Clin. Cancer Res.*, 17: 5695-5704.
131. Andreas, K., R. Georgieva, M. Ladwig, S. Mueller, M. Notter, M. Sittinger and J. Ringe, 2012. Highly efficient magnetic stem cell labeling with citrate-coated superparamagnetic iron oxide nanoparticles for MRI tracking. *Biomaterials*, 33: 4515-4525.
132. Kievit, F.M. and M. Zhang, 2011. Surface engineering of iron oxide nanoparticles for targeted cancer therapy. *Accounts Chem. Res.*, 44: 853-862.

133. Liu, X., J. Cao, H. Li, J. Li, Q. Jin, K. Ren and J. Ji, 2013. Mussel-inspired polydopamine: A biocompatible and ultrastable coating for nanoparticles *in vivo*. *ACS Nano*, 7: 9384-9395.
134. Zhang, M., X. Zhang, X. He, L. Chen and Y. Zhang, 2012. A self-assembled polydopamine film on the surface of magnetic nanoparticles for specific capture of protein. *Nanoscale*, 4: 3141-3147.
135. Tartaj, P., M.D.P. Morales, S. Veintemillas-Verdaguer, T. Gonzalez-Carreno and C.J. Serna, 2003. The preparation of magnetic nanoparticles for applications in biomedicine. *J. Phys. Applied Phys.*, 36: R182-R197.
136. Basuki, J.S., L. Esser, P.B. Zetterlund, M.R. Whittaker, C. Boyer and T.P. Davis, 2013. Grafting of P(OEGA) onto magnetic nanoparticles using Cu(0) mediated polymerization: Comparing grafting "from" and "to" approaches in the search for the optimal material design of nanoparticle MRI contrast agents. *Macromolecules*, 46: 6038-6047.
137. Neuberger, T., B. Schopf, H. Hofmann, M. Hofmann and B. von Rechenberg, 2005. Superparamagnetic nanoparticles for biomedical applications: Possibilities and limitations of a new drug delivery system. *J. Magn. Mater.*, 293: 483-496.
138. Yu, M.K., Y.Y. Jeong, J. Park, S. Park and J.W. Kim *et al.*, 2008. Drug-loaded superparamagnetic iron oxide nanoparticles for combined cancer imaging and therapy *in vivo*. *Angewandte Chemie Int. Edn.*, 47: 5362-5365.
139. Yang, X., H. Hong, J.J. Grailer, I.J. Rowland and A. Javadi *et al.*, 2011. cRGD-functionalized, DOX-conjugated and ⁶⁴Cu-labeled superparamagnetic iron oxide nanoparticles for targeted anticancer drug delivery and PET/MR imaging. *Biomaterials*, 32: 4151-4160.
140. Dobson, J., 2006. Gene therapy progress and prospects: Magnetic nanoparticle-based gene delivery. *Gene Therapy*, 13: 283-287.
141. Mah, C., T.J. Fraitas Jr., I. Zolotukhin, S. Song and T.R. Flotte *et al.*, 2002. Improved method of recombinant AAV2 delivery for systemic targeted gene therapy. *Mol. Therapy*, 6: 106-112.
142. Lubbe, A.S., C. Bergemann, H. Riess and F. Schriever *et al.*, 1996. Clinical experiences with magnetic drug targeting: A phase I study with 4'-Epidoxorubicin in 14 patients with advanced solid tumors. *Cancer Res.*, 56: 4686-4693.
143. Basuki, J.S., H.T. Duong, A. Macmillan, R.B. Erlich and L. Esser *et al.*, 2013. Using fluorescence lifetime imaging microscopy to monitor theranostic nanoparticle uptake and intracellular doxorubicin release. *ACS Nano*, 7: 10175-10189.
144. Laurent, S., D. Forge, M. Port, A. Roch and C. Robic *et al.*, 2008. Magnetic iron oxide nanoparticles: Synthesis, stabilization, vectorization, physicochemical characterizations and biological applications. *Chem. Rev.*, 108: 2064-2110.
145. Shen, J.M., X.M. Guan, X.Y. Liu, J.F. Lan, T. Cheng and H.X. Zhang, 2012. Luminescent/magnetic hybrid nanoparticles with folate-conjugated peptide composites for tumor-targeted drug delivery. *Bioconj. Chem.*, 23: 1010-1021.
146. Kebede, A., A.K. Singh, P.K. Rai, N.K. Giri, A.K. Rai, G. Watal and A.V. Gholap, 2013. Controlled synthesis, characterization and application of iron oxide nanoparticles for oral delivery of insulin. *Lasers Med. Sci.*, 28: 579-587.
147. Shen, J.M., L. Xu, Y. Lu, H.M. Cao, Z.G. Xu, T. Chen and H.X. Zhang, 2012. Chitosan-based luminescent/magnetic hybrid nanogels for insulin delivery, cell imaging and antidiabetic research of dietary supplements. *Int. J. Pharm.*, 427: 400-409.
148. Perez, J.M., J. Grimm, L. Josephson and R. Weissleder, 2008. Integrated nanosensors to determine levels and functional activity of human telomerase. *Neoplasia*, 10: 1066-1072.
149. Lee, H., T.J. Yoon, J.L. Figueiredo, F.K. Swirski and R. Weissleder, 2009. Rapid detection and profiling of cancer cells in fine-needle aspirates. *Proc. Natl. Acad. Sci. USA.*, 106: 12459-12464.
150. Grimm, J., J.M. Perez, L. Josephson and R. Weissleder, 2004. Novel nanosensors for rapid analysis of telomerase activity. *Cancer Res.*, 64: 639-643.
151. Bautista, M.C., O. Bomati-Miguel, X. Zhao, M.P. Morales and T. Gonzalez-Carreno *et al.*, 2004. Comparative study of ferrofluids based on dextran-coated iron oxide and metal nanoparticles for contrast agents in magnetic resonance imaging. *Nanotechnology*, Vol. 15.
152. Perez, J.M., L. Josephson, T. O'Loughlin, D. Hogemann and R. Weissleder, 2002. Magnetic relaxation switches capable of sensing molecular interactions. *Nat. Biotechnol.*, 20: 816-820.
153. Liu, Y., Y. Gao and C. Xu, 2013. Using magnetic nanoparticles to manipulate biological objects. *Chin. Phys. B*, Vol. 22.
154. Zhang, K., L.L. Zhang, X.S. Zhao and J. Wu, 2010. Graphene/polyaniline nanofiber composites as supercapacitor electrodes. *Chem. Mater.*, 22: 1392-1401.
155. Latil, S. and L. Henrard, 2006. Charge carriers in few-layer graphene films. *Phys. Rev. Lett.*, Vol. 97. 10.1103/PhysRevLett.97.036803
156. Liu, Z., J.T. Robinson, X. Sun and H. Dai, 2008. PEGylated nanographene oxide for delivery of water-insoluble cancer drugs. *J. Am. Chem. Soc.*, 130: 10876-10877.
157. Jiang, H., 2011. Chemical preparation of graphene-based nanomaterials and their applications in chemical and biological sensors. *Small*, 7: 2413-2427.
158. Guo, S. and S. Dong, 2011. Graphene nanosheet: Synthesis, molecular engineering, thin film, hybrids and energy and analytical applications. *Chem. Soc. Rev.*, 40: 2644-2672.
159. Hummers, Jr. W.S. and R.E. Offeman, 1958. Preparation of graphitic oxide. *J. Am. Chem. Soc.*, 80: 1339-1339.
160. Loh, K.P., Q. Bao, G. Eda and M. Chhowalla, 2010. Graphene oxide as a chemically tunable platform for optical applications. *Nat. Chem.*, 2: 1015-1024.
161. Kovtyukhova, N.I., P.J. Ollivier, B.R. Martin, T.E. Mallouk, S.A. Chizhik, E.V. Buzaneva and A.D. Gorchinskiy, 1999. Layer-by-layer assembly of ultrathin composite films from micron-sized graphite oxide sheets and polycations. *Chem. Mater.*, 11: 771-778.

162. Sun, X., Z. Liu, K. Welsher, J.T. Robinson, A. Goodwin, S. Zaric and H. Dai, 2008. Nano-graphene oxide for cellular imaging and drug delivery. *Nano Res.*, 1: 203-212.
163. Shan, C., H. Yang, D. Han, Q. Zhang, A. Ivaska and L. Niu, 2009. Water-soluble graphene covalently functionalized by biocompatible poly-L-lysine. *Langmuir*, 25: 12030-12033.
164. Lei, H., L. Mi, X. Zhou, J. Chen, J. Hu, S. Guo and Y. Zhang, 2011. Adsorption of double-stranded DNA to graphene oxide preventing enzymatic digestion. *Nanoscale*, 3: 3888-3892.
165. Zhang, J., F. Zhang, H. Yang, X. Huang, H. Liu, J. Zhang and S. Guo, 2010. Graphene oxide as a matrix for enzyme immobilization. *Langmuir*, 26: 6083-6085.
166. Zhang, F., B. Zheng, J. Zhang, X. Huang, H. Liu, S. Guo and J. Zhang, 2010. Horseradish peroxidase immobilized on graphene oxide: Physical properties and applications in phenolic compound removal. *J. Phys. Chem. C*, 114: 8469-8473.
167. Lee, D.Y., Z. Khatun, J.H. Lee, Y.K. Lee and I. In, 2011. Blood compatible graphene/heparin conjugate through noncovalent chemistry. *Biomacromolecules*, 12: 336-341.
168. Dong, H., W. Gao, F. Yan, H. Ji and H. Ju, 2010. Fluorescence resonance energy transfer between quantum dots and graphene oxide for sensing biomolecules. *Anal. Chem.*, 82: 5511-5517.
169. Chen, W., P. Yi, Y. Zhang, L. Zhang, Z. Deng and Z. Zhang, 2011. Composites of aminodextran-coated Fe₃O₄ nanoparticles and graphene oxide for cellular magnetic resonance imaging. *ACS Applied Mater. Interf.*, 3: 4085-4091.
170. Shen, J., M. Shi, N. Li, B. Yan, H. Ma, Y. Hu and M. Ye, 2010. Facile synthesis and application of Ag-chemically converted graphene nanocomposite. *Nano Res.*, 3: 339-349.
171. Liu, Z., J.T. Robinson, S.M. Tabakman, K. Yang and H. Dai, 2011. Carbon materials for drug delivery & cancer therapy. *Mater. Today*, 14: 316-323.
172. Dai, H., 2002. Carbon nanotubes: Opportunities and challenges. *Surf. Sci.*, 500: 218-241.
173. Depan, D., J. Shah and R.D.K. Misra, 2011. Controlled release of drug from folate-decorated and graphene mediated drug delivery system: Synthesis, loading efficiency and drug release response. *Mater. Sci. Eng. C*, 31: 1305-1312.
174. Zhang, L., J. Xia, Q. Zhao, L. Liu and Z. Zhang, 2010. Functional graphene oxide as a nanocarrier for controlled loading and targeted delivery of mixed anticancer drugs. *Small*, 6: 537-544.
175. Rana, V.K., M.C. Choi, J.Y. Kong, G.Y. Kim and M.J. Kim *et al.*, 2011. Synthesis and drug-delivery behavior of chitosan-functionalized graphene oxide hybrid nanosheets. *Macromol. Mater. Eng.*, 296: 131-140.
176. Yang, X., X. Zhang, Z. Liu, Y. Ma, Y. Huang and Y. Chen, 2008. High-efficiency loading and controlled release of doxorubicin hydrochloride on graphene oxide. *J. Phys. Chem. C*, 112: 17554-17558.
177. Naldini, L., U. Blomer, P. Gallay, D. Ory and R. Mulligan *et al.*, 1996. *In vivo* gene delivery and stable transduction of nondividing cells by a lentiviral vector. *Science*, 272: 263-267.
178. Mintzer, M.A. and E.E. Simanek, 2008. Nonviral vectors for gene delivery. *Chem. Rev.*, 109: 259-302.
179. Feng, L., S. Zhang and Z. Liu, 2011. Graphene based gene transfection. *Nanoscale*, 3: 1252-1257.
180. Chen, B., M. Liu, L. Zhang, J. Huang, J. Yao and Z. Zhang, 2011. Polyethylenimine-functionalized graphene oxide as an efficient gene delivery vector. *J. Mater. Chem.*, 21: 7736-7741.
181. Bao, H., Y. Pan, Y. Ping, N.G. Sahoo and T. Wu *et al.*, 2011. Chitosan-functionalized graphene oxide as a nanocarrier for drug and gene delivery. *Small*, 7: 1569-1578.
182. Yang, K., S. Zhang, G. Zhang, X. Sun, S.T. Lee and Z. Liu, 2010. Graphene in mice: Ultrahigh *in vivo* tumor uptake and efficient photothermal therapy. *Nano Lett.*, 10: 3318-3323.
183. Zhang, L., Z. Lu, Q. Zhao, J. Huang, H. Shen and Z. Zhang, 2011. Enhanced chemotherapy efficacy by sequential delivery of siRNA and anticancer drugs using PEI-grafted graphene oxide. *Small*, 7: 460-464.
184. Markovic, Z.M., L.M. Harhaji-Trajkovic, B.M. Todorovic-Markovic, D.P. Kepic and K.M. Arsinin *et al.*, 2011. *In vitro* comparison of the photothermal anticancer activity of graphene nanoparticles and carbon nanotubes. *Biomaterials*, 32: 1121-1129.
185. Zhang, W., Z. Guo, D. Huang, Z. Liu, X. Guo and H. Zhong, 2011. Synergistic effect of chemo-photothermal therapy using PEGylated graphene oxide. *Biomaterials*, 32: 8555-8561.
186. Huang, P., C. Xu, J. Lin, C. Wang and X. Wang *et al.*, 2011. Folic acid-conjugated graphene oxide loaded with photosensitizers for targeting photodynamic therapy. *Theranostics*, 1: 240-250.
187. Tian, B., C. Wang, S. Zhang, L. Feng and Z. Liu, 2011. Photothermally enhanced photodynamic therapy delivered by nano-graphene oxide. *ACS Nano*, 5: 7000-7009.
188. Hu, W., C. Peng, W. Luo, M. Lv and X. Li *et al.*, 2010. Graphene-based antibacterial paper. *ACS Nano*, 4: 4317-4323.
189. Akhavan, O. and E. Ghaderi, 2010. Toxicity of graphene and graphene oxide nanowalls against bacteria. *ACS Nano*, 4: 5731-5736.
190. Liu, S., T.H. Zeng, M. Hofmann, E. Burcombe and J. Wei *et al.*, 2011. Antibacterial activity of graphite, graphite oxide, graphene oxide and reduced graphene oxide: Membrane and oxidative stress. *ACS Nano*, 5: 6971-6980.
191. Cheung, R.C.F., T.B. Ng, J.H. Wong and W.Y. Chan, 2015. Chitosan: An update on potential biomedical and pharmaceutical applications. *Mar. Drugs*, 13: 5156-5186.
192. Zare, Y. and I. Shabani, 2016. Polymer/metal nanocomposites for biomedical applications. *Mater. Sci. Eng.: C*, 60: 195-203.

193. Dev, A., N.S. Binulal, A. Anitha, S.V. Nair, T. Furuike, H. Tamura and R. Jayakumar, 2010. Preparation of poly(lactic acid)/chitosan nanoparticles for anti-HIV drug delivery applications. *Carbohydr. Polym.*, 80: 833-838.
194. Jemal, A., F. Bray, M.M. Center, J. Ferlay, E. Ward and D. Forman, 2011. Global cancer statistics. *CA: Cancer J. Clin.*, 61: 69-90.
195. Douroumis, D., 2011. Mesoporous silica nanoparticles as drug delivery system. *J. Nanomed. Nanotechnol.*, Vol. 2.
196. Wu, J.C., 2015. Silica three-dimensional biosensors. *Biosens. J.*